

1 Article

2 Explore the potential mechanism of 3 bufadienolides-like chemicals on breast cancer 4 through Bioinformatics analysis

5 Yingbo Zhang^{1,4,#}, Xiaomin Tang^{2,#}, Yuxin Pang^{2,3,*}, Luqi Huang^{3,*}, Dan Wang^{1,4}, Chao Yuan^{1,4}
6 and Xuan Hu^{1,4}

7 ¹ Tropical Crops Genetic Resources Institute, Chinese Academy of Tropical Agricultural Sciences, Danzhou
8 571737, P. R. China

9 ² Guangdong Pharmaceutical University, Guangzhou 510006, P. R. China;

10 ³ National Resource Center for Chinese Materia Medica, China Academy of Chinese Medical Sciences, Beijing
11 100700, P. R. China;

12 ⁴ Hainan Provincial Engineering Research Center for *Blumea Balsamifera*, Danzhou 571737, P. R. China.

13 [#] The author had the same contribution to this work

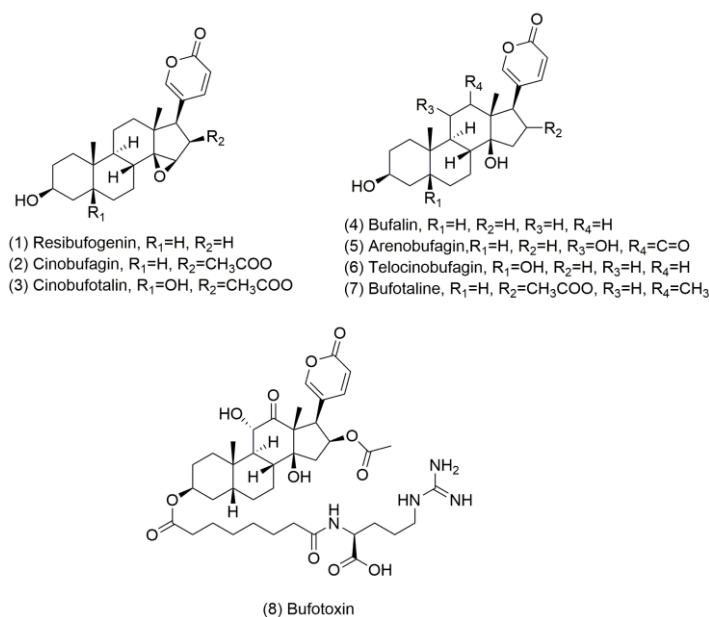
14 ^{*} Correspondence: Yuxin Pang, pyxmarx@126.com; Tel.: +86-898-2330-0268; Luqi Huang,
15 huangluqi01@126.com

16
17 **Abstract:** Bufadienolides-like chemicals, which mostly composed the active ingredient of Chansu,
18 had been widely discovered to possess anti-inflammatory, tumor-suppressing and antipain
19 activity, but the mechanisms of action were not clearly illuminated. In this research, in order to
20 explore the potential mechanism of bufadienolides-like chemicals on breast cancer, a series of
21 bioinformatics analysis, included (1) differentially expressed genes identification combined with
22 gene set variation analysis, (2) tissue specific co-expression network construction, (3) differentially
23 regulated sub-networks detection with disease phenome, (4) hub gene selection and it's relation to
24 survival probability, and (5) similar small molecule detection were performed with gene
25 expression profiles of bufadienolides-like chemicals. Results indicated bufadienolides-like
26 chemicals had the most same target with valproic, estradiol and etc, could disturbed the pathways
27 in RNA splicing, apoptotic process, cell migration, extracellular matrix organization, adherens
28 junction organization, synaptic transmission, Wnt signaling, AK-STAT signaling, BMP signaling
29 pathway and unfolded protein response, and had the potential ability to be used as anticancer,
30 hormones and vasoprotectives agents.

31 **Keywords:** Bufadienolides-like chemicals; Molecular mechanism; Anti-cancer; Bioinformatics
32

33 1. Introduction

34 Despite considerable efforts to the early diagnosis and treatment in the last decade, Breast cancer is still one
35 of the most common malignancies for female worldwide, representing approximately 22% of women's
36 malignancies that pose a threat to women's health [1-4]. In addition to the improvements of early diagnosis,
37 new chemotherapeutic agents and more effective therapies for the treatment become an essential task for
38 improving the mortality of cancer worldwide. Chinese Traditional medicine, had existed and experienced
39 thousands of years of development, and is one of the important sources for antitumor active components
40 screening. Chansu was one of the most famous traditional Chinese medicine, it has been used for centuries
41 in various aspects, such as anaesthesia, antitumor, anti-inflammation and antiarrhythmia [5-8]. The
42 Chansu, mostly come from the glandular secretion dried product of *Bufo gargarizans* Cantor or *B.*
43 *melanostictus* Schneider [6], and including with several group of mixture, such as the resibufogenin, bufalin,
44 arenobufagin, cinobufagin, bufotoxin, telocinobufagin, bufotaline, cinobufotalin, etc [5-7](Figure 1).



45

46

Figure 1. The structural formula of eight bufadienolides-like chemicals

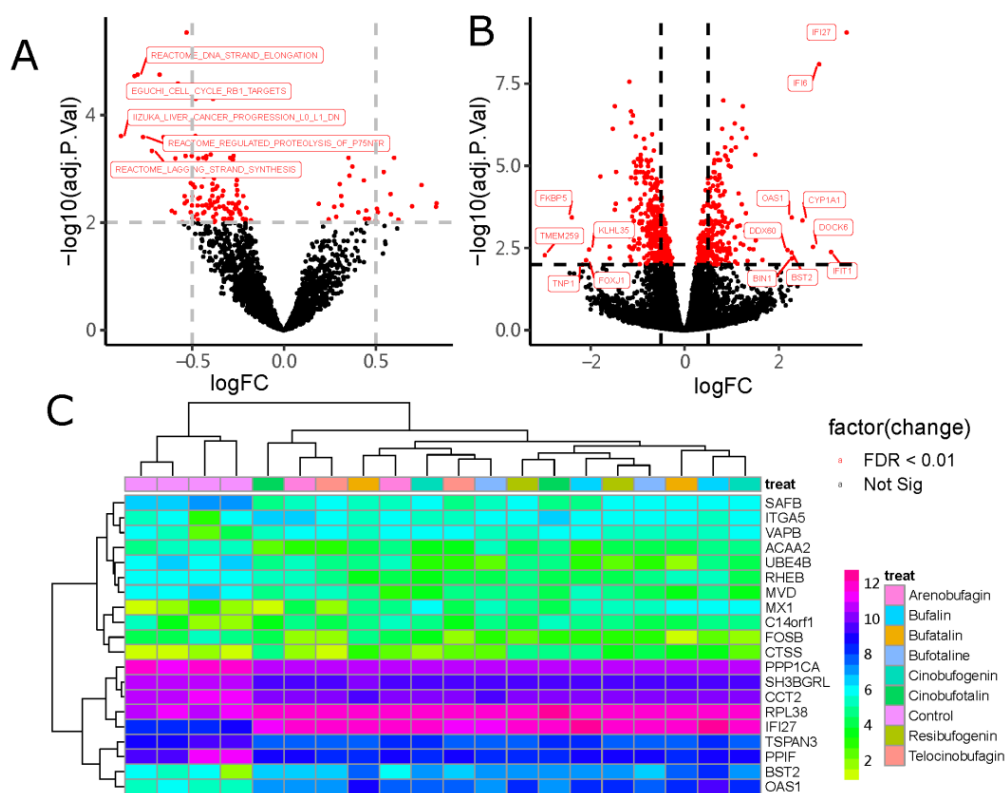
47 In the last decade, lots of research focused on the pharmacological activities and antitumor
 48 activity of bufadienolides-like chemicals. For example, Li et al [9] had reported that cinobufagin has
 49 significant cancer-killing capacity for range of cancers, including HCT116 cells, HT29 cells, A431
 50 cells, PC3 cells, A549 cells, and MCF-7 cells, mechanism analysis showed cinobufagin can induced
 51 tumor cells apoptosis is likely modulated by the hypoxia-inducing factor-1 alpha subunit (HIF-1 α) .
 52 Yeh et al [10] and Yu et al [11] had reported that the bufalin and cinobufagin has a potent inhibiting
 53 effect on androgen dependent and independent prostate cancer cells, also the same results had been
 54 reported by Dong et al [12], Wang et al [13] and Ko et al [14] through HepG2 cells, T24 cells, HeLa
 55 cells, and other cells.

