

1 *Review*

# 2 **Current Status of the Sm14/GLA-SE Schistosomiasis** 3 **Vaccine: Overcoming Barriers and Paradigms** 4 **towards the First Anti-parasitic Human(itarian)** 5 **Vaccine**

6 **Miriam Tendler<sup>1,\*</sup>, Marília S. Almeida<sup>1</sup>, Monica M. Vilar<sup>1</sup>, Patrícia M. Pinto<sup>1</sup> and Gabriel**  
7 **Limaverde-Sousa<sup>1</sup>**

8 <sup>1</sup> Laboratório de Esquistossomose Experimental, Instituto Oswaldo Cruz, FIOCRUZ - Av. Brasil, 4365,  
9 Manguinhos, 21045-900, Rio de Janeiro, Brazil

10 \* Correspondence: mtendler@ioc.fiocruz.br; Tel.: +55-21-2562-1320

11

## 12 **Abstract:**

13 Schistosomiasis, a disease historically associated with poverty, lack of sanitation and social inequalities, is a  
14 chronic, debilitating parasitic infection, affecting hundreds of millions of people in endemic countries.  
15 Although schistosomiasis control approach has shown that chemotherapy is capable of reducing morbidity in  
16 humans, rapid re-infection is a reminder that the impact of drug treatment on transmission control or  
17 elimination initiatives is marginal. In addition, and regardless of more than two decades of well-executed  
18 control activities based on large-scale chemotherapy, the disease is expanding in many areas including Brazil.  
19 The development of the Sm14/GLA-SE schistosomiasis vaccine is an emblematic open knowledge innovation  
20 that has successfully completed Phase I and Phase IIa clinical trials, with Phase II/III trials underway in the  
21 African continent and to be followed in Brazil. Discovery and experimental phases were long term  
22 achievements leading to a robust collection of data that are strongly supporting the presently ongoing Clinical  
23 Phase. This paper reviews the development of the Sm14 vaccine formulated with GLA-SE (Glucopyranosyl  
24 Lipid A), from the earlier experimental developments to clinical trials including the recent status of Phase II  
25 studies.

26 **Keywords:** Schistosomiasis; vaccine; Sm14; FABP

27

## 28 **1. Introduction**

29 Schistosomiasis is the second-most socioeconomically devastating parasitic disease after malaria.  
30 It is a chronic and debilitating endemy with an estimated 200 million people infected, out of which  
31 120 million are symptomatic, with 20 million presenting severe disease symptoms, and most of them  
32 (85%) live in Africa [1]. These estimates may error on the low side as a meta-analysis has found the  
33 number of people at risk to be closer to 800 million [2]. The global Schistosomiasis remains as high as  
34 ever and the estimated number of Disability-Adjusted Life Years (DALYs) has increased with the  
35 inclusion of previously under-recognized morbidities not counted for the DALY index before (eg.,  
36 growth stunting, anaemia associated to retarded intellectual development) in infants, toddlers and  
37 school age children, the part of the population whose physical health and intellectual capacity are  
38 fundamental to nation development and sustainability[3,4]. Under this more realistic scenario, the  
39 impact of schistosomiasis comes second in the list of the 18 World Health Organization (WHO)  
40 neglected tropical diseases (NTDs) [5]. In Brazil, the largest endemic country for schistosomiasis, 6  
41 million individuals are estimated to be infected, 25 million are at risk of contracting the disease [6,7].

42 Mass chemotherapy has been the strategy of choice in an attempt to control schistosomiasis and  
 43 intestinal helminthiasis with the support of international health funding agencies. Estimates show  
 44 that at least 206.5 million people required treatment in 2016 [8]. However, the strategy of large-scale  
 45 treatment, based on chemotherapy also equivocally called “prophylactic treatment” failed to control  
 46 transmission for more than thirty years. Approximately 300 millions of US dollars are still being spent  
 47 annually in treatments to be applied to the same populations year after year, with no prospect to  
 48 prevent reinfections and subsequent treatments, in addition to overloading children and young  
 49 population of endemic countries with chemical drugs [9]. “Deworming” initiatives, originally  
 50 applied to animal species only, were proposed as a tool for schistosomiasis control programs  
 51 addressed to school children in endemic countries [10,11].

52 Contrastingly, under One Health policies for the control of veterinary helminth infections, such  
 53 as *Fasciola hepatica* the major parasitic infection of livestock worldwide, there is a strong demand for  
 54 the replacement of anti-helminthic drugs - that implies in significant amount of chemical residues  
 55 detected in meat, milk and added-value products – for safe vaccines, considered as the most  
 56 environment and human health friendly methods for the control of Fascioliasis in livestock [12].

57 The insertion of vaccines in the context of programs towards effective control of schistosomiasis  
 58 brings hope to a future scenario for the poor. The Brazilian Sm14 Schistosomiasis Vaccine Platform  
 59 was launched and strongly pushed in the context of a formal WHO program, specifically structured  
 60 towards the Development of Anti Schistosomiasis Vaccine in the 1990's. The main outcome of this  
 61 initiative was the selection of six priority antigen candidates out of which only Sm14, continued to be  
 62 developed (Table 1, adapted from [13]).

