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Review

# Pharmacological Significance, Medicinal Use and Toxicity of Extracted and Isolated Compounds from *Euphorbia* Species Found in Southern Africa: A Review

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**Abstract:** This study documents Euphorbiaceae plants in Southern Africa, focusing on their traditional medicinal uses, pharmacological properties, toxicity, and active secondary metabolites. Literature from scientific journals, books, dissertations, and conference papers from 1962 to 2023 was reviewed for 15 *Euphorbia* species. Recent findings indicate that certain compounds in *Euphorbia* plants have significant biological and pharmacological properties. However, the white sticky latex sap they contain is highly toxic yet can also have medicinal applications. Phytochemical analysis has shown these plants exhibit beneficial effects, including antibacterial, antioxidant, antiproliferative, anticancer, anti-inflammatory, antiviral, antifungal, and anti-HIV activities. Key phytochemicals such as euphol, cycloartenol, tirucallol, and triterpenoids contribute to their therapeutic efficacy, along with various proteins like lectin and lysozyme. Despite some Euphorbiaceae species being screened for medicinal compounds, many have not been thoroughly examined, highlighting a critical gap. Given their historical usage, further research is essential to evaluate the medicinal significance of *Euphorbia* species through detailed studies of isolated compounds and their pharmacokinetics and pharmacodynamics. This research will serve as a valuable resource for future inquiries into the benefits of lesser-studied *Euphorbia* species.

**Keywords:** *Euphorbia* species; toxicity; secondary metabolites; pharmacological properties; medicinal plants; cancer

## 1. Introduction

The Euphorbiaceae is a highly varied group of flowering plants, having over 300 genera and 8 000 species [1]. Within this family, the *Euphorbia* genus stands out as one of the largest, with more than 2 000 species [2]. These species can take the form of herbs, shrubs and trees, sometimes resembling succulents or cacti. They are mainly found in tropical and subtropical regions of Africa and America, as noted by Adedapo et al. [3]. Many species in this family produce a toxic white sap, but this sap can also have medicinal properties [4]. The genus contains economically significant species, making it a vital genus with high research potential.

South Africa has a rich collection of *Euphorbia* species, with at least 188 indigenous to the country (SANBI). The *Euphorbia* genus has attracted the attention of many researchers due to its diverse chemical compositions, which include euphol, triterpenoids, diterpene ester and tirucallol. These compositions have been found to have remarkable therapeutic properties such as antimicrobial, antidiabetic, antiviral and anticancer properties, as noted by Betancur-Galvis et al. [5] and also Mwine and Damme [6]. However, there is a lack of scientific documentation on the anticancer properties of many *Euphorbia* species in Southern Africa. This is due to a lack of research on isolated compounds, as well as a lack of testing and discovery of new compounds. Furthermore, there is limited research on unexplored *Euphorbia* species that may have potential anticancer properties. Additionally, the majority of published articles on *Euphorbia* focus on already explored species, with little to no information on the unexplored ones [7,8]. It is worth noting that there is currently only one comprehensive review available on the *Euphorbia* species found in Southern Africa. This review highlights the limited scientific documentation available on these plants and emphasizes the need for further research to unlock their full potential. The review, however, did not focus on the compounds found in the selected plants and anticancer properties [9]. Therefore, the current study focuses on gathering more data on the use of isolated compounds as anticancer lead agents in traditional medicine to advance research in the field of discovery and development of anticancer agents.

For this reason, the purpose of this review is to provide a comprehensive overview of the pharmacological activities of the isolated compounds from various *Euphorbia* species found in Southern Africa. This review also aims to highlight the diversity of the Euphorbiaceae family and its potential for further research. The study focuses on several plants, including *Euphorbia trigona* (*E. trigona*), *Euphorbia tirucalli* (*E. tirucalli*), *Euphorbia ammak* (*E. ammak*), *Euphorbia bupleurifolia* (*E. bupleurifolia*), *Euphorbia enopla* (*E. enopla*), *Euphorbia polygona* (*E. polygona*), *Euphorbia cooperi* (*E. cooperi*), *Euphorbia stellata* (*E. stellata*), *Euphorbia ferox* (*E. ferox*), *Euphorbia clavarioides* (*E. clavarioides*), *Euphorbia gorgonis* (*E. gorgonis*), *Euphorbia coerulescens* (*E. coerulescens*), *Euphorbia horrida* (*E. horrida*), *Euphorbia arabica* (*E. arabica*), and *Euphorbia ledienii* (*E. ledienii*). These plants were selected for their pharmacological activities and toxicological evaluation on breast cancer cells as part of the authors' recent research project. This review serves as a valuable resource for other researchers to explore the remaining *Euphorbia* species for their potentially active compounds and anticancer therapeutic properties.

## 2. Data Collection

To conduct this review, original articles from journals indexed in PubMed Central, Springer Link, Scopus, Science Direct, Hindawi, and Google scholars on *Euphorbia* species, their medicinal uses, toxicity and pharmacological properties, were analysed. For this review, only texts published in English between 1962 and 2023 were considered. The years of publication were selected due to the limited data available on *Euphorbia* plants. Throughout the search process, 881 reports were found across various databases. After reviewing the titles and abstracts, the search was refined and narrowed down to 293 articles, which were fully examined and are now being discussed in this study.

The research criteria for inclusion was based on the guidelines set by Lu et al. [10] with a few modifications. The criteria included: (1) studies on the isolated compounds of *Euphorbia* species conducted in vitro and in vivo; (2) evaluation of their biological activities; (3) studies published in peer-reviewed journals written in English; and (4) studies that provided full-text papers. There were no limitations on the study's location. The study excluded (1) human studies, (2) a combination of isolated compounds and (3) non-original articles.

The retrieved data was carefully examined to identify possible medical applications, biological activities, isolated compounds, toxicity and pharmacological properties. Additional confirmation of the correct plant names was verified from the plantlist.org website. After analyzing all the data collected, conclusive results were obtained.

The cytotoxic tendency of selected compounds on various cancer cell lines and human tumor types was computed using the Cell Line Cytotoxicity Predictor (CLCPred) webserver (<https://www.way2drug.com/Cell-line/>) a free online service for in silico evaluation of human cell line cytotoxicity tendency of bioactive compounds [11]. This prediction was based on Prediction of Activity Spectra for Substances (PASS) technology (<https://www.way2drug.com/PASSonline>) where the training set was generated based on cytotoxicity data derived from ChEMBLdb (version 23)

(<https://www.ebi.ac.uk/chembl/db/>). Compounds with robust anticancer potential were selected and subjected to further analyses to extract physicochemical and pharmacokinetics parameters using the SwissADME (<http://www.swissadme.ch/index.php>) [12] and pKCSM (<http://biosig.unimelb.edu.au/pkcsml/>) [13] web servers as demonstrated earlier [14,15].

### 3. Results

The current study reports on 15 *Euphorbia* species, their traditional and pharmacological properties, and also isolated compounds. *Euphorbia* species have been used in folk medicine for various ailments, which is why scientists have taken an interest in investigating this genus and fully documenting the secondary metabolites responsible for these properties, see Table 1.1.

Out of the 15 plant species examined, 8 have been traditionally used as medicine for treating 7 distinct diseases. The most commonly used plants are for treating cancer (7), followed by warts and wounds (5), as well as other ailments, see Table 1.1.

**Table 1.1.** *Euphorbia* spp of Southern Africa and its neighboring countries, their toxicity and traditional use.

Botanical name  Synonym in bold	Vernacular name  (E)= English;  (S)= Sotho  (Z)= Zulu; (X)= Xhosa; (A)= Afrikaans  (N)= Ndebele  (T)= Tsonga  (Ts)= Tswana  (V)= Venda	Location	Growing condition	Traditional use	Bioactive property	Toxicity	Reference
<i>Euphorbia trigona</i>  <i>Euphorbia hermentiana</i> Lem	African milk tree (E)	West Africa, tropical Asia, India	Dry tropical forests and semiarid environment. Direct sunlight	Respiratory infections, urinary tract infections, gonorrhoea, tumors and warts,	Anticancer	Skin irritation	[16–21]

				intestinal parasites, rheumatoid arthritis, hepatitis, inflammation, piles, constipation, epileptic attack.			
<b><i>Euphorbia cooperi</i></b> <i>E. cooperi</i> var. <i>cooperi</i> <i>E. cooperi</i> var. <i>calidicola</i>	Candelabra <i>Euphorbia</i> (E), Umhlonhlo (N), Tshikondengala (V), Mokhoto (T) Mohlohlo (S)	South Africa (KwaZulu-Natal, Swaziland, North West Province, Mpumalanga and Limpopo) Mozambique, Zimbabwe, Botswana	Well-drained soil. Direct sunlight.	Sore stomach, bloatedness, paralysis, wound healing	Breast cancer inhibitor	Skin irritation, blindness, throat burning sensation	[22–28]
<b><i>Euphorbia ammak</i></b>	African candelabra /Desert candelabra (E)	Saudi Arabia, Yemen peninsula	No data reported	No data reported	Anticancer Antileishmanial activity, (H1N1) influenza virus, MDCK cells, antiparasitic activity	Skin and eye irritation. Normal melanocyte (HFB4).	[29–33]

<p><i>Euphorbia tirucalli</i> <i>Euphorbia laro</i> Drake</p>	<p>Rubber-hedge <i>Euphorbia</i> (E), umSululu (Z), umHlonthlo (X), motsetse/ setlhareseto la (Ts), mahumbana (T), muṭungu (V) kraalmelkboos (A)</p>	<p>Eastern tropical Africa, South Africa, Indian Ocean Island</p>	<p>Rock garden, well-drained soils, mild to warm climates.</p>	<p>Snakebites, sexual impotence, warts, wounds, skin complaints, swollen glands, oedema, haemorrhoids, rheumatism, epilepsy toothache, earache and tumor.</p>	<p>Antibacterial, molluscicide, antiherpetic, antimutagenic antibacterial, myelomodulating activity, antitumor effect on different cell lines. Tumor promotor</p>	<p>Skin irritation, blindness, fish poisoning, gastrointestinal haemorrhage, fatality, Burkitt's lymphoma, Epstein-Barr virus</p>	<p>[5,6,34-52]</p>
<p><i>Euphorbia clavarioides</i> <i>Euphorbia basutica</i></p>	<p>lions Spoor (E), melkpol (A), iSantilele/isihlekehleke (Z), sehlehle/sehloko (S)</p>	<p>South Africa (Eastern Cape, KwaZulu-Natal, Northern Cape, Western Cape), Lesotho</p>	<p>Direct sunlight, mineral soil, grassland gardens</p>	<p>Skin rash in children, acne, sores, bruises, burns, eczema, ulcer, cracked heels and wounds, cancerous sores, warts, swollen feet, leprosy remedy, herpes, HIV related infections, high</p>	<p>No data reported</p>	<p>Skin irritation, bird lime</p>	<p>[53-57]</p>

				blood pressure, diabetes			
<i>Euphorbia gorgonis</i>	Nkalimasan e (Z) melkbol (A)	South Africa (Eastern Cape)	Well-drained soil, Direct sunlight	Wounds, swelling, skin problems.	Antibacterial, antimicrobial activity, cancer cell line (HuTu), hepatoma cells (H411E)	Skin irritation	[58–60]
<i>Euphorbia bupleurifolia</i> <i>Tithymalus bupleurifolius</i>	Pine cone plant (E), melkbol (A), intsele (X), insema (Z)	South Africa (Eastern Cape Province, Transkei and Natal)	Warm and moist conditions, moderate sunlight.	Cancerous sores, for painful cracked feet, eczema, pimples, rashes and wounds in topical formulation, clean teeth, swellings of the lower limbs, cancer, retained placenta.	No data reported	Skin irritation	[61–64]
<i>Euphorbia enopla</i> <i>Euphorbia enopla</i> var. <i>enopla</i>	Milk-barrel/Pincushion <i>Euphorbia enopla</i> (E)	South Africa (Eastern Cape in the Noorsveld, in arid to	Well-drained soil, direct sunlight	No data reported	No data reported	Skin irritation, toxic to Vero cell line	[65]

		semi-arid Karoo)					
<i>Euphorbia coerulescens</i> <i>Euphorbia virosa</i>	Sweetnoors (A)	South Africa (Cape province)	Sunny to half-shady, rocky and gritty-sandy soils	No data reported	No data reported	Inflammation in mice, skin irritation, blindness, throat irritation if swollen	[66]
<i>Euphorbia polygona</i>	African Milk Barrel (E)	South Africa (Eastern Cape)	Well-drained soil, direct sunlight and high temperature environment.	No data reported	No data reported	Eye irritation, skin irritation, paralysis	[65]
<i>Euphorbia horrida</i> Var <i>Euphorbia horrida</i> Boiss.	African Milk Barrel (E)	South Africa (Wittepourt/Karoo)	Well-drained soil, direct sunlight and dry conditions.	No data reported	No data reported	Inhibition of Vero cell line	[65]
<i>Euphorbia arabica</i> <i>Euphorbia neopolycnemoides</i>	Klein Bont <i>Euphorbia</i> (E), umhlonhlo (Z)	Botswana, southern Mozambique, Zimbabwe, Limpopo, Mpumalanga and KwaZulu-Natal, South Africa	Stony grassland area.	Warts, stomach ache, skin infections	Antibacterial	Inhibition of Vero cell line	[67,68]

<i>Euphorbia ledienii</i> <i>Euphorbia ledienii</i> A. Berger var. <i>ledienii</i>	Crested <i>Euphorbia</i> (E)	South Africa (Western Cape)	Well-drained soil, sunny conditions	No data reported	No data reported	Skin irritation	[69]
<i>Euphorbia ferox</i> <i>Euphorbia capitosa</i>	Milkweed (E)	South Africa (Western Cape)	Drained sandy soil, rocky outcrops and in sunny environment	No data reported	No data reported	Poisonous latex	
<i>Euphorbia stellata</i> <i>Euphorbiasquarrosa</i> <i>Euphorbia radiata</i> <i>Euphorbia scolopendrea</i> <i>Euphorbia uncinata</i>	Spurge (E)	South Africa (Eastern Cape)	Well-drained soil and sunny environment	No data reported	No data reported	Skin and eye irritation.	

**Table 1.2.** Comprehensive overview of Isolated Compounds from Individual Plants.

Plant	Compounds	Reference
<i>Euphorbia trigona</i>	Euphol , Cycloartenol, Cycloartanol, Lupeol, $\alpha$ amyryrin, $\beta$ – amyryrin, Betulinic acid, Taraxerol , $\beta$ -sitosterol, Taraxerol acetate, Friedelin, Friedelan 3 $\alpha$ - and 3 $\beta\alpha$ –ols, 24-ethylene cycloartanol, Epi-friedelinyl acetate, 3 $\beta$ , friedelinol, Rhoiptlenone	[70,71]
<i>Euphorbia ledienii</i>	Isobutyric, 2-Methylbutyric acid, 12-Deoxyphorbol-13-isobutyrate-20-acetate, 12-Deoxyphorbol-13-(2-methylbutyrate)-20-acetate, Phorbol, 12-Deoxy phorbol, 12-Deoxy-16-hydroxy phorbol, Ingol-7,8,12-acetate,ditiglate	[72–74]
<i>Euphorbia tirucalli</i>	Triterpenes, Euphol, Diterpene esters, Euphorbiane, 12-Deoxyphorbol esters, Ingenol , $\beta$ -Sitosterol, Euphorbol, hexacosonate, 12-Deoxy-4 $\beta$ -hydroxyphorbol-13-phenyl acetate-20-acetate, 12, 20-Dideoxyphorbol-13-isobutyrate,	[8,9,21,35,36,75–92]

	Glut-5-en-3- $\beta$ -ol, Tirucalicine, Tri-methyl ellagic acid, Terpenic alcohol, Isoeuphorol, Taraxasterol, Tirucallol, Ketone euphorone, Resin, Ellagic acid, Taraxerol, 3,3'-Di-O-methylellagic acid, Euphorbin A, Euphorbin B, Tirucallin A, Tirucallin B, Euphorbol, Cycloartenol, 24-Methylenecycloartenol, Ingenol triacetate, $\beta$ -amyrin, Rhoiptlenone, 3 $\beta$ -friedelinol, Epi-friedelinyl acetate, 24-ethylene cycloartanol, Friedelan 3 $\alpha$ - and 3 $\beta\alpha$ -ols, Friedelin, Taraxerol acetate, Betulinic acid, $\beta$ -amyrin, $\alpha$ amyrin, Lupeol, Cycloartanol, $\beta$ -amyrone, glutinone, taraxerone, Glut-5-en-3- $\beta$ -ol and cycloart-23-ene-3- $\beta$ , 25-diol, Euphorcinol, Euphorginol, 12, 13,20-tri- O-acetylphorbol, 3,5,20-tri-O-acetlingenol 20-acetate	
<i>Euphorbia enopla</i>	Euphol, Tirucallol	[93]
<i>Euphorbia coerulescens</i>	Angelate acetate isobutyrate, Fatty acids, Acetate laurate, $\alpha$ -Methyl butyrate, Heptanoate, Laurate, Euphol, Tirucallol, Euphorbol	[66,89]
<i>Euphorbia ammak</i>	$\alpha$ -glutinol, Stigmasterol, Euphol, Euphorbol	[93,94]
<i>Euphorbia cooperi</i>	16-Hydroxy-12-desoxyphorbol, Euphol, Obtusifoliol, 12-Deoxyphorbol-13-isobutyrate-16-angelate-20-acetate, Euphorbilactone, Norsesquiterpenoid, Arachiside A, Glutinol, 16-Angeloyloxy-13 $\alpha$ -isobutanoyloxy-4 $\beta$ ,9 $\alpha$ ,20-trihydroxytiglia-1,5-diene-3,7-dione, 20-Acetoxy-16-angeloyloxy13 $\alpha$ -isobutanoyloxy-4 $\beta$ ,9 $\alpha$ ,20-tetrahydroxytiglia-1,5-diene-3-one, Gallic acid, Bervifolin, carboxylic acid, Kampferol-3-O- $\beta$ -D-rutinoside, 1-O-Galloyl-3,6-hexahydroxydiphenyl- $\beta$ -D glucopyranoside, 3, 3' Dimethoxy ellagic acid 3, 4, 4' Trimethoxyellagic acid, Ellagic acid, Kaempferol, 7-galloyl catechin, kaempferol 3-O- $\beta$ -(6''-O-galloyl)-glucopyranoside, triesters-16-hydroxy-12-desoxy-phorbol	[9,24,95–97]
<i>Euphorbia horrida</i>	17-Hydroxyingenol-17-benzoate-20-angelate, Diterpene esters	[98]

**Table 1.3.** Phytochemicals extracted from the selected *Euphorbia* spp. This table outlines the diverse array of crude phytochemicals extracted from carefully chosen *Euphorbia* species, shedding light on the rich chemical composition inherent in these selected plants.

Plant	Phytochemical/s	Reference
<i>Euphorbia trigona</i>	Saponins, alkaloids, flavonoids, glycosides, sterols and triterpenoids, tannins	[17,71,99]
<i>Euphorbia cooperi</i>	Triterpenoid	[95]
<i>Euphorbia ammak</i>	alkaloids, saponins, glycosides	[31]
<i>Euphorbia tirucalli</i>	Triterpenoid, phenols, flavonoids, tannins, alkaloids, saponins, glycosides, and steroids.	[37,100]
<i>Euphorbia clavarioides</i>	Alkaloids, flavonoids, saponins, tannins, terpenoids, phytosterols, glycosides, triterpenoids, anthraquinone	[65,101]
<i>Euphorbia gorgonis</i>	Phytosterols, glycosides, triterpenoids, flavonoids, alkaloids, saponins	[60,65]
<i>Euphorbia bupleurifolia</i>	Phytosterols, tannins, glycosides, triterpenoids, saponins, flavonoids and alkaloids	[62,65]
<i>Euphorbia enopla</i>	Phytosterols, glycosides, triterpenoids, flavonoids, alkaloids, tanins, anthraquinone	[65]
<i>Euphorbia polygona</i>	Phytosterols, tannins, glycoside, triterpenoids, flavonoids, alkaloids	[65]
<i>Euphorbia horrida</i> Var	phytosterols, pentose, tannins, glycosides, triterpenoids, anthraquinones, saponins, flavonoids	[65]
<i>Euphorbia arabica</i>	Phytosterols, tannins, glycosides, triterpenoids, anthraquinones, flavonoids	[65]
<i>Euphorbia ledienii</i>	No data reported	
<i>Euphorbia ferox</i>	No data reported	
<i>Euphorbia stellata</i>	No data reported	
<i>Euphorbia coerulescens</i>	No data reported	

An investigation into the chemical composition of various *Euphorbia* species unveiled distinctive profiles of isolated compounds. *E. tirucalli* exhibited the highest diversity with a total of 30 isolated compounds, spanning triterpenoids (Lupane, Oleanane, Tirucallane, Phorbol-type), phenolic compounds (Gallic Acid Derivatives, Ellagic Acid Derivatives, Flavonoids), phytosterols (Sterols), glycosides (Triterpene Glycosides, Other Glycosides), and diterpene esters [9,37,100]. Following closely, *E. cooperi* presented 18 compounds, including triterpenoids (Lupane, Oleanane, Phorbol-type), phytosterols (Sterols), and glycosides (Triterpene Glycosides) [95]. *E. trigona* showcased 16 compounds, including triterpenoids (Lupane, Oleanane, Phorbol-type), phenolic compounds (Gallic Acid Derivatives, Ellagic Acid Derivatives, Flavonoids), phytosterols (Sterols), and glycosides (Other Glycosides) [17,70,99]. *E. coerulescens* and *E. ledienii* exhibited 9 and 8 compounds, respectively, encompassing triterpenoids, phenolic compounds and phytosterols [66,71], see Table 1.2 and 1.3. Notably, certain plants demonstrated fewer or no isolated compounds.

The breakdown of these compounds into subclasses revealed diverse chemical categories, including Euphane-type Triterpenoids (Euphol, Euphorbol), Cycloartane-type Triterpenoids (Cycloartenol, Cycloartanol, 24-ethylene cycloartanol), Lupane-type Triterpenoids (Lupeol), Oleanane-type Triterpenoids ( $\alpha$ -amyirin,  $\beta$ -amyirin), Pentacyclic Triterpenes (Betulinic acid, Glut-5-

en-3- $\beta$ -ol), Taraxarane-type Triterpenoids (Taraxerol, Taraxerol acetate), Phorbol-type Diterpenoids (12-Deoxyphorbol-13-isobutyrate-20-acetate, Phorbol, 12-Deoxy phorbol, 12-Deoxy-16-hydroxy phorbol, 12-Deoxyphorbol esters, 12, 20-Dideoxyphorbol-13-isobutyrate), Tirucallane-type Triterpenoids (Tirucallicine, Tirucallol, Tirucallin A, Tirucallin B), Steroidal Triterpenoids (Terpenic alcohol), Ingenane-type Triterpenoids (Ingol-7,8,12-acetate, ditiglate, Ingenol, Ingenol triacetate, Angelate acetate isobutyrate), Flavonoids (Tri-methyl ellagic acid, 17-Hydroxyingenol-17-benzoate-20-angelate, Kampferol-3-O- $\beta$ -D-rutinoside), Sterols ( $\beta$ -sitosterol, Stigmasterol, Taraxasterol), Ellagic Acid Derivatives (Ellagic acid, 3,3'-Di-O-methylellagic acid), Medium-chain Fatty Acids (Laurate) and no specific subclass records (Rhoiptlenone, 2-Methylbutyric acid, Euphorbin A, Euphorbin B, Euphorbol hexacosonate, 12-Deoxy-4 $\beta$ -hydroxyphorbol-13-phenyl acetate-20-acetate, 20-Acetoxy-16-angeloyloxy-13 $\alpha$ -isobutanoyloxy-4 $\beta$ ,9 $\alpha$ ,20-tetrahydroxytiglic-1,5-diene-3-one, Resin, Arachiside A, 3, 4, 4' Trimethoxyellagic acid, 12-Deoxyphorbol-13-(2-methylbutyrate)-20-acetate, 2-Methylbutyric acid) (see Table 1.4).

*E. tirucalli* as well as *E. ammak* are among the *Euphorbia* species that received substantial research attention. These findings are in agreement with studies conducted by Mavundza et al. [9], which stated that *E. tirucalli* was among the most studied species.

**Table 1. 4.** Classification of Crude and Isolated Compounds from 15 *Euphorbia* Specie. This table delineates the categorization of both crude extracts and individually isolated compounds obtained from 15 distinct *Euphorbia* species. The compounds are organized based on their respective classes, providing a comprehensive overview of the chemical constituents present in each species.

Compounds	Class	subclass
Euphol (Tetracyclic Triterpene), Cycloartanol (Sterol Lipid), Lupeol, $\alpha$ -amyrin, $\beta$ -amyrin, Betulinic acid, Taraxerol, Taraxerol acetate, Friedelin, Friedelan-3- $\beta$ -ol, 3 $\beta$ -Friedelinol	Terpenoids	Triterpenoids
Cycloartenol, $\beta$ -sitosterol, 24-Methylenecycloartanol, Tirucallol (Tetracyclic Triterpene), Obtusifoliol, Glutanol, Triterpene euphol (Steroidal Alcohol), 24-Methylene cycloartenol, Ingenol triacetate (Diterpene), Terpenic alcohol (Terpene), Taraxasterol (Phytosterol), Stigmasterol		Phytosterol
12-Deoxyphorbol-13-isobutyrate-20-acetate (Phorbol Ester), Phorbol (Diterpenoid), 12-Deoxyphorbol ester (Diterpenoid), 12-Deoxy-16-hydroxy-phorbol (Component of DHPB), 16-Hydroxy-12-desoxyphorbol (Diterpenoid), 12-Deoxyphorbol-13-isobutyrate-16-angelate-20-acetate (Diterpene), 12-Deoxy phorbol esters (Phorbol Ester), Ingenol (Diterpenoid), Glut-5-en-3- $\beta$ -ol (Triterpenoid)		Other Terpenoids
Gallic acid (Phenolic Acid), Bervifolin carboxylic acid (Tannin), 3, 3'-Dimethoxy ellagic acid (Tannin), Ellagic acid (Tannin), Kaempferol (Flavonoid), Kampferol-3-O- $\beta$ -D-rutinoside (Flavonoid), 1-O-Galloyl-3,6-hexahydroxydiphenyl- $\beta$ -D-glucopyranoside/corilagin	Phenolic	

(Tannin), Tri-methyl ellagic acid (Tannin), 3'-Di-O-methylellagic acid (Phenol)		
Laurate	Fatty acids	
Diterpene esters	Miscellaneous	
Flavonoids		Polyphenolic compounds
Alkaloids		Organic compounds
Saponins		glycosides
Tannins		Polyphenols
Glycosides		Acetal Derivatives of Monosaccharides
Anthraquinones		Phenolics

The investigation into the chemical composition of various *Euphorbia* species are presented in Table 1.4, and has revealed a diverse array of isolated compounds, each showcasing unique pharmacological and biological activities. Euphol, classified within the Euphane subclass, demonstrates a diverse profile, including anticancer, cytotoxicity, anti-nociceptive, anti-inflammatory and HIV-1 reverse transcriptase inhibitor activities [76,102,103]. The Cycloartenol and Cycloartanol compounds, falling under the Cycloartane subclass, exhibit a wide range of effects such as anti-inflammatory, antitumor, antioxidant, antibiosis, anti-Alzheimer's disease, apoptotic, analgesic and antifungal activities [104–110]. Lupeol, belonging to the Lupane subclass, displays diverse properties like anticancer, anti-inflammatory, antimicrobial, antiprotozoal, antiproliferative, antiangiogenic and cholesterol-lowering effects [40,111–117]. Oleanane-type triterpenoids, represented by  $\alpha$ -amyrin and  $\beta$ -amyrin, demonstrate activities such as cytotoxicity, antifungal, anti-inflammatory, nitric oxide inhibition, reactive oxygen species activation and anticancer effects [118–125]. Pentacyclic triterpenes, including Betulinic acid, exhibit antitumor, antidiabetic, anti-inflammatory, HIV-1 reverse transcriptase inhibition, antiviral and hepatoprotective activities [126–135]. The Taraxarane subclass, represented by compounds like Taraxerol and Taraxerol acetate, showcases properties like anticancer, anti-inflammatory, apoptotic, antioxidative, antimicrobial, antifungal and antidiabetic effects [136–141]. Sterols, represented by  $\beta$ -sitosterol, display a spectrum of activities, including anti-inflammatory, anticancer, antiproliferative, analgesic and antimicrobial effects [125,142–145]. Tirucallane-type triterpenoids, such as Tirucallicine and Tirucallol, currently have no specific records of biological activity [146,147]. Terpenic alcohol, a steroidal triterpenoid, demonstrates antibacterial and irritant effects [148,149]. Ellagic Acid Derivatives, including Tri-methyl ellagic acid, exhibit anticancer properties [148,150,151]. Ingenane-type Triterpenoids, represented by Ingenol and its derivatives, showcase cytotoxicity and HIV-1 reverse transcriptase inhibition [102,152]. Various subclasses such as Taraxarane, Euphane-type, Sterol, Tirucallane and Phorbol-type exhibit diverse pharmacological activities, including anti-inflammatory, antifungal, antibacterial, hepatoprotective, antioxidant, antiproliferative and anti-HIV effects, see Table 1.5.

**Table 1.5.** Bioactive compounds isolated from 15 *Euphorbia* species alongside their biological activities and subclasses. This table provides an in-depth exploration of bioactive compounds isolated from 15 distinct *Euphorbia* species, highlighting their diverse biological activities.