56 These results demonstrate that Chansu is a potent anticancer agent for a range of cancers, but
 57 it's potential anticancer mechanisms has been little reported. The increasing public expression
 58 profile treat with Chinese Traditional medicine, make it possible to extract the most robust
 59 information or potential molecular mechanism from them. In this paper, the gene set variation
 60 analysis (GSVA) algorithm [15] was first used for identifying the differentially expressed genes
 61 (DEGs) and relative enrichment pathways underlying with eight bufadienolides-like chemicals, and
 62 then a serious of bioinformatics analysis, including gene enrichment analysis, tissue specific
 63 co-expression network construction, differentially regulated sub-networks detection relate to breast
 64 cancer phenome, hub gene selection and it's relation to survival probability, and similar small
 65 molecule detection were conducted with the DEGs in the relative enrichment pathways. This work
 66 may reveal the potential mechanism of bufadienolides-like chemicals on breast cancer, especially
 67 differentially regulated sub-networks relate to breast cancer and hub genes disturbed by
 68 bufadienolides-like chemicals, and this work may highlight the potential application of
 69 bufadienolides-like chemicals on Breast cancer, especially as a novel agent for cancer therapy.

70

71 2. Results

72 2.1. Identification of DEGs



73

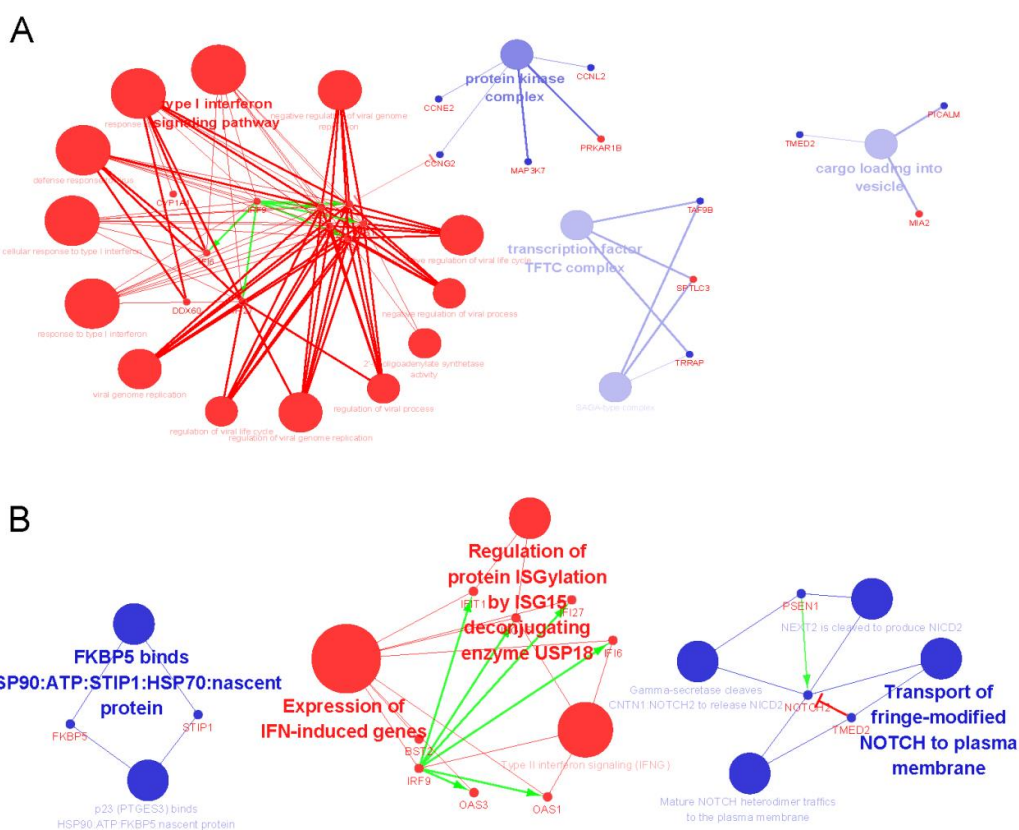
74 **Figure. 2** The DEGs disturbed by bufadienolides-like chemicals through gene set variation analysis (GSVA)
 75 algorithm. (A) The differentially expressed gene sets disturbed by bufadienolides-like chemicals ($|\log_{2}(\text{FC})| \geq$
 76 $\log_{2}(2)$ and $\text{adjPvalue} < 0.001$). (B) The differentially expressed genes (DEGs) relate to differentially expressed
 77 gene sets ($|\log_{2}(\text{FC})| \geq \log_{2}(2)$ and $\text{adjPvalue} < 0.001$). (C) The heatmap of top 20 DEGs disturbed by
 78 bufadienolides-like chemicals.

79 Based on the differentially expressed genes analysis associated with gene sets enrichment
 80 variation analysis strategy, a total of 80 differentially expressed genes (DEGs) involved in the 44
 81 MSigDB C2 curated gene sets were identified (Fig. 1A and Fig. 1B), the top 20 DEGs expression
 82 heatmap is shown in Fig. 1C. Of which, 38 genes involved in the Singh NFE2L2 targets gene sets,
 83 Chang dominant negative gene sets immortalized by HPV31 and Lin silenced gene sets by tumor
 84 microenvironment were up-regulated (Table S1 and Table S2), including IFI6 (interferon-inducible
 85 protein 6), IRF9 (interferon regulatory factor 9), IFIT1 (IFN-induced protein 1 with tetratricopeptide
 86 repeats), ISG15 (Interferon-stimulated gene 15), BST2 (bone marrow stromal cell antigen 2), OAS3
 87 (2'-5'-oligoadenylate synthetase 3), OAS1 (2'-5'-oligoadenylate synthetase 1), DDX60 (DEAD box
 88 polypeptide 60), CYP1A1 (cytochrome P450 1A1), CEACAM6 (carcinoembryonic antigen-related
 89 cell adhesion molecule 6), keratin genes KRT81, and so on.

90 Among the differentially expressed genes associated with enrichment gene sets, 42 genes
 91 involved in the 41 gene sets were down-regulated (Table S1 and Table S2), such as the genes involved
 92 in Iizuka (Table S1) Liver cancer progression pathway, including PPIF (peptidylprolyl isomerase F),
 93 TMED2 (transmembrane trafficking protein 2 with emp24 domain), SAFB (scaffold attachment
 94 factor B), SQLE (squalene epoxidase), PICALM (phosphatidylinositol binding clathrin assembly
 95 protein), STIP1 (stress-induced phosphoprotein 1), CYB561 (cytochromes b561), CCT2 (chaperonin
 96 2β with TCP1 domain), the genes involved in Thum systolic heart failure pathway, including CCNG2
 97 (cyclin G2), TMED2 (transmembrane emp24 domain trafficking protein 2), FH (fumarate hydratase),
 98 TAF9B (ATA-box binding protein associated factor 9b), CCT2 (chaperonin-containing t-complex

99 polypeptide 1 beta), transmembrane receptor NOTCH2, PICALM (subfamily A (MS4A) and
 100 CCNL2 (cyclin L2) , also Reactome DNA strand elongation, Reactome regulated proteolysis of
 101 P75NTR, and other gene sets were downregulated with logFC form - 0.89 ~ - 0.27.

102 In order to obtain a biological interpretation of those genes in GO and KEGG pathway
 103 functional groups, GO and KEGG enrichment analysis were performed with clueGO plug [16] in
 104 Cystoscape [17]. Results indicated, those genes with up-regulated were rich in the terms of type I
 105 interferon signaling response to virus, defense to other organism, regulation of viral genome
 106 replication and 2'-5'-oligoadenylate synthetase activity, and those activate may cause by IRF9, IFI6,
 107 IFI27, ISG15, IFIT1, OAS1 and OAS3 (Fig3a), also the KEGG pathway enrichment analysis those
 108 genes could cause the activate of IFN-induced pathway, type II interferon signaling pathway and
 109 regulation of protein ISGylation by ISG15 deconjugating enzyme USP18 pathway (Fig3B). Those
 110 genes with down-regulated were rich in the terms of protein kinase complex, transcription factor
 111 TFTC complex-1, SAGA- complex and cargo loading into vesicle (Fig3A), further KEGG pathway
 112 enrichment analysis those may negative the transport of fringe-modified NOTCH to plasma
 113 membrane pathway (Fig3B).