63 With strategic support of WHO, this initiative is moving forward, emerging from an endemic  
 64 country, to the final development of Sm14 Schistosomiasis Vaccine based on a recombinant protein.  
 65 It is being developed under the most sophisticated and modern technological platforms and  
 66 professionally conducted in a network of outstanding companies and collaborators. This was the  
 67 result of long-term scientific developments carried under the coordination of FIOCRUZ, a public  
 68 institution linked to the Brazilian Ministry of Health. Sm14 based vaccine is protected by strong  
 69 patents owned by FIOCRUZ in all countries of interest worldwide.  
 70

71 **Table 1.** Schistosomiasis priority antigens selected by WHO for independent testing (adapted from  
 72 [13]).

Antigen	Size (kDa)	Stage expressed	Description	Protection (%)	Place of Development
Glutathione S-transferase (P28/GST)	28	Adult/somula/egg	Enzyme	30-60	Institut Pasteur, Lille, France
Paramyosin (Sm97)	97	Adult/somula	Muscle protein	30	Case Western Reserve University/ National Institute of Health/Cornell University, USA
IrV-5	62	Adult/somula/egg	Muscle protein	50-70	Johns Hopkins School of Medicine, Baltimore, USA
Triose phosphate isomerase (TPI)	28	Adult/somula/egg	Enzyme	30-60	Harvard School of Public Health, Boston, USA
Sm23	23	Adult/somula/egg	Integrated membrane protein	40-50	Johns Hopkins School of Medicine/Harvard School of Public Health, USA
Sm14	14	Adult/somula	Fatty acid-binding protein	65	Instituto Oswaldo Cruz, Rio de Janeiro, Brazil

73  
 74 Recently, the Consultative Expert Working Group on Research and Development: Financing and  
 75 Coordination (CEWG/WHO) selected the Sm14 Vaccine as one of the six Demonstration Projects,  
 76 globally, under an extensive selection and several rounds of shortlists as it has demonstrated to be  
 77 fully adherent and in accordance with the principles of CEWG, such as to demonstrate clear  
 78 mechanisms of Delinkage of costs from investments in R&D from costs of final product, Accessibility,  
 79 Affordability, Viability and to be an Open Knowledge Innovation. The CEWG recognized that the  
 80 Sm14 vaccine may become a key tool for the implementation of effective programs, based not only  
 81 on chemotherapy, but yet in infection reduction and transmission control of Schistosomiasis [14].

82 Over the last years it was possible to overcome important bottlenecks in the process of new  
83 product/vaccine development: scaling up production process from laboratory bench to production  
84 scale and successful conclusion of two Phase I human trials in healthy adults (man and woman) living  
85 in a Brazilian non-endemic area (2011-2014) [15] and the first Phase II trial in 30 male adults living in  
86 highly endemic area for both *Schistosoma mansoni* and *S. haematobium* at the Senegal River Basin (2015-  
87 2017). Safety was extensively confirmed and strong and long-lasting immunogenicity was also  
88 demonstrated (manuscript in preparation). Process development and master cell bank generation  
89 were recently completed and GMP manufacture of Sm14 lot is currently ongoing, under the  
90 coordination of the Infectious Disease Research Institute (IDRI, Seattle, US). This article is an  
91 overview of Sm14 Schistosomiasis vaccine development from the antigen discovery to the current  
92 human studies (Fig. 1). The need to overcome barriers for the rise of a truly humanitarian vaccine - to  
93 the needs of the developing world - will be discussed.