Compounds	Subclasses	Pharmacological/ biological activity	Reference
Euphol	Euphane	Anticancer Cytotoxicity Anti-nociceptive Antiinflammatory HIV-1 reverse transcriptase inhibitor	[76,102,103]
Cycloartenol	Cycloartane	Antiinflammatory Antitumor Antioxidant Antibiosis Anti-alzheimer's disease Apoptotic Analgesic Bactericidal	[106–109]
Cycloartanol	Cycloartane	Antifungal Vasodepressor Antitumor	[104,105,110]
Lupeol	Lupane	Anticancer Antiinflammatory Antimicrobial Antiprotozoal Antiproliferative Antiangiogenic Anti-invasive	[40,111–117]

		Cholesterol lowering	
$\alpha$ amyirin	Oleanane	Cytotoxicity Antifungal Antiinflammatory	[120,122–124]
$\beta$ -amyirin	Oleanane	Antiinflammatory Nitric oxide inhibitor Reactive oxygen species activator Anticancer	[118,119,121,122,125]
Betulinic acid	Pentacyclic triterpenes	Antitumor Antidiabetic Antiinflammatory HIV-1 reverse transcriptase inhibitor Antiviral Hepatoprotective activity	[126–135]
Taraxerol	Taraxarane	Anticancer Antiinflammatory Apoptotic Anti-oxidative activity Antimicrobial Antifungal Antidiabetic	[138–141]

$\beta$ -sitosterol	Sterols	Antiinflammatory Anticancer Antiproliferation Analgesic Antimicrobial	[125,142–145]
Taraxerol acetate	Taraxarane	Antiinflammatory Cyclooxygenases inhibitor	[136,137]
Friedelin	Friedelane	Cytotoxicity Antibacterial	[153,154]
Friedelan 3 $\alpha$ - and 3 $\beta$ -ols	Friedelane	Anticancer	[154]
24-ethylene cycloartanol	Cycloartane	Antiinflammatory Antifungal	[125,155]
Epi-friedelinyl acetate	Friedelane		[156]
3 $\beta$ -friedelinol	Friedelane	Antibacterial Cytotoxicity	[156–159]
Rhoiptlenone	No data reported	No data reported	[156]
Isobutyric	Carboxylic acid	irritant	[73,160]
2-Methylbutyric acid	No data reported	No data reported	[73]
12-Deoxyphorbol-13- isobutyrate-20-acetate	Phorbol	Antifungal	[161]
12-Deoxyphorbol-13- (2-methylbutyrate)-20- acetate	Phorbol	No data reported	[72]
Phorbol	Phorbol	Tumor promoter Apoptosis	[7,162]

12-Deoxy phorbol	Phorbol	Antitumor Apoptotic	[163–166]
12-Deoxy-16-hydroxy phorbol	Phorbol	Irritant Tumor promoter	[167]
Ingol-7,8,12-acetate,ditiglate	Ingenane	No data reported	[160]
Triterpenes euphol	Glycosides	Anticancer Antiinflammatory	[52,88,168]
Diterpene esters	Terpenoids	Anticancer Cytotoxicity Tumor promoter Irritant Pro-inflammatory	[7,80,169–171]
12 Deoxyphorbol esters	Phorbol	Irritant Pro-inflammatory Tumor promoter	[172–175]
Ingenol	Ingenane	Cytotoxicity HIV- reverse transcriptase Inhibitor	[102,152]
Euphorbol hexacosonate	No data reported	No data reported	[148]
12-Deoxy-4 $\beta$ -hydroxyphorbol-13-phenyl acetate-20-acetate	No data reported	No data reported	[148]
12, 20-Dideoxyphorbol-13-isobutyrate	Phorbol	No data reported	[148]
Glut-5-en-3- $\beta$ -ol	Pentacyclic triterpene	Antibacterial	[176]

Tirucalicine	Tirucallane	No data reported	[81,148,177]
Tri-methyl ellagic acid	Ellagic Acid Derivatives	Anticancer	[148,150,151]
Terpenic alcohol	Steroidal Triterpenoids	Antibacterial Irritant	[148,149]
Isoeuphorol	Ingenane-type Triterpenoids	No data reported	[148]
Taraxasterol	Sterol	Inhibition of tumor promotion Antiproliferation	[178]
Tirucallol	Tirucallane	Antiinflammatory HIV-Inhibition reverse transcriptase	[146,147]
Ketone euphorone	Euphane-type Triterpenoids	No data reported	[179]
Resin	No data reported	Digestive enzyme Antioxidant Antispasmodic Hypotensive Hepatoprotective Antiviral Antifungal Anticancer Anxiolytics Anthelmintic	[180]
Ellagic acid	Ellagic Acid Derivatives	Hepatoprotective activity Antiproliferative activity Antioxidant	[181]

3,3'-Di-O-methylellagic acid	Ellagic Acid Derivatives	Antioxidant Moderate antibacterial activity Antimicrobial activity Anticancer	[182–185]
Euphorbin A	No data reported	No data reported	[186]
Euphorbin B	No data reported	No data reported	[186]
Tirucallin A	Tirucallane	No data reported	[79]
Tirucallin B	Tirucallane	No data reported	[79]
Euphorbol	Euphane-type Triterpenoids	Antibacterial Antiinflammatory	[94,187]
24-Methylenecycloartenol	Cycloartane	Anti-oxidant Antiinflammatory	[188,189]
Ingenol triacetate	Ingenane-type Triterpenoids	Antimicrobial Antitumor	[190,191]
Angelate acetate isobutyrate	Ingenane-type Triterpenoids	No data reported	[66]
Fatty acids	No data reported	No data reported	[66]
Acetate laurate	No data reported	No data reported	[66]
$\alpha$ -Methyl butyrate	No data reported	No data reported	[66]
Heptanoate	No data reported	No data reported	[66]
Laurate	Medium-chain fatty acids	Antibacterial	[66,192]
$\alpha$ -glutinol	Pentacyclic triterpenes	Ant-proliferation Cytotoxicity	[94,193,194]
Stigmasterol	Phytosterol	Cytotoxicity Antioxidant	[94,195]

		Hypoglycemic Thyroid inhibitor	
16-Hydroxy-12-desoxyphorbol	Phorbol	Antitumor Tumor promoter	[97,196]
Obtusifoliol	Lanostane	Cytotoxicity	[24,95,197,198]
12-Deoxyphorbol-13-isobutyrate-16-angelate-20-acetate	Phorbol-type diterpenoid	Cytotoxicity	[24,95,197]
Euphorbilactone	Ingenane-type Triterpenoids	No data reported	[95]
Norsesquiterpenoid	No data reported	No data reported	[95]
Arachiside A	Ingenane-type Triterpenoids	No data reported	[95]
Glutinol	Pentacyclic triterpenes	Antiproliferation	[193]
16-Angeloyloxy-13 $\alpha$ -isobutanoyloxy-4 $\beta$ ,9 $\alpha$ ,20-trihydroxytiglic-1,5-diene-3,7-dione	No data reported	No data reported	[95]
20-Acetoxy-16-angeloyloxy-13 $\alpha$ -isobutanoyloxy-4 $\beta$ ,9 $\alpha$ ,20-tetrahydroxytiglic-1,5-diene-3-one	No data reported	No data reported	[95]
Gallic acid	Gallic Acid Derivatives	Hepatoprotective activity Antioxidant	[181]
Bervifolin carboxylic acid	Gallic Acid Derivatives	Hepatoprotective activity Antioxidant	[181]

Kampferol-3-O-β-D-rutinoside	Flavonoids	Hepatoprotective activity Antioxidant	[181]
1-O-Galloyl-3,6-hexahydroxydiphenyl-β-D-glucopyranoside	Gallic Acid Derivatives	Antitumor Antiinflammation Antioxidant Hepatoprotective activity Antimicrobial Antihypertensive Antidiabetic Anti-HIV Antifungal	[197,199–206]
3, 3' Dimethoxy ellagic acid	Ellagic Acid Derivatives	Hepatoprotective activity Antioxidant	[181]
3, 4, 4' Trimethoxyellagic acid	Ellagic Acid Derivatives	Hepatoprotective activity Antioxidant	[181]
Kaempferol	Flavonoids	Hepatoprotective activity Antiproliferative activity Antioxidant	[197]
17-Hydroxyingenol-17-benzoate-20-angelate	Flavonoids	No data reported	[98]

*E. trigona* has the most isolated anticancer compounds (14), followed by *E. tirucalli* (13), *E. cooperi* (8), and the others have fewer or none. The *Euphorbia* species found in Southern Africa are rich sources of various types of bioactive compounds, including triterpenoids, phorbol esters, alkaloids, flavonoids, phytosterols, glycosides and saponins [9,60,65]. Furthermore, minor classes that were isolated from *Euphorbia* species from Southern Africa are: anthraquinone, polyphenols and tannins [9,60,65].

It has further been reported that most spurge contain an acidic and burning vesicant juice, as well as cyanoglycosides, which can be toxic [207]. Reports have shown that ingestion of a large quantity of the latex may cause gastro-intestinal haemorrhage and even result in death [41]. Although incidents of poisoning in children and animals are rare, it is important to handle these plants with great caution [208]. The latex from these plants can also cause blisters on the skin and temporary blindness [34]. Furthermore, they have been used as fish poison and bird-lime; see Table 1.6.

**Table 1.6.** Type of toxicity caused by some *Euphorbia* species.

Type of toxicity	<i>Euphorbia</i> species	References
<b>Fish poisoning</b>	<i>Euphorbia scheffleri</i> Pax, <i>Euphorbia tirucalli</i> L., and <i>Euphorbia inaequilatera</i> Sond	[208]
<b>Human poisoning</b>	<i>Euphorbia ledienii</i> A. Berger, <i>Euphorbia heterophylla</i> L., <i>Euphorbia cooperi</i> N.E.Br. ex A. Berger, <i>Euphorbia candelabrum</i> Kotschy, <i>Euphorbia virosa</i> Willd., <i>Euphorbia poissonii</i> Pax, <i>Euphorbia unispina</i> N.E.Br. and <i>Euphorbia venenifera</i> Tremaux ex Kotschy;	[208]
<b>Domestic animals poisoning</b>	<i>Euphorbia caput-medusae</i> L., <i>Euphorbia silenifolia</i> (Haworth) Sweet, <i>Euphorbia ingens</i> E. Mey. Ex Boiss; as well as irritating ones: <i>E. tirucalli</i> , <i>Euphorbia poissonii</i> , <i>Euphorbia unispina</i> and <i>E. venenifera</i> . I	[208]
<b>Carcinogen/promotor of cell division</b>	<i>E. tirucalli</i> , <i>Euphorbia leuconeura</i> , <i>J. curcas</i>	[167,209,210]
<b>Conjunctivitis</b>	<i>E. tirucalli</i> and <i>Euphorbia royleana</i>	[39,211]

Taxonomic classification of all *Euphorbia* plants discussed in this review:

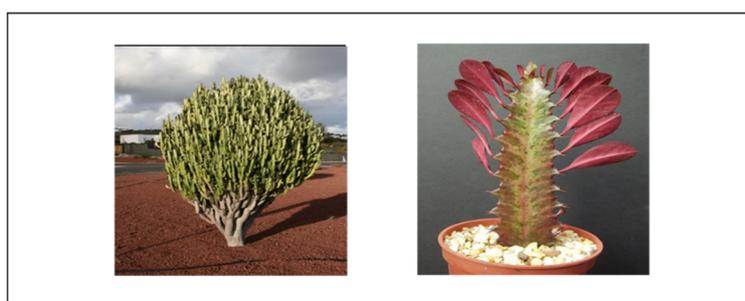
Domain: Eukaryote; Kingdom: Plantae; Order: Malpighiales; Family: *Euphorbiaceae*; Genus: *Euphorbia* (<https://www.mindat.org/taxon-4691.html>)

## 4. Discussion

### 4.1. Ethnopharmacological Use, Phytochemistry and Toxicity

#### 4.1.1. *Euphorbia trigona*

*E. trigona*, a plant native to Central Africa, tropical Africa and India has been used in traditional Indian Ayurvedic medicine to treat respiratory and urinary tract infections and gonorrhoea; Figure 1.1 [18]. Studies by Nashikkar et al. [17] have also shown that *E. trigona* is effective in treating various ailments, such as tumors, warts, intestinal parasites, rheumatoid arthritis, hepatitis and inflammation. A combination of the roots and ginger can be consumed in the morning to alleviate piles [19]. Additionally, some people take latex drops in palm wine for severe constipation or during an epileptic attack [20].



**Figure 1.1.** *Euphorbia trigona* tree and pot plant (source: <http://tropical.theferns.info/image.php?id=Euphorbia+trigona;> <https://laidbackgardener.blog/2018/12/09/when-a-red-euphorbia-turns-green/>).

## Phytochemistry

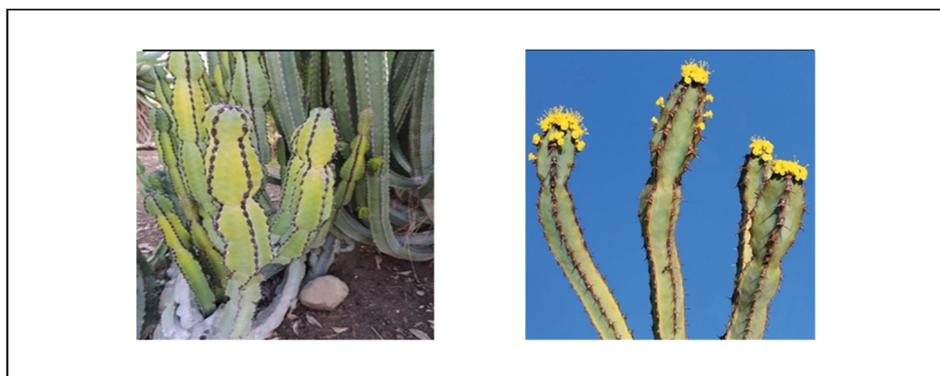
The plant *E. trigona* was analyzed for its phytochemical composition and was found to contain saponins, alkaloids, flavonoids, glycosides, sterols and triterpenoids [99]. Additional studies by Nashikkar et al. [17] also identified the presence of sterols, alkaloids, flavonoids and saponins, as well as tannins that were not detected by [99]. The absence of tannins may be due to environmental factors. Nielsen et al. [71] found that the latex of *E. trigona* contains a high level of sterols, with the main components being euphol and cycloartenol. Anjaneyulu and Rao [70] isolated several triterpenoids from the latex, including euphol, cycloartanol, cycloartenol, lupeol,  $\alpha$ -amyrin and  $\beta$ -amyrin. They also identified five diterpenes ester skin-irritants. *E. trigona* is also a good source of lectin, which has been shown to have a potency in human erythrocyte agglutination [21].

## Toxicity

A study, conducted by EL-Hawary et al. [98], tested the cytotoxic effects of a methanolic extract of *E. trigona* against HEPG2, MCF-7 and CACO2 cell lines. The findings revealed a pronounced cytotoxic effect on MCF-7 and Caco-2 cell lines, with IC50 values of 16.1 and 15.6  $\mu\text{g/mL}$ , respectively. In 2022, Anju and Rameshkumar evaluated a methanol extract's cytotoxic effect on HeLa and H9C2 cell lines but found no notable cytotoxic effect on either cell line [156]. Another study evaluated the cytotoxicity effect of Hex, DCM, MeOH and EtoAc extract against Vero cell lines [65]. The results indicated that none of the four extracts from *E. trigona* exhibited cytotoxicity, as they did not hinder 50% of the cell growth at concentrations of 10  $\mu\text{g/mL}$  and below.

### 4.1.2. *Euphorbia ledienii*

*E. ledienii* is a plant species that is indigenous to the Western Cape, South Africa (see Figure 1.2). The plant does not have any traditional or pharmacological data.



**Figure 1.2.** *Euphorbia ledienii* plant with stem and flowers (source: <https://www.agaveville.org/viewtopic.php?f=54&dt=3571>, [http://www.llifile.com/Encyclopedia/SUCCULENTS/Family/Euphorbiaceae/14292/Euphorbia\\_ledienii](http://www.llifile.com/Encyclopedia/SUCCULENTS/Family/Euphorbiaceae/14292/Euphorbia_ledienii)).

## Phytochemistry

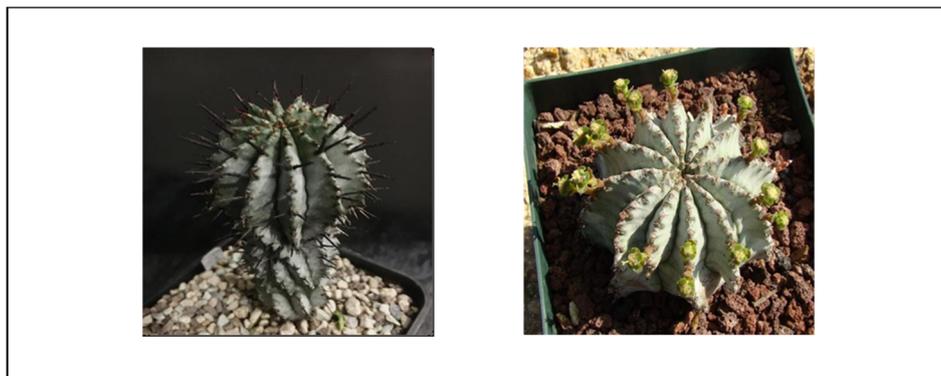
Despite a lack of traditional or pharmacological data, several compounds have been identified from the plant, including 12-Deoxyphorbol-13-isobutyrate-20-acetate and 12-Deoxyphorbol-13-(2-methylbutyrate)-20-acetate [72]. Evans and Kinghorn [74] discovered three variations of taglines which are phorbol, 12-Deoxyphorbol, and 12-Deoxy-16-hydroxy phorbol. *E. ledienii* has been reported to contain Ingol-7,8,12-acetate and ditiglate [160]. Redei et al. [73] identified Isobutyric and 2-methylbutyric acid. Other researchers discovered several hydrolytic proteins in *E. ledienii*, including N-acetyl-b-glucosamidase, chitobiosidase, endochitinase, and lysozyme activity [212]. According to Domsalla et al. [213], the *E. ledienii* plant has been found to have high proteolytic activity.

## Toxicity

It is crucial to highlight that the plant is extremely toxic to humans, leading to potential skin irritation [69]. It is important to note that none of the isolated compounds have undergone toxicity assessments to date.

### 4.1.3. *Euphorbia horrida*

*E. horrida* is a plant species that can be found in Wittepoort/Karoo, South Africa (see Figure 1.3). There is currently no known traditional or pharmacological information about this plant.



**Figure 1.3.** *Euphorbia horrida* plant with stem and flowers (source: <http://pza.sanbi.org/euphorbia-horrida>).

## Phytochemistry

In a study by El-Hawary et al. [98], 17-Hydroxyingenol-17-benzoate-20-angelate from the plant was isolated. The plant also contains diterpene esters that can cause skin irritation. In addition, Mampa et al. [65] found various other classes of compounds in the plant, including phytosterols, pentose, tannins, glycosides, triterpenoids, anthraquinones, saponins, flavonoids and alkaloids.

## Toxicity

The cytotoxic effect of dichloromethane (DCM) extract derived from *E. horrida* var. was evaluated on a Vero cell line. The results demonstrated notable cytotoxicity, as evidenced by its achievement of an IC<sub>50</sub> value at a concentration of 10 µg/mL, highlighting its potential cytotoxic effects [65]. A study conducted by El-Hawary et al. [98] evaluated the cytotoxicity effect of methanol extract of *E. horrida* against HeLa and H9C2 cell lines and no significant effect was observed.

### 4.1.4. *Euphorbia enopla*

*E. enopla*, a plant that is native to the Eastern Cape and semi-arid Karoo, South Africa, has no documented traditional or pharmacological data (see Figure 1.4).



**Figure 1.4.** *Euphorbia enopla* plant with stem and flowers (source: [https://www.cactus-art.biz/schede/EUPHORBIA/Euphorbia\\_enopla/Euphorbia\\_enopla/Euphorbia\\_enopla.htm](https://www.cactus-art.biz/schede/EUPHORBIA/Euphorbia_enopla/Euphorbia_enopla/Euphorbia_enopla.htm)).

### Phytochemistry

The plant contains euphol and tirucallol, which were isolated by Ponsinet and Ourisson [93]. Furthermore, Mampa et al. [65] extracted phytosterols, glycosides, triterpenoids, flavonoids, alkaloids, tanins and anthraquinone. Sytwala et al. [212] isolated N-acetyl-b-glucosamidase, chitobiosidase, endochitinase and lysozyme hydrolytic proteins.

### Toxicity

A study conducted by Mampa et al. [65] assessed the toxicity of the hexane extract of *E. enopla* on the Vero cell line. The results indicated that the extract had a substantial inhibitory effect on cell growth, particularly at a concentration of 10 µg/mL. Notably, the highly non-polar hexane fraction exhibited the most potent effects [65].

#### 4.1.5. *Euphorbia coerulescens*

*E. coerulescens* which is native to the Cape province, South Africa, has no documented traditional or pharmacological data (see Figure 1.5). However, it has a number of compounds isolated from it.



**Figure 1.5.** *Euphorbia coerulescens* plant with stem and flowers (source: <https://www.ebay.com.au/itm/euphorbia-coerulescens-POT-10> CM /233323039902?nma=true&si=DenzQanzHzMK8eHcDW6MHzA0s9s%253D&orig\_cvip=true&nordt=true&rt=nc&\_trksid=p2047675.l2557.

### Phytochemistry

Studies by Evans [66] resulted in the isolation of angelate acetate isobutyrate, acetate a-methyl butyrate, acetate laurate, α-Methyl butyrate, heptanoate and laurate. In other studies, euphol, tirucallol and euphorbol were also isolated [89]. In addition, Sytwala et al. [212] isolated several hydrolytic proteins, including N-acetyl-b-glucosamidase, chitobiosidase, endochitinase and lysozyme activity. Lynn and Clevette-Radford [214] also isolated homogeneous lectins.

### Toxicity

During an irritancy test conducted by Evans [66], it was observed that the latex of *E. coerulescens* induced ear inflammation in mice. Furthermore, this latex has the potential to cause skin irritation and, if ingested, may result in a burning sensation in the throat. Direct contact with the eyes can lead to severe consequences, including the risk of blindness. It is worth noting that none of the extracted compounds have been subjected to toxicity evaluations at this time.

#### 4.1.6. *Euphorbia cooperi*

*E. cooperi* is a plant that grows naturally in KwaZulu Natal and Limpopo province, South Africa (see Figure 1.6). Research has shown that it may have potential in treating various health conditions. It has been reported that the liquid from the soaked roots and stem has been used as an enema for sore stomachs and bloatedness [25]. In South Africa, the Venda tribe uses this plant to cure paralysis and apply it to infected wounds [25]. Farmers have historically used this plant to treat various

bacterial infections in their livestock [22,215]. Furthermore, the latex is utilized for poisoning fish in South Africa's Limpopo province [38,216].



**Figure 1.6.** *Euphorbia cooperi* tree and fruits (source: <http://pza.sanbi.org/euphorbia-cooperi>).

### Phytochemistry

Phytochemical analysis of *E. cooperi* revealed that the latex of *E. cooperi* comprises of numerous diesters and triesters [97]. The chloroform fraction of the plant's latex was found to contain three compounds that had never been isolated before. These were euphol, obtusifoliol and 12-Deoxyphorbol-13-isobutyrate-16-angelate-20-acetate, which belong to the triterpene, steroid and diterpenoid families, respectively [24]. A study by El-Toumy et al. [96] isolated 7-galloyl catechin, kaempferol 3-O-β-(6''-O-galloyl)-glucopyranoside and triesters-16-hydroxy-12-desoxy-phorbol from the flower of *E. cooperi*.

A study of the aerial part of *E. cooperi* has uncovered some interesting findings. Hlengwa [95] has discovered a unique norsesquiterpenoid called euphorbilactone, as well as its glycoside, arachiside A. The researcher further identified a triterpenoid, glutinol, a known phorbol ester (16-Angeloyloxy-13α-isobutanoyloxy-4β,9α,20-trihydroxytiglia-1,5-diene-3,7-dione) and a new phorbol esters (20-Acetoxy-16-angeloyloxy-13α-isobutanoyloxy-4β,9α,20-tetrahydroxytiglia-1,5-diene-3-one). Upon a thorough review of existing literature, it has come to light that certain compounds such as Euphol, Obtusifoliol, 12-Deoxyphorbol-13-isobutyrate-16-angelate-20-acetate, Euphorbilactone, Norsesquiterpenoid, Arachiside A, Glutinol, 16-Angeloyloxy-13α-isobutanoyloxy-4β,9α,20-trihydroxytiglia-1,5-diene-3,7-dione, 20-Acetoxy-16-angeloyloxy-13α-isobutanoyloxy-4β,9α,20-tetrahydroxytiglia-1,5-diene-3-one, Bervifolin, carboxylic acid, Kampferol-3-O-β-D-rutinoside, 1-O-Galloyl-3,6-hexahydroxydiphenyl-β-D glucopyranoside, 3, 3' Dimethoxy ellagic acid and 3, 4, 4' Trimethoxyellagic acid have not been previously discussed.

### Toxicity

A study conducted by El-Sherei et al. [24] evaluated the cytotoxic effect of the chloroform extract of *E. Cooperi*. Their findings revealed that the chloroform extract demonstrated significant cytotoxic effects against MCF-7, HepG2 and HeLa cell lines. The IC<sub>50</sub> values for these cell lines were 4.23, 10.80 and 26.6, respectively. The results align with Mavundza et al.'s [9] findings, as no further research has been conducted to date.

#### 4.1.7. *Euphorbia tirucalli*

*E. tirucalli* is a plant that is indigenous to Eastern tropical Africa, South Africa and the Indian Ocean Island (see Figure 1.7). It has been proven to be effective in treating various illnesses. Hargreaves [38] stated that *E. tirucalli* is used to induce vomiting for treating snakebites. Other authors have reported the use of its latex for various purposes such as treating sexual impotence, skin disorders, swollen glands, edema, haemorrhoids, rheumatoid arthritis, epilepsy, tooth and ear pain, and tumor [39,40,217]. In addition, the latex has pharmacological properties that include antibacterial, molluscicidal, antiherpetic and antimutagenic effects [5,40,44–49,177]. Studies have demonstrated that extracts from *E. tirucalli* have myelomodulating activity and suppress the formation of colonies [50]. Additionally, the latex of the plant contains compounds that have antitumor effects on various

cell lines [51,52]. In fact, *E. tirucalli* has been patented as a possible medicinal treatment for prostate cancer [6].



**Figure 1.7.** *Euphorbia tirucalli* plant and flower (source: <https://za.pinterest.com/pin/298785756510953587/>; <https://za.pinterest.com/pin/474426141998696380/>).

### Phytochemistry

Extensive studies have analyzed the chemical makeup of *E. tirucalli* and diterpenes have been identified as the primary isolated compound in all parts of the plant [9]. The latex of *E. tirucalli* contains several phytoconstituents, including triterpenes euphol, diterpene esters of phorbol, 12-Deoxyphorbol esters and ingenol,  $\beta$ -sitosterol, euphorbol hexacosonate, 12-Deoxy-4 $\beta$ -hydroxyphorbol-13-phenyl acetate-20-acetate, 12, 20-Dideoxyphorbol-13-isobutyrate, glut-5-en-3- $\beta$ - and euphol, 12-O-2Z-4E-octadienoyl-4-deoxyphorbol-13-acetate, cycloart-23-ene-3- $\beta$ -, 25-diol, Euphorcinol, 4-Deoxyphorbol di-ester, cycloeuphordenol, cyclotirucanenol, diterpene ester, serine proteases, euphol, steroids, tirucallicine, tri-methyl ellagic acid, terpenic alcohol, isoeuphorol, taraxasterol, tirucallol (fresh latex), ketone euphorone and resin [8,21,75,76,78,80–84,86–91]. Researchers have identified several compounds in the stem of *E. tirucalli*, including ellagic acid, taraxerol, 3,3'-Di-O-methylellagic acid,  $\beta$ -sitosterol, euphorbin A (a type of polyphenol), euphorbin F (dimers), tirucallin A (a type of tannin) and tirucallin B. [79,80,92]. Euphorbiane which is a triterpenoid was isolated from the stem [77]. Rasool et al. [85] isolated euphorginol from the stem bark of *E. tirucalli*. The bark, on the other hand, was found to contain phorbol,  $\beta$ -sitosterol, cycloartenol, 24-Methylene cycloartenol and ingenol triacetate [35]. Other studies have revealed that  $\beta$ -amyrin is present in the leaves of this plant, as reported by Kajikawa et al. [36]. Additionally, Shivkumar [37] found various compounds such as phenols, flavonoids, tannins, alkaloids, saponins, glycosides, triterpenes and steroids. However, Kgosiemang et al. [100] isolated only tannins, glycosides, triterpenoids and saponins from the same plant. Upon review, it was noted that several compounds, such as diterpene esters, 12-Deoxyphorbol esters, ingenol, hexacosonate, 12-Deoxy-4 $\beta$  hydroxyphorbol-13- phenyl acetate -20- acetate, 12, 20 Dideoxyphorbol-13 isobutyrate, tirucallicine, tri-methyl ellagic acid, terpenic alcohol, isoeuphorol, taraxasterol, tirucallol, ketone euphorone, resin, ellagic acid, 3,3'-Di-O-methylellagic acid, euphorbin A, euphorbin B, tirucallin A, tirucallin B, cycloartenol, 24-Methylenecycloartenol, ingenol triacetate, rhoiptlenone, 3 $\beta$ -friedelinol, epi-friedelinyl acetate, 24-ethylene cycloartanol, friedelan 3  $\alpha$  - and 3  $\beta$   $\alpha$  -ols, taraxerol acetate, betulinic acid,  $\alpha$  amyryl, lupeol, Cycloartanol and  $\beta$ -amyryl were not discussed in other reviews compared to the current review [9].