114

115 **Figure 3.** The GO and KEGG enrichment result of DEGs disturbed by bufadienolides-like chemicals. (A)

116 Representative biomolecular network of GO enrichment term, the nodes with red colour and bigger size
 117 means the enrichment GO terms with up-regulated genes, the nodes with blue colour and bigger size
 118 means the enrichment GO terms with down-regulated genes, the nodes with red colour and smaller size
 119 means up-regulated genes, the nodes with blue colour and small size means down-regulated genes, undirected edges
 120 means enrichment, the green directed edges means activate from the evidence generated by String database.
 121 the red directed edges means suppressive from the evidence generated by String database, (B) Representative
 122 biomolecular network of KEGG enrichment term, the nodes and edges also had the same means with Figure
 123 3A.

124

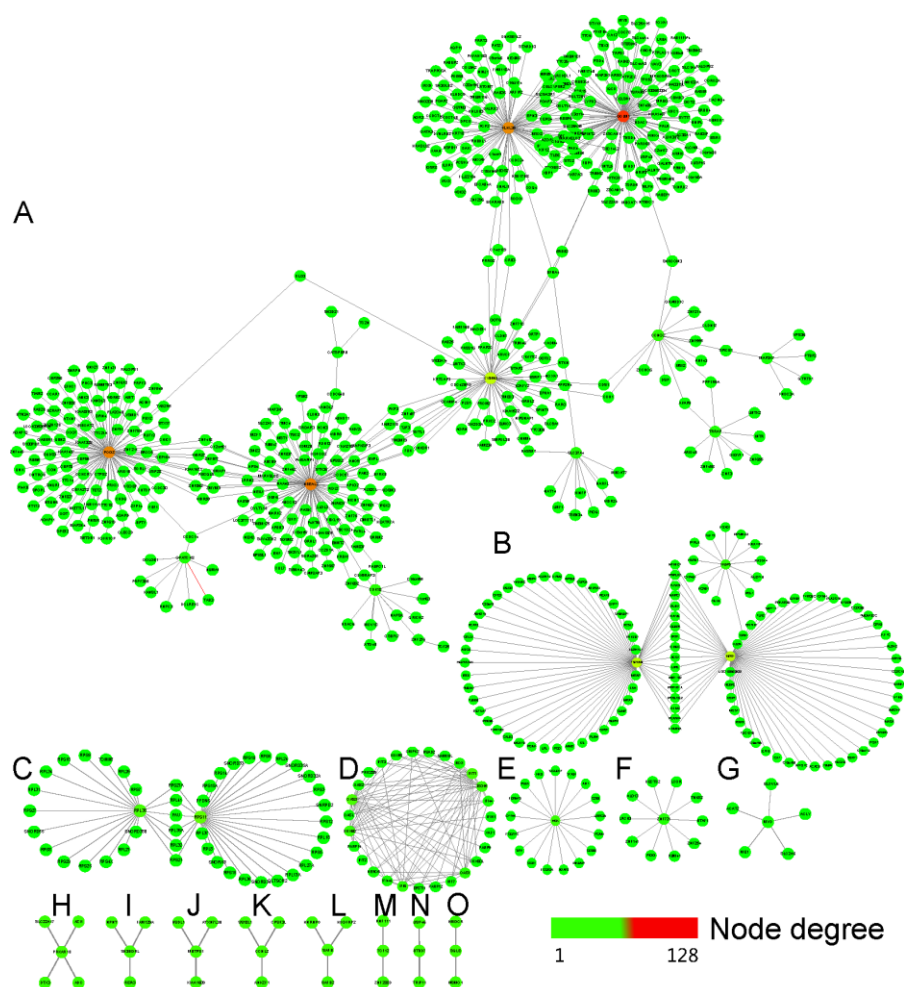
125 **2.2. The tissue specific co-expression network and breast cancer associated subnetwork regulated** 126 **by bufadienolides-like chemicals**

127 It is clear that most of genes exert their function by collaborating with other genes in network which
128 represent rigid molecular machines, cellular structures, or dynamic signaling pathways [18]. In this research, in
129 order to comprehensive understanding the potential function of DEGs involed in Breast cancer, a breast tissue
130 specific co-expression network with DEGs were generated with TCSBN database [19] through
131 NetworkAnalyst web serve [20]. Results indicated the co-expression networks were consisted of 743 nodes and
132 876 edges (Figure 4 and Table1). Furthermore, a functional enrichment analysis with KEGG pathways revealed
133 that the co-expression networks network with DEGs were enriched in pathways related to tight junction,
134 PPAR signaling pathway, mTOR signaling pathway, influenza A, tuberculosis, N-Glycan biosynthesis,
135 terpenoid backbone biosynthesis, Notch signaling pathway, regulation of cyclin-dependent protein kinase
136 activity and steroid biosynthesis (Table 1). Also the GO BP term enrich analysis, those genes mostly involved
137 in Establishment or maintenance of cell polarity, triglyceride metabolic process, protein targeting to membrane,
138 defense response to virus, tuberculosis, post-translational protein modification, coenzyme biosynthetic process,
139 gamete generation, transcription, DNA-dependent, positive regulation of translation, endoplasmic reticulum
140 unfolded protein response, regulation of cyclin-dependent protein kinase activity, steroid biosynthetic process,
141 regulation of the transcription of DNA-dependent, intra-Golgi vesicle-mediated transport term., and other
142 rigid molecular machines in biological process.

143

Table 1. The tissue specific co-expression network regulated by bufadienolides-like chemicals and it's enrichment with GO and KEGG

| Subnetwork Number | Nodes | Edges | Seeds | KEGG Enrichment | | GO Enrichment | |
|-------------------|-------|-------|-------|--|----------|--|----------|
| | | | | KEGG Pathway | P-value | BP term | P-value |
| A | 492 | 558 | 13 | Tight junction | 4.19E-04 | Establishment or maintenance of cell polarity | 2.83E-04 |
| B | 113 | 128 | 3 | PPAR signaling pathway | 7.75E-06 | Triglyceride metabolic process | 1.25E-07 |
| C | 46 | 50 | 2 | mTOR signaling pathway | 9.62E-03 | Protein targeting to membrane | 4.93E-67 |
| D | 27 | 86 | 6 | Influenza A | 3.04E-10 | Defense response to virus | 1.24E-22 |
| E | 18 | 17 | 1 | Tuberculosis | 2.01E-04 | Tuberculosis | 2.01E-04 |
| F | 11 | 10 | 1 | N-Glycan biosynthesis | 9.19E-03 | Post-translational protein modification | 6.33E-03 |
| G | 6 | 5 | 1 | Terpenoid backbone biosynthesis | 1.72E-04 | Coenzyme biosynthetic process | 1.55E-05 |
| H | 5 | 4 | 1 | Notch signaling pathway | 2.98E-02 | Gamete generation | 1.34E-02 |
| I | 4 | 3 | 1 | NA | NA | Transcription, DNA-dependent | 1.31E-02 |
| J | 4 | 3 | 1 | NA | NA | Positive regulation of translation | 1.17E-02 |
| K | 4 | 3 | 1 | NA | NA | Endoplasmic reticulum unfolded protein response | 6.51E-03 |
| L | 4 | 3 | 1 | Regulation of cyclin-dependent protein kinase activity | 1.24E-02 | Regulation of cyclin-dependent protein kinase activity | 1.24E-02 |
| M | 3 | 2 | 1 | Steroid biosynthesis | 7.68E-03 | Steroid biosynthetic process | 2.07E-06 |
| N | 3 | 2 | 1 | NA | NA | Regulation of transcription, DNA-dependent | 1.84E-02 |
| O | 3 | 2 | 1 | NA | NA | Intra-Golgi vesicle-mediated transport | 4.47E-03 |



144

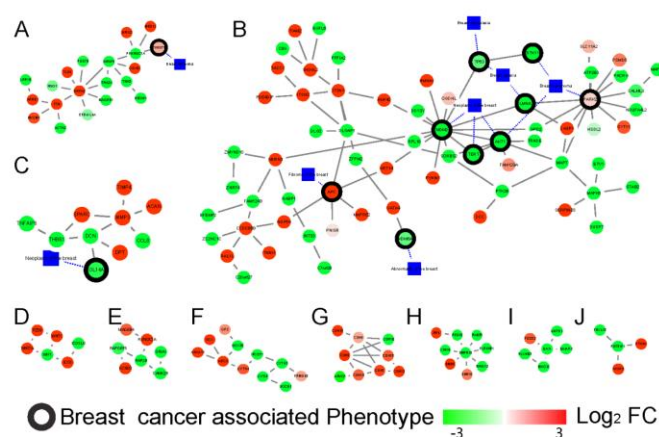
145 **Figure 4.** The breast tissue specific co-expression network with DEGs generated by TCSBN database through
 146 NetworkAnalyst web serve. (A) ~ (O), the Subnetworks of co-expression network origin from the seeds of
 147 DEGs.