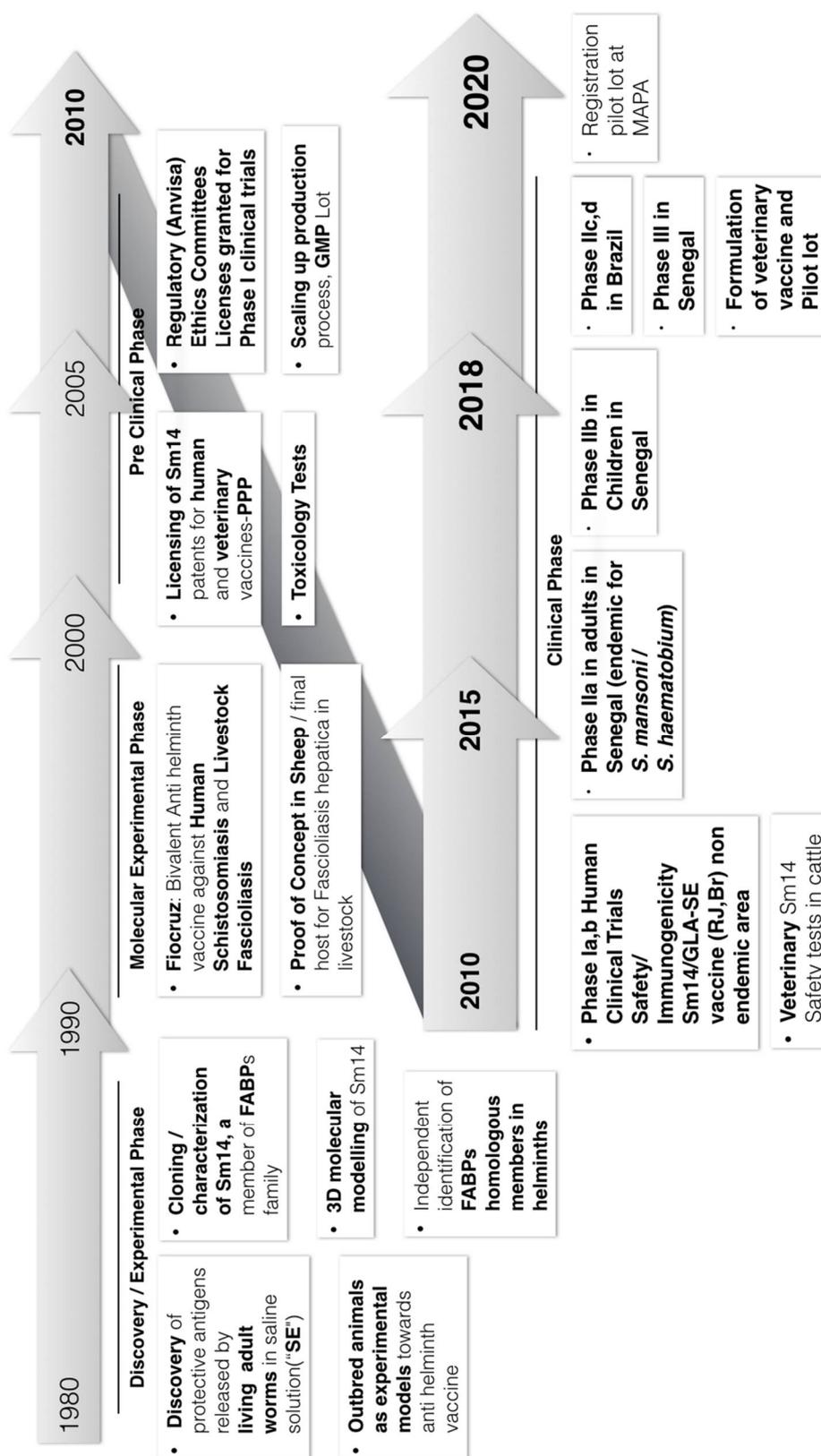
## 94 2. Innovative Strategies Adopted for Antigen Discovery and Early Development Phases

95 Biotechnological advances in various areas of vaccine research have contributed to the  
96 development of safer and more effective formulations. Efforts to develop anti-helminth vaccines  
97 lasted for many years and are continuing to progress in identification of candidate antigens, which  
98 have recently been aided with the generation of a number of helminth genomes [16]. The  
99 implementation of a vaccine against schistosomiasis represents an important step in a context of  
100 research and development in public health for poor populations infected and exposed to  
101 Schistosomes. There have been initiatives from research groups in different countries to develop a  
102 vaccine against schistosomiasis. The Brazilian Sm14-based anti-schistosomiasis vaccine is the sole  
103 technology that is at an advanced stage of development toward a safe highly innovative product [17].

104 In contrast to the current "OMICS" strategies, in which high-throughput screenings of potential  
105 antigens are being processed in parallel by automated "discovery protocols and platforms", the Sm14  
106 project was borne by gathering observations from animal models of infection, and a sequential and  
107 specific rational thinking of development and experimental design based on evidences.

108 The first original approach was in how the worm extract was obtained for subsequent  
109 assessments of protection. Instead of lyophilized parasites, generally used by other groups, a saline  
110 extract containing secreted/excreted antigens derived from **living adult schistosomes** was obtained  
111 for the initial immunizations. The restricted number of potential protective antigens released in this  
112 saline extract, allowed a more direct identification of strong candidates for the following molecular  
113 phase of the development [18–22]. Innovative methods were also adopted in the experimental phase,  
114 when the use of outbred models, in contrast to the commonly employed inbred animals, allowed a  
115 better representation of the ultimate target population and provided a unique opportunity to develop  
116 an alternative strategy for the protection assessment, improving the overall understanding of the  
117 outcomes. The analyses were stratified based on the measurement of frequencies of worm burden  
118 distribution of vaccinated–challenged animal population over non-vaccinated infected controls, as  
119 opposed to evaluation of mean values of parasite loads, as usually adopted. A solid base of pre-  
120 clinical data was raised establishing the immunization protocols that would be adopted on the  
121 following steps of the development [23–31].