### Toxicity

Silva et al. [218] assessed the antitumor effect of euphol from *E. tirucalli* against a wide range of human cancer cell lines. The study demonstrated that euphol exhibits cytotoxic properties against various cancer cell lines, exhibiting IC<sub>50</sub> values ranging from 1.41 to 38.89  $\mu$ M. The highest impact was observed in esophageal squamous cell lines (11.08  $\mu$ M) and pancreatic carcinoma cells (6.84  $\mu$ M), with notable effects also observed in prostate, melanoma and colon cancer cells. Letícia et al. [219] evaluated the antiproliferative efficacy of *E. tirucalli* extracts against leukaemia (HL-60), lymphoma

(Daudi) and melanoma (B16F10) cell lines using methyl thiazol tetrazolium assay (MTT) at concentrations of 62, 125, 250 and 500  $\mu\text{g}/\text{mL}$ . There was a notable regional variation in the cytotoxicity of the extracts, displaying a dose-dependent pattern. The extracts exhibited comparable effectiveness against the leukaemia cell line HL-60, resulting in a reduction of cell viability to approximately 60–70%. In a separate study, other researchers assessed anti-proliferation effects of highly diluted latex and *E. tirucalli* homeopathic remedies on melanoma cells *In vitro*. The researchers created solutions of 0.5% and 5% concentration in 70%GL EtOH, which were then prepared for use. The findings indicated that the 0.5% latex solution at 30cH reduced melanoma cell growth by 19.7%, while the 0.5% *E. tirucalli* solution at 30cH showed a 32.1% reduction [220]. In another study, Waczuk et al. [217] conducted an evaluation of the cytotoxic effect of an aqueous extract obtained from *E. tirucalli* on human leukocytes. The results revealed that exposure to high concentrations of the extract induced a notable reduction in cell viability. In a study by Abdel-Aty et al. [221], the cytotoxicity of phenol content from *E. tirucalli* was evaluated against various cancer cell lines (HepG2, MCF7, A549, HL-60, HCT116) and human normal melanocyte, HFB4. The results showed that low concentrations of the phenolic content exhibited significant potent cytotoxicity against HL-60, with an  $\text{IC}_{50}$  value of  $22.76 \pm 2.85 \mu\text{g}/\text{ml}$ . Moreover, the extract demonstrated moderate cytotoxicity against MCF-7 and A549 cells, with  $\text{IC}_{50}$  values of  $31.65 \pm 3.67$  and  $35.36 \pm 3.82 \mu\text{g}/\text{ml}$ , respectively. Upon thorough evaluation of the anti-proliferative potential of the methanolic extract on MiaPaCa-2 cancer cell line, it was unequivocally observed that the extract exhibited a remarkable ability to significantly impede the growth of MiaPaCa-2 cancer cells [222]. The presented toxicity is in line with what was presented by Mavundza et al. [9].

#### 4.1.8. *Euphorbia ammak*

*E. ammak* is a plant that grows in Saudi Arabia and the Yemen peninsula (see Figure 1.8) [29].



**Figure 1.8.** *Euphorbia ammak* tree and flower (source: <http://luirig.altervista.org/cpm/albums/bot-units07/euphorbia-ammak16454.jpg>).

#### Phytochemistry

Abdel-Sattar et al. [94] screened the leaves of *E. ammak* and found three compounds: euphol,  $\alpha$ -glutinous and stigmasterol. Furthermore, Ponsinet and Ourisson [93] isolated euphol and euphorbol. Alkaloids, saponins and glycosides were detected by Al-Hajj et al. [31]. According to Al-Hajj et al. [31], it has been found to have good anti-leishmanial activity against *Cutaneous leishmaniasis*.

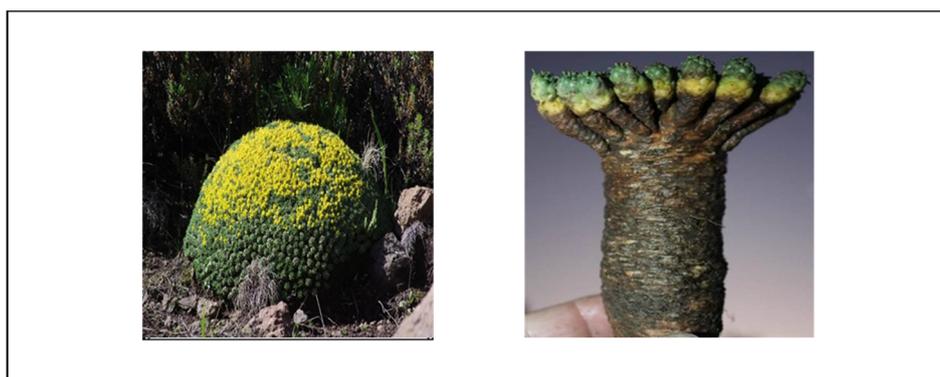
#### Toxicity

Studies have also shown that the methanolic extract of *E. ammak* could potentially reduce the H1N1 influenza virus and exhibits considerable cytotoxic activity against MDCK cells [32]. Additionally, Abdel-Satter et al. [33] found that the methanolic extract has anti-parasitic properties and that the euphol compound, which was also isolated, demonstrated significant cytotoxic effect against various human cancer cell lines *in vitro*. According to research conducted by Mampa et al. [65], the effect of four distinct *E. ammak* extracts on cell proliferation was evaluated. The extracts were obtained using various solvents such as Hexane, DCM, MeOH and EtOAc. The findings of the study indicated that the DCM extract exhibited the most significant inhibition of cell growth at

concentrations as low as 1 µg/mL. Almehdar et al. [30] evaluated the cytotoxic effect of *E. ammak* latex on MCF-7 breast cancer cells. The study found that the latex showed significant toxicity against MCF-7 cells, with an IC<sub>50</sub> value of 14.3.

#### 4.1.9. *Euphorbia clavarioides*

*E. clavarioides* is a plant that originates from South Africa and Lesotho (see Figure 1.9). It has been traditionally used to treat various skin conditions, such as skin rashes in children, acne, sores, bruises, burns, eczema, ulcers, cracked heels and wounds [53,54,223]. According to a study by Mbhele [224], *E. clavarioides* is effective in wound healing, confirming its traditional use. Additionally, in Lesotho, *E. clavarioides* is used for bathing swollen feet and, when mixed with *Berkheya onopordifolia*, it is used as a remedy for leprosy [224]. Other reports suggest that it can be used to treat herpes, HIV-related infections, high blood pressure, and diabetes [55–57].



**Figure 1.9.** *Euphorbia clavarioides* plant with flowers and stem (source: <https://www.inaturalist.org/observations/38217609>; [http://www.llifile.com/Encyclopedia/SUCCULENTS/Family/Euphorbiaceae/32951/Euphorbia\\_clavarioides\\_var.\\_truncata](http://www.llifile.com/Encyclopedia/SUCCULENTS/Family/Euphorbiaceae/32951/Euphorbia_clavarioides_var._truncata)).

#### Phytochemistry

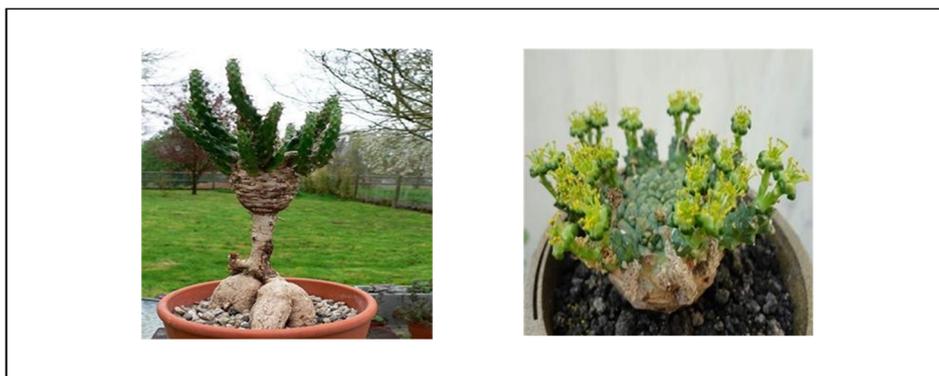
In 2019, Moteetee et al. conducted a phytochemical analysis of *E. clavarioides* and found that it contains alkaloids, flavonoids, saponins, terpenoids and tannins [225]. In 2020, Mampa et al. conducted a study on the same plant and isolated phytosterols, glycosides, triterpenoids, anthraquinone, flavonoids and alkaloids. Despite these findings, there is still a lack of research on this plant [65].

#### Toxicity

In a study conducted by Mampa et al. [65], the effect of Hex, DCM, MeOH and EtOAc derived from *E. clavarioides* extracts on cell proliferation was evaluated against the Vero cell line. The Hex and DCM extracts exhibited the highest cell growth inhibition of Vero cells.

#### 4.1.10. *Euphorbia gorgonis*

*E. gorgonis* is a plant species that can only be found in the Eastern Cape of South Africa (see Figure 1.10). Studies have shown that it has medicinal properties that can be used to treat cancer, wounds, swelling and skin problems [58,59]. The plant has also been found to have high antibacterial and antimicrobial activity [60].



**Figure 1.10.** *Euphorbia gorgonis* plant with flowers and stem (source: <http://www.bihrmann.com/caudiciforms/SUBS/eup-gor-sub.asp>).

### Phytochemistry

Studies have identified the presence of phytosterols, glycosides, triterpenoids, flavonoids and alkaloids (Mampa et al. [65]. While Tiwani [60] reported the presence of tannins, saponins, alkaloids and flavonoids.

### Toxicity

When tested on human intestinal cancer cells, the aqueous extracts of *E. gorgonis* extract showed cell reduction, while the acetone and ethanol extracts showed cytotoxic and reduced cell viability patterns, respectively. In another study, the acetone extract was found to reduce cell viability in rat hepatoma cells, while the aqueous extract showed a high percentage of cell viability [60]. Mampa et al. [65], evaluated the cytotoxicity effect of Hex, DCM, MeOH and EtOAc extracts on a Vero cell line. It was observed that none of the extracts displayed toxicity against the cell line.

#### 4.1.11. *Euphorbia bupleurifolia*

*E. bupleurifolia* is a plant native to South Africa and is commonly found in the Eastern Cape Province and Natal (see Figure 1.11). The milky latex of this plant has been historically used to treat various ailments including cancerous sores, painful cracked feet, eczema, pimples, rashes and wounds [61]. Additionally, the twigs of *E. bupleurifolia* have been utilized as a teeth-cleaning agent [61]. It has also been used to treat swelling in the lower limbs and cancer [62,63]. Some have even used it to help with retained placenta [64].



**Figure 1.11.** *Euphorbia bupleurifolia* plant with stem and flowers (source: <https://davesgarden.com/guides/pf/showimage/238493/#b>; <https://planetdesert.com/products/euphorbia-bupleurifolia-large>).

### Phytochemistry

Previous studies have revealed that *E. bupleurifolia* contains various secondary metabolites such as phytosterols, tannins, glycosides, triterpenoids, saponins, flavonoids and alkaloids [65]. Van Wyk et al. [62] specifically reported the presence of triterpenes in this plant.

### Toxicity

An investigation conducted by Mampa et al. [65] examined the potential cytotoxicity of *E. bupleurifolia* extracts on the Vero cell line. The findings indicated that both the Hex and DCM extracts had an antiproliferative effect on the Vero cell line. It is worth noting that the Hex and DCM extracts showed significant efficacy at concentrations of 1 and 10 µg/mL respectively.

#### 4.1.12. *Euphorbia polygona*

*E. polygona* is a plant species that originates in the Eastern Cape, South Africa (see Figure 1.12). There is no recorded information on its traditional and pharmacological use.



**Figure 1.12.** *Euphorbia polygona* plant with stem and flowers (source: [http://www.biodiversityexplorer.info/plants/euphorbiaceae/euphorbia\\_polygona.htm](http://www.biodiversityexplorer.info/plants/euphorbiaceae/euphorbia_polygona.htm)).

### Phytochemistry

A study conducted by Mampa et al. [65] discovered the presence of various compounds in the plant, including phytosterols, tannins, glycosides, triterpenoids, flavonoids and alkaloids.

### Toxicity

The study conducted by Mampa et al. [65] evaluated the antiproliferative effects of the Hex, DCM, MeOH and EtOAc extracts of *E. polygona* against the Vero cell line. The results of the study indicated that the Hex and DCM extracts exhibited the highest antiproliferative effect compared to the MeOH and EtOAc extracts.

#### 4.1.13. *Euphorbia arabica*

*E. arabica* can be found in various regions, including Botswana, southern Mozambique, Zimbabwe and South Africa (see Figure 1.13). This plant has been utilized in the past as an antibacterial agent, according to El-Shanwani [68]. It has also been used to treat ailments such as warts and stomachaches, while the juice has been used to treat skin infections [67,68].



**Figure 1.13.** *Euphorbia arabica* plant with stem and flowers (source: [http://www.biodiversityexplorer.info/plants/euphorbiaceae/images/1362101\\_658w.jpg](http://www.biodiversityexplorer.info/plants/euphorbiaceae/images/1362101_658w.jpg), [http://www.biodiversityexplorer.info/plants/euphorbiaceae/images/136210-2\\_658w.jpg](http://www.biodiversityexplorer.info/plants/euphorbiaceae/images/136210-2_658w.jpg)).

#### Phytochemistry

A recent study by Mampa et al. [65] has revealed the presence of phytosterols, tannins, glycosides, triterpenoids, anthraquinones and flavonoids.

#### Toxicity

When the cytotoxic effect of Hex, DCM, MeOH and EtOAc extracts of *E. arabica* were evaluated against a Vero cell line. The findings of the study indicate that the hexane extract of *E. arabica* inhibited cell growth of the Vero cell line, achieving IC<sub>50</sub> at all concentrations tested. It is worth noting that the DCM extract showed IC<sub>50</sub> at a concentration of 10 µg/mL.

#### 4.1.14. *Euphorbia ferox*

*E. ferox* is a plant species that is indigenous to the Western Cape region of South Africa (see Figure 1.14). Currently, there is no available information on the traditional and pharmacological activities and isolated compounds of *E. ferox*.



**Figure 1.14.** *Euphorbia ferox* plant with stem and flowers (source: [communities/southern-africa/view/observation/586073](http://communities/southern-africa/view/observation/586073), <http://gadi.agric.za/Grootfontein%20plants/Euphorbia%20ferox.html>).

#### 4.1.15. *Euphorbia stellata*

*E. stellata* is a plant species that can be found in the Eastern Cape region of South Africa (see Figure 1.15). There is currently no recorded information on the traditional and pharmacological uses or the isolated compounds of *E. stellata*.



**Figure 1.15.** *Euphorbia stellata* plant with stem and flowers (source:communities/southern-africa/view/observation/586073, <http://gadi.agric.za/Grootfontein%20plants/Euphorbia%20ferox.html>).

#### 4.2. Pharmacological Activities

The use of medicinal plants has gained attention due to their effectiveness in treating various ailments, and some of these claims have been supported by scientific evidence. Natural products are significant because of their diverse biological activities and drug-like properties, making them useful in the development of new lead compounds, natural drugs, pharmacological tools and herbal remedies [226]. The therapeutic properties of these plants are attributed to their unique bioactive chemicals.

Many species of *Euphorbia* are known for their medicinal properties, with over 5% of them used to treat various ailments such as warts, wounds, skin complications, tumor, respiratory disorder, sexually transmitted diseases, urinary tract infections and intestinal parasites throughout the world [17,24,25,28]. This is likely due to the presence of unique secondary metabolites and isolated compounds [227]. However, it is worth noting that the latex found in most *Euphorbia* species is toxic and can cause serious skin irritation and blindness. Despite the toxicity, latex also contains crucial biological active compounds such as terpenes, diterpenoids and triterpenes [228].

Several species of *Euphorbia* have demonstrated medicinal properties, as their extracts have been patented as prescription drugs. For example, *E. lathyris* extract (US 5707631) is used to treat arthritis, high cholesterol, Alzheimer's and blood pressure. Extracts from *E. peplus*, *E. hirta* and *E. drummondii* (US 6844013) have shown selective cytotoxicity against various cancer cell lines and their compounds are used to treat malignant melanomas and squamous cell carcinomas. *E. aaron-rossii*, *E. tirucalli*, *E. tomentella* and *E. tomentosa* (US 2003/0171334 A1) are used in treating prostate cancer. Additionally, *E. tirucalli* latex (US 2009/0142421 A1) has been found to be potent in treating diseases related to cell proliferation or angiogenesis [229]. Extract from *E. obesa* (US 6923993) has been shown to stimulate apoptosis and inhibit the growth of cancer cells. *E. hirta* (US 2007/0248694 A1) is used to reduce inflammation and *E. antiquorum* (US 2003/0165579 A1) has been found to inhibit tumor growth [6].

This study has analyzed 15 different species of *Euphorbia* and found a diverse range of secondary metabolites, which are important in biomedical sciences. The analysis identified several phytochemicals, including diterpenoids, triterpenes, sesquiterpenoids, phloracetophenones, cerebrosides, glycerols, flavonoids, and steroids, as well as isolated compounds (listed in Table 1.5).

##### 4.2.1. Flavonoids

a type of polyphenolic compound, were found in 9 species of *Euphorbia*, including *E. horrida*, *E. trigona*, *E. clavarioides*, *E. enopla*, *E. gorgonis*, *E. bupleurifolia*, *E. polygona*, *E. arabica* and *E. tirucalli*. Additionally, these have been observed in other species of the *Euphorbiaceae* family, including *E. microsciadia*, *E. heterophylla*, *E. hirta*, *E. neriifolia*, *E. paralysis*, *E. lunulata* and *E. larica*, indicating their prevalence within the *Euphorbiaceae* family. Studies have shown that flavonoids have medicinal properties that make them useful for treating different health conditions. These properties include antiinflammatory, enzyme inhibition, antimicrobial, estrogenic activity, antiallergic and antioxidant activity [230–232]. According to Mai et al. [233], *E. helioscopia* contains a significant amount of flavonoids and has shown to have a cytotoxic effect on triple-negative breast

cancer cells. In addition, according to research by Yang et al. [234] found that there was a decrease in the differentiation of 3T3-L1 preadipocytes, a reduction in triglyceride accumulation in mature adipocytes and a decrease in nitric oxide production in RAW 264.7 cells. According to a report by Galleggiante et al. [235], flavonoids can prevent the degradation of cAMP by phosphodiesterases and extend cAMP signaling by the enzyme. This leads to the development of antiinflammatory properties. According to El-Hawary et al. [98], the methanol extracts from *E. trigona* were found to cause cell death in MCF-7 and Caco-2 cells with IC50 values of 16.1 and 15.6 µg/mL, respectively. Previous research has shown that the butanol extract derived from *E. tirucalli* demonstrated effective cytotoxicity against MCF-7 cells and MDA-MB231, with IC50 values of 15 and 30 µg/mL [236]. Hence, researchers have been drawn towards using this secondary metabolite for pharmaceutical purposes.

#### 4.2.2. Alkaloids

are a type of organic compound that occur naturally and have at least one nitrogen atom [237]. It was found in 9 plant species, including *E. horrida*, *E. trigona*, *E. clavarioides*, *E. gorgonis*, *E. enopla*, *E. bupleurifolia*, *E. ammak*, *E. polygona* and *E. tirucalli*. The fact that alkaloids were found in nearly all of the chosen plants should not come as a surprise since previous studies have documented the presence of alkaloids in species from the Euphorbiaceae family [238–240]. Prescription medicines derived from plants with alkaloids have been used for many years. Their potent effects are attributed to the presence of alkaloids. Morphine, currently used as an analgesic, was the first alkaloid from Opium poppy [241]. Furthermore, other alkaloids such as vinblastine, quinine, morphine, atropine, nicotine, caffeine, ephedrine and strychnine are also used in medicine production. Alkaloids have several physiological effects on humans, such as antibacterial, antimitotic, antiinflammatory, local anesthetic, hypnotic, antitumor activity and several others [242].

#### 4.2.3. Saponins

which are glycosides, were present in 7 plants, namely; *E. horrida*, *E. trigona*, *E. clavarioides*, *E. gorgonis*, *E. bupleurifolia*, *E. polygona* and *E. ammak*. According to a report by Bigoniya and Rana [243], saponins have been found in *E. neriifolia*, *E. paralias* and *E. terracina* [244]. Saponins possess pharmacological and medicinal properties such as cell membrane permeability, hemolytic activity, antiviral, antifungal, antiinflammatory and antiallergic [245–247]. In addition, they have shown potential for cancer treatment by reducing cell invasiveness, induce cell cycle arrest and apoptosis, and also for suppressing angiogenesis [248]. Along with other antitumor medicines, saponins have been added to enhance their cytotoxicity in tumor treatment [249,250]. A study conducted by Xiao et al. [251] discovered that triterpene saponin found in the root bark of *Aralia dasyphylla* Miq. had the ability to inhibit cancer cells in two types of cell lines: KB and HeLa-S3. Another study discovered that eight steroidal saponins found in *Allium porrum* L. were able to inhibit WEHI 164 and J774 cells [252]. In 2001, Tran et al. conducted a study on *Dracaena angustifolia* Roxb. roots and rhizomes, testing spirostanol and furostanol-type saponins for their antiproliferative activity against murine colon 26-L5 carcinoma, human HT-1080 fibrosarcoma and B-16 BL6 melanoma cells [253]. The study found that three of the tested compounds were highly effective in inhibiting the growth of HT-1080 fibrosarcoma cells. According to a study by Yokosuka et al. [254], two types of saponins (ruscogenin glycoside and 26-glycosyloxyfurostanol saponin) demonstrated cytostatic activity against HL-60 human leukaemia cells. Researchers have discovered two new triterpenoid saponins, namely glycoside A and B, in the aerial parts of *Glinus oppositifolius* L. These saponins were found to be effective against *Plasmodium falciparum*, a type of protozoan [255]. In a study conducted by Iorizzi et al. [256], three new furostanol saponins and seven previously known saponins were extracted from the seeds of *Capsicum annuum* L. var. *acuminatum* Fingerh. However, the analysis showed that these saponins had little to no effect on the growth of Gram-positive and Gram-negative bacteria.

#### 4.2.4. Tannins

which are polyphenols, were found in 9 plants. These plants are *E. horrida*, *E. trigona*, *E. clavarioides*, *E. gorgonis*, *E. bupleurifolia*, *E. enopla*, *E. polygona*, *E. arabica* and *E. tirucalli*. These findings are in agreement with a study by Aleksandrov et al. [257], which also observed high levels of tannins in *E. hirta*. According to Fraga-Corral et al. [258], applying tannin topically can help remove irritants from the skin, reduce inflammation and be beneficial in treating burns and wounds due to its

antihaemorrhagic and antiseptic properties. Tannins have been used in various studies for treating different diseases due to their high antioxidant content, ability to scavenge free radicals and their antimicrobial, antiviral properties, and they have also been found to be effective in cancer chemotherapy [259–263]. Tannins were also reported to inhibit various coronavirus strains [264].

#### 4.2.5. Glycosides

which are acetal derivatives of monosaccharides, were present in 10 *Euphorbia* spp including *E. horrida*, *E. trigona*, *E. enopla*, *E. clavarioides*, *E. gorgonis*, *E. bupleurifolia*, *E. polygona*, *E. arabica*, *E. tirucalli* and *E. ammak*. Studies by Mshvildadze et al. [265] discovered that glycosides from the bark of *Betula papyrifera* and they had a significant cytotoxic effect against lung carcinoma, colorectal adenocarcinoma and normal skin fibroblast. Liu et al. [266] further reported the cytotoxic activity of glycosides from *Antiaris toxicaria* against human NIH460 lung cancer cells.

#### 4.2.6. Anthraquinones

which are phenolics, were present in 4 *Euphorbia* spp. namely; *E. horrida*, *E. enopla*, *E. clavarioides* and *E. arabica*. Anthraquinones are potent compounds found in various plant-based medicines that possess numerous health benefits, including acting as laxatives, diuretics, estrogenic agents and immunomodulators. Additionally, they are also utilized in cancer treatment. Moreover, they exhibit antibacterial, antiparasitic, insecticidal, fungicidal and antiviral properties as well [267].

According to studies by Hanson [268] and Berdy [269], several plants including *E. bupleurifolia*, *E. gorgonis*, *E. horrida*, *E. polygona* and *E. coerulescens* contain alkaloids, saponins and terpenoids. These compounds are believed to have pharmacological effects such as anticancer and antibacterial properties [270]. In addition, the report found that a single plant can treat multiple illnesses and many recorded plants have similar secondary metabolites. According to the data, seven different *Euphorbia* plants were used to treat cancer, these include *E. trigona*, *E. tirucalli*, *E. clavarioides*, *E. gorgonis*, *E. bupleurifolia* and *E. cooperi*. Five plants were used for wound healing and warts, which included *E. trigona*, *E. tirucalli*, *E. clavarioides*, *E. arabica* and *E. bupleurifolia*. Three plants, *E. trigona*, *E. bupleurifolia* and *E. gorgonis*, were used for inflammation, while *E. arabica* and *E. cooperi* were used for stomach aches, skin infections and cracked heels. The seven plants with cancer-fighting properties all contained common secondary metabolites, including phytosterols, tannins, glycosides, triterpenoids, saponins, upholds, flavonoids and alkaloids. It has been reported that the latex from *Euphorbia* species can be highly toxic if ingested and can cause severe skin irritation [34,41].

#### 4.2.7. Terpenoids

##### 4.2.7.1. Triterpenoids

Triterpenoids which are triterpenes, are present in most *Euphorbia* spp. In this study, it was discovered in 10 plants, including *E. horrida*, *E. enopla*, *E. trigona*, *E. clavarioides*, *E. gorgonis*, *E. bupleurifolia*, *E. polygona*, *E. cooperi*, *E. arabica* and *E. tirucalli*. It has been found that these secondary metabolites have medicinal properties, including being anticarcinogenic, antimalarial, antiulcer, antiinflammatory, cytotoxic, antiHIV, antiangiogenic, hepatocidal, antimicrobial and antiviral [271–273]. According to Munro et al. [222], the tetracyclic triterpene euphol found in *E. tirucalli* latex has been proven to have anticancer properties. According to Yasukawa et al. [103], *E. kansui* contains lanostane-type triterpenes that can inhibit inflammation caused by TPA. The main triterpene in this plant, euphol, was found to effectively prevent tumor promotion induced by TPA.

**Euphol**, a tetracyclic triterpene, is commonly found in the *Euphorbiaceae* family, including *E. trigona*, *E. enopla*, *E. tirucalli*, *E. coerulescens* and *E. ammak*. Popplewell et al. [72] discovered that this compound showed moderate activity against the HepG2 cells. Ahmed et al. [181] also reported remarkable cytotoxic effect of euphol from *E. bothae* against the MCF-7 cells. Silva et al. [102] evaluated the antitumor effect of euphol on a number of human cancer cell lines and discovered that euphol from *E. tirucalli* was cytotoxic to several cancer cell lines, including esophageal squamous and pancreatic carcinoma cells. Meanwhile, Abdel-Sattar et al. [94] found that euphol from *E. ammak* exhibited notable cytotoxic effect against HeLa cells. In other studies, euphol from *E. umbellata* was found to exhibit noteworthy cytotoxic effect against HL-60, K-562 and B16F10 cells, while Yasukawa et al. [103] discovered that topical application of euphol from *E. kansui* greatly reduced the cancer-

promoting impact of TPA in mouse skin, lowering it by 90%. Akihisa et al. [147] reported that euphol was a successful inhibitor of HIV-1 reverse transcriptase. Additionally, euphol has shown potential in reducing inflammation and pain by blocking PGE2 and protein C kinase epsilon mediators [168]. However, data has yet to be documented on the euphol of *E. trigona*, *E. enopla*, *E. tirucalli*, *E. coerulescens* and *E. ammak*.

**Cycloartanol** is a type of sterol lipid that has been found in various plants including *E. trigona*, *E. glareosa* Pall. Ex M. Bieb., *E. amygdaloides* L. and *E. palustris* L. [105]. According to a recent report by Salome-Abarca et al. [105], cycloartenol extracted from these plants was noted to have antifungal properties against *B. cinerea*. Barla et al. [110] also discovered cycloartanol in *E. helioscopia*, but observed no vasodepressor activity. Meanwhile, Heliawati et al. [104] conducted a study on the cytotoxicity of cycloartanol extracted from the bark of *Corypha utan* Lamk on leukemia cells. The research indicated that cycloartanol was able to restrict the growth of these cells. Nevertheless, there is currently no information available on the effects of cycloartanol from *E. trigona*.

**Lupeol**, a triterpenoid, is found in many plants including *E. trigona*, *Tamarindus indica*, *Celastrus paniculatus*, *Zanthoxylum riedelianum*, *Allanblackia monticola*, *Himatanthus sucuuba*, *Leptadenia hastata*, *Crataeva nurvala*, *Bombax ceiba*, *Sebastiania adenophora*, *Aegle marmelos* and *Emblica officinalis* [114,274–277]. You et al. [116] studied the effects of Lupeol from *Bombax ceiba* on various cells such as HUVEC, SK-MEL-2, A549, and B16-F10 melanoma cells. Their findings showed that lupeol effectively inhibited the formation of HUVEC tubes by over 80% at a concentration of 50 µg/mL. However, there was no notable cytotoxic effect observed in the three cancer cell lines. Nguemfo et al. [278] conducted a study on the antiinflammatory properties of lupeol found in *Allanblackia monticola* Staner. The findings demonstrated that within just 30 minutes, lupeol effectively reduced paw edema by approximately 57.14%. Zhang et al. [111] conducted a study on lupeol's ability to inhibit the growth of liver cancer cells called HCCLM3. It was found that lupeol effectively stopped liver cancer cell growth by suppressing the secretion of Brain-Derived Neurotrophic Factor, phosphatidylinositol 3-kinase and Wnt pathways. Borgati et al. [113] conducted a study on the effectiveness of lupeol from *Parahancornia fasciculata* against chloroquine-resistant W2 clones in treating malaria. The study found that the compound had limited activity. Other studies have explored the antidiabetic properties of lupeol found in *Solanum xanthocarpum* [279]. The findings showed that lupeol can effectively hinder the advancement of diabetes by decreasing glucose levels, nitric oxide, further increases serum insulin and antioxidant levels. Sudhakar et al. [280] conducted a study on the impact of lupeol from the *Crataeva nurvala* plant on cholesterol levels. They discovered that this compound can reduce oxidative stress and inflammatory cytokines, which in turn leads to a decrease in nitric oxide production and ultimately results in lower cholesterol levels. However, data has yet to be documented on the lupeol of *E. trigona*.