148 Based on the novel differentially regulated sub-networks detection tool, PhenomeScape [21], which could
 149 combined the fold changes of genes into the knowledge of networks and disease phenotypes, and then a series
 150 of differentially regulated sub-networks associated with phenotypes were identified with random walk
 151 algorithm. In this research, 7 phenotypes relate to breast cancer were chosen as the seed phenotypes,
 152 subsequently a total of 23 differentially regulated sub-networks enriched in breast cancer phenotype related
 153 subnetwork were identified (Table 2). The Sub-networks distributed by bufadienolides-like chemicals included
 154 RNA splicing (p-value=2.00E-03), apoptotic process (p-value=2.00E-03), extracellular matrix organization
 155 (p-value=1.00E-03), canonical Wnt signaling pathway (p-value=2.20E-02), synaptic transmission
 156 (p-value=1.40E-02), negative regulation of JAK-STAT cascade (p-value=4.20E-02), adherens junction
 157 organization (p-value=3.80E-02), BMP signaling pathway (p-value=4.10E-02), negative regulation of cell
 158 migration (p-value=1.30E-02), activation of signaling protein activity involved in unfolded protein response
 159 (p-value=1.90E-02) (Fig. 4). The subnetwork A (Fig. 5A), relate to the RNA splicing function, was the first
 160 identified dysregulation subnetwork, it could observe the genes involved in mRNA splicing spliceosome were
 161 down regulated, included the serine and arginine rich splicing factor members SRSF4, SRSF5, SRSF6 and
 162 peroxisome proliferator activated receptor gamma coactivator PPARGC1A. The apoptotic process (Fig. 5B),
 163 also could be dysregulated by bufadienolides-like chemicals, and this dysregulation were performed with
 164 the increase expression of SYT11, PARK2, PYHIN1, APC, RNF40, SERPINB3, TIAM2, ITSN1, SH3GL2, CASP1,
 165 GATA4, ITSN2 and PDE4DIP. Several cancer signaling pathway included Wnt signaling pathway, JAK-STAT
 166 signaling pathway and BMP signaling pathway also could had been dysregulated by bufadienolides-like
 167 chemicals (Fig. 5D, 5F and 5H), this may gave further evidence of bufadienolides-like chemicals could increase
 168 the apoptotic process through a series of pathways or regulation network. The Subnetwork C (Fig. 5C), mostly

169 related to the extracellular matrix organization were upregulated, included the genes TIMP4, MMP3, SPARC,
 170 DPT and ACAN, also in this subnetwork the genes referred to the regulation of cell migration were
 171 downregulated, included the genes TNFAIP6, DCN, SPARC, THBS1 and CCL8, this means the increase of
 172 extracellular matrix may hindered the migration of tumor, also the negative of synaptic transmission, adherens
 173 junction organization and regulation of cell migration could find in subnetwork E, G and I (Fig. 5E, 5G and 5I).
 174 Several metabolic process also had been discovered, included the drug metabolic process, xenobiotic metabolic
 175 process, oligosaccharide metabolic process and etc. All other PhenomeScape networks can be found in
 176 Supplementary Fig. 1.

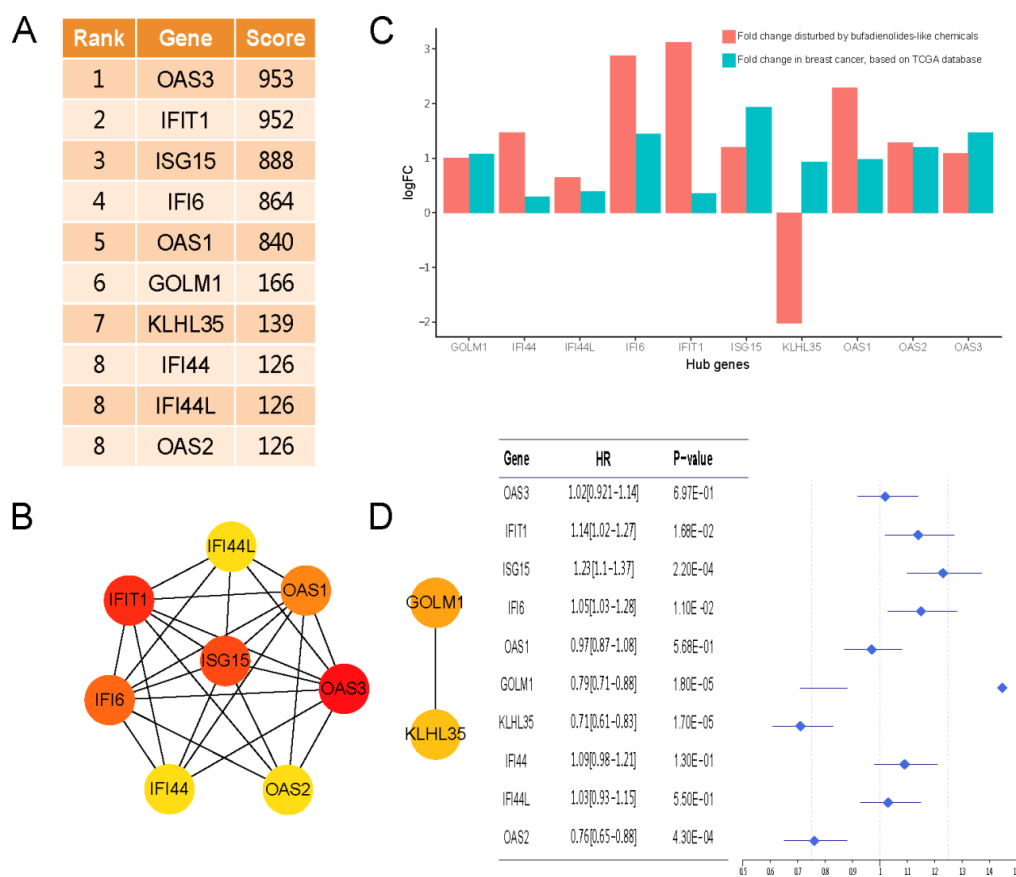
177 **Table 2** Summary of differentially regulated sub-networks disturbed by bufadienolides-like
 178 chemicals

| Subnetwork number | No. of nodes | GO-BP | Empirical P-value |
|-------------------|--------------|--|-------------------|
| A | 21 | RNA splicing | 2.00E-03 |
| B | 73 | apoptotic process | 2.00E-03 |
| C | 11 | extracellular matrix organization | 1.00E-03 |
| D | 6 | canonical Wnt signaling pathway | 2.20E-02 |
| E | 7 | synaptic transmission | 1.40E-02 |
| F | 11 | negative regulation of JAK-STAT cascade | 4.20E-02 |
| G | 9 | adherens junction organization | 3.80E-02 |
| H | 9 | BMP signaling pathway | 4.10E-02 |
| I | 6 | negative regulation of cell migration | 1.30E-02 |
| J | 4 | activation of signaling protein activity involved in unfolded protein response | 1.90E-02 |
| K | 12 | drug metabolic process | 1.20E-02 |
| L | 6 | negative regulation of lipid storage | 4.50E-02 |
| M | 6 | xenobiotic metabolic process | 1.70E-02 |
| N | 8 | relaxation of cardiac muscle | 4.80E-02 |
| O | 5 | very long-chain fatty acid metabolic process | 1.70E-02 |
| P | 4 | oligosaccharide metabolic process | 3.10E-02 |
| Q | 4 | collagen catabolic process | 2.50E-02 |
| R | 4 | response to cocaine | 2.70E-02 |
| S | 4 | behavioral response to nicotine | 4.20E-02 |



180 **Figure 5.** The differentially expressed networks regulated by bufadienolides-like chemicals, and generated by
 181 PhenomeScope plug. Sub-networks linked to breast cancer, RNA splicing(2.00E-03)(A), apoptotic
 182 process(2.00E-03)(B), extracellular matrix organization(1.00E-03)(C), canonical Wnt signaling
 183 pathway(2.20E-02)(D), synaptic transmission(1.40E-02)(E), negative regulation of JAK-STAT
 184 cascade(4.20E-02)(F), adherens junction organization(3.80E-02)(G), BMP signaling pathway(4.10E-02) (H) ,
 185 negative regulation of cell migration(1.30E-02) (I), activation of signaling protein activity involved in unfolded
 186 protein response(1.90E-02) (J). The fold change of the proteins is shown by the node colour and breast cancer
 187 associated phenotype annotated proteins used to generate the sub-networks are shown with a black border.