122 As molecular biology tools evolved and gene-cloning techniques became available, several  
123 antigens released in the saline culture medium of live schistosomes, were cloned and sequenced.  
124 Antiserum from rabbits immunized with the "saline extract" was used to screen an adult *S. mansoni*  
125 cDNA library and the most promising antigen was identified as a member of the Fatty Acid Binding  
126 Protein (FABP) family, termed Sm14 [32]. Molecular modeling studies from our group predicted the  
127 beta-barrel structure of the Sm14 tridimensional structure [33] that was later experimentally  
128 confirmed by crystallography [34] and Nuclear Magnetic Resonance [35]. Such analyses allowed the  
129 engineering of a stabilizing mutation that rendered this antigen a remarkable long-term stability,  
130 while maintaining its function and immunogenicity [36].



131  
132  
133  
134  
135

**Figure 1.** Timeline: Sm14/GLA-SE Anti Schistosomiasis Vaccine - From Discovery Phase to Final Product. MAPA: Brazilian Ministry of Agriculture.

136 Sm14 was shown to be particularly important to helminths, that are not capable of synthesizing  
137 fatty acids themselves, which ultimately are provided by the host species. Lipids, apart from being  
138 constituents of membranes, have important role in development of different lifecycle stages and  
139 evasion of immune responses by adults and larvae [37]. After the first publication on the presence of  
140 a FABP family member in the *Schistosoma mansoni* [32], different groups published on the  
141 identification of homologous protein members from FABP family in many helminths of human and  
142 veterinary importance. The first one published after Sm14, was the *Fasciola hepatica* FABP, the main  
143 parasite of livestock worldwide [38]. We have managed to successfully test with Sm14 vaccination  
144 against *F. hepatica* in mice followed by two independent experiments in sheep, one of the final host  
145 species for fascioliasis [39]. It was thus demonstrated that Sm14 is also protective against *Fasciola*  
146 *hepatica* infection and it is therefore being also developed in parallel as the molecular basis for a  
147 veterinary vaccine by FIOCRUZ in collaboration with the private Brazilian company Ourofino  
148 Animal Health.

### 149 3. Clinical Studies

150 The licensing of the Sm14 patents for veterinary use gave birth to a Public-Private Partnership  
151 (PPP) model of product development that rendered significant visibility to the Sm14 vaccine project.  
152 Such gain in momentum was followed by a strong support of the human vaccine development branch  
153 by the Brazilian government project financing agency (FINEP) that allowed the use of Contract  
154 Research Organizations (CROs) for antigen production, quality control and fill-finish in GMP world-  
155 class facilities based in the United States of America, under the coordination of the Infectious Disease  
156 Research Institute (IDRI, Seattle, USA).

157 In order to have a consistent, stable and defined final product for clinical human use, the Sm14  
158 antigen was formulated with the synthetic adjuvant Glucopyranosyl Lipid A (GLA-SE), a clinically  
159 approved molecule already used in a number of commercially available human vaccines.

160 In December of 2010, the Brazilian Health Regulatory Agency (ANVISA) approved the Phase Ia  
161 Clinical Trial in 20 healthy male volunteers in non-endemic area (Rio de Janeiro, Brazil) to evaluate  
162 the safety of the investigational product. The study was conducted by the Brazilian National Institute  
163 of Infectious Diseases (INI/FIOCRUZ). The results of this first trial attested the safety of the vaccine  
164 on the studied population, showing no systemic reactogenicity and no adverse event was associated  
165 to the investigational product [15]. The Phase Ib Clinical Study, to evaluate the safety and  
166 immunogenicity of the vaccine preparation in 10 healthy women volunteers, was successfully  
167 concluded in 2012 (manuscript in preparation).

168 In 2015-2016, already under the scope of CEWG Demonstration Project, it was developed and  
169 concluded the first Phase II trial (Phase IIa) in 30 male adults living in highly endemic area for both  
170 *Schistosoma mansoni* and *S. haematobium* at Senegal River Basin, conducted by specialized team from  
171 Espoir Pour La Santé (EPLS), linked to the Pasteur Institute of Lille (IPL, France), headed by Dr. Gilles  
172 Riveau (IPL) in conjunction with Brazilian group guidance (ClinicalTrials.gov Identifier:  
173 NCT03041766) [40]. Main objectives of safety and immunogenicity in the context of vaccination with  
174 Sm14/GLA-SE vaccine were fully achieved in Phase II a trial. The investigational product rSm14 (50  
175 µg) formulated with GLA-SE in two dosages (2.5 µg and 5 µg/dose, denominated groups 1 and 2,  
176 respectively) and administered IM was shown to be safe with no observed serious adverse events in  
177 either group. The most common reactions were local pain and heaviness of vaccinated arm, that were  
178 transitory and mild. Seroconversion of 92% of individuals after second dose was observed, in an  
179 analogous pattern as described in Phase I trials [15]. Immunogenicity based on additional cellular  
180 response, memory cells and T cell activation markers was analyzed at IDRI in an extensive panel  
181 focusing on the identification of vaccine related immune response.