**α-amyirin**, a type of triterpenoid, can be found in several plants including *E. trigona*, *E. tirucalli*, *E. aphylla* Brouss, *E. schimperi* C. Presl and *E. hirta* [123,125,281]. Abdel-Monem et al. [123] also discovered that α-amyirin obtained from *E. schimperi* C. Presl displayed moderate cytotoxicity, resulting in a 40-50% reduction in u251 and MCF-7 cell lines at a concentration of 10 µg/mL. Other studies reported insignificant cytotoxicity against HCT 116 cell line [281]. Jabeen et al. [120] conducted an analysis of the antifungal properties of α-amyirin from *Melia azedarach* L. in vitro. The results indicated that the compound was effective against *Ascochyta rabiei*, with a MIC of 0.0156 mg mL<sup>-1</sup>. However, there is currently no available data on the effects of α-amyirin from *E. trigona* and *E. tirucalli*.

**β-amyirin**, a triterpenoid, was found in *E. trigona* and *E. hirta* and has demonstrated anticancer properties against Hep-G2 cancer [118]. The same compound also exhibited weak cytotoxic effect against NTUB1, A549 and HL-60 [119]. According to Ragasa and Cornelio [281] insignificant cytotoxicity against HCT 116 cell line was observed. According to Vazquez et al. [125], a study showed that the antiinflammatory activity in mice ears was effective. Another study by Shih et al. [121] found that β-amyirin from *E. hirta* could treat arthritis inflammation by blocking the function of the nitric oxide pathways. According to Lin et al. [119], the use of β-amyirin together with cisplatin caused reactive oxygen species to trigger cell cycle arrest and apoptosis in NTUB1 cells. However, there is currently no documented information on the β-amyirin found in *E. trigona*.

**Betulinic acid**, a triterpenoid, has been discovered in various plants such as *E. trigona*, *Tetracarpidium conophorum* seeds, *Uapaca paludosa*, *Manniophyton fulvum* (*Euphorbiaceae*) and *Agathosma*

*betulina* [282–285]. A study conducted by Zhang et al. [111] showed that betulinic acid has the potential to inhibit the growth of more than 20 different cancer cells. According to studies conducted by Damle et al. [128], betulinic acid was found to have cytotoxic effects on MCF-7 cells with an IC<sub>50</sub> value of 13.5 mg/mL. Additionally, the compound was found to reduce the level of specificity of protein transcription factors, which are known to be overproduced in cancer cells compared to normal cells [130]. A study conducted by Mbeunkeu et al. [283] found that betulinic acid from *Manniophyton fulyum* had a substantial impact on HeLa cells, resulting in a cell viability rate of only 4%. A study conducted by Foo et al. [127] discovered that the DCM fraction of *Dillenia suffruticosa* can cause cell cycle arrest and apoptosis of MCF-7 cells through the p35/p21 pathway. The compound was found to inhibit the growth and ability of all human melanoma cell lines to form colonies [135]. According to Oriakhi et al. [282], there was a 54% inhibition of HepG2 cells as well as hepatoprotective activity, whereby a binding energy of -11.2 kcal/mol to the Hepatitis B virus DNA was observed. Other studies reported antidiabetic activity, where the pancreatic  $\alpha$ -amylase is inhibited [131,286]. According to Bernard et al. [134], betulinic acid was found to have an antiinflammatory effect by inhibiting phospholipase A2. Additionally, Mukherjee et al. [287] reported that betulinic acid reduced rat paw edema caused by carrageenan and serotonin. According to additional research, *Hypericum hircinum* L. contains betulinic acid which can prevent HIV-1 from replicating by stopping reverse transcriptase-associated DNA polymerase. This is achieved by either hindering the fusion of HIV or disrupting a certain stage of its maturation process [129,133]. However, data has yet to be documented on the betulinic acid of *E. trigona*.

**Taraxerol**, a triterpenoid, has previously been isolated from *E. trigona*, *E. neriifolia* Linn, *Artemisia roxburghiana*, *Taraxacum japonicum* and Fruits of *Dregea volubilis* [136,139,141,288]. Taraxerol isolated from *Vepris punctate* showed limited activity against the A2780 cells [289]. Studies by Cao et al. [290] also showed minimal inhibition of cell growth of A2780 cells at the highest tested concentration. Taraxerol from *Conyza canadensis* was found to have a maximum antiproliferative effect against A431. However, taraxerol displayed no activity against HeLa, MCF-7 and MRC-5 [291]. In addition, it was found that taraxerol effectively hindered the growth of AGS cells by causing G(2)/M arrest and stimulating cell apoptosis [138]. According to Takasaki et al. [141], taraxerol has been found to be highly effective in preventing tumors in mice during two-stage carcinogenesis tests. In addition, Singh et al. [140] reported that taraxerol has antiinflammatory properties, reducing paw edema by 48.61%. Furthermore, it was also observed that taraxerol exhibited moderate antimicrobial activity against certain gram negative and gram positive bacteria [140]. According to a study conducted by Min et al. [292], taraxerol found in *Styrax japonica* showed low effectiveness in scavenging free radicals based on the DPPH assay. Sangeetha et al. [293] investigated the use of taraxerol from *Mangifera indica* as an antidiabetic agent. According to the findings, the compound has dual activity as a glucose activator and stimulator of glycogen, making it a potential treatment for type 2 diabetes. However, there is currently no documented information available on the taraxerol of *E. trigona*.

**Taraxerol acetate**, a triterpenoid, was previously isolated from *E. trigona*, *E. pubescens* and *Artemisia roxburghiana* [294]. Studies have shown that taraxerol acetate from *A. roxburghiana* has the ability to significantly reduce edema in mice caused by carrageen [136]. In addition, Rehman et al. [137] discovered that taraxerol acetate can inhibit Cyclooxygenases enzymes 1 and 2. However, there is currently no documented data on the taraxerol acetate found in *E. trigona*.

**Friedelin**, which is a type of triterpenoid, has been found in plants such as *E. trigona*, *E. tortilis* Rottler, *Mangifera indica* and *Lentinus edodes* [153,295,296]. Previous studies have shown that when investigated for its antibacterial properties against certain gram positive and gram negative bacteria, friedelin had weak activity with a minimum inhibitory concentration (MIC) value of over 250  $\mu$ g/mL [153]. However, it was discovered that friedelin from *Mangifera indica* had anti-colorectal cancer activity [154]. Although studies have yet to be conducted on friedelin from *E. trigona*, it is an area of interest for future research.

**Friedelan-3- $\beta$ -ol**, a triterpenoid, was also isolated from *E. trigona* and *Mangifera indica*. The compound exhibited significant inhibition of thymidylate synthase, thereby displaying its anti-colorectal potential [154]. However, data has yet to be documented on the friedelan-3- $\beta$ -ol of *E. trigona*.

**3 $\beta$ -Friedelinol**, a triterpenoid, has been found in *E. trigona*, *E. vajravelui*, *E. kamerunica* and *Maytenus robusta* [157,158]. Sousa et al. [158] discovered that 3 $\beta$ -friedelinol from *Maytenus robusta*

exhibited cytotoxicity against 4T1 cells. Meanwhile, Ogunnusi et al. [157] investigated the antibacterial activity of 3 $\beta$ -Friedelinol from *E. kamerunica* against certain bacteria and found that the compound had inhibitory activity. However, there is currently no available data on the 3 $\beta$ -Friedelinol of *E. trigona*. The latex of *E. trigona* has ingenol esters that are highly irritating to the skin. Studies have shown that it can cause dermatitis in open patch tests and blisters in closed patch tests [16]. Two compounds isolated from *E. trigona*, Epi-Friedelinyl acetate and Rhoiptlenone, have no known biological effects.

#### 4.2.8. Phytosterols

Phytosterols which are sterols, were present in 9 plants, including *E. horrida*, *E. trigona*, *E. clavarioides*, *E. gorgonis*, *E. enopla*, *E. bupleurifolia*, *E. cooperi*, *E. polygona* and *E. arabica*. Research suggests that phytosterols possess bioactive properties that offer multiple health benefits, such as reducing inflammation, preventing oxidative stress, fighting against cancer and lowering cholesterol levels [297,298]. A mixture of phytosterols inhibited tumor development in various cancer forms, including cholangiocarcinoma and breast cancer, at physiological doses [299,300].

**Cycloartenol** is a phytosterol compound found in certain plants such as *E. trigona*, *E. nicaeensis*, *E. broteri*, *E. macrosteigia*, and *E. boetica* (NHI). It serves as a precursor to various sterol compounds and has several pharmacological benefits such as antiinflammatory, antitumor, antioxidant, antibacterial and antiAlzheimer's properties [108,109]. Niu et al. [107] conducted a study on the potential anticancer properties of cycloartenol in relation to glioma U87 cells. The findings indicated that cycloartenol was able to hinder the growth and ability of the glioma U87 cells to form colonies. This was due to the induction of Sub-G1 cell cycle arrest and apoptosis, both of which contributed to the antiproliferative effects observed. According to Zare et al. [109], cycloartenol has displayed anticancer, analgesic and bactericidal properties. Meanwhile, Sawale et al. [106] discovered that cycloartenol from *E. neriifolia*, at concentrations of 10, 20, 40, 60, 80 and 100 $\mu$ g /mL, exhibited antioxidant activity ranging from 34.56% to 72.87%. However, data has yet to be documented on the Cycloartenol of *E. trigona*.

**$\beta$ -sitosterol**, a sterol, was previously isolated from *E. trigona*, *E. abyssinica*, *Pinellia ternate* and *Nyctanthes arbortristis* [142,143,145].  $\beta$ -sitosterol was investigated for its effect on the growth of Caski and HeLa cells; investigating its potential antiproliferation and anticancer effects. The results revealed that the compound was found to reduce the expression of antigen responsible for cell proliferation in both cervical carcinoma cells [142]. In their 2015 study, Cheng et al. discovered that elevated levels of P53 and decreased levels of HPVE6 viral oncogenes resulted in anticancer activity in Caski and HeLa cells [142,153]. Previous research has indicated that  $\beta$ -sitosterol derived from *Nyctanthes arbortristis* leaves had a considerable antiinflammatory impact on the paw edema of rats [143].  $\beta$ -sitosterol derived from *E. abyssinica* exhibits notable antimicrobial properties against *Candida albicans* [145]. According to reports,  $\beta$ -sitosterol found in *E. hirta* was able to prevent inflammation caused by TPA [125]. However, there is currently no documented data on the  $\beta$ -sitosterol of *E. trigona*.

**24-Methylenecycloartanol**, a type of sterol, was previously isolated from *E. hirta*, *E. heteradena*, *E. trigona*, *E. broteri*, *E. palustris*, *Moroccan propolis* and *E. aleppica* [155,301–303]. According to Krstić et al. [155], it was found that 24-Methylenecycloartanol from *E. palustris* has a stronger antifungal effect against *Fusarium sporotrichioides* and *Alternaria alternata*. Previous research has shown that 24-Methylenecycloartanol found in *E. hirta* can reduce inflammation caused by TPA [125]. However, there is currently no available data on the effects of 24-Methylenecycloartanol of *E. trigona*.

**Tirucallol**, a tetracyclic triterpene, was found in *E. enopla* and *E. lacteal* latex. This compound has antiinflammatory effects. Other studies have shown that mice models experienced a decrease in ear edema and that nitrate production in stimulated macrophages was inhibited [146,304]. Furthermore, it was discovered that it inhibits HIV-1 reverse transcriptase [147]. However, there is currently no available data on the tirucallol of *E. enopla*.

**Obtusifoliol** is a type of sterol that is present in various plants, including *E. cooperi*, *E. bothae*, *E. chamaesyce* and *E. sogdiana* [72,305]. A study by Aghaei et al. [198] found that obtusifoliol from *E. Sogdiana* has reduced the cytotoxic effect against MCF-7 and MDA-231, with an IC<sub>50</sub> value of 29.33  $\pm$  1.52 and 41.81  $\pm$  2.42  $\mu$ M; respectively. Similarly, Ahmed [197] reported weak cytotoxicity of obtusifoliol from *E. cooperi* against MCF-7.

**Glutinol**, a triterpenoid, was isolated from *E. cooperi*, *E. ammak*, *E. chamaesyce* and *Scoparia dulcis* [94,306]. This compound exhibited a notable cytotoxic effect against HeLa cells [94]. A study conducted by Ding et al. [194] investigated the potential toxicity of glutinol from *Acer mandshuricum* on different types of cells; including leukaemia, ovary, lung and human colon. The findings showed that the cell lines were significantly inhibited, with GI<sub>50</sub> values ranging from 11.6 to 16.0. Another research by Chen and Li [307] also reported the antiproliferation of ovarian cells. However, there is currently no available data on the effects of glutinol from *E. cooperi* and *E. ammak*.

**Triterpene euphol**, a steroidal alcohol, is found in plants such as *E. tirucalli*, *E. umbellata* and *Synadenium grantii* [308,309]. Studies have shown that triterpene euphol, the primary compound present in *E. umbellata* and *E. tirucalli*, shows potential as a supplementary cancer treatment [52,308]. According to Lin et al. [310], triterpene euphol exhibited moderate cytotoxic activity against gastric adenocarcinoma cells. Additionally, Silva et al. [102] reported that triterpene euphol had a cytotoxic effect on glioblastoma cells. de Oliveira et al. [309] conducted a study on the antitumoral effect of triterpene euphol from *Synadenium grantii*. The study found that the compound did not exhibit any antitumoral effect on B16F10 melanoma cells. A study conducted by Dutra et al. [168] focused on the potential of triterpene euphol found in *E. tirucalli* in preventing and treating inflammation in the colons of mice. Their findings showed that this compound was highly effective in reducing pro-inflammatory mediators in vitro.

**24-Methylene cycloartenol**, a sterol, was previously extracted from the *E. tirucalli* and *E. neriifolia*. The plant was tested for its pharmacological activities, rather than the compound itself. Additionally, 24-Methylene which was isolated from *E. hirta*, has been shown to have significant anti-inflammatory effects when used to treat ear inflammation caused by acetate anti-inflammatory [189]. There is currently no information available regarding the anticancer properties of 24-Methylene cycloartenol from *E. tirucalli*.

**Ingenol triacetate**, a diterpene, has shown to have an antimicrobial effect against different pathogens that cause infectious diseases [190]. Ingenol triacetate was also found to be a non-tumor promotor [190]. However, a separate study by Tilabi and Upadhyay [191] discovered that when ingenol triacetate was applied topically to female NMRI mice, it resulted in a high occurrence of lung adenoma. There is currently no information available regarding the anticancer properties of Ingenol triacetate from *E. tirucalli*.

**Terpenic alcohol** which is a terpene, from *E. tirucalli*, has demonstrated antibacterial properties against *S. aureus*. This is due to its ability to damage cell membrane [148,311]. However, this study was not directly done on terpenic alcohol [149]. More studies need to be performed on this compound, especially that which is isolated from *E. tirucalli*.

**Taraxasterol**, which is a phytosterol was extracted from *E. tirucalli*, *Carthamus tinctorius*, *Chrysanthemum morifolium* and *Helianthus annuus* [178,312]. Taraxasterol was effective in reducing ear inflammation caused by TPA in mice and also in preventing tumor growth on mouse skin. When taraxasterol was tested on the skin of mice, it demonstrated strong antitumor activity during two-stage carcinogenesis tests. Furthermore, the research showed that taraxasterol has the ability to suppress spontaneous mammary tumors in C3H/OuJ mice. The experiment was conducted with taraxasterol, which was not extracted from *Euphorbia* species [178]. Further pharmacological testing is required on *Euphorbia* species, especially on *E. tirucalli*, as suggested by Ovesnà et al. [178].

**Stigmasterol** which falls under the sterol class, was extracted from *E. ammak* and *Butea monosperma* [195]. In a separate study by Abdel-Sattar et al. [94], the cytotoxicity activity of stigmasterol from *E. ammak* was examined. The study showed that the compound was effective against HeLa cells. In 2009, Panda and colleagues conducted a study on the effects of stigmasterol from *Butea monosperma* [195]. Their findings showed that stigmasterol had properties that could inhibit the thyroid and lower blood glucose levels. The study further showed that stigmasterol has the potential to reduce liver damage caused by oxidative stress, as it decreased the level of harmful lipid peroxidation and increased the activity of protective enzymes. Studies have shown that this compound exhibits cytotoxicity against MCF-7 breast cancer cells [24,28]. Although some *Euphorbia* diterpenoids (phorbol ester) are toxic, recent research indicates that certain compounds in this plant have noteworthy bioactivity [26,27].

#### 4.2.9. Other terpenoids

**12-Deoxyphorbol-13-isobutyrate-20-acetate** is a type of phorbol ester that can be found in various plants such as *E. ledienii*, *E. coerulescens*, *E. tirucalli*, *E. triangularis*, *E. resinifera* and *E. bothae*. Studies by Ourhizif et al. [161] reported cytotoxicity of 12-Deoxyphorbol-13-isobutyrate-20-acetate from *E. resinifera* latex which inhibited the growth of *A. carbonarius*. However, there is currently no available data on the 12-Deoxyphorbol-13-isobutyrate-20-acetate found in *E. ledienii*, *E. coerulescens* and *E. tirucalli*.

**Phorbol**, a diterpenoid, was extracted from *E. ledienii* and *Croton tiglium*, a plant belonging to the *Euphorbiaceae* family. It was also discovered in *E. tirucalli* by Fürstenberger and Hecker [7]. While some research suggests that phorbol can irritate and promote tumor growth, other studies have found that it may induce apoptosis in tumor cells by activating protein kinase C [162]. However, there is currently no available information on the phorbol found in *E. ledienii* and *E. tirucalli*.

**12-Deoxyphorbol ester**, a diterpenoid, was isolated from *E. ledienii* and *E. grandicornis*. According to a study by Zayed et al. [163], this compound was found to activate protein kinase C instead of promoting tumor growth. Previous research has also shown that this activation can have antiproliferative effects on certain types of cancer cells. According to research conducted by Shen et al. [164], 12-Deoxyphorbol ester was found to prevent the growth of myeloid leukemia cells. Additionally, other studies showed that 12-Deoxyphorbol ester increased cell death in breast cancer cells compared to normal breast epithelial cells [165]. According to a study by Tsai et al. [166], 12-Deoxyphorbol ester has the capacity to stop the growth and cause cell death in human lung cancer cells by activating the PKC- $\delta$ /PKD/ERK Signaling Pathway. However, there is currently no information available on the effects of this compound from *E. ledienii*.

**12-Deoxy-16-hydroxy-phorbol**, is a component of DHPB which has been found in the latex of several plants including *E. ledienii*, *E. poisonii* Pax, *E. cooperi*, *Jatropha curcas* and *Jatropha gossypifolia* [313,314]. It was found to stimulate ornithine decarboxylase in mouse skin, restrict the binding of [3H]-12-O-tetradecanoylphorbol-13-acetate to phorbol ester receptors and activate protein kinase C in vitro [167]. However, data has yet to be documented on the 12-Deoxy-16-hydroxy-phorbol of *E. ledienii* and *E. cooperi*.

**16-Hydroxy-12-desoxyphorbol**, a diterpenoid, was isolated from *E. cooperi*, *E. bothae*, *E. triangularis*, *E. ingens* and *Croton rhanmmifolius*, and *E. rowlandii* [196,315]. Dinala et al. [196] documented that this compound showed activity against HCC70 and MCF-7 with EC<sub>50</sub> values of 0.592 and 1.003 $\mu$ M; respectively. Contrary to this, Gschwendt and Hecker [97] reported the compound as a tumor promoter. However, data has yet to be documented on the 16-Hydroxy-12-desoxyphorbol of *E. cooperi*.

**12-Deoxyphorbol-13-isobutyrate-16-angelate-20-acetate**, which belongs to the diterpene class, was discovered in *E. cooperi* and *E. bothae* [72]. Ahmed [197] reported that 12-Deoxyphorbol-13-isobutyrate-16-angelate-20-acetate from *E. cooperi* demonstrated noteworthy cytotoxicity against MCF-7 cells.

**12-Deoxy phorbol esters**, which fall under the phorbol ester class, were previously isolated from *E. tirucalli*, *E. triangularis*, *E. resinifera* and *Excoecaria bicolor* [97,163,316]. It was reported that 12-Deoxy phorbol esters are carcinogenic and a tumor promotor [97,175]. Studies by Driedger and Blumberg [174] reported inflammation in mice ears after exposure to 12-Deoxy phorbol esters.

**Ingenol**, a diterpenoid, was discovered in *E. tirucalli* and *E. sikkimensis*. In 2019, Silva and colleagues conducted research on the impact of ingenol, derived from *E. tirucalli*, on multiple human cancer cell lines [102]. The findings showed that the compound had varying degrees of effectiveness, ranging from weak to potent, against the different cell lines. In other studies, ingenol effectively prevented the replication of HIV-1 subtype B and C in both MT-4 cells and human PBMCs [152].

**Glut-5-en-3- $\beta$ -ol**, a triterpenoid, was discovered in *E. tirucalli*, *E. pseudocactus* Berger [176]. Abdel-Monem and Abdelrahman [176] conducted a study on Glut-5-en-3- $\beta$ -ol's ability to fight against various microorganisms. However, the results showed that Glut-5-en-3- $\beta$ -ol did not exhibit any activity against any of the tested bacteria.

#### 4.2.10. Phenolic Compounds

**Gallic acid**, a phenolic acid from *E. cooperi*, is effective in protecting the liver and preventing liver damage caused by paracetamol. It also has antioxidant properties that can improve the levels of

serum alanine aminotransferase, aspartate aminotransferase, alkaline phosphatase and bilirubin antioxidant [181].

**Bervifolin carboxylic acid**, a tannin from *E. cooperi*, has a strong hepatoprotective effect. It also acts as an antioxidant, reducing the levels of serum alanine aminotransferase, aspartate aminotransferase, alkaline phosphatase and bilirubin in rats with paracetamol-induced hepatotoxicity [181].

**3, 3'-Dimethoxy ellagic acid** a tannin, from *E. cooperi* displayed significant hepatoprotective effect as well as antioxidant effect. In particular, it was able to lower levels of serum alanine aminotransferase, aspartate aminotransferase, alkaline phosphatase and bilirubin in rats with paracetamol-induced hepatotoxicity. In addition, the compound showed moderate activity against HepG2 cells [181].

**Ellagic acid**, a tannin from *E. cooperi*, has been shown to have many benefits for liver health in rats. In the same study, it has been found to protect the liver, prevent the growth of cells and act as an antioxidant [181,197]. It also reduces the levels of certain enzymes and bilirubin in the blood, which are commonly elevated in cases of liver damage caused by paracetamol antioxidant [181,197].

**Kaempferol and Kampferol-3-O-β-D-rutinoside**, which are a class of flavonoids from *E. cooperi*. They were found to be effective in protecting the liver, preventing excessive cell growth and reducing the levels of certain enzymes and bilirubin in the blood, all of which are associated with liver damage caused by paracetamol antioxidant [181,197].

**1-O-Galloyl-3,6-hexahydroxydiphenyl-β-D-glucopyranoside/corilagin** a tannin, was extracted from various plants including *E. cooperi*, *E. prostrata*, *Phyllanthus amarus* L, *Phyllanthus niruri* L and *Euphoria longana* Lam (*Euphorbiaceae*) [200,203,206,317,318]. In 2013, Ming and colleagues conducted a study on the potential antitumor effects of corilagin from *Phyllanthus niruri* L. on three types of cells - Chang-liver, SMMC7721 and Bel7402 [201]. The findings showed that the compound had a moderate level of effectiveness against SMMC7721 and Bel7402 cells. However, it only exhibited weak activity against Chang-liver cells. In a study by Bai et al. [202], corilagin extracted from *Dimocarpus longan* Lour was found to have a cytotoxic effect on A549 cells. Another research by Tong et al. [199] discovered that corilagin from *Phyllanthus urinaria* induced apoptosis through reactive oxygen species and autophagy. During autophagy mediation, the Akt/mTOR/p70S6K pathway was suppressed and there was an increase in the formation of autophagic vacuoles and conversion of LC3-I to LC3-II. According to a report by Ahmed [197], corilagin showed notable toxicity against MCF-7 cells. Kolodziej et al. [203] studied the effects of corilagin from the *Phyllanthus amarus* L. plant on various genes related to inflammation including iNOS. Their findings showed that corilagin increased the mRNA levels of iNOS and cytokines in parasitized cells. In 2013, Jin and colleagues conducted a study on animals to explore the effects of corilagin on liver protection and inflammation reduction [205]. The findings indicated that corilagin was able to inhibit the NF-κB pathway, leading to an increase in the level of superoxide dismutase, a key defense mechanism against superoxide radical toxicity and nitric oxide, which plays an important role in liver metabolism. Previous research has shown that corilagin, derived from *E. longana* Lam, can decrease blood pressure by reducing the release of plasma noradrenaline and inducing direct vasorelaxation [206]. Yang et al. [319] studied the impact of corilagin from *Terminalia bellerica* Roxb on HepG2 cells; specifically, its potential to treat diabetes. Their findings showed that corilagin increased PPARγ signaling, which led to an increase in adipogenesis. According to studies by Latté and Kolodziej [320], from the *Zeitschrift für Naturforschung*, corilagin has a strong antifungal effect against *Candida glabrata*. Notka et al. [321] reported HIV-1 replication inhibition from corilagin of *Phyllanthus amarus*.

**Tri-methyl ellagic acid** is a tannin which was previously isolated from *E. tirucalli* and *E. sorori*. In 2008, Zhang et al. conducted a study to assess the antibacterial effect of tri-methyl ellagic acid on various types of bacteria [151]. The results indicated that the compound had a moderate effect on *B. subtilis* and *S. aureus*, but did not show any inhibitory activity against *E. coli*. There is currently no information available regarding the anticancer properties of tri-methyl ellagic acid from *E. tirucalli*.

**3'-Di-O-methylellagic acid**, which is a phenol, has been isolated from *E. tirucalli*, *E. lunulata* and *E. schimperiana* [322]. Researchers have tested the cytotoxicity of this compound from *E. schimperiana* extract on four human cancer cell lines. The results showed that 3,3'-Di-O-methylellagic acid had promising cytotoxicity against PC3 cells [184,323], with an IC50 of 5.5 μg/mL. This depicts the likely anticancer activity of the compound [184]. Research has found that 3'-di-O-methylellagic acid, which

was extracted from *E. schimperiana*, has exhibited promising antibacterial properties. This is reported by the same author. Aljubiri et al. [184] confirmed the antiinflammatory activity of 3'-Di-O-methylellagic acid from the *E. lunulate* isolate. Guo et al. [185] discovered that *E. hylonoma* contains 3'-Di-O-methylellagic acid, which has a strong antioxidant effect. A great efficacy of a potential antimicrobial agent by the compound was observed from the isolate of *E. thymifolia*.

#### 4.2.11. Fatty Acids

**Laurate**, a fatty acid, has been found in *E. coerulescens* and has limited antimicrobial activity against beneficial lactic acid bacteria but strong antimicrobial activity against harmful *Bacteroides* and *Clostridium* bacteria [192,324]. However, there is currently no available data on the effect of laurate of *E. coerulescens*.

#### 4.2.12. Miscellaneous

**Diterpene esters**, a terpene, were extracted from the latex of *E. horrida*, *E. periplus*, *E. tirucalli* and *E. cauducifolia* [7,325]. It is a skin irritant and tumor promoter [7,325]. Contrary, studies by Kedei et al. [326] reported that diterpene ester is an anticancer agent as it activates the protein kinase C, which has previously been proven to induce an anticancer effect on cells. According to research conducted by Ogbourne et al. [327], applying high doses of diterpene ester topically cured skin cancer in mice through protein kinase C. Another study by Hampson et al. [328] found that applying a much lower dose of diterpene ester resulted in cell death through a protein kinase C mechanism in leukemia cells. Nothias-Scaglia et al. [329] reported that diterpene ester could also inhibit HIV replication at the nanomolar level. However, there is currently no available data on the effects of diterpene esters from *E. horrida* and *E. tirucalli*.

Out of the 15 *Euphorbia* plants that were studied, some of the compounds were left unexplored. These include Ingol-7,8,12-acetate and ditiglate from *E. ledienii*, as well as 12-Deoxyphorbol-13-(2-methyl butyrate)-20-acetate. One compound from *E. horrida* was identified as 17-Hydroxyingenol-17-benzoate-20-angelate. Additionally, five compounds were identified from *E. coerulescens*: Angelate acetate isobutyrate, Fatty acids, Acetate laurate,  $\alpha$ -Methyl butyrate and Heptanoate. *E. cooperi* had five compounds identified: Euphorbilactone, Norsesquiterpenoid, Arachiside A, 16-Angeloyloxy-13 $\alpha$ -isobutanoyloxy-4 $\beta$ ,9 $\alpha$ ,20-trihydroxytiglia-1,5-diene-3,7-dione and 20-Acetoxy-16-angeloyloxy-13 $\alpha$ -isobutanoyloxy-4 $\beta$ ,9 $\alpha$ ,20-tetrahydroxytiglia-1,5-diene-3-one. Lastly, *E. tirucalli* had ten compounds identified: Euphorbol hexacosonate, 12-Deoxy-4 $\beta$ -hydroxyphorbol-13-phenyl acetate-20-acetate, 12, 20-Dideoxyphorbol-13-isobutyrate, tirucallicine, isoeuphorol, ketone euphorone, euphorbin A, euphorbin B, tirucallin A and tirucallin B.