188 Hub genes, mostly the highly connected nodes in network, were identified by node degree and MCC
 189 algorithm with Cytoscape plugin cytoHubba [22]. Based on the threshold of degree (degree > 5) and MCC
 190 algorithm, 10 genes with MCC scores ranged from 126~953 were identified as hub genes (Figure 6A, Figure 6B).
 191 Among the 10 hub genes, included 3 2'-5'-oligoadenylate synthetase genes OAS1, OAS2 and OAS3, included 5
 192 interferon-induced genes ISG15, IFIT1, IFI6, IFI44 and IFIL44L, also two other genes included the kelch-like
 193 family member 35 (KLHL35) and Golgi Membrane Protein 1 (GOLM1) were selected as the hub genes. Further
 194 investigated with TCGA [23] and Kaplan-Meier database [24] indicates, 10 hub genes except KLHL35, were
 195 increased both in treat with bufadienolides-like chemicals and TCGA Breast cancer sample (Figure 6C) , 6 hub
 196 genes, included IFIT1, ISG15, IFI6, GOLM5, KLHL35 and OAS2 were associated the total survival probability
 197 in Breast cancer patients (Figure 6D). Further analyzed with the correlation between the hub genes and total
 198 survival time in Breast cancer, the high expression of GOLM5, KLHL35 and OAS2 were associated with better
 199 survival probability.



200

201 **Figure. 6** The 10 hub genes and it's correlation with total survival probability in Breast cancer. (A) The 10 hub
 202 genes and it's MCC score. (B) The network of hub genes. (C) The expression correlation with breast cancer,
 203 validated by TCGA database. (D) The total survival probability correlation with breast cancer, validated by
 204 Kaplan-Meier (KM) plotter database.

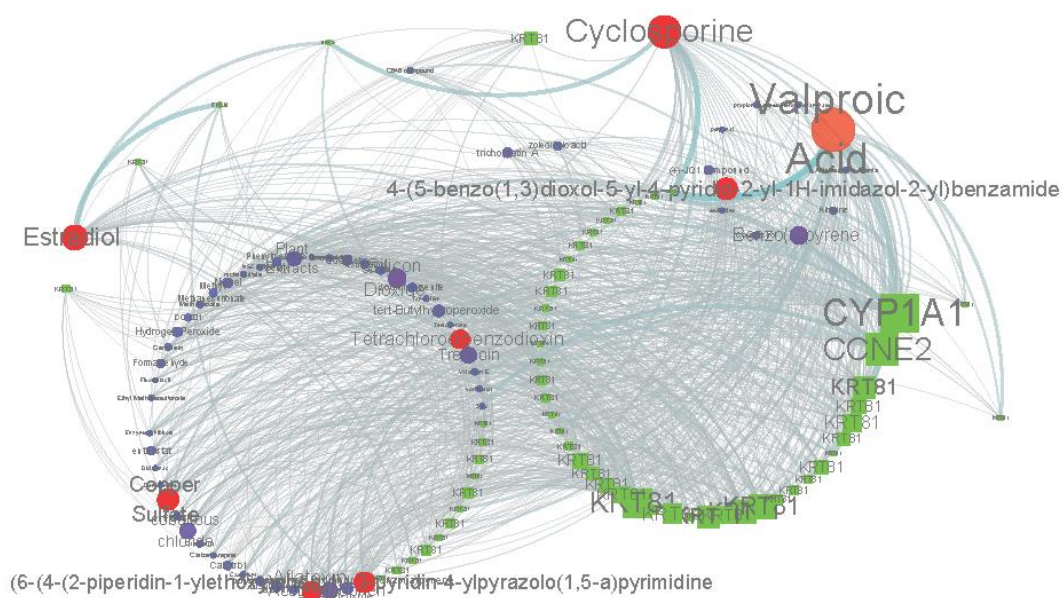
205

206 2.3. Similar small molecule detection

207 Detect the similar small molecule with Comparative Toxicogenomics Database (CTD) [25] and
 208 connectivity map (CMAP2) [26, 27] database will provide a better understanding the molecular mechanism
 209 of bufadienolides-like chemicals, and its potential value of novel agent for cancer therapy. Based on the results
 210 with detecting CTD Database, valproic, cyclospoine and estradiol had the most same target with
 211 bufadienolides-like chemicals (Figure 7). Valproic, a histone deacetylase inhibitor, which once had been widely
 212 used as antiepileptic, and recently also had been proved with anti-cancer activity in vitro/vivo model [28] [29].
 213 Estradiol, a sex hormones with anticancer activity, and also widely used for the treatment of Breast cancer,
 214 especially the postmenopausal women [30-32].

215 Based on the results from connectivity map (CMAP2) database (Table2) [26, 27], the type of V03AF,
 216 G03GB, C05AX and C05CX were the top matching drugs with bufadienolides-like chemicals, V03AF, one type
 217 of detoxifying agents for antineoplastic treatment, had an opposing effect on expression of the
 218 bufadienolides-like chemicals, this result indicated the evidence of bufadienolides-like chemicals had an
 219 potential value of novel agent for cancer therapy. G03GB, one type of sex hormones and modulators of the
 220 genal system, had the mostly same expression profile with bufadienolides-like chemicals, this means the
 221 bufadienolides-like chemicals had the same use of estradiol, epimestrol, cyclofenil in Breast cancer. C05AX and
 222 C05CX, two types of vasoprotectives agents, this means the bufadienolides-like chemicals also had the
 223 potential use of vasoprotectives-like drugs.

224 From the evidence from detecting the similar small molecule with CTD database and CMAP2 database,
 225 indicated bufadienolides-like chemicals were one kinds of steroids with the same physiological activity as
 226 estradiol and G03GB (ATC code), had the potential value for cancer, especially the Breast cancer.



227
 228 **Figure 7.** Chemicals-gene interaction network for the DEGs disturbed by bufadienolides-like chemicals. Square
 229 nodes represent for the DEGs. Circle nodes represent for the chemicals predicted by Comparative
 230 Toxicogenomics Database. The size of nodes represent for the degree. Circle nodes with red represent the
 231 Similar small molecule predicted by degree (degree ≥ 30)

232

233

Table 3. Top 20 CMAP hits correlated with bufadienolides-like chemicals treatment

| Rank | ATC code | Mean score | enrichment | p-value | specificity |
|------|----------|------------|------------|----------|-------------|
| 1 | V03AF | -0.471 | -0.71 | 4.45E-03 | 3.82E-02 |
| 2 | G03GB | 0.449 | 0.655 | 3.29E-02 | 7.47E-02 |
| 3 | C05AX | 0.41 | 0.689 | 1.95E-02 | 4.76E-02 |
| 4 | C05CX | 0.41 | 0.689 | 1.95E-02 | 4.76E-02 |
| 5 | D07XC | -0.372 | -0.661 | 1.44E-03 | 8.10E-03 |
| 6 | N05BE | -0.359 | -0.719 | 1.26E-02 | 1.22E-02 |
| 7 | C08EA | 0.292 | 0.539 | 1.87E-02 | 1.45E-01 |
| 8 | N05AC | 0.259 | 0.365 | 2.32E-03 | 3.90E-01 |
| 9 | D06BB | -0.252 | -0.405 | 9.39E-03 | 1.44E-01 |
| 10 | D06BX | -0.249 | -0.72 | 3.74E-03 | 1.38E-02 |
| 11 | N02BB | 0.244 | 0.404 | 2.71E-03 | 1.75E-02 |
| 12 | N02CX | 0.189 | 0.481 | 3.16E-02 | 4.43E-02 |
| 13 | A07EA | -0.186 | -0.343 | 6.96E-03 | 2.55E-02 |
| 14 | S02BA | -0.167 | -0.383 | 5.03E-03 | 1.31E-02 |
| 15 | B01AC | 0.152 | 0.243 | 2.71E-02 | 1.19E-01 |
| 16 | S03BA | -0.144 | -0.366 | 2.02E-02 | 4.80E-02 |
| 17 | R03BA | -0.141 | -0.29 | 1.19E-02 | 4.00E-02 |
| 18 | S01CB | -0.136 | -0.326 | 1.21E-02 | 2.61E-02 |
| 19 | R01AD | -0.113 | -0.266 | 4.30E-03 | 4.83E-02 |
| 20 | C07AA | -0.109 | -0.262 | 1.14E-02 | 2.22E-01 |

234

235 3. Discussion

236 Recently, the gene expression profile technology, included the microarray and RNA-seq, had
 237 been widely to detect the potential mechanism of chemicals, but an central problem still perplex the
 238 researchers on pharmacology and biology, that is the chemicals how to disturb pathways and
 239 phenotypes through gene and its co-expression network. In this research, with use of the
 240 bioinformatics tools, especially the differentially regulated sub-networks detection tools
 241 PhenomeScape [21], comparative toxicogenomics database [25] and connectivity map [26, 27]
 242 database, revealed several differentially regulated sub-networks treat with bufadienolides-like
 243 chemicals, also the hub genes in co-expression network and its relation to survival probability of
 244 breast cancer, similar small molecule detection and other results may highlight the potential
 245 molecular mechanism and application of bufadienolides-like chemicals on cancer, especially as a
 246 novel agent for Breast cancer.