182 After the closure of the Phase IIa, based on the good results of induced strong and lasting  
183 immune response, an extension study to assess the possible persistence and profile of this response  
184 beyond the initial schedule of the trial was duly authorized by Senegalese Ministry of Health (MoH)  
185 and carried out between August and December 2017 with the inclusion of two additional time points,  
186 9 and 12 months, after the first vaccine injection. ELISA preliminary tests were performed at Centre

187 de Recherche Biomédicale/Espoir Pour La Santé (EPLS) and showed the persistence of significant  
188 specific antibody titers for until 12 months after first vaccination dose with Sm14/GLA-SE  
189 (manuscript in preparation).

190 Phase IIb trial design and protocol were defined in January 2018 based on results of the Phase  
191 IIa trial in adults. Organization of Phase IIb clinical trial with 95 school children from 07-11 years old  
192 living at the same endemic area for both *Schistosoma mansoni* and *S. hamatobium* of the Senegal River  
193 Basin Region was concluded, Ethical Committee approval and Regulatory License granted by  
194 Senegalese MoH. Trial is already ongoing under conduction of EPLS, at the same region of precedent  
195 Phase IIa trial in adults, using the same Lot of GMP Sm14 under a regimen of three IM doses of 50  
196 µg/dose formulated with 2.5 µg of GLA-SE, 30 days apart. The Phase IIa and b trials are mainly being  
197 sponsored by the private Brazilian partner Orygen Biotechnology and governmental institution  
198 FIOCRUZ with support of CEWG-WHO platform and Financial Fund.

#### 199 4. Sm14 +GLA-SE anti-Schistosomiasis Humanitarian Vaccine

200 During the process of analysis by member state representatives under WHO regional structure  
201 and posterior extensive questioning by technical experts and ad hoc committees, Sm14  
202 Schistosomiasis vaccine, was selected as one of the present list of six Demonstration Projects,  
203 confirmed by WHO Executive Board. In June of 2015, a Stake Holders Meeting was organized by  
204 WHO at Genève headquarters, before the first installment was granted. During the process of  
205 selection and shortlisting of presented projects; discussions on the scientific merits, state of the art  
206 of the project and demanded full adherence to CEWG principles, much was learned on the  
207 mandatory need to assure the Accessibility and Affordability of this vaccine to the poor endemic  
208 countries to which it is ultimately addressed [14].

209 From its inception, the Sm14 based Schistosomiasis vaccine was conducted targeting an effective  
210 and low-cost product as the final objective. To achieve this goal, several innovations to vaccine  
211 development were implemented and a strong effort of choosing IP free components was prioritized  
212 [36] successfully leading to very low cost of a stable end product after recombinant protein  
213 purification process for large scale production.

214 Delinkage of final product price from the costs of the long R&D phases was privileged by Sm14  
215 being developed at a governmental scientific foundation (FIOCRUZ) from the Brazilian MoH and  
216 strongly supported by funds from public Brazilian financial institutions (FINEP and FAPERJ).

217 After 2005, licensing of FIOCRUZ patent rights to a private company was ruled by specific  
218 contracts designed to protect the accessibility, affordability and supply strategies to lower- and  
219 middle-income countries (LMIC) that are the target areas to receive the anti-Schistosomiasis vaccine.  
220 Presently licensee company, Orygen Biotechnology, a startup owned by Brazilian giant players from  
221 Pharmaceutical sector, is highly contributing to final development of the Sm14 human vaccine under  
222 contracts based on cost plus strategy of pricing vaccines, as adopted by WHO [41].

223 In parallel, the veterinary anti-fasciola vaccine that is being developed in keeping with current  
224 European guidelines to reduce the presence of anti-helminthic drug chemical residues in milk and  
225 meat of livestock, is addressed to rich countries and markets and designed to contribute/support  
226 future potential large scale delivery programs.

227 We are not anymore at a time when anti-Schistosomiasis vaccine is to be discussed, attacked or  
228 delayed, as it was for decades along with all anti-parasite vaccines. Our knowledge about vaccines  
229 improved enormously, as well as technical resources available.

230 Vaccines represent the intervention strategy with the best cost-benefit ratio so far applied in  
231 public health. Likewise, transmission control of infectious/transmissible diseases has only been  
232 achieved through vaccination. Sanitation, chemotherapy and health education are not sufficient to  
233 eliminate parasitic diseases that affect disproportionately people living in endemic areas of poor  
234 countries. Immunization with a safe and effective vaccine, can contribute to a long-term reduction of  
235 egg excretion from the host, truly controlling transmission. So far, there are no vaccines against  
236 parasites that afflict countries fighting to emerge from poverty and reach better conditions of health  
237 and overall development.

238 The Sm14 vaccine against Schistosomiasis is being developed as a humanitarian vaccine to be  
239 included in effective Schistosomiasis transmission control programs and hopefully invert the  
240 paradigm for north-to-south route for technology generation, contributing to the broad use of the  
241 most safe, effective and environment- and human-friendly health-promoting strategy.