### 4.3. In Silico Evaluation of Selected Phytochemicals

*Euphorbia* species are well reported for various biological activities and pharmacological potential indicating their potential as anticancer agents. Some extracts and isolates of the plants are well reported to show cytotoxic effects, as reviewed in this study. However, the phytochemical constituents that may account for the cytotoxic effects and other biological activities are largely unexplored. In silico techniques, which are valuable for resource maximization and search space minimization, may help to provide insightful information on the biological activities, pharmacokinetics, pharmacodynamics and pharmacotherapeutics of phytochemicals reported from *Euphorbia* species; and thereby serve as a fundamental resource for researchers to design more targeted experiments and manage future studies into the medicinal benefits of the unexplored and underutilized *Euphorbia* species.

#### 4.3.1. Cell Line Cytotoxicity Tendency of Selected Compounds

Promising *Euphorbia*-derived compounds as shown in Table 1.7 were evaluated in silico for cytotoxicity and anticancer tendency using the Cell Line Cytotoxicity Predictor (CLCPred) webserver [11]. Among 74 compounds subjected to prediction, 40 compounds were prioritized based on the availability of information and high cytotoxic activity. The compounds predicted to have high cytotoxic activity exhibited high anticancer potential activity when the probability cut-off was set as both P(a)ctive > P(i)nactive and Pa > 0.50. These compounds exhibit sufficient cytotoxic tendencies against multiple cancer cell lines and tumor types in human. Among these are the several *Euphorbia*

derived top compounds (EDCs) viz: Euphol, cycloartenol, lupeol,  $\alpha$ -amyrin, betulinic acid, 24-Methylene-cycloartanol, 12 Deoxyphorbol 13 tiglate 20 acetate, Ingol 7,8,12 acetate, diterpene glycoside, Tirucallol, Isobutyl angelate, Kaempferol-3-O-rutinoside exhibit high cytotoxic probability against 3 or more cell lines (Table 1.7). Considering the huge potential of plant-based natural products in drug discovery programs and the contribution of terpenoids in the production of anti-cancer compounds, terpenoids from *Euphorbia* species can be exploited for more reliable economical and environmentally safe bioactive molecules. Collectively, the terpenoids feature cytotoxicity tendencies against the various cell lines of stomach carcinoma, kidney carcinoma, skin melanoma, liver hepatoblastoma, lung carcinoma, brain glioma, colon adenocarcinoma and thyroid carcinoma (see Table 1.7).

**Table 1.7:** Compounds with high cytotoxic probability against 3 or more cancer cell lines.

Compounds	Pa	Pi	Cancer cell line	Cell line full name	Tissue	Tumor type
Euphol (C1)	0.591	0.002	MKN-7	Gastric carcinoma	Stomach	Carcinoma
	0.555	0.014	UO-31	Renal carcinoma	Kidney	Carcinoma
	0.554	0.014	SK-MEL-2	Melanoma	Skin	Melanoma
Cycloartenol (C2)	0.783	0.001	MKN-7	Gastric carcinoma	Stomach	Carcinoma
	0.577	0.015	HepG2	Hepatoblastoma	Liver	Hepatoblastoma
	0.523	0.029	DMS-114	Lung carcinoma	Lung	Carcinoma
	0.501	0.017	U-251	Glioma	Brain	Glioma
Lupeol (C3)	0.785	0.003	8505C	Thyroid-gland-undifferentiated carcinoma	Thyroid	Carcinoma

	0.62	0.001	FaDu	Hypopharyngeal squamous carcinoma	Upper aerodigestive tract	Carcinoma
	0.563	0.013	SK-MEL- 2	Melanoma	Skin	Melanoma
	0.539	0.004	DLD-1	Colon adenocarcinoma	Colon	Adenocarcinoma
	0.526	0.005	SW480	Colon adenocarcinoma	Colon	Adenocarcinoma
	0.52	0.005	PANC-1	Pancreatic carcinoma	Pancreas	Carcinoma
	0.632	0.004	8505C	Thyroid-gland- undifferentiated (anaplastic) carcinoma	Thyroid	Carcinoma
Alpha-amyrin (C4)	0.523	0.003	MKN-7	Gastric carcinoma	Stomach	Carcinoma
Betulinic acid (C5)	0.695	0.003	8505C	Thyroid-gland- undifferentiated (anaplastic) carcinoma	Thyroid	Carcinoma

	0.667	0.001	FaDu	Hypopharyngeal-squamous carcinoma	Upper aerodigestive tract	Carcinoma
	0.562	0.013	SK-MEL-2	Melanoma	Skin	Melanoma
24-Methylene-cycloartanol (C6)	0.555	0.003	MKN-7	Gastric carcinoma	Stomach	Carcinoma
	0.539	0.023	HL-60	Promyeloblast leukemia	Haematopoietic and lymphoid tissue	Leukemia
	0.52	0.031	DMS-114	Lung carcinoma	Lung	Carcinoma
	0.505	0.026	HL-60	Promyeloblast leukemia	Haematopoietic and lymphoid tissue	Leukemia
	0.638	0.033	A549	Lung carcinoma	Lung	Carcinoma
12-Deoxyphorbol-13-tiglate-20-acetate (C7)	0.568	0.024	NCI-H838	Non-small cell lung cancer. 3 stage	Lung	Carcinoma
	0.556	0.021	HL-60	Promyeloblast leukemia	Haematopoietic and lymphoid tissue	Leukemia

Ingol-7,8,12- acetate,ditiglate (C8)	0.663	0.012	NCI-H838	Non-small cell lung cancer. 3 stage	Lung	Carcinoma
	0.592	0.005	SK-MEL-1	Metastatic melanoma	Skin	Melanoma
	0.513	0.033	DMS-114	Lung carcinoma	Lung	Carcinoma
Diterpene glycoside (C9)	0.682	0.008	SK-MEL-2	Melanoma	Skin	Melanoma
	0.614	0.015	HL-60	Promyeloblast leukemia	Haematopoietic and lymphoid tissue	Leukemia
	0.534	0.034	NCI-H838	Non-small cell lung cancer. 3 stage	Lung	Carcinoma
Tirucallol (C10)	0.591	0.002	MKN-7	Gastric carcinoma	Stomach	Carcinoma
	0.555	0.014	UO-31	Renal carcinoma	Kidney	Carcinoma
	0.554	0.014	SK-MEL-2	Melanoma	Skin	Melanoma
Isobutyl angelate (C11)	0.616	0.012	HepG2	Hepatoblastoma	Liver	Hepatoblastoma
	0.545	0.03	NCI-H838	Non-small cell lung cancer. 3 stage	Lung	Carcinoma

	0.544	0.052	A549	Lung carcinoma	Lung	Carcinoma
	0.546	0.004	Caco-2	Colon adenocarcinoma	Colon	Adenocarcinoma
	0.548	0.016	SK-MEL-1	Metastatic melanoma	Skin	Melanoma
	0.55	0.022	HL-60	Promyeloblast leukemia	Haematopoietic and lymphoid tissue	Leukemia
Kaempferol-3-O-rutinoside (C12)	0.542	0.031	NCI-H838	Non-small cell lung cancer. 3 stage	Lung	Carcinoma

\* Pa - Probability to be Active, Pi - Probability to be Inactive.

#### 4.3.2. Physicochemical and Drug-Like Properties

The physicochemical parameters and drug-likeness of the EDCs were computed based on their chemical structures using the SwissADME platform as presented in Table 1.8 (physicochemical properties; drug-like properties). Table 1.7 and Table 1.8 show that most of the compounds show good and desirable physicochemical properties. All the EDCs except C8, C9 and C12 have molecular weight less than 500. Other desirable physicochemical properties exhibited by several compounds are lipophilicity (LogP), solubility, Polar Surface Area (PSA), Hydrogen Bond Donors and Acceptors. Moderate molecular weight is preferred for drug-likeness. Extremely large or small molecules might have issues with absorption, distribution, metabolism and excretion (ADME) properties. Balanced lipophilicity is desirable. Too hydrophilic or too hydrophobic molecules might face issues with absorption or distribution in the body. Good solubility in physiological fluids is essential for a drug candidate to be effective. Poorly soluble compounds might have bioavailability issues. Smaller polar surface area often correlates with better oral absorption. Fewer hydrogen bond donors and acceptors often indicate better drug-like properties.

Drug-likeness refers to the set of characteristics that make a molecule more likely to become an effective and safe drug candidate. Understanding physicochemical properties helps in predicting a molecule's behavior in biological systems and its potential as a drug. Many of these properties are essential for drug development and violation of any of these properties may result in delay or failure of the drug candidate to reach market operation. All the compounds except C9 and C12 exhibit desirable Lipinski and Veber features. They also feature Synthetic accessibility. The Lipinski's Rule-of-Five (RO5) is a guideline used to assess drug-likeness based on physicochemical properties. It suggests that for a compound to have good oral bioavailability, it should have: Molecular weight <500 Daltons, LogP <5, Hydrogen bond donors <5, Hydrogen bond acceptors <10 [330,331]. While adherence to the RO5 is not a strict requirement for a compound to become a drug, it serves as a



<b>Drug-likeness</b>						
Lipinski	Yes	Yes	Yes	Yes	Yes	Yes
Bioavailability	0.55	0.55	0.55	0.55	0.85	0.55
PAINS	0 alert					
Brenk	1 alert					
Synthetic accessibility	Yes	Yes	Yes	Yes	Yes	Yes
<b>Absorption</b>						
Caco2 permeability(log Papp in 10 <sup>-6</sup> cm/s)	1.203	1.194	1.226	1.227-J	1.175	1.221
Human intestinal absorption (% Absorbed)	93.119	95.248	95.782	94.062	99.763	95.309
P-glycoprotein SUBstrate	Neg.	Neg.	Neg.	Neg.	Neg.	Neg.
P-glycoprotein Inhibitor	Pos.	Pos.	Pos.	Pos.	Neg.	Neg.
P-glycoprotein II inhibitor	Pos.	Pos.	Pos.	Pos.	Neg.	Pos.
<b>Distribution</b>						
VD <sub>ss</sub> (human) (log L/kg)	0.661	-0.075	0	0.266	-1.18	-0.072

BBB permeability (Log BB)	0.683	0.794	0.726	0.674,	-0.322	0.845
CNS permeability (Log PS)	-2.254	-1.714	-1.714	-1.773	-1.343	-1.462
<b>Metabolism</b>						
CYP2D6 substrate	Neg.	Neg.	Neg.	Neg.	Neg.	Neg.
CYP3A4 substrate	Pos.	Pos.	Pos.	Pos.	Pos.	Pos.
CYP1A2 inhibition	Neg.	Neg.	Neg.	Neg.	Neg.	Neg.
CYP2C19 inhibitor	Neg.	Neg.	Neg.	Neg.	Neg.	Neg.
CYP2C9 inhibitor	Neg.	Neg.	Neg.	Neg.	Neg.	Neg.
CYP2D6 inhibitor	Neg.	Neg.	Neg.	Neg.	Neg.	Neg.
CYP3A4 inhibitor	Neg.	Neg.	Neg.	Neg.	Neg.	Neg.
<b>Excretion</b>						
Total Clearance (log ml/min/kg)	0.403	0.262	0.153	0.119	0.116	0.255
Renal OCT2 substrate	Neg.	Neg.	Neg.	Neg.	Neg.	Neg.
<b>Toxicity</b>						
Max. tolerated dose (human) (log mg/kg/day)	-0.568	-0.46	0.502	-0.571	0.144	0.303

hERG Inhibitor	Neg.	Neg.	Neg.	Neg.	Neg.	Neg.
Oral Rat Acute Toxicity (LOSO)(mol/ka)	1.906	2.627	2.563	2.4	2.256	2.542
Oral Rat Chronic Toxicity (LOAEL)(log mg/ka bw/day)	0.788	0.806	0.8	0.856	2.206	0.802
Hepatotoxicity	Neg.	Neg.	Neg.	Neg.	Pos.	Neg.

[Neg. – Negative; Pos. – Positive].

The predictions regarding the intestinal absorption of Euphorbia-derived terpenoids in humans, based on computed molecular descriptors, indicate that all the compounds exhibit good human intestinal absorption (%Absorbed). Additionally, most of these compounds are predicted as P-glycoprotein inhibitors (PGIs). PGIs interfere with glycoprotein activity, preventing the pumping of drug compounds out of cells. This inhibition can lead to the maintenance of high intracellular drug concentration, potentially improving therapeutic efficacy. Notably, several FDA-approved drugs and natural compounds are recognized as PGIs [332].

Human volume of distribution (VD<sub>ss</sub>) is a pivotal parameter for determining the human dose. While some Euphorbia-derived compounds feature low VD<sub>ss</sub>, the majority exhibit a low plasma binding tendency. A drug with strong plasma protein binding may struggle to dissociate from plasma proteins and bind to target receptors, even if present in the bloodstream. Thus, a candidate with very high plasma levels due to very low VD<sub>ss</sub> may appear promising, but it is essential to examine the magnitude of the pharmacological effect to avoid potential misinterpretations [333].

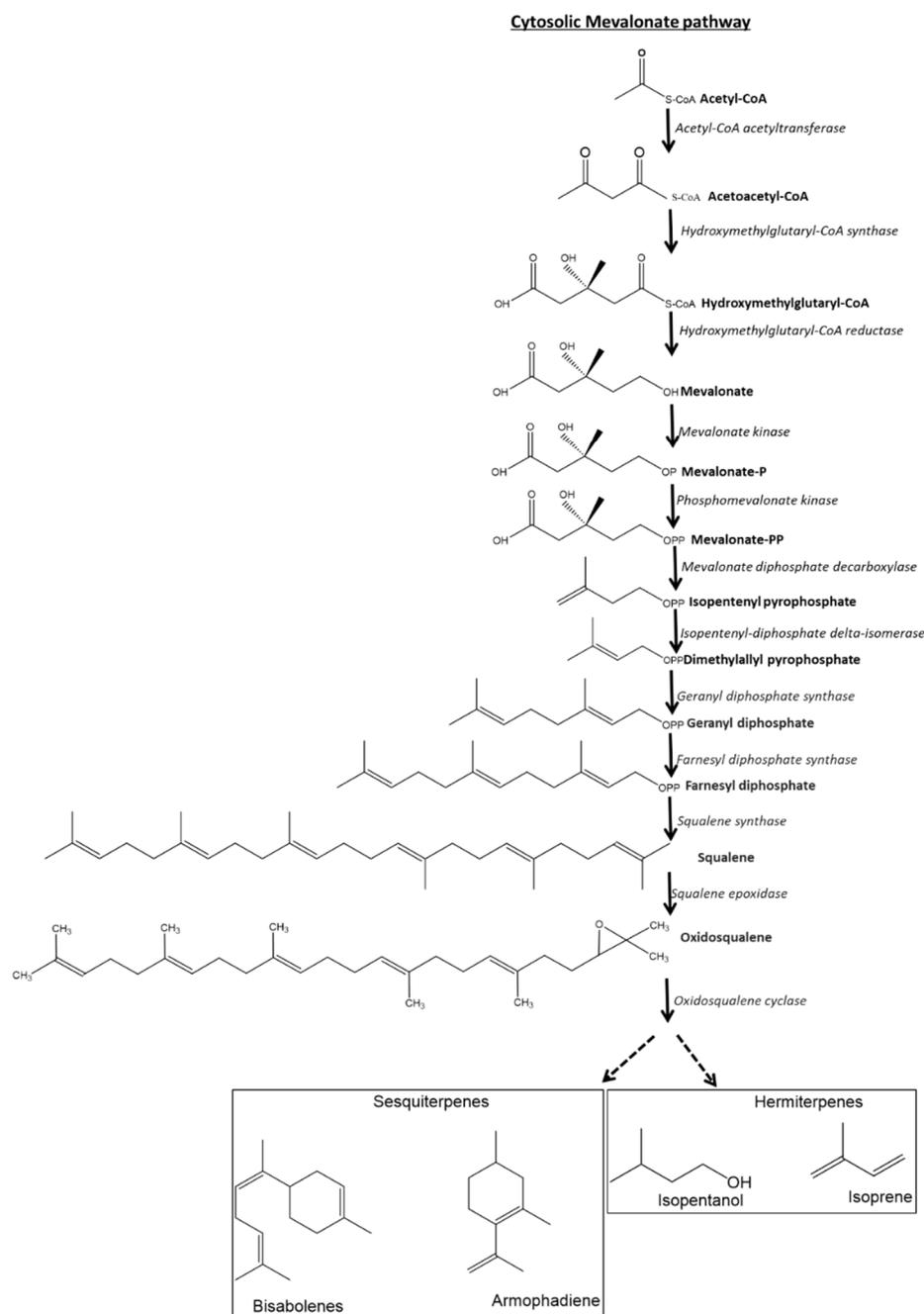
The computed interactions of the compounds with liver phase I drug metabolism, using various cytochrome P450 descriptors, reveal that most of the top compounds do not exhibit inhibitory tendencies against various cytochrome P450s. This suggests that these compounds may not adversely affect phase I drug metabolism in the liver.

Organic cation transporter 2 (OCT2) mediates the initial step in the renal secretion of organic cations. While several compounds exhibit a high clearance rate tendency, all are predicted to be non-substrates of OCT2.

Computed molecular descriptors assessing the toxicity tendency of the compounds indicate that the majority may not be toxic, as indicated by AMES toxicity and maximum dose (human) predictions. hERG I, which blocks the hERG channel, may cause cardiotoxicity and oral rat acute toxicity. However, none of the six phytocompounds in this study exhibit the potential to be hERG channel blockers, suggesting that they may not cause hERG channel-related cardiotoxicity [330,334].

#### 4.3.4. Biosynthesis of Terpenoid Class

The Euphorbia species contains a diverse array of compounds, including terpenoids, steroids, fatty acids and phenolic compounds. Each compound has its own specific biosynthetic pathway. However, the current study will only focus on the biosynthesis pathway of terpenoid, as it is the main class of compounds isolated from Euphorbia species. A schematic diagram of the biosynthesis is presented in Figure 1.16.



**Figure 1.16.** Terpene biosynthesis in the cytosol through the mevalonate pathway. Starting with acetyl-CoA, this process produces terpene precursors (isopentenyl pyrophosphate and dimethylallyl pyrophosphate), serving as building blocks for various terpenes with diverse biological functions. The diagram was adopted from Qiao et al. (2018).

## 5. Conclusions and Future Perspective

This review focuses on the medicinal properties of 15 species of *Euphorbia*, particularly their potential as alternative treatments for cancer. These plants contain unique compounds such as triterpenoids, tannins, diterpene esters and sterols. Several studies have reported that the *Euphorbiaceae* family contains phytochemicals that lead to the isolation of various classes of triterpenoids. Some of these triterpenoids include euphol (found in *E. trigona*, *E. tirucalli*, *E. enopla*, *E. coerulescens*, *E. ammak*, *E. cooperi*), cycloartenol, lupeol, amyrins, taraxerols, friedelin and tirucallos (found in *E. trigona*). Tannins classes include ellagic acid, 1-O-Galloyl-3,6-hexahydroxydiphenyl- $\beta$ -D-, tri methyl ellagic acid and unknown structures Tirucallin A and Tirucallin B. Sterols classes such as  $\beta$ -sitosterol, Stigmasterol and Obtusifoliol are also present. Diterpene classes such as diterpene esters, 12-Deoxy phorbol, phorbol, ingenol, ingenol triacetate, 16-Hydroxy-12-desoxyphorbol, 12-

Deoxyphorbol-13-isobutyrate-16-angelate-20-acetate are also found. These compounds have shown cytotoxic effects through various mechanisms of action, including cell proliferation and differentiation, and also apoptosis. Out of all the *Euphorbia* species studied, *E. trigona* had the most anti-cancer isolated compounds with a total of 14. *E. tirucalli* followed closely behind with 13, while *E. cooperi* had 8 and the others had fewer or none at all. While some studies have reported promising results, others have found no significant inhibitory activity against certain cell lines. The insilico results further revealed that several *Euphorbia*-derived top compounds show good cytotoxic potential against multiple cancer cell lines indicating anticancer tendency against stomach carcinoma, kidney carcinoma, skin melanoma, liver hepatoblastoma, lung carcinoma, brain glioma, colon adenocarcinoma and thyroid carcinoma, etc. Most of these compounds are drug-like as indicated by Lipinski screening and Veber parameters which were derived from good physicochemical properties. Most of the EDCs were predicted to possess good pharmacokinetic tendencies as indicated by the ADMET properties.

Therefore, building upon the diverse compounds identified, such as triterpenoids, tannins, diterpene esters and sterols, requires further in-depth pharmacological studies to unravel the precise mechanisms of action and potential synergies among these compounds. Subsequent clinical trials would be pivotal in assessing the safety and efficacy of these compounds in human subjects. Additionally, exploring combination therapies with existing cancer treatments, investigating the bioavailability of isolated compounds and identifying new, yet-to-be-evaluated compounds like Euphorbin A, Euphorbin B and Tirucallin A are crucial areas of focus. Development of optimized formulations, understanding the factors contributing to variability in study results and fostering collaboration between diverse research fields are imperative for advancing the potential of *Euphorbia* species in cancer therapeutics. Furthermore, public awareness initiatives and the integration of ethnobotanical knowledge can contribute to a holistic understanding of the historical and contemporary significance of these plants in medicinal contexts. This comprehensive and multidisciplinary approach underscores the need for ongoing research to harness the full therapeutic potential of *Euphorbia* species for cancer treatment. Overall, this report aims to provide scientific credibility to the traditional use of *Euphorbia* species for medicinal purposes.

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## References

1. Bijekar, S. R.; Gayatri, M. C. Ethanomedicinal Properties of Euphorbiaceae Family—A Comprehensive Review. *Int. J. Phytomedicine* 2014, 6 (2), 144. <http://www.arjournals.org/index.php/ijpm/article/view/1280>
2. Aleksandrov, M.; Maksimova, V.; Koleva Gudeva, L. Review of the anticancer and cytotoxic activity of some species from genus *Euphorbia*. *Agric. Consp. Sci.* 2019, 84(1), 1-5. <https://doi.org/10.1016/j.ejps.2006.04.006>
3. Adedapo, A. A.; Saba, A. B.; Dina, O. A.; Oladejo, G. M. A. Effects of dexamethasone on the infectivity of *Trypanosoma vivax* Y486 and the haematological changes in Nigerian domestic chickens (*Gallus gallus domesticus*). *Vet. Arhiv* 2004, 74(5), 371-381. <https://hrcak.srce.hr/68647>
4. Villanueva, J.; Quirós, L. M.; Castañón, S. Purification and Partial Characterization of a Ribosome-Inactivating Protein from the Latex of *Euphorbia Trigona* Miller with Cytotoxic Activity toward Human Cancer Cell Lines. *Phytomedicine* 2015, 22 (7-8), 689-695. <https://doi.org/10.1016/j.phymed.2015.04.006>
5. Betancur-Galvis, L. A.; Morales, G. E.; Forero, J. E.; Roldan, J. Cytotoxic and antiviral activities of Colombian medicinal plant extracts of the *Euphorbia* genus. *Mem. Inst. Oswaldo Cruz* 2002, 97, 541-546. <https://doi.org/10.1590/S0074-02762002000400017>
6. Mwine, T. J.; Damme, V. P. Why Do Euphorbiaceae Tick as Medicinal Plants? A Review of Euphorbiaceae Family and Its Medicinal Features. 2011. <https://doi.org/10.5897/JMPR.9001294>

7. Fürstenberger, G.; Hecker, E. On the Active Principles of the Spurge Family (Euphorbiaceae) XI. The Skin Irritant and Tumor Promoting Diterpene Esters of *Euphorbia tirucalli* L. Originating from South Africa. *Z. Naturforsch. C* 1985, 40(9-10), 631-646. <https://doi.org/10.1515/znc-1985-9-1008>
8. Cataluna, P.; Rates, S. M. K. The traditional use of the latex from *Euphorbia tirucalli* Linnaeus (Euphorbiaceae) in the treatment of cancer in South Brazil. In II WOCMAP congress medicinal and aromatic plants, part 2: pharmacognosy, pharmacology, phytomedicine, toxicology 1997, 501, 289-296. <https://doi.org/10.17660/ActaHortic.1999.501.46>
9. Mavundza, E. J.; Street, R.; Baijnath, H. A Review of the Ethnomedicinal, Pharmacology, Cytotoxicity, and Phytochemistry of the Genus *Euphorbia* in Southern Africa. *S. Afr. J. Bot.* 2022, 144, 403-418. <https://doi.org/10.1016/j.sajb.2021.08.029>
10. Lu, L.; Zhang, J.; Xie, Y.; Gao, F.; Xu, S.; Wu, X.; et al. Wearable health devices in health care: narrative systematic review. *J. Med. Internet Res.* 2020, 8(11), e18907. <https://doi.org/10.2196/18907>
11. Lagunin, A. A.; Dubovskaja, V. I.; Rudik, A. V.; Pogodin, P. V.; Druzhilovskiy, D. S.; Glorizova, T. A.; Poroikov, V. V. CLC-Pred: A Freely Available Web-Service for In Silico Prediction of Human Cell Line Cytotoxicity for Drug-Like Compounds. *PLOS ONE* 2018, 13 (1), e0191838. <https://doi.org/10.1371/journal.pone.0191838>
12. Daina, A.; Michielin, O.; Zoete, V. SwissADME: A Free Web Tool to Evaluate Pharmacokinetics, Drug-Likeness, and Medicinal Chemistry Friendliness of Small Molecules. *Sci. Rep.* 2017, 7 (1), 42717. <https://doi.org/10.1038/srep42717>
13. Pires, D. E. V.; Blundell, T. L.; Ascher, D. B. pkCSM: Predicting Small-Molecule Pharmacokinetic and Toxicity Properties Using Graph-Based Signatures. *J. Med. Chem.* 2015, 58 (9), 4066-4072. DOI:10.1021/acs.jmedchem.5b00104.
14. Adetunji, J. A.; Ogunyemi, O. M.; Gyebi, G. A.; Adewumi, A. E.; Olaiya, C. O. Atomistic Simulations Suggest Dietary Flavonoids from Beta Vulgaris (Beet) as Promising Inhibitors of Human Angiotensin-Converting Enzyme and 2-Alpha-Adrenergic Receptors in Hypertension. *Bioinform. Adv.* 2023, 3 (1). <https://doi.org/10.1093/bioadv/vbad133>
15. Ogunyemi, O. M.; Gyebi, G. A.; Ibrahim, I. M.; Esan, A. M.; Olaiya, C. O.; Soliman, M. M.; Batiha, G. E.-S. Identification of Promising Multi-Targeting Inhibitors of Obesity from *Vernonia Amygdalina* through Computational Analysis. *Mol. Divers.* 2022. <https://doi.org/10.1007/s11030-022-10397-6>
16. Tada, M.; Seki, H. Toxic Diterpenes from *Euphorbia trigona* (Saiunkaku: An Indoor Foliage Plant in Japan). *Agric. Biol. Chem.* 1989, 53 (2), 425-430. <https://doi.org/10.1080/00021369.1989.10869285>
17. Nashikkar, N.; Begde, D.; Bundale, S.; Mashitha, P.; Rudra, J.; Upadhyay, A. Evaluation of the Immunomodulatory Properties of *Euphorbia trigona*—An In Vitro Study. *Int. J. Inst. Pharm. Life Sci.* 2012, 2, 88-105.
18. Marathe, K.; Nashikkar, N.; Bundale, S.; Upadhyay, A. Analysis of Quorum Quenching Potential of *Euphorbia trigona* Mill. *Int. J. Pharm. Sci. Res.* 2019, 10 (3), 1372-1386. [http://dx.doi.org/10.13040/IJPSR.0975-8232.10\(3\).1372-86](http://dx.doi.org/10.13040/IJPSR.0975-8232.10(3).1372-86)
19. Siddique, N. A.; da Silva, J. A. T.; Bari, M. A. Preservation of Indigenous Knowledge Regarding Important and Endangered Medicinal Plants in Rajshahi District of Bangladesh. *J. Plant Sci.* 2010, 5 (2), 201-215. <https://www.cabdirect.org/cabdirect/abstract/20113194288>
20. Bouquet, A. J. Natural products as an alternative remedy. 24th ed. Royal Botanic Garden Kew 1969, 166 - 179. [https://www.researchgate.net/publication/242213547\\_Acute\\_effect\\_of\\_administration\\_of\\_ethanol\\_extract\\_s\\_of\\_Ficus\\_exasperata\\_vahl\\_on\\_kidney\\_function\\_in\\_albino\\_rats](https://www.researchgate.net/publication/242213547_Acute_effect_of_administration_of_ethanol_extract_s_of_Ficus_exasperata_vahl_on_kidney_function_in_albino_rats)
21. Lynn, K. R.; Clevette-Radford, N. A. Four serine proteases from the latex of *Euphorbia tirucalli*. *Can. J. Biochem. Cell Biol.* 1985, 63(10), 1093-1096. [doi.org/10.1139/o85-136](https://doi.org/10.1139/o85-136)
22. Luseba, D.; Van der Merwe, D. Ethnoveterinary medicine practices among Tsonga speaking people of South Africa. *Onderstepoort J. Vet. Res.* 2006, 73(2), 115-122. <https://hdl.handle.net/10520/EJC86248>
23. Gildenhuis, S. The three most abundant tree *Euphorbia* species of the Transvaal (South Africa). *Euphorbia World* 2006, 2(1), 9 -14. [https://www.Euphorbia international.org/journal/pdf\\_files/EW2-1-sample.pdf](https://www.Euphorbia international.org/journal/pdf_files/EW2-1-sample.pdf)
24. El-Sherei, M. M.; Islam, W. T.; El-Dine, R. S.; El-Toumy, S. A.; Ahmed, S. R. Phytochemical investigation of the cytotoxic latex of *Euphorbia cooperi* NE Br. *Aust. J. Basic Appl. Sci.* 2015, 9(11), 488-493. <http://ajbasweb.com/old/ajbas/2015/May/488-493.pdf>
25. Hedberg, I.; Staugård, F. *Traditional Medicinal Plants*; Ipelegeng Publishers: 1989; Vol. 3. <https://www.sciencedirect.com/science/article/pii/S0378874108004637>
26. Gundidza, M.; Sorg, B.; Hecker, E. A skin irritant phorbol ester from *Euphorbia cooperi* NE Br. *Cent. Afr. J. Med.* 1992, 38(12), 444-447. [https://hdl.handle.net/10520/AJA00089176\\_156](https://hdl.handle.net/10520/AJA00089176_156)
27. Engi, H.; Vasas, A.; Redei, D.; Molnár, J.; Hohmann, J. New MDR Modulators and Apoptosis Inducers from *Euphorbia* Species. *Anticancer Res.* 2007, 27 (5A), 3451-3458. <https://ar.iijournals.org/content/anticancer/27/5A/3451.full.pdf>