247 First, during the process of differentially expressed genes identification, in contrast to use the
248 conventional method of differentially expressed genes selection with significance in statistics, a
249 non-parametric unsupervised method of gene set variation analysis were used for differentially
250 expressed genes identification. Results indicated a total of 80 differentially expressed genes (DEGs)
251 involved in the 44 MSigDB C2 curated gene sets were identified (Fig. 1A and Fig. 1B). Further
252 analysis with enrichment of GO and KEGG pathway, we found the genes with up-regulated most
253 rich in interferon signaling response to virus, defense to other organism, regulation of viral genome
254 replication and 2'-5'-oligoadenylate synthetase activity, KEGG pathway enrichment analysis showed
255 those genes could cause the activate of IFN-induced pathway, type II interferon signaling pathway
256 and regulation of protein ISGylation. But the genes with down-regulated were rich in the terms of
257 protein kinase complex, transcription factor TF1C complex-1, SAGA- complex and cargo loading
258 into vesicle, KEGG pathway enrichment analysis showed those genes may involve in negative the
259 transport of fringe-modified NOTCH to plasma membrane pathway. Compare the method with
260 statistical significance, those differentially expressed genes in gene set variation maybe much less,
261 but with more same participate in same pathway or biology function, also the same results had been
262 proved by the examples of GSEA package [15].

263 Second, during the process of co-expression network reconstruction and dysregulated
264 sub-networks detection, a novel plug of PhenomeScape was used, which could combine the data of
265 gene expression into the knowledge of protein-protein interaction networks and disease phenotype
266 [21]. During the analysis with damaged osteoarthritic cartilage gene expression profile, several
267 significant sub-networks related to damaged osteoarthritic cartilage were identified, including
268 mitotic cell cycle, Wnt signalling, apoptosis and matrix organisation [33, 34]. In this research, with
269 PhenomeScape tool [21], a total of 23 differentially regulated sub-networks were identified, and 10
270 sub-networks had been proved to relate to breast cancer by evidence, included RNA splicing,
271 apoptotic process, cell migration, extracellular matrix organization, adherens junction organization,
272 synaptic transmission and so on.

273 Third, during the process of similar small molecule detection, Comparative Toxicogenomics
274 Database (CTD) [25] and connectivity map (CMAP2) [26, 27] database were used. Results indicated
275 bufadienolides-like chemicals had the same effect with valproic and estradiol, valproic, a histone
276 deacetylase inhibitor, it had been proved to inhibit proliferation through Wnt/ β catenin signalling
277 activation. The estradiol, also had been proved to with anticancer activity, especially the
278 postmenopausal women. Also the evidence form connectivity map database indicated
279 bufadienolides-like chemicals had the potential ability to be used as anticancer, hormones and
280 vasoprotectives agents.

281 During the hub gene selection and it's relation to survival probability indicated 10 hub genes
282 except KLHL35, were increased both Breast cancer and samples treat with bufadienolides-like
283 chemicals, further analysis with relation to total survival probability, 6 hub genes, included IFIT1,
284 ISG15, IFI6, GOLM5, KLHL35 and OAS2 were associated the total survival time and high expression
285 of GOLM5, KLHL35 and OAS2 were associated with better survival probability.

286 4. Materials and Methods

287 4.1 Microarray data information

288 The gene expression profiles of GSE85871 (<https://www.ncbi.nlm.nih.gov/gds/>), which is an
289 gene expression profile treat with 102 Chinese traditional medicine, and it was based on Affymetrix
290 GPL571 platform (Affymetrix Human Genome U133A 2.0 Array), was submitted by Lv et al [35].

291 In this study, the raw data of 4 controls and 14 samples treat with bufadienolides-like
292 chemicals, including resibufogenin, bufalin, arenobufagin, cinobufagin, bufotoxin, telocinobufagin,
293 bufotaline and cinobufotali, were downloaded from GEO database through GEOquery [36]
294 packages in R3.5.1 [37] environment.
295

296 4.2 Identification of DEGs associated with relative enrichment pathways

297 In order to obtain the biological interpretation of differentially expressed genes (DEGs)
298 disturbed by bufadienolides-like chemicals, a novel R package GSVA [15] was employed, which
299 allows the assessment of the DEGs underlying pathway activity variation by transforming the gene
300 expression profile into the prior knowledge of gene set. In accordance with MIAME standards [38,
301 39], the differentially expressed genes (DEGs) disturbed by bufadienolides-like chemicals were
302 identified by a series of standard flow with R environment. First, the quality assessments,
303 background correction and normalization were preprocessed and normalization with affy [40] and
304 gcrma [41] packages. Then, the batch effects were examined and removed out with combat and sva
305 functions in SVA package [42]. Subsequently, a non-specific probes filtering step were carried out
306 with nsFilter function in the genefilter package [43], the quality control probes of Affymetrix,
307 probesets without Entrez ID annotation, probesets whose associated Entrez ID is duplicated in the
308 annotation and the top 20% with smaller variability were removed. Finally, the GSVA [15],
309 GSEABase [44], limma [45] package and c2BroadSets from Molecular Signatures Database
310 (MSigDB) [46, 47] were used for the selection for DEGs with relative enrichment pathways.

311 During the process of DEGs selection with relative enrichment sets, the gene expression profile
312 was first transformed into the prior knowledge gene set of c2BroadSets and the enrich gene sets
313 were selection with the screening criteria of $FDR < 0.01$. Then the different genes enrichment in the
314 c2BroadSets gene sets were selected with limma [45] package, and the screening criteria were set
315 with $FDR < 0.01$ and $|\log FC| > 1$, and those DEGs associated with relative enrichment pathways
316 were used for further analyzed and validated

317 During the process of DEGs identification, the Biobase [48] package and GSVAdata [49] package
318 were also applied. The results were visualized with ggplot2 [50], ggpubr [51], pheatmap [52] and
319 cowplot [53] package.

320 4.3 Gene enrichment analysis

321 On the bias of the DEGs selection associated with relative enrichment sets, in order to obtain a
322 comprehensive understanding of those genes involved in the prior knowledge of gene sets, GO and
323 KEGG enrichment analysis were performed with clueGO plug [16] in Cytoscape [17]. The
324 significantly enrich GO terms and KEGG pathways were calculated by the hypergeometric test [54],
325 and cut-off criteria was set as $FDR < 0.05$. Another statistical parameter of Kappa Score were set as
326 middle stringency, its means the terms in network were combined with middle related terms as
327 based on their overlapping genes. The min percentage and min genes enriched in GO terms or
328 KEGG pathways were set as 1.0% and 2, also the term fusion was chosen. Other options, including
329 the statistical options, reference options, grouping options and visual options were set with default
330 setting.

331

332 4.4 Gene co-expression network analysis and disease phenotype association

333 In order to comprehensive understanding the potential mechanism of DEGs in involved in
 334 Breast cancer, co-expression network analysis, phenome association and survival correlation
 335 analysis were investigated with NetworkAnalyst database [20] and PhenomeScape plug [21] in
 336 Cytoscape [17], also other plugs and databases including the cytoHubba [22] , TCSBN database
 337 [19], TCGA database [23] and Kaplan-Meier (KM) plotter database [24] and Phenomiser [55] web
 338 tool were also used for hub genes selection and survival correlation analysis. First, the breast
 339 mammary tissue specific co-expression networks were investigated with TCSBN database through
 340 NetworkAnalyst web server, also the GO and KEGG enrichment terms of networks were also
 341 investigated with NetworkAnalyst web server. Subsequently, the differentially regulated
 342 sub-networks enriched in genes associated with breast cancer phenotype were identified by
 343 random sampling (10,000 sub-networks) methods with PhenomeScape plug and Phenomiser web
 344 tool. First, through the search with Phenomiser web tool and the manual of UberPheno ontology
 345 [55], six phenotypes (Table 4) were chosen as the breast cancer association phenotype, and then
 346 with the parameters of maximum initial sub-network size of 7 and an empirical P-value threshold
 347 of 0.05 were used for filtering the differentially regulated sub-networks enriched in genes associated
 348 with breast cancer phenotype.