## 242 References

- 243 1. Chitsulo, L.; Engels, D.; Montresor, A.; Savioli, L. The global status of schistosomiasis and its control. *Acta*  
244 *Trop.* **2000**, *77*, 41–51, doi:10.1016/S0001-706X(00)00122-4.
- 245 2. Steinmann, P.; Keiser, J.; Bos, R.; Tanner, M.; Utzinger, J. Schistosomiasis and water resources development:  
246 systematic review, meta-analysis, and estimates of people at risk. *Lancet Infect. Dis.* **2006**, *6*, 411–425,  
247 doi:10.1016/S1473-3099(06)70521-7.
- 248 3. Osakunor, D. N. M.; Woolhouse, M. E. J.; Mutapi, F. Paediatric schistosomiasis: What we know and what  
249 we need to know. *PLoS Negl. Trop. Dis.* **2018**, *12*, e0006144, doi:10.1371/journal.pntd.0006144.
- 250 4. Poole, H.; Terlouw, D. J.; Naunje, A.; Mzembe, K.; Stanton, M.; Betson, M.; Lalloo, D. G.; Stothard, J. R.  
251 Schistosomiasis in pre-school-age children and their mothers in Chikhwawa district, Malawi with notes on  
252 characterization of schistosomes and snails. *Parasit. Vectors* **2014**, *7*, 153, doi:10.1186/1756-3305-7-153.
- 253 5. Bergquist, R.; Zhou, X.-N.; Rollinson, D.; Reinhard-Rupp, J.; Klohe, K. Elimination of schistosomiasis: the  
254 tools required. *Infect. Dis. Poverty* **2017**, *6*, doi:10.1186/s40249-017-0370-7.
- 255 6. Coura, J. R.; Amaral, R. S. Epidemiological and control aspects of schistosomiasis in Brazilian endemic  
256 areas. *Mem. Inst. Oswaldo Cruz* **2004**, *99*, 13–19, doi:10.1590/S0074-02762004000900003.
- 257 7. Barbosa, C. S.; Araújo, K. C.; Antunes, L.; Favre, T.; Pieri, O. S. Spatial distribution of schistosomiasis foci  
258 on Itamaracá Island, Pernambuco, Brazil. *Mem. Inst. Oswaldo Cruz* **2004**, *99*, 79–83, doi:10.1590/S0074-  
259 02762004000900014.
- 260 8. Schistosomiasis Available online: <http://www.who.int/news-room/fact-sheets/detail/schistosomiasis>  
261 (accessed on Sep 14, 2018).
- 262 9. Ahuja, A.; Baird, S.; Hicks, J. H.; Kremer, M.; Miguel, E. Economics of Mass Deworming Programs. In *Child*  
263 *and Adolescent Health and Development*; Bundy, D. A. P., Silva, N. de, Horton, S., Jamison, D. T., Patton, G. C., Eds.;  
264 The International Bank for Reconstruction and Development / The World Bank: Washington (DC), 2017 ISBN  
265 978-1-4648-0423-6.
- 266 10. Guidelines for school-based deworming programs Available online:  
267 <https://www.globalpartnership.org/content/guidelines-school-based-deworming-programs> (accessed on Sep  
268 25, 2018).
- 269 11. WHO | School deworming at a glance Available online:  
270 [http://www.who.int/intestinal\\_worms/resources/joint\\_statement\\_WHO\\_World\\_Bank/en/](http://www.who.int/intestinal_worms/resources/joint_statement_WHO_World_Bank/en/) (accessed on Sep 25,  
271 2018).
- 272 12. Molina-Hernández, V.; Mulcahy, G.; Pérez, J.; Martínez-Moreno, Á.; Donnelly, S.; O'Neill, S. M.; Dalton, J.  
273 P.; Cwiklinski, K. Fasciola hepatica vaccine: We may not be there yet but we're on the right road. *Vet. Parasitol.*  
274 **2015**, *208*, 101–111, doi:10.1016/j.vetpar.2015.01.004.
- 275 13. Bergquist, N. R.; Colley, D. G. Schistosomiasis Vaccine: Research to Development. *Parasitol. Today* **1998**, *14*,  
276 99–104, doi:10.1016/S0169-4758(97)01207-6.
- 277 14. WHO | CEWG Demonstration Projects: Background and Process Available online:  
278 [http://www.who.int/phi/implementation/cewg\\_background\\_process/en/](http://www.who.int/phi/implementation/cewg_background_process/en/) (accessed on Sep 25, 2018).