28. Morsi, S. R. A. Phytochemical and Biological Study of Certain Euphorbia Species Cultivated in Egypt. Doctoral Dissertation, Cairo University, 2015. <http://erepository.cu.edu.eg/index.php/cuttheses/article/view/5355>
29. Carter, S. A preliminary classification of Euphorbia subgenus Euphorbia. *Ann. Mo. Bot. Gard.* 1994, 368-379. doi.org/10.2307/2992103
30. Almehdar, H.; Abdallah, H. M.; Osman, A. M. M.; Abdel-Sattar, E. A. In vitro cytotoxic screening of selected Saudi medicinal plants. *J. Nat. Med.* 2012, 66, 406-412. doi.org/10.1007/s11418-011-0589-8
31. Al-Hajj, M. M. A.; Al-Shamahy, H. A.; Alkhatib, B. Y.; Moharram, B. A. In vitro anti-leishmanial activity against cutaneous Leishmania parasites and preliminary phytochemical analysis of four Yemeni medicinal plants. *Univ. J. Pharm. Res.* 2018, 3, 48-54. <http://ujpr.org>
32. Kiyohara, H.; Ichino, C.; Kawamura, Y.; Nagai, T.; Sato, N.; Yamada, H.; et al. In vitro anti-influenza virus activity of a cardiotoxic glycoside from *Adenium obesum* (Forssk.). *Phytomedicine* 2012, 19(2), 111-114. doi.org/10.1016/j.phymed.2011.07.004
33. Abdel-Sattar, E.; Maes, L.; Salama, M. M. In Vitro Activities of Plant Extracts from Saudi Arabia Against Malaria, Leishmaniasis, Sleeping Sickness, and Chagas Disease. *Phytother. Res.* 2010, 24, 1322-1328. <https://doi.org/10.1002/ptr.3071>
34. Watt, J. M.; Breyer-Brandwijk, M. G. The Medicinal and Poisonous Plants of Southern and Eastern Africa Being an Account of Their Medicinal and Other Uses, Chemical Composition, Pharmacological Effects and Toxicology in Man and Animal; Edn. 2, 1962. <https://www.cabdirect.org/cabdirect/abstract/19622704780>
35. Baslas, R. K.; Gupta, N. C. Chemical constituents of the bark of *Euphorbia tirucalli*. *Indian J. Pharm. Sci.* 1982. [https://scholar.google.com/scholar\\_lookup?title=Chemical%20constituents%20of%20the%20bark%20of%20Euphorbia%20tirucalli&publication\\_year=1982&author=R.K.%20Baslas&author=N.C.%20Gupta](https://scholar.google.com/scholar_lookup?title=Chemical%20constituents%20of%20the%20bark%20of%20Euphorbia%20tirucalli&publication_year=1982&author=R.K.%20Baslas&author=N.C.%20Gupta)
36. Kajikawa, M.; Yamato, K. T.; Fukuzawa, H.; Sakai, Y.; Uchida, H.; Ohyama, K. Cloning and characterization of a cDNA encoding  $\beta$ -amyrin synthase from petroleum plant *Euphorbia tirucalli* L. *Phytochemistry* 2005, 66(15), 1759-1766. doi.org/10.1016/j.phytochem.2005.05.021
37. Shivkumar, S. P. Ethnopharmacological Validation of Medicinal Plants Treating Skin Diseases in Hyderabad Karnataka Region. <http://hdl.handle.net/10603/37479>, 2015.
38. Hargreaves, B. J. The spurges of Botswana. *Botswana Notes Rec.* 1991, 23(1), 115-130. [https://www.jstor.org/stable/40980848?seq=1#metadata\\_info\\_tab\\_contents](https://www.jstor.org/stable/40980848?seq=1#metadata_info_tab_contents)
39. Van Damme, P. Studie van *Euphorbia Tirucalli* L.: Morfologie, Fysiologie, Teeltvoorwaarden; Doctoral Dissertation, Ghent University, 1989. <http://hdl.handle.net/1854/LU-8551967>
40. Gupta, N.; Vishnoi, G.; Wal, A.; Wal, P. Medicinal value of *Euphorbia tirucalli*. *Syst. Rev. Pharm.* 2013, 4(1), 40. doi: 10.4103/0975-8453.135843/ [www.sysrevpharm.org](http://www.sysrevpharm.org)
41. Voigt, W. E. *Euphorbia Tirucalli* L. (Euphorbiaceae). [Online] Available: <http://pza.sanbi.org/Euphorbia-tirucalli>. Accessed March 9, 2023.
42. MacNeil, A.; Sumba, O. P.; Lutzke, M. L.; Moormann, A.; Rochford, R. Activation of the Epstein-Barr virus lytic cycle by the latex of the plant *Euphorbia tirucalli*. *Br. J. Cancer* 2003, 88(10), 1566-1569. doi.org/10.1038/sj.bjc.6600929
43. Van den Bosch, C.; Griffin, B. E.; Kazembe, P.; Dziweni, C.; Kadzamira, L. Are Plant Factors a Missing Link in the Evolution of Endemic Burkitt's Lymphoma? *Br. J. Cancer* 1993, 68 (6), 1232-1235. doi.org/10.1038/bjc.1993.510
44. Kajikawa, M.; Yamato, K. T.; Fukuzawa, H.; Sakai, Y.; Uchida, H.; Ohyama, K. Cloning and characterization of a cDNA encoding  $\beta$ -amyrin synthase from petroleum plant *Euphorbia tirucalli* L. *Phytochemistry* 2005, 66(15), 1759-1766. doi.org/10.1016/j.phytochem.2005.05.021
45. Lirio, L. G.; Hermano, M. L.; Fontanilla, M. Q. Antibacterial activity of medicinal plants from the Philippines. *Pharm. Biol.* 1998, 36(5), 357-359. doi.org/10.1076/phbi.36.5.357.4656
46. Tiwari, S.; Singh, P.; Singh, A. Toxicity of *Euphorbia Tirucalli* Plant against Freshwater Target and Non-Target Organisms. *Pak. J. Biol. Sci.* 2003. doi.org/10.3923/pjbs.2003.1423.1429
47. Rezende, E. L.; Bozinovic, F.; Garland, T., Jr. Climatic Adaptation and the Evolution of Basal and Maximum Rates of Metabolism in Rodents. *Evolution* 2004, 58 (6), 1361-1374. doi.org/10.1111/j.0014-3820.2004.tb01714.x
48. Parekh, J.; Chanda, S. In Vitro Antimicrobial Activity and Phytochemical Analysis of Some Indian Medicinal Plants. *Turk. J. Biol.* 2007, 31 (1), 53-58. <https://journals.tubitak.gov.tr/biology/issues/biy-07-31-1/biy-31-1-9-0610-4.pdf>
49. Altamimi, M.; Jaradat, N.; Alham, S.; Al-Masri, M.; Bsharat, A.; Alsaleh, R.; Sabobeh, R. Antioxidant, anti-enzymatic, antimicrobial and cytotoxic properties of *Euphorbia tirucalli* L. *bioRxiv* 2019, 2019-12. doi.org/10.1101/2019.12.17.879692
50. Valadares, M. C.; Carrucha, S. G.; Accorsi, W.; Queiroz, M. L. *Euphorbia Tirucalli* L. Modulates Myelopoiesis and Enhances the Resistance of Tumor-Bearing Mice. *Int. Immunopharmacol.* 2006, 6 (2), 294-299. doi.org/10.1016/j.intimp.2005.07.013

51. Silva, V. A.; Rosa, M. N.; Miranda-Gonçalves, V.; Costa, A. M.; Tansini, A.; Evangelista, A. F., et al. Euphol, a Tetracyclic Triterpene, from *Euphorbia tirucalli* Induces Autophagy and Sensitizes Temozolomide Cytotoxicity on Glioblastoma Cells. *Invest. New Drugs* 2019, 37, 223–237. doi.org/10.1007/s10637-018-0620-y
52. De Souza, L. S.; Puziol, L. C.; Tosta, C. L.; Bittencourt, M. L.; Santa Ardisson, J.; et al. Analytical methods to access the chemical composition of an *Euphorbia tirucalli* anticancer latex from traditional Brazilian medicine. *J. Ethnopharmacol.* 2019, 237, 255-265. doi.org/10.1016/j.jep.2019.03.041
53. Moteetee, A.; Moffett, R. O.; Seleteng-Kose, L. A Review of the Ethnobotany of the Basotho of Lesotho and the Free State Province of South Africa (South Sotho). *S. Afr. J. Bot.* 2019, 122, 21–56. doi.org/10.1016/j.sajb.2017.12.012
54. Shale, T. L.; Stirk, W. A.; van Staden, J. Screening of Medicinal Plants Used in Lesotho for Antibacterial and Anti-inflammatory Activity. *J. Ethnopharmacol.* 1999, 67 (3), 347–354. doi.org/10.1016/S0378-8741(99)00035-5
55. Maliehe, E. B. Medicinal plants and herbs of Lesotho. Mafeteng Development Project 1997. doi.org/10.1016/j.apjtb.2017.06.002
56. Kose, L. S.; Moteetee, A.; Van Vuuren, S. Ethnobotanical survey of medicinal plants used in the Maseru district of Lesotho. *J. Ethnopharmacol.* 2015, 170, 184-200. doi.org/10.1016/j.jep.2015.04.047
57. Cock, I. E.; Ndlovu, N.; Van Vuuren, S. F. The use of South African botanical species for the control of blood sugar. *J. Ethnopharmacol.* 2021, 264, 113234. doi.org/10.1016/j.jep.2020.113234
58. Johnson, T. CRC ethnobotany desk reference. CRC Press 2019. <https://books.google.co.za/books?hl=en&lr=&id=uTS0DwAAQBAJ&oi=fnd&pg=PA2&dq=Johnson>
59. Komoreng, L.; Thekiso, O.; Lehasa, S.; Tiwani, T.; Mzizi, N.; Mokoena, N.; et al. An ethnobotanical survey of traditional medicinal plants used against lymphatic filariasis in South Africa. *S. Afr. J. Bot.* 2017, 111, 12-16. doi.org/10.1016/j.sajb.2017.03.005
60. Tiwani, T. Phytochemical Screening, Cytotoxicity, Antimicrobial and Anthelmintic Activity of Medical Plants Used in the Treatment of Lymphatic Filariasis in the Eastern Cape, South Africa; Doctoral Dissertation, University of the Free State, 2017. <http://hdl.handle.net/11660/10002>
61. Afolayan, A. J.; Grierson, D. S.; Mbeng, W. O. Ethnobotanical survey of medicinal plants used in the management of skin disorders among the Xhosa communities of the Amathole District, Eastern Cape, South Africa. *J. Ethnopharmacol.* 2014, 153(1), 220-232. doi.org/10.1016/j.jep.2014.02.023
62. Van Wyk, B. E.; Oudtshoorn, B. V.; Gericke, N. Medicinal Plants of South Africa; Briza, 1997. <https://www.cabdirect.org/cabdirect/abstract/20006782435>
63. Iwalewa, E. O.; McGaw, L. J.; Naidoo, V.; Eloff, J. N. Inflammation: The Foundation of Diseases and Disorders. A Review of Phytomedicines of South African Origin Used to Treat Pain and Inflammatory Conditions. *Afr. J. Biotechnol.* 2007, 6 (25), 2868–2885. DOI: 10.5897/AJB2007.000-2457
64. Assefa, A.; Bahiru, A. Ethnoveterinary botanical survey of medicinal plants in Abergelle, Sekota and Lalibela districts of Amhara region, Northern Ethiopia. *J. Ethnopharmacol.* 2018, 213, 340-349. doi.org/10.1016/j.jep.2017.11.024
65. Mampa, S. T. M.; Mashele, S. S.; Sekhoacha, M. P. Applications of Chromatographic Techniques for Fingerprinting of Toxic and Non-toxic *Euphorbia* Species. *Pak. J. Biol. Sci.* 2020, 23(4), 552-560. doi.org/10.3923/pjbs.2020.552.560
66. Evans, F. J. The irritant toxins of Blue *Euphorbia* (*Euphorbia coerulescens* Haw.). *Toxicon* 1978, 16(1), 51-57. doi.org/10.1016/0041-0101(78)90060-0
67. Al-Harbi, N. A. Diversity of medicinal plants used in the treatment of skin diseases in Tabuk region, Saudi Arabia. *J. Med. Plants Res.* 2017, 11(35), 549-555. doi.org/10.5897/JMPR2017.6438
68. El-Shanwani, M. A. A. Al-nibat al-mustakhdima fi al-tibb al-sha'abi al-Saudi [Plants used in Saudi folk medicine]. General Directorate of Research Grants Program, KACST, Riyadh 1996. [scholar.google.com/scholar\\_lookup?title=Plants+Used+in+Saudi+Folk+Medicine&author=Al-Shanwani,+M.&](https://scholar.google.com/scholar_lookup?title=Plants+Used+in+Saudi+Folk+Medicine&author=Al-Shanwani,+M.&)
69. Abdel-Fattah, M. R. The Chemical Constituents and Economic Plants of the Euphorbiaceae. *Bot. J. Linn. Soc.* 1987, 94, 293–326. doi.org/10.1111/j.1095-8339.1987.tb01858.x
70. Anjaneyulu, V.; Rao, G. S.; Connolly, J. D. Occurrence of 24-epimers of cycloart-25-ene-3 $\beta$ , 24-diols in the stems of *Euphorbia trigona*. *Phytochemistry* 1985, 24(7), 1610-1612. DOI: 10.1016/0031-9422(85)83599-2
71. Nielsen, P. E.; Nishimura, H.; Otvos, J. W.; Calvin, M. Plant Crops as a Source of Fuel and Hydrocarbon-Like Materials. *Science* 1977, 198(4320), 942–944. DOI: 10.1126/science.198.4320.942.
72. Popplewell, W. L.; Marais, E. A.; Brand, L.; Harvey, B. H.; Davies-Coleman, M. T. Euphorbias of South Africa: Two New Phorbol Esters from *Euphorbia bothae*. *S. Afr. J. Chem.* 2010, 63, 175–179. [http://www.scielo.org.za/scielo.php?script=sci\\_arttext&pid=S0379-43502010000100027&lng=en&tlng=en](http://www.scielo.org.za/scielo.php?script=sci_arttext&pid=S0379-43502010000100027&lng=en&tlng=en).

73. Rédei, D.; Forgo, P.; Hohmann, J. New Tiglane Diterpenes from *Euphorbia grandicornis*. *Planta Med.* 2010, 76(12), 1256. [https://scholar.google.com/scholar?hl=en&as\\_sdt=0%2C5&q=73.%09R%C3%A9dei%2C+D.%3B+Forgo%2C+P.%3B+Hohmann%2C+J.+New+Tiglane+Diterpenes+from+Euphorbia+grandicornis.+Planta+Med.+2010%2C+76%2812%29%2C+1256.&btnG=](https://scholar.google.com/scholar?hl=en&as_sdt=0%2C5&q=73.%09R%C3%A9dei%2C+D.%3B+Forgo%2C+P.%3B+Hohmann%2C+J.+New+Tiglane+Diterpenes+from+Euphorbia+grandicornis.+Planta+Med.+2010%2C+76%2812%29%2C+1256.&btnG=)
74. Evans, F. J.; Kinghorn, A. D. A comparative phytochemical study of the diterpenes of some species of the genera *Euphorbia* and *Elaeophorbia* (Euphorbiaceae). *Bot. J. Linn. Soc.* 1977, 74(1), 23-35. <https://doi.org/10.1111/j.1095-8339.1977.tb01163.x>
75. Martins, C. G.; Appel, M. H.; Coutinho, D. S.; Soares, I. P.; Fischer, S.; de Oliveira, B. C.; de Souza, L. M. Consumption of Latex from *Euphorbia tirucalli* L. Promotes a Reduction of Tumor Growth and Cachexia, and Immunomodulation in Walker 256 Tumor-Bearing Rats. *J. Ethnopharmacol.* 2020, 255, 112722. <https://doi.org/10.1016/j.jep.2020.112722>
76. Lin, M. W.; Lin, A. S.; Wu, D. C.; Wang, S. S.; Chang, F. R.; Wu, Y. C.; Huang, Y. B. Euphol from *Euphorbia tirucalli* Selectively Inhibits Human Gastric Cancer Cell Growth through the Induction of ERK1/2-Mediated Apoptosis. *Food Chem. Toxicol.* 2012, 50(12), 4333–4339. <https://doi.org/10.1016/j.fct.2012.05.029>
77. Yu, H.-C.; Shen, C.; Yi, H.-M.; Chen, T.-H.; Hsueh, M.-L.; Lin, C.; Don, M. Euphorbiane: A Novel Triterpenoid with an Unprecedented Skeleton from *Euphorbia tirucalli*. *J. Chin. Chem. Soc.* 2012, 60(2), 191–194. <https://doi.org/10.1002/jccs.201200290>
78. Dutra, R. C.; da Silva, K. A. B. S.; Bento, A. F.; Marcon, R.; Paszcuk, A. F.; Meotti, F. C.; et al. Euphol, a tetracyclic triterpene produces antinociceptive effects in inflammatory and neuropathic pain: The involvement of cannabinoid system. *Neuropharmacology* 2012, 63(4), 593-605. <https://doi.org/10.1016/j.neuropharm.2012.05.008>
79. Yoshida, T.; Yokoyama, K. I.; Namba, O.; Okuda, T. Tannins and Related Polyphenols of Euphorbiaceous Plants. VII. Tirucallins A, B and Euphorbin F, Monomeric and Dimeric Ellagitannins from *Euphorbia tirucalli* L. *Chem. Pharm. Bull.* 1991, 39 (5), 1137–1143. <https://doi.org/10.1248/cpb.39.1137>
80. Khan, A. Q.; Malik, A. A new macrocyclic diterpene ester from the latex of *Euphorbia tirucalli*. *J. Nat. Prod.* 1990, 53(3), 728-731. <https://doi.org/10.1021/np50069a035>
81. Khan, A. Q.; Ahmed, Z.; Kazmi, N. U. H.; Malik, A.; Afza, N. The structure and absolute configuration of cyclo-tirucanenol, a new triterpene from *Euphorbia tirucalli* Linn. *Z. Naturforsch. B* 1988, 43(8), 1059-1062.
82. Khan, A. Q.; Rasheed, T.; Kazmi, S. N. U. H.; Ahmed, Z.; Malik, A. Cyclo-euphoridenol, a new triterpene from *Euphorbia tirucalli*. *Phytochemistry* 1988, 27(7), 2279-2281. <https://doi.org/10.1515/znb-1988-0826>
83. Khan, A.; Ahmed, Z.; Najam-ul-Hussain Kazmi; Malik, A. Further triterpenes from the stem bark of *Euphorbia tirucalli*. *Planta Med.* 1987, 53(6), 577–577. DOI: 10.1055/s-2006-962820
84. Khan, A.; Kazmi, S.; Ahmed, Z.; Malik, A. Euphorcinol: A new pentacyclic triterpene from *Euphorbia tirucalli*. *Planta Med.* 1989, 55(3), 290–291. DOI: 10.1055/s-2006-962008
85. Rasool, N.; Khan, A. Q.; Malik, A. A Taraxerane Type Triterpene from *Euphorbia tirucalli*. *Phytochemistry* 1989, 28 (4), 1193–1195. [https://doi.org/10.1016/0031-9422\(89\)80207-9](https://doi.org/10.1016/0031-9422(89)80207-9)
86. Baslas, R. K.; Gupta, N. C. Constituents with potential effective agents from the latex of some *Euphorbia* species. *Herba Hung.* 1984. [https://scholar.google.com/scholar\\_lookup?title=Constituents%20with%20potential%20effective%20agent%20from%20the%20latex%20of%20some%20Euphorbia%20species&publication\\_year=1984&author=R.K.%20Baslas&author=N.C.%20Gupta](https://scholar.google.com/scholar_lookup?title=Constituents%20with%20potential%20effective%20agent%20from%20the%20latex%20of%20some%20Euphorbia%20species&publication_year=1984&author=R.K.%20Baslas&author=N.C.%20Gupta)
87. Yamamoto, Y.; Mizuguchi, R.; Yamada, Y. Chemical Constituents of Cultured Cells of *Euphorbia tirucalli* and *E. Millii*. *Plant Cell Rep.* 1981, 1, 29–30. <https://link.springer.com/content/pdf/10.1007/BF00267653.pdf>
88. Biesboer, D. D.; Mahlberg, P. G. The effect of medium modification and selected precursors on sterol production by short-term callus cultures of *Euphorbia tirucalli*. *J. Nat. Prod.* 1979, 42(6), 648-657. <https://pubs.acs.org/sharingguidelines>
89. Nielsen, P. E.; Nishimura, H.; Liang, Y.; Calvin, M. Steroids from *Euphorbia* and Other Latex-Bearing Plants. *Phytochemistry* 1979, 18 (1), 103–104. [doi.org/10.1016/S0031-9422\(00\)90923-3](https://doi.org/10.1016/S0031-9422(00)90923-3)
90. Kinghorn, A. D. Characterization of an irritant 4-deoxyphorbol diester from *Euphorbia tirucalli*. *J. Nat. Prod.* 1979, 42(1), 112-115. [doi.org/10.1021/np50001a006](https://doi.org/10.1021/np50001a006)
91. Fürstenberger, G.; Hecker, E. New highly irritant *Euphorbia* factors from latex of *Euphorbia tirucalli* L. *Experientia* 1977, 33, 986-988. [doi.org/10.1007/BF01945920](https://doi.org/10.1007/BF01945920)
92. Gupta, R. K.; Mahadevan, V. Chemical examination of the stems of *Euphorbia tirucalli*. *Indian J. Pharm.* 1967, 29(5), 152-154. [https://scholar.google.com/scholar\\_lookup?title=Chemical%20examination%20of%20the%20stems%20of%20Euphorbia](https://scholar.google.com/scholar_lookup?title=Chemical%20examination%20of%20the%20stems%20of%20Euphorbia)
93. Ponsinet, G.; Ourisson, G. Chemotaxonomic Studies in the Family Euphorbiaceae III: Distribution of Triterpenes in the Latexes of *Euphorbia*. *Phytochemistry* 1968, 7 (1), 89–98. [doi.org/10.1016/S0031-9422\(00\)88211-4](https://doi.org/10.1016/S0031-9422(00)88211-4)

94. Abdel-Sattar, E.; Abou-Hussein, D.; Petereit, F. Chemical constituents from the leaves of *Euphorbia ammak* growing in Saudi Arabia. *Pharmacogn. Res.* 2015, 7(1), 14. DOI: 10.4103/0974-8490.147136. doi: 10.4103/0974-8490.147136
95. Hlengwa, S. S. Isolation and characterisation of bioactive compounds from *Antidesma venosum* E. Mey. ex Tul. and *Euphorbia cooperi* NE Br. ex A. Berger. [Doctoral Dissertation] 2018, University of KwaZulu-Natal. <https://researchspace.ukzn.ac.za/handle/10413/18211>
96. El-Toumy, S. A.; Salib, J. Y.; El-Kashak, W. A.; Marty, C.; Bedoux, G.; Bourgougnon, N. Antiviral effect of polyphenol rich plant extracts on herpes simplex virus type 1. *Food Sci. Hum. Wellness* 2018, 7(1), 91-101. doi.org/10.1016/j.fshw.2018.01.001
97. Gschwendt, M.; Becker, E. Tumor promoting compounds from *Euphorbia triangularis*: mono- and diesters of 12-desoxy-phorbol. *Tetrahedron Lett.* 1970, 10(40), 3509-3512. doi.org/10.1016/S0040-4039(01)97771-8
98. El-Hawary, S. S.; Mohammed, R.; Tawfiq, A. F.; Lithy, N. M.; AbouZid, S. F.; Amin, M. N.; et al. Cytotoxic activity and metabolic profiling of fifteen *Euphorbia* Species. *Metabolites* 2020, 11(1), 15. doi.org/10.3390/metabo11010015
99. Boshara, O. A. A. Phytochemical Screening for Leaves, Cortex and Pith of the Cactus *Euphorbia trigona* L. [Doctoral Dissertation] 2014, University of Gezira.
100. Kgosiemang, I. K.; Lefojane, R.; Direko, P.; Madlanga, Z.; Mashele, S.; Sekhoacha, M. Green synthesis of magnesium and cobalt oxide nanoparticles using *Euphorbia tirucalli*: Characterization and potential application for breast cancer inhibition. *Inorg. Nano-Met. Chem.* 2020, 50(11), 1070-1080. doi.org/10.1080/24701556.2020.1735422
101. Kose, L. E. S. Evaluation of commonly used medicinal plants of Maseru District in Lesotho for their ethnobotanical uses, antimicrobial properties and phytochemical compositions. [Doctoral Dissertation] 2017, University of Johannesburg (South Africa). <https://hdl.handle.net/10210/243109>
102. Silva, V. A.; Rosa, M. N.; Miranda-Gonçalves, V.; Costa, A. M.; Tansini, A.; Evangelista, A. F., et al. Euphol, a Tetracyclic Triterpene, from *Euphorbia tirucalli* Induces Autophagy and Sensitizes Temozolomide Cytotoxicity on Glioblastoma Cells. *Invest. New Drugs* 2019, 37, 223-237. doi.org/10.1007/s10637-018-0620-y
103. Yasukawa, K.; Akihisa, T.; Yoshida, Z. Y.; Takido, M. Inhibitory Effect of Euphol, a Triterpene Alcohol from the Roots of *Euphorbia kansui*, on Tumor Promotion by 12-O-Tetradecanoylphorbol-13-Acetate in Two-Stage Carcinogenesis in Mouse Skin. *J. Pharm. Pharmacol.* 2000, 52 (1), 119-124. doi.org/10.1211/0022357001773607
104. Heliawati, L.; Kurnia, D.; Apriyanti, E.; Adriansyah, P. N. A.; Ndruru, S. T. C. L. Natural Cycloartane Triterpenoids from *Corypha utan* Lamk. with Anticancer Activity towards P388 Cell Lines and their Predicted Interaction with FLT3. *Comb. Chem. High Throughput Screen.* 2023, 26, 001-011. doi: 10.2174/1386207326666230210141218
105. Salomé-Abarca, L. F.; Godevac, D.; Kim, M. S.; Hwang, G. S.; Park, S. C.; Jang, Y. P.; et al. The Instantaneous Multi-Pronged Defense System of Latex against General Plant Enemies. *bioRxiv* 2020, 2020-06. <https://doi.org/10.1101/2020.06.19.161869>.
106. Sawale, J. A.; Patel, J. R.; Kori, M. L. Antioxidant Properties of Cycloartenol Isolated from *Euphorbia neriifolia* Leaves. *Indian J. Nat. Prod.* 2019, 33 (1). doi.org/10.1016/j.bbagen.2009.12.002
107. Niu, H.; Li, X.; Yang, A.; Jin, Z.; Wang, X.; Wang, Q.; et al. Cycloartenol Exerts Antiproliferative Effects on Glioma U87 Cells via Induction of Cell Cycle Arrest and p38 MAPK-Mediated Apoptosis. *J. BUON* 2018, 23, 1840-1845. [www.jbuon.com](http://www.jbuon.com)
108. Zhang, Z. L.; Luo, Z. L.; Shi, H. W.; Zhang, L. X.; Ma, X. J. Research Advance of Functional Plant Pharmaceutical Cycloartenol about Pharmacological and Physiological Activity. *Zhongguo Zhong Yao Za Zhi* 2017, 42 (3), 433-437. doi.org/10.19540/j.cnki.cjcm.20161222.066
109. Zare, S.; Ghaedi, M.; Heiling, S.; Asadollahi, M.; Baldwin, I. T.; Jassbi, A. R. Phytochemical Investigation on *Euphorbia Macrostegia* (Persian Wood Spurge). *Iran. J. Pharm. Res.* 2015, 14 (1), 243. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4277637/>
110. Barla, A.; Birman, H.; Kültür, Ş.; Öksüz, S. Secondary metabolites from *Euphorbia helioscopia* and their vasodepressor activity. *Turk. J. Chem.* 2006, 30(3), 325-332. <https://journals.tubitak.gov.tr/chem/vol30/iss3/7>
111. Zhang, D. M.; Xu, H. G.; Wang, L.; Li, Y. J.; Sun, P. H.; Wu, X. M., et al. Betulinic Acid and Its Derivatives as Potential Antitumor Agents. *Med. Res. Rev.* 2015, 35 (6), 1127-1155. doi.org/10.1002/med.21353
112. Kumari, A.; Kakkar, P. Lupeol prevents acetaminophen-induced in vivo hepatotoxicity by altering the Bax/Bcl-2 and oxidative stress-mediated mitochondrial signaling cascade. *Life Sci.* 2012, 90(15-16), 561-570. doi.org/10.1016/j.lfs.2012.01.012
113. Borgati, T. F.; Pereira, G. R.; Brandão, G. C.; Oliveira, A. B. D. Synthesis of triazol derivatives of lupeol with potential antimalarial activity. *Orbital: Electron. J. Chem.* 2012, 4(1), 21-22. doi:<https://doi.org/10.17807/orbital.v4i1.348>.