349 **Table 4.** UberPheno phenotype terms selected for differentially regulated sub-network detection in

350

co-expression network

| Phenotype ID | Phenotype Description |
|--------------|---------------------------|
| HP:0100783 | Breast aplasia |
| HP:0100013 | Neoplasm of the breast |
| HP:0003002 | Breast carcionma |
| HP:0003187 | Breast hypoplasia |
| HP:0000769 | Abnormality of the breast |
| HP:0010619 | Fibroma of the breast |

351 Hub genes, highly interconnected with nodes in network, have been considered functionally
 352 significant in network. In our study, the top 10 hub genes were defined by node degree and MCC
 353 algorithm in Cytoscape plugin cytoHubba [22]. Reference the previously described workflow of
 354 selection the essential proteins from the yeast protein interaction network with MCC algorithm [22].
 355 First, the degrees of nodes were computed by NetworkAnalyzer [56] in Cytoscape. Then the node
 356 with degree greater than a threshold were choosed as potential candidate Hub genes, and the
 357 threshold is the maximum integer as $2 \times \sum_{v \in V, Deg(v) > t} Deg(v) > \sum_{v \in V, Deg(v)}$, where v is the
 358 collection of nodes within the network V , $Deg(v)$ is the degree of node v . Last, the top 10 hub
 359 genes were ranked by MCC algorithm in cytoHubba plugin. Hub genes common in breast tissue
 360 co-expression networks were chosen as the candidates to be further analyzed and validated with

361 TCGA [23] and Kaplan-Meier (KM) plotter database (<http://kmplot.com/analysis/>) [24].

362 4.5 Similar small molecule detection

363 In order to detect the similar small molecule with bufadienolides-like chemicals, the DEGs with
364 up or down were respectively submitted to the comparative toxicogenomics database (CTD) [25]
365 and connectivity map (CMAP2, <http://www.broadinstitute.org/cMAP/>) database [26, 27]. During
366 the process of detection similar small molecule with CTD Database, the threshold of degree in the
367 degree filter network was set as 10. During the process of detection similar small molecule with
368 connectivity map database, the enrichment score and p-value of were choose as similarity index
369 between the gene expression profile of the query signature and that of chemicals in CMAP2.

370 Also the potential toxicity same as bufadienolides-like chemicals were also detected by CEBS
371 database (<https://manticore.niehs.nih.gov/cebssearch/>) [57], but there is no evidence to prove the
372 bufadienolides-like chemicals with obvious toxicity.

373 5. Conclusions

374 In this research, with a serious of bioinformatics analysis, we take notice the bufadienolides-like
375 chemicals may perform anticancer activity through RNA splicing, apoptotic process, cell migration,
376 extracellular matrix organization, adherens junction organization, synaptic transmission, Wnt
377 signaling, AK-STAT signaling, BMP signaling pathway and unfolded protein response, and those
378 may highlight the potential molecular mechanism of bufadienolides-like chemicals on Breast cancer,
379 but still there are several problem had better solution, the toxicity of bufadienolides-like chemicals,
380 especially the cardiotoxicity, which had been widely observe from clinic. The second problem, the
381 difference of potential molecular mechanism among bufadienolides-like chemicals also had been
382 clear illuminated in this research.

383 **Supplementary Materials:** The following are available online, Figure S1: Other differentially expressed
384 networks regulated by bufadienolides-like chemic, Table S1: The DEGs disturbed by bufadienolides-like
385 chemicals, Table S2: The different gene sets disturbed by bufadienolides-like chemicals.

386 **Author Contributions:** Y. P., L. H., and Y. Z. designed the flow of this analysis; D. W., and X. H. collected the
387 gene expression profile information; X. T., and Y. Z. performed the analysis; Y. Z., and C. Y. Discussed the
388 results. Y. Z., and X. T., wrote the manuscript. All of the authors made important suggestions to the manuscript
389 and reviewed and approved the manuscript.

390 **Funding:** This work was supported by grants from National Natural Science Foundation of China (No.
391 81374065 and No. 81403035), and Basal Research Fund of Central Public-interest Scientific Institution (No.
392 1630032015039).

393 **Acknowledgments:** We thank Doctor Fulai Yu and Xiaolu Chen, for the advice and review of the manuscript.

394 **Conflicts of Interest:** The following authors report no conflicts of interest.

395 References

- 396 1. Mai, F.T. and H.A. Omar, *Immunotherapy, an evolving approach for the management of triple negative breast*
397 *cancer: Converting non-responders to responders*. *Critical Reviews in Oncology/hematology*, 2018. **122**.
- 398 2. Zhou, H., et al., *RING1 and YY1 binding protein suppresses breast cancer growth and metastasis*. *International*
399 *Journal of Oncology*, 2016. **49**(6): p. 2442.
- 400 3. Pan, Z., et al., *SATB1 is Correlated with Progression and Metastasis of Breast Cancers: A Meta-Analysis*. *Cellular*
401 *Physiology & Biochemistry*, 2016. **38**(5): p. 1975-1983.

- 402 4. Xu, H., et al., *CD44 correlates with clinicopathological characteristics and is upregulated by EGFR in breast cancer*.
403 *International Journal of Oncology*, 2016. **49**(4): p. 1343-1350.
- 404 5. Akiko, E., et al., *Inhibitory effects of bufadienolides on interleukin-6 in MH-60 cells*. *Journal of Natural Products*,
405 2004. **67**(12): p. 2070-2072.
- 406 6. Hong, Z., ., K. Chan, ., and H.W. Yeung, *Simultaneous determination of bufadienolides in the traditional Chinese*
407 *medicine preparation, liu-shen-wan, by liquid chromatography*. *Journal of Pharmacy & Pharmacology*, 2011.
408 **44**(12): p. 1023-1026.
- 409 7. Tian-Jie, et al., *Efficacy and safety of gemcitabine-oxaliplatin combined with huachansu in patients with advanced*
410 *gallbladder carcinoma*. *World Journal of Gastroenterology*, 2008. **14**(33): p. 5210-5216.
- 411 8. Wang, J., et al., *Involvement of caspase-3 activity and survivin downregulation in cinobufocini-induced apoptosis in*
412 *A 549 cells*. *Experimental Biology & Medicine*, 2009. **234**(5): p. 566-572.
- 413 9. Chun, L., et al., *The mechanisms of chansu in inducing efficient apoptosis in colon cancer cells*. *Evidence-Based*
414 *Complementray and Alternative Medicine*, 2013,(2013-5-30), 2013. **2013**(12): p. 849054.
- 415 10. Jiun-Yih, Y., et al., *Effects of bufalin and cinobufagin on the proliferation of androgen dependent and independent*
416 *prostate cancer cells*. *Prostate*, 2010. **54**(2): p. 112-124.
- 417 11. Yu, C., et al., *Apoptotic signaling in bufalin- and cinobufagin-treated androgen-dependent and -independent human*
418 *prostate cancer cells*. *Cancer Science*, 2010. **99**(12): p. 2467-2476.
- 419 12. Q., D.Y., et al., *Effect of cinobufagin on nuclear factor-kappa B pathway in HepG2 cells*. *ournal of Southern Medical*
420 *University*, 2010. **30**(01): p. 137-139+142.
- 421 13. Ko, W.S., et al., *Induction of apoptosis by Chansu, a traditional Chinese medicine, in human bladder carcinoma T24*
422 *cells*. *Oncology Reports*, 2005. **14**(2): p. 475-480.
- 423 14. Wang, L., et al., *Pilot study on the mechanisms of growth inhibitory effect of cinobufagin on HeLa cells*. *Chinese*
424 *Journal of Oncology*, 2005. **27**(12): p. 717.
- 425 15. Haenzelmann and Sonja, *GSVA: gene set variation analysis for microarray and RNA-Seq data*. *Bmc*
426 *Bioinformatics*, 2013. **14**(1): p. 7-7.
- 427 16. Bindea, G., et al., *ClueGO: a Cytoscape plug-in to decipher functionally grouped gene ontology and pathway*
428 *annotation networks*. *Bioinformatics*, 2009. **25**(8): p. 1091-1093.
- 429 17. Su, G., et al., *Biological network exploration with cytoscape 3*. *Curr Protoc Bioinformatics*, 2014. **47**: p. 8.13.1.
- 430 18. Barabási, A.L. and Z.N. Oltvai, *Network biology: Understanding the cell's functional organization*. *Nature*
431 *Reviews Genetics*, 2004. **5**(2): p. 101- 113.
- 432 19. Lee, S., et al., *TCSBN: a database of tissue and cancer specific biological networks*. *Nucleic Acids Research*, 2017.
433 **46**(Database issue): p. gkx994.
- 434 20. Xia, J., E.E. Gill, and R.E.W. Hancock, *NetworkAnalyst for statistical, visual and network-based meta-analysis of*
435 *gene expression data*. *Nature Protocols*, 2015. **10**(6): p. 823-844.
- 436 21. Soul, J., et al., *PhenomeScope: a cytoscape app to identify differentially regulated sub-networks using known disease*
437 *associations*. *Bioinformatics*, 2016. **32**(24): p. 3847–3849.
- 438 22. Chin, C.H., et al., *cytoHubba: identifying hub objects and sub-networks from complex interactome*. *Bmc Systems*
439 *Biology*, 2014. **8 Suppl 4**(S4): p. S11.
- 440 23. Kosinski M, B.P., *RTCGA: The Cancer Genome Atlas Data Integration*. *R package version 1.12.0*. 2018.
- 441 24. Balazs, G., et al., *An online survival analysis tool to rapidly assess the effect of 22,277 genes on breast cancer prognosis*
442 *using microarray data of 1,809 patients*. *Breast Cancer Res Treat*, 2010. **123**(3): p. 725-731.
- 443 25. Davis A. P, G.C.J., Johnson R. J., Sciaky D., King B. L., McMorran R., Wiegiers J., Wiegiers T. C., Mattingly C.
444 J., *The Comparative Toxicogenomics Database: update 2017*. *Nucleic Acids Res.*, 2017. **45**(D1): p. D972-D978.