- 279 15. Santini-Oliveira, M.; Coler, R. N.; Parra, J.; Veloso, V.; Jayashankar, L.; Pinto, P. M.; Ciol, M. A.; Bergquist,  
280 R.; Reed, S. G.; Tendler, M. Schistosomiasis vaccine candidate Sm14/GLA-SE: Phase 1 safety and immunogenicity  
281 clinical trial in healthy, male adults. *Vaccine* **2016**, *34*, 586–594, doi:10.1016/j.vaccine.2015.10.027.
- 282 16. Lustigman, S.; Geldhof, P.; Grant, W. N.; Osei-Atweneboana, M. Y.; Sripa, B.; Basáñez, M.-G. A Research  
283 Agenda for Helminth Diseases of Humans: Basic Research and Enabling Technologies to Support Control and  
284 Elimination of Helminthiasis. *PLoS Negl. Trop. Dis.* **2012**, *6*, e1445, doi:10.1371/journal.pntd.0001445.
- 285 17. Tendler, M.; Almeida, M.; Simpson, A. Development of the Brazilian Anti Schistosomiasis Vaccine Based  
286 on the Recombinant Fatty Acid Binding Protein Sm14 Plus GLA-SE Adjuvant. *Front. Immunol.* **2015**, *6*,  
287 doi:10.3389/fimmu.2015.00218.
- 288 18. Scapin, M.; Tendler, M. Immunoprecipitins in human schistosomiasis detected with adult worm antigens  
289 released by 3M KC1. *J. Helminthol.* **1977**, *51*, 71–72, doi:10.1017/S0022149X00007276.
- 290 19. Tendler, M.; Scapin, M. The presence of *Schistosoma mansoni* antigens in solutions used for storing adult  
291 worms. *Rev. Inst. Med. Trop. Sao Paulo* **1979**, *21*, 293–296.
- 292 20. Scapin, M.; Tendler, M.; Messineo, L.; Katz, N. Preliminary studies with a *Schistosoma mansoni* saline  
293 extract inducing protection in rabbits against the challenge infection. *Rev. Inst. Med. Trop. Sao Paulo* **1980**, *22*, 164–  
294 172.
- 295 21. Tendler, M.; Scapin, M.; Tendler, M.; Scapin, M. *Schistosoma mansoni* antigenic extracts obtained by  
296 different extraction procedures. *Mem. Inst. Oswaldo Cruz* **1981**, *76*, 103–109, doi:10.1590/S0074-  
297 02761981000200001.
- 298 22. Tendler, M.; Lima, A. O.; Pinto, R. M.; Cruz, M. Q.; Brascher, H. M.; Katz, N.; Tendler, M.; Lima, A. O.;  
299 Pinto, R. M.; Cruz, M. Q.; Brascher, H. M.; Katz, N. Immunogenetic and protective activity of an extract of  
300 *Schistosoma mansoni*. *Mem. Inst. Oswaldo Cruz* **1982**, *77*, 275–283, doi:10.1590/S0074-02761982000300006.
- 301 23. Tendler, M.; Pinto, R. M.; Bambirra, E. A.; Cruz, M. Q.; Lima, A. O.; Tendler, M.; Pinto, R. M.; Bambirra, E.  
302 A.; Cruz, M. Q.; Lima, A. O. Acquired resistance of mice against *S. mansoni* and lung granulomatous reaction  
303 induced by BCG. *Mem. Inst. Oswaldo Cruz* **1983**, *78*, 147–151, doi:10.1590/S0074-02761983000200003.
- 304 24. Tendler, M.; Magalhães Pinto, R.; Côrtes, M.; Gebara, G. *Schistosoma mansoni*: comparative evaluation of  
305 different routes of experimental infection. *Rev. Inst. Med. Trop. São Paulo* **1985**, *27*, 111–114, doi:10.1590/S0036-  
306 46651985000300001.
- 307 25. Tendler, M.; Pinto, R. M.; Lima, A. O.; Gebara, G.; Katz, N. *Schistosoma mansoni*: Vaccination with adult  
308 worm antigens. *Int. J. Parasitol.* **1986**, *16*, 347–352, doi:10.1016/0020-7519(86)90113-X.
- 309 26. Tendler, M. *Schistosoma mansoni*: protective antigens. *Mem. Inst. Oswaldo Cruz* **1987**, *82*, 125–128,  
310 doi:10.1590/S0074-02761987000800021.
- 311 27. Almeida, M. S. S.; Pinto, R. M.; Noronha, D.; Tendler, M.; Katz, N.; Almeida, M. S. S.; Pinto, R. M.; Noronha,  
312 D.; Tendler, M.; Katz, N. *Schistosoma mansoni* - NZ rabbit-model: resistance due to infection and active  
313 immunization with adult worm antigen. *Mem. Inst. Oswaldo Cruz* **1987**, *82*, 233–233, doi:10.1590/S0074-  
314 02761987000800043.
- 315 28. Almeida, M. S.; Pinto, R. M.; Noronha, D.; Katz, N.; Tendler, M. Curative and protective activity in rabbits  
316 after reinfection with *Schistosoma mansoni*: a new model of immunity? *J. Parasitol.* **1989**, *75*, 308–310.
- 317 29. Tendler, M.; Almeida, M. S.; Pinto, R. M.; Noronha, D.; Katz, N. *Schistosoma mansoni*-New Zealand rabbit  
318 model: resistance induced by infection followed by active immunization with protective antigens. *J. Parasitol.*  
319 **1991**, *77*, 138–141.