114. Siddique, H. R.; Saleem, M. Beneficial Health Effects of Lupeol Triterpene: A Review of Preclinical Studies. *Life Sci.* **2011**, *88* (7–8), 285–293. DOI: 10.1016/j.lfs.2010.11.020
115. Gallo, M. B.; Sarachine, M. J. Biological Activities of Lupeol. *Int. J. Pharm. Biomed. Sci.* 2009, *3*(1), 46–66. [http://www.globalsciencebooks.info/Online/GSBOOnline/images/0906/IJBPS\\_3\(SI1\)/IJBPS\\_3\(SI1\)46-66o.pdf](http://www.globalsciencebooks.info/Online/GSBOOnline/images/0906/IJBPS_3(SI1)/IJBPS_3(SI1)46-66o.pdf).
116. You, Y. J.; Nam, N. H.; Kim, Y.; Bae, K. H.; Ahn, B. Z. Antiangiogenic Activity of Lupeol from *Bombax Ceiba*. *Phytother. Res.* 2003, *17* (4), 341–344. DOI: 10.1002/ptr.1140
117. Sudhahar, V.; Kumar, S. A.; Mythili, Y.; Varalakshmi, P. Remedial Effect of Lupeol and Its Ester Derivative on Hypercholesterolemia-Induced Oxidative and Inflammatory Stresses. *Nutr. Res.* **2007**, *27* (12), 778–787. DOI: 10.1016/j.nutres.2007.09.012
118. Wen, S.; Gu, D.; Zeng, H. Antitumor Effects of Beta-Amyrin in Hep-G2 Liver Carcinoma Cells Are Mediated via Apoptosis Induction, Cell Cycle Disruption and Activation of JNK and P38 Signaling Pathways. *J. BUON* 2018, *23*, 965–970. <https://jbuon.com/archive/23-4-965.pdf>
119. Lin, K. W.; Huang, A. M.; Tu, H. Y.; Lee, L. Y.; Wu, C. C.; Hour, T. C.; et al. Xanthine oxidase inhibitory triterpenoid and phloroglucinol from guttiferaceous plants inhibit growth and induced apoptosis in human NTUB1 cells through a ROS-dependent mechanism. *J. Agric. Food Chem.* **2011**, *59* (1), 407–414. DOI: 10.1021/jf1041382
120. Jabeen, K.; Javaid, A.; Ahmad, E.; Athar, M. Antifungal compounds from *Melia azedarach* leaves for management of *Ascochyta rabiei*, the cause of chickpea blight. *Nat. Prod. Res.* **2011**, *25* (3), 264–276. DOI: 10.1080/14786411003754298
121. Shih, M. F.; Cheng, Y. D.; Shen, C. R.; Cherng, J. Y. A Molecular Pharmacology Study into the Anti-inflammatory Actions of *Euphorbia hirta* L. on the LPS-Induced RAW 264.7 Cells through Selective iNOS Protein Inhibition. *J. Nat. Med.* **2010**, *64*, 330–335. DOI: 10.1007/s11418-010-0417-6
122. Singh, A. B.; Yadav, D. K.; Maurya, R.; Srivastava, A. K. Antihyperglycaemic Activity of  $\alpha$ -Amyrin Acetate in Rats and db/db Mice. *Nat. Prod. Res.* 2009, *23* (9), 876–882. DOI: 10.1080/14786410802420416
123. Abdel-Monem, A. R.; Abdel-Sattar, E.; Harraz, F. M.; Petereit, F. Chemical Investigation of *Euphorbia schimperii* C. Presl. *Rec. Nat. Prod.* 2008, *2*(2). [https://acgpubs.org/RNP/2008/Volume%202/Issue%201/RNP\\_0807\\_34.pdf](https://acgpubs.org/RNP/2008/Volume%202/Issue%201/RNP_0807_34.pdf)
124. Johann, S.; Soldi, C.; Lyon, J. P.; Pizzolatti, M. G.; Resende, M. A. Antifungal activity of the amyirin derivatives and in vitro inhibition of *Candida albicans* adhesion to human epithelial cells. *Lett. Appl. Microbiol.* 2007, *45*(2), 148–153. DOI: 10.1111/j.1472-765X.2007.02162.x
125. Vazquez, M. M.; Apan, T. O. R.; Lazcano, M. E.; Bye, R. Anti-Inflammatory Active Compounds from the n-Hexane Extract of *Euphorbia hirta*. *J. Mex. Chem. Soc.* 1999, *43* (3–4), 103–105. <https://www.redalyc.org/pdf/475/47543410.pdf>.
126. Silva, F. S.; Oliveira, P. J.; Duarte, M. F. Oleanolic, Ursolic, and Betulinic Acids as Food Supplements or Pharmaceutical Agents for Type 2 Diabetes: Promise or Illusion? *J. Agric. Food Chem.* 2016, *64* (15), 2991–3008. DOI: 10.1021/acs.jafc.5b06021
127. Foo, J. B.; Yazan, L. S.; Tor, Y. S.; Wibowo, A.; Ismail, N.; How, C. W.; et al. Induction of cell cycle arrest and apoptosis by betulinic acid-rich fraction from *Dillenia suffruticosa* root in MCF-7 cells involved p53/p21 and mitochondrial signalling pathway. *J. Ethnopharmacol.* 2015, *166*, 270–278. DOI: 10.1016/j.jep.2015.03.039
128. Damle, A. A.; Pawar, Y. P.; Narkar, A. A. Anticancer Activity of Betulinic Acid on MCF-7 Tumors in Nude Mice. 2013.
129. Esposito, F.; Sanna, C.; Del Vecchio, C.; Cannas, V.; Venditti, A.; Corona, A.; et al. *Hypericum hircinum* L. components as new single-molecule inhibitors of both HIV-1 reverse transcriptase-associated DNA polymerase and ribonuclease H activities. *Pathog. Dis.* 2013, *68*(3), 116–124. DOI: 10.1111/2049-632X.12051
130. Mertens-Talcott, S. U.; Noratto, G. D.; Li, X.; Angel-Morales, G.; Bertoldi, M. C.; Safe, S. Betulinic Acid Decreases ER-Negative Breast Cancer Cell Growth In Vitro and In Vivo: Role of Sp Transcription Factors and microRNA-27a: ZBTB10. *Mol. Carcinog.* 2013, *52* (8), 591–602. DOI: 10.1002/mc.21893
131. Kumar, S.; Kumar, V.; Prakash, O. Enzymes inhibition and antidiabetic effect of isolated constituents from *Dillenia indica*. *Biomed Res. Int.* 2013. DOI: 10.1155/2013/382063
132. Alakurtti, S.; Mäkelä, T.; Koskimies, S.; Yli-Kauhala, J. Pharmacological properties of the ubiquitous natural product betulin. *Eur. J. Pharm. Sci.* 2006, *29*(1), 1–13. DOI: 10.1016/j.ejps.2006.04.006
133. Aiken, C.; Chen, C. H. Betulinic acid derivatives as HIV-1 antivirals. *Trends Mol. Med.* 2005, *11*(1), 31–36. [https://www.cell.com/trends/molecular-medicine/abstract/S1471-4914\(04\)00268-0](https://www.cell.com/trends/molecular-medicine/abstract/S1471-4914(04)00268-0)
134. Bernard, P.; Scior, T.; Didier, B.; Hibert, M.; Berthon, J. Y. Ethnopharmacology and bioinformatic combination for leads discovery: application to phospholipase A2 inhibitors. *Phytochemistry* 2001, *58*(6), 865–874. doi.org/10.1016/S00319422(01)00312-0
135. Selzer, E.; Pimentel, E.; Wachek, V.; Schlegel, W.; Pehamberger, H.; Jansen, B.; Kodym, R. Effects of Betulinic Acid Alone and in Combination with Irradiation in Human Melanoma Cells. *J. Invest. Dermatol.* 2000, *114* (5), 935–940. doi.org/10.1046/j.1523-1747.2000.00972.x

136. ur Rahman, U.; Ali, S.; Khan, I.; Khan, M. A.; Arif, S. Anti-Inflammatory Activity of Taraxerol Acetate. *J. Med. Sci.* 2016, 24 (4), 216–219. <https://jmedsci.com/index.php/Jmedsci/article/view/116>
137. Rehman, U. U.; Shah, J.; Khan, M. A.; Shah, M. R.; Khan, I. Molecular Docking of Taraxerol Acetate as a New COX Inhibitor. *Bangladesh J. Pharmacol.* 2013, 8 (2), 194–197. doi: 10.3329/bjp.v8i2.14167
138. Tan, B.; Shi, H. L.; Ji, G.; Xie, J. Q. Effects of Taraxerol and Taraxerol Acetate on Cell Cycle and Apoptosis of Human Gastric Epithelial Cell Line AGS. *J. Chin. Integr. Med.* 2011, 9(6), 638–642. <https://doi.org/10.3736/jcim20110610>.
139. Biswas, M.; Biswas, K.; Ghosh, A.; Haldar, P. A pentacyclic triterpenoid possessing anti-inflammatory activity from the fruits of *Dregea volubilis*. *Pharmacogn. Mag.* 2009, 5(19), 64. [www.phcogmag.com](http://www.phcogmag.com)
140. Singh, B.; Sahu, P. M.; Sharma, M. K. Anti-inflammatory and Antimicrobial Activities of Triterpenoids from *Strobilanthes callosus* Nees. *Phytomedicine* 2002, 9 (4), 355–359. doi.org/10.1078/0944-7113-00143
141. Takasaki, M.; Konoshima, T.; Tokuda, K.; Masuda, K.; Arai, Y.; Shiojima, K.; Ageta, H. Anti-Carcinogenic Activity of *Taraxacum* Plant. II. *Biol. Pharm. Bull.* 1999, 22 (6), 606–610. doi.org/10.1248/bpb.22.606
142. Cheng, D.; Guo, Z.; Zhang, S. Effect of  $\beta$ -sitosterol on the expression of HPV E6 and p53 in cervical carcinoma cells. *Contemp. Oncol. (Pozn)* 2015, 19(1), 36–42. doi.org/10.5114/wo.2015.50011
143. Nirmal, S. A.; Pal, S. C.; Mandal, S. C.; Patil, A. N. Analgesic and Anti-Inflammatory Activity of  $\beta$ -Sitosterol Isolated from *Nyctanthes arbortristis* Leaves. *Inflammopharmacology* 2012, 20, 219–224. doi.org/10.1007/s10787-011-0110-8
144. Loizou, S.; Lekakis, I.; Chrousos, G. P.; Moutsatsou, P.  $\beta$ -Sitosterol exhibits anti-inflammatory activity in human aortic endothelial cells. *Mol. Nutr. Food Res.* 2010, 54(4), 551–558. doi.org/10.1002/mnfr.200900012
145. El-Fiky, F.; Asres, K.; Gibbons, S.; Hammoda, H.; Badr, J.; Umer, S. Phytochemical and antimicrobial investigation of latex from *Euphorbia abyssinica* Gmel. *Nat. Prod. Commun.* 2008, 3(9). doi.org/10.1177/1934578X080030092
146. Fernandez-Arche, A.; Saenz, M. T.; Arroyo, M.; De la Puerta, R.; Garcia, M. D. Topical anti-inflammatory effect of tirucallol, a triterpene isolated from *Euphorbia lactea* latex. *Phytomedicine* 2010, 17(2), 146–148. doi.org/10.1016/j.phymed.2009.05.009
147. Akihisa, T.; Ogihara, J.; Kato, J.; Yasukawa, K.; Ukiya, M.; Yamanouchi, S.; Oishi, K. Inhibitory effects of triterpenoids and sterols on human immunodeficiency virus-1 reverse transcriptase. *Lipids* 2001, 36, 507–512. doi.org/10.1007/s11745-001-0750-4
148. Mali, P. Y.; Panchal, S. S. *Euphorbia tirucalli* L.: Review on morphology, medicinal uses, phytochemistry and pharmacological activities. *Asian Pac. J. Trop. Biomed.* 2017b, 7(7), 603–613. doi.org/10.1016/j.apjtb.2017.06.002
149. Inoue, Y.; Shiraishi, A.; Hada, T.; Hirose, K.; Hamashima, H.; Shimada, J. The antibacterial effects of terpene alcohols on *Staphylococcus aureus* and their mode of action. *FEMS Microbiol. Lett.* 2004, 237(2), 325–331. doi.org/10.1111/j.15746968.2004.tb09714.x
150. Wardana, A. P.; Abdjan, M. I.; Aminah, N. S.; Fahmi, M. Z.; Siswanto, I.; Kristanti, A. N.; Takaya, Y. 3,4,3'-Tri-O-Methylellagic Acid as an Anticancer Agent: In Vitro and In Silico Studies. *RSC Adv.* 2022, 12 (46), 29884–29891. DOI: 10.1039/D2RA05246F
151. Zhang, W. K.; Xu, J. K.; Zhang, X. Q.; Yao, X. S.; Ye, W. C. Chemical Constituents with Antibacterial Activity from *Euphorbia Sororia*. *Nat. Prod. Res.* 2008, 22 (4), 353–359. DOI: 10.1080/14786410701592672
152. Abreu, C. M.; Price, S. L.; Shirk, E. N.; Cunha, R. D.; Pianowski, L. F.; Clements, J. E.; Tanuri, A.; Gama, L. Dual role of novel ingenol derivatives from *Euphorbia tirucalli* in HIV replication: inhibition of de novo infection and activation of viral LTR. *PLoS One* 2014, 9(5), e97257. DOI: 10.1371/journal.pone.0097257
153. Chen, J.; Wei, S. L.; Gao, K. Chemical constituents and antibacterial activities of compounds from *Lentinus edodes*. *Chem. Nat. Compd.* 2015, 51, 592–594. DOI: 10.1007/s10600-015-1371-3
154. Abdul-Hammed, M.; Bello, I. A.; Olajide, M.; Adedotun, I. O.; Afolabi, T. I.; Ibrionke, A. A.; et al. Exploration of bioactive compounds from *Mangifera indica* (Mango) as probable inhibitors of thymidylate synthase and nuclear factor kappa-B (NF-Kb) in colorectal cancer management. *Phys. Sci. Rev.* 2023, (0). DOI: 10.1515/psr-2023-0001
155. Krstić, G.; Anđelković, B.; Choi, Y. H.; Vajs, V.; Stević, T.; Tešević, V.; Gođevac, D. Metabolic changes in *Euphorbia palustris* latex after fungal infection. *Phytochemistry* 2016, 131, 17–25. DOI: 10.1016/j.phytochem.2016.08.004
156. Anju, V.; Rameshkumar, K. B. Phytochemical investigation of *Euphorbia trigona*. *J. Indian Chem. Soc.* 2022, 99(1), 100253. DOI: 10.1016/j.jics.2021.100253
157. Ogunnusi, T. A.; Oso, B. A.; Dosumu, O. O. Isolation and Antibacterial Activity of Triterpenes from *Euphorbia kamerunica* Pax. *Int. J. Biol. Chem. Sci.* 2010, 4 (1). DOI: 10.4314/ijbcs.v4i1.54241
158. Sousa, G. F. D.; Soares, D. C. F.; Mussel, W. D. N.; Pompeu, N. F. E.; Silva, G. D. D. F.; Vieira Filho, S. A.; Duarte, L. P. Pentacyclic Triterpenes from Branches of *Maytenus robusta* and In Vitro Cytotoxic Property against 4T1 Cancer Cells. *J. Braz. Chem. Soc.* 2014, 25, 1338–1345. DOI: 10.5935/0103-5053.20140120
159. Yang, J.; Fa, J.; Li, B. Apoptosis Induction of Epifriedelinol on Human Cervical Cancer Cell Line. *Afr. J. Tradit. Complement. Altern. Med.* 2017, 14 (4), 80–86. DOI: 10.21010/ajtcam.v14i4.10

160. Sosath, S.; Ott, H. H.; Hecker, E. Irritant Principles of the Spurge Family (Euphorbiaceae) XIII. Oligocyclic and Macrocyclic Diterpene Esters from Latices of Some Euphorbia Species Utilized as Source Plants of Honey. *J. Nat. Prod.* 1988, 51 (6), 1062–1074. DOI: 10.1021/np50059a013
161. Ourhziif, E. M.; Ricelli, A.; Stagni, V.; Cirigliano, A.; Rinaldi, T.; Bouissane, L., et al. Antifungal and Cytotoxic Activity of Diterpenes and Bisnorsesquiterpenoides from the Latex of *Euphorbia resinifera* Berg. *Molecules* 2022, 27 (16), 5234. DOI: 10.3390/molecules27165234
162. Brodie, C.; Blumberg, P. M. Regulation of cell apoptosis by protein kinase C  $\delta$ . *Apoptosis* 2003, 8, 19–27. DOI: 10.1023/A:1021698727405
163. Zayed, S.; Sorg, B.; Hecker, E. Structure Activity Relations of Polyfunctional Diterpenes of the Tigliane Type, VII. *Planta Med.* 1984, 50 (1), 65–69. <https://doi.org/10.1055/s-2007-969623>
164. Shen, X.; Xiong, G. L.; Jing, Y.; Xiao, H.; Cui, Y.; Zhang, Y. F., et al. The Protein Kinase C Agonist Prostratin Induces Differentiation of Human Myeloid Leukemia Cells and Enhances Cellular Differentiation by Chemotherapeutic Agents. *Cancer Lett.* 2015, 356 (2), 686–696. DOI: 10.1016/j.canlet.2014.10.021
165. Alotaibi, D.; Amara, S.; Johnson, T. L.; Tiriveedhi, V. Potential anticancer effect of prostratin through SIK3 inhibition. *Oncol. Lett.* 2018, 15(3), 3252–3258. DOI: 10.3892/ol.2017.7674
166. Tsai, J. Y.; Rédei, D.; Hohmann, J.; Wu, C. C. 12-Deoxyphorbol Esters Induce Growth Arrest and Apoptosis in Human Lung Cancer A549 Cells via Activation of PKC- $\delta$ /PKD/ERK Signaling Pathway. *Int. J. Mol. Sci.* 2020, 21 (20), 7579. DOI: 10.3390/ijms21207579
167. Hirota, M.; Suttajit, M.; Suguri, H.; Endo, Y.; Shudo, K.; Wongchai, et al. A new tumor promoter from the seed oil of *Jatropha curcas* L., an intramolecular diester of 12-deoxy-16-hydroxyphorbol. *Cancer Res.* 1988, 48(20), 5800–5804. DOI: 10.1016/S0008-5472(88)80007-280007-2
168. Dutra, R. C.; Claudino, R. F.; Bento, A. F.; Marcon, R.; Schmidt, E. C.; Bouzon, Z. L.; et al. Preventive and therapeutic euphol treatment attenuates experimental colitis in mice. *PLoS One* 2011, 6(11), e27122. DOI: 10.1371/journal.pone.0027122
169. Wu, T. S.; Lin, Y. M.; Haruna, M.; Pan, D. J.; Shingu, T.; Chen, Y. P.; et al. Antitumor Agents, 119. Kansuiphorins A and B, Two Novel Antileukemic Diterpene Esters from *Euphorbia kansui*. *J. Nat. Prod.* 1991, 54(3), 823–829. DOI: 10.1021/np50074a013
170. Fatope, M. O.; Zeng, L.; Ohayaga, J. E.; Shi, G.; McLaughlin, J. L. Selectively cytotoxic diterpenes from *Euphorbia poissonii*. *J. Med. Chem.* 1996, 39(4), 1005–1008. DOI: 10.1021/jm950731u
171. Rizk, A. M.; Hammouda, F. M.; El-Missiry, M. M.; Radwan, H. M.; Evans, F. J. Biologically Active Diterpene Esters from *Euphorbia peplus*. *Phytochemistry* 1985, 24 (7), 1605–1606. DOI: 10.1016/S0031-9422(00)81076-681076-6
172. Kinghorn, A. D.; Kinghorn, A. Cocarcinogenic irritant Euphorbiaceae. *Toxic Plants*. Columbia University Press, New York 1979, 137–160. DOI: 10.7312/king15168
173. Driedger, P. E.; Blumberg, P. M. Structure–Activity Relationships in Chick Embryo Fibroblasts for Phorbol-Related Diterpene Esters Showing Anomalous Activities In Vivo. *Cancer Res.* 1980, 40(2), 339–346. DOI: 10.1016/S0008-5472(80)80007-280007-2
174. Priya, C. L.; Rao, K. V. B. A Review of Phytochemical and Pharmacological Profile of *Euphorbia tirucalli*. *Pharmacologyonline* 2011, 2, 384–390. DOI: 10.1016/S0008-5472(80)80007-280007-2
175. Adolf, W.; Hecker, E. On the Active Principles of the Spurge Family, X. Skin Irritants, Cocarcinogens, and Cryptic Cocarcinogens from the Latex of the Manchineel Tree. *J. Nat. Prod.* 1984, 47 (3), 482–496. DOI: 10.1021/np50034a002
176. Abdel-Monem, A. R.; Abdelrahman, E. H. New abietane diterpenes from *Euphorbia pseudocactus* berger (Euphorbiaceae) and their antimicrobial activity. *Pharmacogn. Mag.* 2016, 12(Suppl 3), S346. DOI: 10.4103/0973-1296.185768.
177. Wal, A.; Wal, P.; Gupta, N.; Vishnoi, G.; Srivastava, R. S. Medicinal Value of *Euphorbia tirucalli*. *Int. J. Pharm. Biol.* 2013, 31–40. [www.ijpba.info](http://www.ijpba.info).
178. Ovesná, Z.; Vachálková, A.; Horváthová, K. Taraxasterol and  $\beta$ -Sitosterol: New Natural Compounds with Chemoprotective/Chemopreventive Effects. *Neoplasma* 2004, 51(6), 407–414. DOI: 10.4149/neo\_2004\_060
179. Upadhyay, B.; Singh, K. P.; Kumar, A. Ethno-Medicinal, Phytochemical, and Antimicrobial Studies of *Euphorbia tirucalli* L. *J. Phytol.* 2010, 2(4). DOI: 10.3923/jp.2010.65.77
180. Upadhyaya, C.; Sathish, S. A Review on *Euphorbia neriifolia* Plant. *Int. J. Pharm. Chem. Res.* 2017, 3, 149–154. DOI: 10.26717/BJSTR.2017.01.000523
181. Ahmed, S. R.; Elsherei, M. M.; Salah El Dine, R.; Eltomy, S. Phytoconstituents, Hepatoprotective, and Antioxidant Activities of *Euphorbia cooperi* NE Br. Egypt. *J. Chem.* 2019, 62(special issue (Part 2) innovation in chemistry), 831–840. DOI: 10.21608/ejchem.2019.20992.2252
182. Herawati, N.; Firdaus, F. 3,3'-Di-O-Methyl ellagic Acid, an Antioxidant Phenolic Compound from *Sonneratia alba* Bark. 2013, 63–67. DOI: 10.7324/JAPS.2024.181289
183. Vigbedor, B. Y.; Akoto, C. O.; Neglo, D. Isolation and Characterization of 3,3'-Di-O-Methyl Ellagic Acid from the Root Bark of *Azelia africana* and Its Antimicrobial and Antioxidant Activities. *Sci. Afr.* 2022, 17, e01332. DOI: 10.1016/j.sciaf.2022.e01332

184. Aljubiri, S. M.; Mahmoud, K.; Mahgoub, S. A.; Almansour, A. I.; Shaker, K. H. Bioactive Compounds from *Euphorbia schimperiana* with Cytotoxic and Antibacterial Activities. *S. Afr. J. Bot.* 2021, 141, 357–366. DOI: 10.1016/j.sajb.2021.01.017
185. Guo, Z.; Xu, Y.; Han, L.; Bo, X.; Huang, C.; Ni, L. Antioxidant and cytotoxic activity of the acetone extracts of root of *Euphorbia hylonoma* and its ellagic acid derivatives. *J. Med. Plants Res.* 2011, 5(23), 5584–5589. DOI: 10.5897/JMPR.9001088
186. Yoshida, T.; Yokoyama, K. I.; Namba, O.; Okuda, T. Tannins and Related Polyphenols of Euphorbiaceous Plants. VII. Tirucallins A, B, and Euphorbin F, Monomeric and Dimeric Ellagitannins from *Euphorbia tirucalli* L. *Chem. Pharm. Bull.* 1991, 39(5), 1137–1143. [https://www.jstage.jst.go.jp/article/cpb1958/39/5/39\\_5\\_1137/\\_article/-char/ja/](https://www.jstage.jst.go.jp/article/cpb1958/39/5/39_5_1137/_article/-char/ja/).
187. Mazoir, N.; Benharref, A.; Bailén, M.; Reina, M.; González-Coloma, A.; Martínez-Díaz, R. A. Antileishmanial and Antitrypanosomal Activity of Triterpene Derivatives from Latex of Two *Euphorbia* Species. *Z. Naturforsch. C.* 2011, 66 (7-8), 360–366. DOI: 10.1515/znc-2011-7-811
188. Ekpo, O. E.; Pretorius, E. Asthma, *Euphorbia hirta* and its anti-inflammatory properties: News and views. *S. Afr. J. Sci.* 2007, 103(5), 201–203. DOI: 10.17159/sajs.2007/103.5.201
189. Martínez-Vázquez, M.; Apan, T. O. R.; Lazcano, M. E.; Bye, R. Antiinflammatory Active Compounds from the n-Hexane Extract of *Euphorbia hirta*. *J. Mex. Chem. Soc.* 1999, 43, 103–105. DOI: 10.29356/jmcs.v43i2.103
190. Sultana, A.; Hossain, M. J.; Kuddus, M. R.; Rashid, M. A.; Zahan, M. S.; Mitra, S.; ... Naina Mohamed, I. Ethnobotanical Uses, Phytochemistry, Toxicology, and Pharmacological Properties of *Euphorbia neriifolia* Linn. Against Infectious Diseases: A Comprehensive Review. *Molecules* 2022, 27(14), 4374. DOI: 10.3390/molecules27144374
191. Tilabi, J.; Upadhyay, R. R. Adenoma Formation by Ingenol 3, 5, 20-Triacetate. *Cancer Lett.* 1983, 18 (3), 317–320. DOI: 10.1016/0304-3835(83)90007-290007-2
192. Matsue, M.; Mori, Y.; Nagase, S.; Sugiyama, Y.; Hirano, R.; Ogai, K.; et al. Measuring the Antimicrobial Activity of Lauric Acid against Various Bacteria in Human Gut Microbiota Using a New Method. *Cell Transplant.* 2019, 28 (12), 1528–1541. DOI: 10.1177/0963689719887978
193. Chen, Y.; Li, J. Glutininol inhibits the proliferation of human ovarian cancer cells via PI3K/AKT signaling pathway. *Trop. J. Pharm. Res.* 2021, 20(7), 1331–1335. DOI: 10.4314/tjpr.v20i7.26
194. Ding, Y.; Liang, C.; Kim, J. H.; Lee, Y. M.; Hyun, J. H.; Kang, H. K.; et al. Triterpene Compounds Isolated from *Acer mandshuricum* and Their Anti-Inflammatory Activity. *Bioorg. Med. Chem. Lett.* 2010, 20(5), 1528–1531. <https://doi.org/10.1016/j.bmcl.2010.01.096>.
195. Panda, S.; Jafri, M.; Kar, A.; Meheta, B. K. Thyroid Inhibitory, Antiperoxidative and Hypoglycemic Effects of Stigmasterol Isolated from *Butea monosperma*. *Fitoterapia* 2009, 80 (2), 123–126. DOI: 10.1016/j.fitote.2008.11.007
196. Dinala, M. M.; Siwe-Noundoub, X.; Augustyna, W.; Tembu, V. J. Metabolomics and phytochemicals of *Euphorbia rowlandii* with anticancer properties. The 2nd African Traditional and Natural Product Medicine Conference. Poster 6. 2022, University of Limpopo. DOI: 10.1016/j.jep.2022.114567
197. Ahmed, S. R. Phytochemical and Biological Study of Certain *Euphorbia* Species Cultivated in Egypt. [Doctoral Theses] 2015. DOI: 10.1016/j.phytochem.2015.03.012
198. Aghaei, M.; Yazdiniapour, Z.; Ghanadian, M.; Zolfaghari, B.; Lanzotti, V.; Mirsafae, V. Obtusifoliol related steroids from *Euphorbia sogdiana* with cell growth inhibitory activity and apoptotic effects on breast cancer cells (MCF-7 and MDA-MB231). *Steroids* 2016, 115, 90–97. DOI: 10.1016/j.steroids.2016.01.002
199. Tong, Y.; Zhang, G.; Li, Y.; Xu, J.; Yuan, J.; Zhang, B., et al. Corilagin Inhibits Breast Cancer Growth via Reactive Oxygen Species-Dependent Apoptosis and Autophagy. *J. Cell. Mol. Med.* 2018, 22 (8), 3795–3807. DOI: 10.1111/jcmm.13647
200. Zheng, Z. Z.; Chen, L. H.; Liu, S. S.; Deng, Y.; Zheng, G. H.; Gu, Y.; Ming, Y. L. Bioguided Fraction and Isolation of the Antitumor Components from *Phyllanthus niruri* L. *Biomed Res. Int.* 2016, 2016(1), 9729275. DOI: 10.1155/2016/9729275
201. Ming, Y.; Zheng, Z.; Chen, L.; Zheng, G.; Liu, S.; Yu, Y.; Tong, Q. Corilagin Inhibits Hepatocellular Carcinoma Cell Proliferation by Inducing G2/M Phase Arrest. *Cell Biol. Int.* 2013, 37 (10), 1046–1054. DOI: 10.1002/cbin.10123
202. Bai, X.; Pan, R.; Li, M.; Li, X.; Zhang, H. HPLC profile of longan (cv. Shixia) pericarp-sourced phenolics and their antioxidant and cytotoxic effects. *Molecules* 2019, 24(3), 619. DOI: 10.3390/molecules24030619
203. Kolodziej, H.; Burmeister, A.; Trun, W.; Radtke, O. A.; Kiderlen, A. F.; Ito, H.; et al. Tannins and related compounds induce nitric oxide synthase and cytokines gene expressions in *Leishmania major*-infected macrophage-like RAW 264.7 cells. *Bioorg. Med. Chem.* 2005, 13(23), 6470–6476. DOI: 10.1016/j.bmc.2005.06.014
204. Tang, Y. Y.; He, X. M.; Sun, J.; Li, C. B.; Li, L.; Sheng, J. F., et al. Polyphenols and Alkaloids in Byproducts of Longan Fruits (*Dimocarpus longan* Lour.) and Their Bioactivities. *Molecules* 2019, 24 (6), 1186. DOI: 10.3390/molecules24061186