- 445 26. Justin, L., et al., *The Connectivity Map: using gene-expression signatures to connect small molecules, genes, and*
446 *disease*. *Science*, 2006. **313**(5795): p. 1929-1935.
- 447 27. Subramanian, A., et al., *A Next Generation Connectivity Map: L1000 Platform and the First 1,000,000 Profiles*.
448 *Cell*, 2017. **171**(6): p. 1437-1452.
- 449 28. Minegaki T., S.A., Mori M., Tsuji S., Yamamoto S., Watanabe A., Tsuzuki T., Tsunoda T., Yamamoto A.,
450 Tsujimoto M., Nishiguchi K., *Histone deacetylase inhibitors sensitize 5-fluorouracil-resistant MDA-MB-468 breast*
451 *cancer cells to 5-fluorouracil*. *Oncology letters*, 2018. **16**(5): p. 6.
- 452 29. Riva G., C.C., Bazzoni R., Cadamuro M., Negroni C., Butta V., Strazzabosco M., Dalprà L., Lavitrano M.,
453 Bentivegna A., *Valproic Acid Inhibits Proliferation and Reduces Invasiveness in Glioma Stem Cells Through Wnt/ β*
454 *Catenin Signalling Activation*. *Genes (Basel)*, 2018. **9**(11): p. E522.
- 455 30. Bennink, H.J.T.C., et al., *The use of high-dose estrogens for the treatment of breast cancer*. *Maturitas*, 2017. **95**: p.
456 11-23.
- 457 31. Hankinson, S.E., et al., *Plasma sex steroid hormone levels and risk of breast cancer in postmenopausal women*. *J Natl*
458 *Cancer Inst*, 1998. **91**(8): p. 1292-1299.
- 459 32. Mdc, B.O., et al., *Use of anastrozole in the chemoprevention and treatment of breast cancer: A literature review*.
460 *Revista Da Associacao Medica Brasileira*, 2017. **63**(4): p. 371.
- 461 33. Soul J., H.T.E., Boot-Handford R. P., Schwartz J. M., *PhenomeExpress: A refined network analysis of expression*
462 *datasets by inclusion of known disease phenotype*. *Scientific Reports*, 2015(5): p. 8117.
- 463 34. Dunn S. L., S.J., Anand S., Schwartz J. M., Boot-Handford R. P., Hardingham T. E., *Gene expression changes in*
464 *damaged osteoarthritic cartilage identify a signature of non-chondrogenic and mechanical responses*. *Osteoarthritis &*
465 *Cartilage*, 2016. **24**(8): p. 9.
- 466 35. Chao, L., et al., *The gene expression profiles in response to 102 traditional Chinese medicine (TCM) components: a*
467 *general template for research on TCMs*. *Sci Rep*, 2017. **7**(1): p. 352.
- 468 36. Sean, D. and P.S. Meltzer, *GEOquery: a bridge between the Gene Expression Omnibus (GEO) and BioConductor*.
469 *Bioinformatics*, 2007. **23**(14): p. 1846-7.
- 470 37. Team, R.C., *R: A language and environment for statistical computing*. *R Foundation for Statistical Computing,*
471 *Vienna, Austria*. . 2018.
- 472 38. A., B., *Minimum Information About a Microarray Experiment (MIAME)--successes, failures, challenges*. *Scientific*
473 *World Journal*., 2009. **9**: p. 3.
- 474 39. Dondrup, M., et al., *EMMA 2 – A MAGE-compliant system for the collaborative analysis and integration of*
475 *microarray data*. *Bmc Bioinformatics*, 2009. **10**(1): p. 50.
- 476 40. Gautier, L., B.M. Cope LBolstad, and R.A. Irizarry, *affy - analysis of Affymetrix GeneChip data at the probe level*.
477 *Bioinformatics*, 2004. **20**(3): p. 307-315.
- 478 41. Gharaibeh, R.Z., A.A. Fodor, and C.J. Gibas, *Background correction using dinucleotide affinities improves the*
479 *performance of GCRMA*. *Bmc Bioinformatics*, 2008. **9**(1): p. 452-452.
- 480 42. Leek, J.T., et al., *The sva package for removing batch effects and other unwanted variation in high-throughput*
481 *experiments*. *Bioinformatics*, 2012. **28**(6): p. 882-883.
- 482 43. Gentleman R, C.V., Huber W, Hahne F, *genefilter: genefilter: methods for filtering genes from high-throughput*
483 *experiments*. *R package version 1.64.0*. 2018.
- 484 44. Martin Morgan, S.F.a.R.G., *GSEABase: Gene set enrichment data structures and methods*. *R package version 1.44.0*.
485 2018.
- 486 45. Ritchie, M.E., et al., *limma powers differential expression analyses for RNA-sequencing and microarray studies*.
487 *Nucleic Acids Research*, 2015. **43**(7): p. e47.

- 488 46. Arthur, L., et al., *Molecular signatures database (MSigDB) 3.0*. *Bioinformatics*, 2011. **27**(12): p. 1739.
- 489 47. Liberzon, A., et al., *The Molecular Signatures Database (MSigDB) hallmark gene set collection*. *Cell Syst*, 2015.
- 490 1(6): p. 417-425.
- 491 48. Wolfgang, H., et al., *Orchestrating high-throughput genomic analysis with Bioconductor*. *Nature Methods*, 2015.
- 492 12(2): p. 115-21.
- 493 49. Castelo, R., *GSVAdata: Data employed in the vignette of the GSVA package*. 2018.
- 494 50. Wickham, H., *ggplot2: Elegant Graphics for Data Analysis*. Springer-Verlag New York., 2016.
- 495 51. Kassambara, A., *ggpubr: 'ggplot2' Based Publication Ready Plots*. 2018.
- 496 52. Kolde, R., *pheatmap: Pretty Heatmaps*. 2018.
- 497 53. Wilke, C.O., *cowplot: Streamlined Plot Theme and Plot Annotations for 'ggplot2'*. 2018.
- 498 54. Berkopec, A., *HyperQuick algorithm for discrete hypergeometric distribution*. *Journal of Discrete Algorithms*,
- 499 2007. **5**(2): p. 341-3472004.
- 500 55. Sebastian, K.H., et al., *Clinical diagnostics in human genetics with semantic similarity searches in ontologies*.
- 501 *American Journal of Human Genetics*, 2009. **85**(4): p. 457-464.
- 502 56. Yassen, A., et al., *Computing topological parameters of biological networks*. *Bioinformatics*, 2008. **24**(2): p. 282-4.
- 503 57. Lea, I.A., et al., *CEBS: a comprehensive annotated database of toxicological data*. *Nucleic Acids Research*, 2017.
- 504 **45**(D1): p. D964.