- 320 30. Tendler, M.; Pinto, R. M.; de Oliveira Lima, A.; Savino, W.; Katz, N. Vaccination in murine schistosomiasis  
321 with adult worm-derived antigens: Variables influencing protection in outbred mice. *Int. J. Parasitol.* **1991**, *21*,  
322 299–306, doi:10.1016/0020-7519(91)90031-2.
- 323 31. Tendler, M.; Pinto, R. M.; Lima, A. de O.; Savino, W.; Katz, N.; Tendler, M.; Pinto, R. M.; Lima, A. de O.;  
324 Savino, W.; Katz, N. Vaccination in murine schistosomiasis with adult worm derived antigens - II. Protective  
325 and immune response in inbred mice. *Mem. Inst. Oswaldo Cruz* **1992**, *87*, 281–286, doi:10.1590/S0074-  
326 02761992000500053.
- 327 32. Moser, D.; Tendler, M.; Griffiths, G.; Klinkert, M. Q. A 14-kDa *Schistosoma mansoni* polypeptide is  
328 homologous to a gene family of fatty acid binding proteins. *J. Biol. Chem.* **1991**, *266*, 8447–8454.
- 329 33. Tendler, M.; Brito, C. A.; Vilar, M. M.; Serra-Freire, N.; Diogo, C. M.; Almeida, M. S.; Delbem, A. C.; Silva,  
330 J. F. D.; Savino, W.; Garratt, R. C.; Katz, N.; Simpson, A. S. A *Schistosoma mansoni* fatty acid-binding protein,  
331 Sm14, is the potential basis of a dual-purpose anti-helminth vaccine. *Proc. Natl. Acad. Sci.* **1996**, *93*, 269–273,  
332 doi:10.1073/pnas.93.1.269.
- 333 34. Angelucci, F.; Johnson, K. A.; Baiocco, P.; Miele, A. E.; Brunori, M.; Valle, C.; Vigorosi, F.; Troiani, A. R.;  
334 Liberti, P.; Cioli, D.; Klinkert, M.-Q.; Bellelli, A. *Schistosoma mansoni* Fatty Acid Binding Protein: Specificity  
335 and Functional Control as Revealed by Crystallographic Structure. *Biochemistry* **2004**, *43*, 13000–13011,  
336 doi:10.1021/bi048505f.
- 337 35. Pertinhez, T. A.; Sforça, M. L.; Alves, A. C.; Ramos, C. R. R.; Ho, P. L.; Tendler, M.; Zanchin, N. I. T.; Spisni,  
338 A. Letter to the Editor: <sup>1</sup>H, <sup>15</sup>N and <sup>13</sup>C resonance assignments of the apo Sm14-M20(C62V) protein, a mutant  
339 of *Schistosoma mansoni* Sm14. *J. Biomol. NMR* **2004**, *29*, 553–554, doi:10.1023/B:JNMR.0000034355.38944.cf.
- 340 36. Ramos, C. R. R.; Spisni, A.; Oyama, S.; Sforça, M. L.; Ramos, H. R.; Vilar, M. M.; Alves, A. C.; Figueredo, R.  
341 C. R.; Tendler, M.; Zanchin, N. I. T.; Pertinhez, T. A.; Ho, P. L. Stability improvement of the fatty acid binding  
342 protein Sm14 from *S. mansoni* by Cys replacement: Structural and functional characterization of a vaccine  
343 candidate. *Biochim. Biophys. Acta BBA - Proteins Proteomics* **2009**, *1794*, 655–662, doi:10.1016/j.bbapap.2008.12.010.
- 344 37. Giera, M.; Kaiser, M. M. M.; Derks, R. J. E.; Steenvoorden, E.; Kruize, Y. C. M.; Hokke, C. H.; Yazdanbakhsh,  
345 M.; Everts, B. The *Schistosoma mansoni* lipidome: Leads for immunomodulation. *Anal. Chim. Acta* **2018**, *1037*,  
346 107–118, doi:10.1016/j.aca.2017.11.058.
- 347 38. Rodríguez-Pérez, J.; Rodríguez-Medina, J. R.; García-Blanco, M. A.; Hillyer, G. V. *Fasciola hepatica*:  
348 molecular cloning, nucleotide sequence, and expression of a gene encoding a polypeptide homologous to a  
349 *Schistosoma mansoni* fatty acid-binding protein. *Exp. Parasitol.* **1992**, *74*, 400–407.
- 350 39. Almeida, M. S.; Torloni, H.; Lee-Ho, P.; Vilar, M. M.; Thaumaturgo, N.; Simpson, A. J. G.; Tendler, M.  
351 Vaccination against *Fasciola hepatica* infection using a *Schistosoma mansoni* defined recombinant antigen,  
352 Sm14. *Parasite Immunol.* **2003**, *25*, 135–137, doi:10.1046/j.1365-3024.2003.00619.x.
- 353 40. Study of Safety and Immune Response of the Sm14 Vaccine in Adults of Endemic Regions - Full Text View  
354 - ClinicalTrials.gov Available online: <https://clinicaltrials.gov/ct2/show/NCT03041766> (accessed on Sep 17, 2018).
- 355 41. WHO *Guideline on Country Pharmaceutical Pricing Policies*; WHO Guidelines Approved by the Guidelines  
356 Review Committee; World Health Organization: Geneva, 2013;
- 357