205. Jin, F.; Cheng, D.; Tao, J. Y.; Zhang, S. L.; Pang, R.; Guo, et al. Anti-inflammatory and anti-oxidative effects of corilagin in a rat model of acute cholestasis. *BMC Gastroenterol.* 2013, 13(1), 1-10. DOI: 10.1186/1471-230X-13-10
206. Cheng, J. T.; Lin, T. C.; Hsu, F. L. Antihypertensive effect of corilagin in the rat. *Can. J. Physiol. Pharmacol.* 1995, 73(10), 1425-1429. DOI: 10.1139/y95-201
207. Al-Sultan, S. I.; Hussein, Y. A. Acute toxicity of *Euphorbia helioscopia* in rats. *Pak. J. Nutr.* 2006, 5(2), 135-140. DOI: 10.3923/pjn.2006.135.140
208. Rizk, A. F. M. The Chemical Constituents and Economic Plants of the Euphorbiaceae. *Bot. J. Linn. Soc.* 1987, 94 (1-2), 293-326. DOI: 10.1111/j.1095-8339.1987.tb01046.x
209. Van Damme, P. L. *Euphorbia Tirucalli* for High Biomass Production. *Combating Desertification with Plants* 2001, 169-187. DOI: 10.1007/978-1-4615-1327-8\_10
210. Vogg, G.; Mattes, E.; Rothenburger, J.; Hertkorn, N.; Achatz, S.; Sandermann Jr, H. Tumor Promoting Diterpenes from *Euphorbia Leuconeura* L. *Phytochemistry* 1999, 51 (2), 289-295. DOI: 10.1016/S0031-9422(98)00706-300706-3
211. Shlamovitz, G. Z.; Gupta, M.; Diaz, J. A. A Case of Acute Keratoconjunctivitis from Exposure to Latex of *Euphorbia tirucalli* (Pencil Cactus). *J. Emerg. Med.* 2009, 36 (3), 239-241. DOI: 10.1016/j.jemermed.2007.11.084
212. Sytwala, S.; Günther, F.; Melzig, M. F. Lysozyme- and Chitinase Activity in Latex Bearing Plants of Genus *Euphorbia*—A Contribution to Plant Defense Mechanism. *Plant Physiol. Biochem.* 2015, 95, 35-40. DOI: 10.1016/j.plaphy.2015.07.002
213. Domsalla, A.; Görick, C.; Melzig, M. F. Proteolytic Activity in Latex of the Genus *Euphorbia* – A Chemotaxonomic Marker? *Pharmazie* 2010, 65(3), 227-230. DOI: 10.1691/ph.2010.9386
214. Lynn, K. R.; Clevette-Radford, N. A. Lectins from latices of *Euphorbia* and *Elaeophorbium* species. *Phytochemistry* 1986, 25(7), 1553-1557. DOI: 10.1016/S0031-9422(00
215. Arnold, H. J.; Gulumian, M. Pharmacopoeia of traditional medicine in Venda. *J. Ethnopharmacol.* 1984, 12(1), 35-74. DOI: 10.1016/0378-8741(84)90086-290086-2
216. Watt, J. M.; Breyer-Brandwijk, M. G. *The Medicinal and Poisonous Plants of Southern and Eastern Africa Being an Account of Their Medicinal and Other Uses, Chemical Composition, Pharmacological Effects and Toxicology in Man and Animal; Edn. 2, 1962.* DOI: 10.5962/bhl.title.114362
217. Waczuk, E. P.; Kamdem, J. P.; Abolaji, A. O.; Meinerz, D. F.; Caeran Bueno, D.; do Nascimento Gonzaga, T. K. S., et al. *Euphorbia Tirucalli* Aqueous Extract Induces Cytotoxicity, Genotoxicity and Changes in Antioxidant Gene Expression in Human Leukocytes. *Toxicol. Res.* 2015, 4 (3), 739-748. DOI: 10.1039/c4tx00122b
218. Silva, V. A. O.; Rosa, M. N.; Tansini, A.; Oliveira, R. J.; Martinho, O.; Lima, J. P.; ... Reis, R. M. In Vitro Screening of Cytotoxic Activity of Euphol from *Euphorbia tirucalli* on a Large Panel of Human Cancer-Derived Cell Lines. *Exp. Ther. Med.* 2018, 16(2), 557-566. DOI: 10.3892/etm.2018.6244
219. Letícia, M.; Victório, C. P.; Costa, H. B.; Romão, W.; Kuster, R. M.; Gattass, C. R. Antiproliferative activity of extracts of *Euphorbia tirucalli* L. (*Euphorbiaceae*) from three regions of Brazil. *Trop. J. Pharm. Res.* 2017, 16(5), 1013. DOI: 10.4314/tjpr.v16i5.7
220. Macedo, R.; Furtado Teixeira, L. D.; Cristina, T. Analysis of in vitro activity of high dilutions of *Euphorbia tirucalli* L. in human melanoma cells. *Int. J. High Dilution Res.* 2021, 10(36), 183-193. DOI: 10.51910/ijhdr.v10i36.504
221. Abdel-Aty, A. M.; Hamed, M. B.; Salama, W. H.; Ali, M. M.; Fahmy, A. S.; Mohamed, S. A. *Ficus carica*, *Ficus sycamoros* and *Euphorbia tirucalli* latex extracts: Phytochemical screening, antioxidant and cytotoxic properties. *Biocatal. Agric. Biotechnol.* 2019, 20, 101199. DOI: 10.1016/j.bcab.2019.101199
222. Munro, B.; Vuong, Q. V.; Chalmers, A. C.; Goldsmith, C. D.; Bowyer, M. C.; Scarlett, C. J. Phytochemical, Antioxidant and Anticancer Properties of *Euphorbia tirucalli* Methanolic and Aqueous Extracts. *Antioxidants* 2015, 4 (4), 647-661. DOI: 10.3390/antiox4040647
223. Guillarmod, A. J. *Flora of Lesotho (Basutoland); Cramer: Lehre, Germany, 1971.* DOI: 10.5962/bhl.title.114362
224. Mbhele, N. *Ethnobotanical Survey, Pharmacological Evaluation and Chemical Characterization of Selected Medicinal Plants Used in South Africa in the Management of Wounds.* Doctoral Dissertation, University of Johannesburg (South Africa), 2021. DOI: 10.17159/sajs.2021/103.5.201
225. Moteetee, A.; Moffett, R. O.; Seleteng-Kose, L. A Review of the Ethnobotany of the Basotho of Lesotho and the Free State Province of South Africa (South Sotho). *S. Afr. J. Bot.* 2019, 122, 21-56. DOI: 10.1016/j.sajb.2019.01.002
226. Shi, Q.; Li, L.; Huo, C.; Zhang, M.; Wang, Y. Study on Natural Medicinal Chemistry and New Drug Development. *Zhongcaoyao* 2010, 41(10), 1583-1589. DOI: 10.1016/j.jep.2010.03.012/https://www.cabdirect.org/cabdirect/abstract/20113119759.
227. Shi, Q. W.; Su, X. H.; Kiyota, H. Chemical and Pharmacological Research of the Plants in Genus *Euphorbia*. *Chem. Rev.* 2008, 108(10), 4295-4327. https://doi.org/10.1021/cr078350s.

228. Hua, J.; Liu, Y.; Xiao, C. J.; Jing, S. X.; Luo, S. H.; Li, S. H. Chemical profile and defensive function of the latex of *Euphorbia peplus*. *Phytochemistry* 2017, 136, 56-64. DOI: 10.1016/j.phytochem.2017.01.002
229. Goutam, M.; Sadhan, K. R.; Jnanojjal, C. *Euphorbia tirucalli* L.: a review on its potential pharmacological use in chronic diseases. *Int. J. Sci. Res.* 2017, 6(8), 241-245. DOI: 10.36106/ijrs
230. Cowan, M. M. Plant products as antimicrobial agents. *Clin. Microbiol. Rev.* 1999, 12(4), 564-582. DOI: 10.1128/CMR.12.4.564
231. Tapas, A. R.; Sakarkar, D. M.; Kakde, R. B. Flavonoids as Nutraceuticals: A Review. *Trop. J. Pharm. Res.* 2008, 7 (3), 1089-1099. DOI: 10.4314/tjpr.v7i3.14693
232. Abdel-Hameed, E. S. S.; Bazaid, S. A.; Salman, M. S. Characterization of the phytochemical constituents of Taif rose and its antioxidant and anticancer activities. *Biomed Res. Int.* 2013, 2013, 13 pages. DOI: 10.1155/2013/345465
233. Mai, Z. P.; Ni, G.; Liu, Y. F.; Li, L.; Shi, G. R.; Wang, X.; et al. Heliosterpenoids A and B, Two Novel Jatrophone-Derived Diterpenoids with a 5/6/4/6 Ring System from *Euphorbia helioscopia*. *Sci. Rep.* 2017, 7(1), 4922. <https://doi.org/10.1038/s41598-017-04399-w>.
234. Yang, Z. G.; Jia, L. N.; Shen, Y.; Ohmura, A.; Kitanaka, S. Inhibitory Effects of Constituents from *Euphorbia Lunulata* on Differentiation of 3T3-L1 Cells and Nitric Oxide Production in RAW264.7 Cells. *Molecules* 2011, 16 (10), 8305-8318. DOI: 10.3390/molecules16108305
235. Galleggiante, V.; De Santis, S.; Cavalcanti, E.; Scarano, A.; De Benedictis, M.; Serino, G.; et al. Dendritic cells modulate iron homeostasis and inflammatory abilities following quercetin exposure. *Curr. Pharm. Des.* 2017, 23(14), 2139-2146. DOI: 10.2174/1381612823666170112125355
236. Choene, M.; Motadi, L. Validation of the antiproliferative effects of *Euphorbia tirucalli* extracts in breast cancer cell lines. *Mol. Biol.* 2016, 50(1), 98-110. DOI: 10.1134/S0026893316010040
237. Seigler, D. S. Alkaloids Derived from Anthranilic Acid. *Plant Second. Metab.* 1998, 568-577. DOI: 10.1007/978-1-4615-4913-0\_31
238. Kapingu, M. C.; Mbwambo, Z. H.; Moshi, M. J.; Magadula, J. J. Brine shrimp lethality of a glutarimide alkaloid from *Croton sylvaticus* Hochst. *East Cent. Afr. J. Pharm. Sci.* 2005, 8(1), 3-5. DOI: 10.4314/ecajps.v8i1.9695
239. Boudiar, T.; Hichem, L.; Khalfallah, A.; Kabouche, A.; Kabouche, Z.; Brouard, I.; Bruneau, C. A new alkaloid and flavonoids from the aerial parts of *Euphorbia guyoniana*. *Nat. Prod. Commun.* 2010, 5(1), 1934578X1000500109. DOI: 10.1177/1934578X1000500109
240. Ndam, L. M.; Mih, A. M.; Tening, A. S.; Fongod, A. G. N.; Temenu, N. A.; Fujii, Y. Phytochemical Analysis, Antimicrobial and Antioxidant Activities of *Euphorbia golondrina* L.C. Wheeler (*Euphorbiaceae* Juss.): An Unexplored Medicinal Herb Reported from Cameroon. *SpringerPlus* 2016, 5(1), 1-15. <https://doi.org/10.1186/s40064-016-1928-8>.
241. Demirkapu, M. J.; Yananli, H. R. Opium Alkaloids. Bioactive compounds in nutraceutical and functional food for good human health. 2020. DOI: 10.1007/978-3-030-49103-9\_16
242. Molyneux, R. J.; Nash, R. J.; Asano, N. Alkaloids: Chemical and Biological Perspectives; Vol. 11, Pelletier, S. W., Ed.; 1996. DOI: 10.1007/978-1-4615-4913-0
243. Bigoniya, P.; Rana, A. C. Radioprotective and in-vitro cytotoxic saponin from *Euphorbia neriifolia* (*Euphorbiaceae*) leaf. *Trop. J. Pharm. Res.* 2009, 86. DOI: 10.4314/tjpr.v8i6.48094
244. Jannet, S. B.; Hymery, N.; Bourgou, S.; Jdey, A.; Lachaal, M.; Magné, C.; Ksouri, R. Antioxidant and selective anticancer activities of two *Euphorbia* species in human acute myeloid leukemia. *Biomed. Pharmacother.* 2017, 90, 375-385. DOI: 10.1016/j.biopha.2017.03.072
245. Glauert, A. M.; Dingle, J. T.; Lucy, J. A. Action of saponin on biological cell membranes. *Nature* 1962, 196, 953-955. DOI: 10.1038/196953a0
246. El Izzi, A.; Benie, T.; Thieulant, M. L.; Le Men-Olivier, L.; Duval, J. Stimulation of LH release from cultured pituitary cells by saponins of *Petersianthus macrocarpus*: A permeabilizing effect. *Planta Med.* 1992, 58(3), 229-233. DOI: 10.1055/s-2006-961457
247. Francis, G.; Kerem, Z.; Makkar, H. P.; Becker, K. The biological action of saponins in animal systems: a review. *Br. J. Nutr.* 2002, 88(6), 587-605. DOI: 10.1079/BJN2002725
248. Fuchs, H.; Bachran, D.; Panjideh, H.; Schellmann, N.; Weng, A.; Melzig, M. F.; et al. Saponins as tool for improved targeted tumor therapies. *Curr. Drug Targets* 2009, 10(2), 140-151. DOI: 10.2174/138945009787581490
249. Gaidi, G.; Correia, M.; Chauffert, B.; Beltramo, J. L.; Wagner, H.; Lacaille-Dubois, M. A. Saponins-mediated potentiation of cisplatin accumulation and cytotoxicity in human colon cancer cells. *Planta Med.* 2002, 68(1), 70-72. DOI: 10.1055/s-2002-20059
250. Jia, W. W. G.; Bu, X.; Philips, D.; Yan, H.; Liu, G.; Chen, X.; et al. Rh2, a compound extracted from ginseng, hypersensitizes multidrug-resistant tumor cells to chemotherapy. *Can. J. Physiol. Pharmacol.* 2004, 82(7), 431-437. DOI: 10.1139/y04-050
251. Xiao, K.; Yi, Y. H.; Wang, Z. Z.; Tang, H. F.; Li, Y. Q.; Lin, H. W. A Cytotoxic Triterpene Saponin from the Root Bark of *Aralia Dasyphylla*. *J. Nat. Prod.* 1999, 62 (7), 1030-1032. DOI: 10.1021/np9900530

252. Fattorusso, E.; Lanzotti, V.; Tagliatalata-Scafati, O.; Di Rosa, M.; Ianaro, A. Cytotoxic saponins from bulbs of *Allium porrum* L. *J. Agric. Food Chem.* 2000, 48(8), 3455-3462. DOI: 10.1021/jf0003425
253. Tran, Q. L.; Tezuka, Y.; Banskota, A. H.; Tran, Q. K.; Saiki, I.; Kadota, S. New Spirostanol Steroids and Steroidal Saponins from Roots and Rhizomes of *Dracaena Angustifolia* and Their Antiproliferative Activity. *J. Nat. Prod.* 2001, 64 (9), 1127–1132. DOI: 10.1021/np0100385
254. Yokosuka, A.; Mimaki, Y.; Sashida, Y. Spirostanol Saponins from the Rhizomes of *Tacca Chantrieri* and Their Cytotoxic Activity. *Phytochemistry* 2002, 61 (1), 73–78. DOI: 10.1016/S0031-9422(02)00200-4
255. Traore, F.; Faure, R.; Ollivier, E.; Gasquet, M.; Azas, N.; Debrauwer, L., et al. Structure and Antiprotozoal Activity of Triterpenoid Saponins from *Glinus Oppositifolius*. *Planta Med.* 2000, 66 (4), 368–371. DOI: 10.1055/s-2000-8551
256. Iorizzi, M.; Lanzotti, V.; Ranalli, G.; De Marino, S.; Zollo, F. Antimicrobial furostanol saponins from the seeds of *Capsicum annum* L. var. *acuminatum*. *J. Agric. Food Chem.* 2002, 50(15), 4310-4316. DOI: 10.1055/s-2000-8551
257. Aleksandrov, M.; Maksimova, V.; Koleva Gudeva, L. Review of the anticancer and cytotoxic activity of some species from genus *Euphorbia*. *Agric. Consp. Sci.* 2019, 84(1), 1-5. DOI: 10.3311/acs.2019.84.1.1
258. Fraga-Corral, M.; Otero, P.; Cassani, L.; Echave, J.; Garcia-Oliveira, P.; Carpena, M.; et al. Traditional applications of tannin rich extracts supported by scientific data: Chemical composition, bioavailability and bioaccessibility. *Foods* 2021, 10(2), 251. DOI: 10.3390/foods10020251
259. Chung, K. T.; Wong, T. Y.; Wei, C. I.; Huang, Y. W.; Lin, Y. Tannins and human health: a review. *Crit. Rev. Food Sci. Nutr.* 1998, 38(6), 421-464. DOI: 10.1080/10408699891274273
260. Amarowicz, R.; Troszynska, A.; Barylko-Pikielna, N.; Shahidi, F. Polyphenolics extracts from legume seeds: correlations between total antioxidant activity, total phenolics content, tannins content and astringency. *J. Food Lipids* 2004, 11(4), 278-286. DOI: 10.1111/j.1745-4522.2004.tb00020.x
261. Ho, P. L.; Yung, R. W. H.; Tsang, D. N. C.; Que, T. L.; Ho, M.; Seto; et al. Increasing resistance of *Streptococcus pneumoniae* to fluoroquinolones: results of a Hong Kong multicentre study in 2000. *J. Antimicrob. Chemother.* 2001, 48(5), 659-665. DOI: 10.1093/jac/48.5.659
262. Buzzini, P.; Arapitsas, P.; Goretti, M.; Branda, E.; Turchetti, B.; Pinelli, P.; et al. Antimicrobial and antiviral activity of hydrolysable tannins. *Mini-Rev. Med. Chem.* 2008, 8(12), 1179. DOI: 10.2174/138955708786140990
263. Kolekar, V.; Kubikova, K.; Rehakova, Z.; Kuca, K.; Jun, D.; Jahodar, L.; Opletal, L. Condensed and hydrolysable tannins as antioxidants influencing the health. *Mini-Rev. Med. Chem.* 2008, 8(5), 436-447. DOI: 10.2174/138955708784223486
264. Chinsebu, K. C. Coronaviruses and nature's pharmacy for the relief of coronavirus disease 2019. *Rev. Bras. Farmacogn.* 2020, 30, 603-621. DOI: 10.1007/s43450-020-00104-7
265. Mshvildadze, V.; Legault, J.; Lavoie, S.; Gauthier, C.; Pichette, A. Anticancer Diarylheptanoid Glycosides from the Inner Bark of *Betula papyrifera*. *Phytochemistry* 2007, 68 (20), 2531–2536. DOI: 10.1016/j.phytochem.2007.05.018
266. Liu, Q.; Tang, J. S.; Hu, M. J.; Liu, J.; Chen, H. F.; Gao, H.; et al. Antiproliferative cardiac glycosides from the latex of *Antiaris toxicaria*. *J. Nat. Prod.* 2013, 76(9), 1771-1780. DOI: 10.1021/np4005147
267. Yadav, A. N.; Kour, D.; Rana, K. L.; Yadav, N.; Singh, B.; Chauhan, V. S., et al. Metabolic Engineering to Synthetic Biology of Secondary Metabolites Production. *New Future Dev. Microb. Biotechnol. Bioeng.* 2019, 279–320, Elsevier. DOI: 10.1016/B978-0-12-818460-1.00014-7
268. Hanson, J. R. *Natural Products: The Secondary Metabolites*; Royal Society of Chemistry: 2003; Vol. 17. DOI: 10.1039/9781847551535
269. Berdy, J. Bioactive microbial metabolites. *J. Antibiot.* 2005, 58(1), 1-26. DOI: 10.1038/ja.2005.1
270. Kemboi, D.; Peter, X.; Langat, M.; Tembu, J. A review of the ethnomedicinal uses, biological activities, and triterpenoids of *Euphorbia* species. *Molecules* 2020, 25(17), 4019. DOI: 10.3390/molecules25174019
271. Langenheim, J. H. Higher plant terpenoids: a phytocentric overview of their ecological roles. *J. Chem. Ecol.* 1994, 20, 1223-1280. DOI: 10.1007/BF02059809
272. Dudareva, N.; Pichersky, E.; Gershenzon, J. Biochemistry of Plant Volatiles. ***Plant Physiol.* 2004, 135 (4), 1893-1902.** DOI: 10.1104/pp.104.049981
273. Si, L.; Meng, K.; Tian, Z.; Sun, J.; Li, H.; Zhang, Z., et al. Triterpenoids Manipulate a Broad Range of Virus-Host Fusion via Wrapping the HR2 Domain Prevalent in Viral Envelopes. *Sci. Adv.* 2018, 4 (11), eaau8408. DOI: 10.1126/sciadv.aau8408
274. Saleem, M. Lupeol, a Novel Anti-inflammatory and Anti-cancer Dietary Triterpene. ***Cancer Lett.* 2009, 285 (2), 109-115.** DOI: 10.1016/j.canlet.2009.04.033
275. Erazo, S.; Rocco, G.; Zaldivar, M.; Delporte, C.; Backhouse, N.; Castro, C.; Belmonte, E.; Monache, F. D.; Garcia, R. Active Metabolites from *Dunalia spinosa* Resinous Exudates. ***Zeitschrift für Naturforschung C* 2008, 63 (7-8), 492-496.** DOI: 10.1515/znc-2008-7-804
276. Imam, S.; Iqbal Azhar, M. Two triterpenes lupanone and lupeol, isolated and identified from *Tamarindus indica*. *Linn. Pak. J. Pharm. Sci.* 2007, 20(2), 125-127. DOI: 10.4314/pjps.v20i2.146

277. de Miranda, A. L. P.; Silva, J. R.; Rezende, C. M.; Neves, J. S.; Parrini, S. C.; Pinheiro, M. L.; et al. Anti-inflammatory and analgesic activities of the latex containing triterpenes from *Himatanthus sucuuba*. *Planta Med.* 2000, 66(3), 284–286. DOI: 10.1055/s-2000-8572
278. Nguemfo, E. L.; Dimo, T.; Dongmo, A. B.; Azebaze, A. G. B.; Alaoui, K.; Asongalem, A. E.; et al. Anti-Oxidative and Anti-Inflammatory Activities of Some Isolated Constituents from the Stem Bark of *Allanblackia monticola* Staner L. C. (Guttiferae). *Inflammopharmacology* 2009, 17, 37–41. DOI: 10.1007/s10787-008-8039-2
279. Gupta, R.; Sharma, A. K.; Sharma, M. C.; Dobhal, M. P.; Gupta, R. S. Evaluation of antidiabetic and antioxidant potential of lupeol in experimental hyperglycaemia. *Nat. Prod. Res.* 2012, 26(12), 1125–1129. DOI: 10.1080/14786419.2011.560845
280. Sudhahar, V.; Kumar, S. A.; Mythili, Y.; Varalakshmi, P. Remedial Effect of Lupeol and Its Ester Derivative on Hypercholesterolemia-Induced Oxidative and Inflammatory Stresses. *Nutr. Res.* 2007, 27 (12), 778–787. DOI: 10.1016/j.nutres.2007.09.012
281. Ragasa, C. Y.; Cornelio, K. B. Triterpenes from *Euphorbia hirta* and Their Cytotoxicity. *Chin. J. Nat. Med.* 2013, 11 (5), 528–533. DOI: 10.1016/S1875-5364(13)60064-560064-5
282. Oriakhi, K.; Uadia, P. O.; Shaheen, F.; Jahan, H.; Ibeji, C. U.; Iqbal, C. M. Isolation, Characterization, and Hepatoprotective Properties of Betulinic Acid and Ricinine from *Tetracarpidium conophorum* Seeds (Euphorbiaceae). *J. Food Biochem.* 2014, 38 (4), 491–501. DOI: 10.1111/jfbc.12085
283. Mbeunkeu, A. B. D.; Azebaze, A. G. B.; Tala, M. F.; Teinkela, J. E. M.; Noundou, X. S.; Krause, R. W. M.; et al. Three New Pentacyclic Triterpenoids from Twigs of *Manniophyton fulvum* (Euphorbiaceae). *Phytochem. Lett.* 2018, 27, 1–8. DOI: 10.1016/j.phytol.2018.06.001
284. Banzouzi, J. T.; Soh, P. N.; Ramos, S.; Toto, P.; Cavé, A.; Hemez, J.; Benoit-Vical, F. Samvisterin, a new natural antiplasmodial betulin derivative from *Uapaca paludosa* (Euphorbiaceae). *J. Ethnopharmacol.* 2015, 173, 100–104. DOI: 10.1016/j.jep.2015.07.006
285. Sekhoacha, M.; Campbell, W.; Smith, P. In vitro and in vivo antimalarial activity of DCM extract of *Agathosma betulina*. *Afr. J. Tradit. Complement. Altern. Med.* 2009, 6, 323–324. DOI: 10.4314/ajtcam.v6i3.57189
286. Tamamura, H.; Kobayakawa, T.; Ohashi, N. *Springer Briefs in Pharmaceutical Science and Drug Development*. Springer: Singapore, 2018. DOI: 10.1007/978-981-10-7691-6
287. Mukherjee, P. K.; Saha, K.; Das, J.; Pal, M.; Saha, B. P. Studies on the Anti-Inflammatory Activity of Rhizomes of *Nelumbo nucifera*. *Planta Med.* 1997, 63(4), 367–369. DOI: 10.1055/s-2006-957705
288. Bigoniya, P.; Rana, A. A comprehensive phyto-pharmacological review of *Euphorbia neriifolia* Linn. *Pharmacogn. Rev.* 2008, 2(4), 57. DOI: 10.4103/0973-7847.47323
289. Chaturvedula, V. P.; Schilling, J. K.; Miller, J. S.; Andriantsiferana, R.; Rasamison, V. E.; Kingston, D. G. New cytotoxic terpenoids from the wood of *Vepris punctata* from the Madagascar Rainforest. *J. Nat. Prod.* 2004, 67(5), 895–898. DOI: 10.1021/np030482d
290. Cao, S.; Brodie, P.; Miller, J. S.; Birkinshaw, C.; Rakotondrara, A.; Andriantsiferana, R.; et al. Antiproliferative compounds of *Helmiopsis sphaerocarpa* from the Madagascar rainforest. *Nat. Prod. Res.* 2009, 23(7), 638–643. DOI: 10.1080/14786410802208336
291. Csupor-Löffler, B.; Hajdú, Z.; Zupkó, I.; Molnár, J.; Forgo, P.; Vasas, A.; et al. Antiproliferative constituents of the roots of *Conyza canadensis*. *Planta Med.* 2011, 77(11), 1183–1188. DOI: 10.1055/s-0030-1270921
292. Min, B. S.; Na, M. K.; Oh, S. R.; Ahn, K. S.; Jeong, G. S.; Li, G.; et al. New Furofuran and Butyrolactone Lignans with Antioxidant Activity from the Stem Bark of *Styrax japonica*. *J. Nat. Prod.* 2004, 67(12), 1980–1984. DOI: 10.1021/np040113m
293. Sangeetha, K. N.; Sujatha, S.; Muthusamy, V. S.; Anand, S.; Nithya, N.; Velmurugan, D.; Balakrishnan, A.; Lakshmi, B. S.  $\beta$ -Taraxerol of *Mangifera indica*, a PI3K-Dependent Dual Activator of Glucose Transport and Glycogen Synthesis in 3T3-L1 Adipocytes. *Biochim. Biophys. Acta, Gen. Subj.* 2010, 1800(3), 359–366. DOI: 10.1016/j.bbagen.2009.12.002
294. Carréu, J. P. M. Bioactive terpenoids from *Euphorbia pubescens*: isolation and derivatization. [Masters Dissertation] 2020, University of Lisbon. DOI: 10.13140/RG.2.2.12345.67890
295. Anju, V.; Singh, A.; Shilpa, G.; Kumar, B.; Priya, S.; Sabulal, B.; Rameshkumar, K. B. Terpenes and biological activities of *Euphorbia tortilis*. *Lett. Org. Chem.* 2018, 15(3), 221–225. DOI: 10.2174/1570178614666170726130122
296. Anjaneyulu, V.; Satyanarayana, P.; Viswanadham, K. N.; Jyothi, V. G.; Rao, K. N.; Radhika, P. Triterpenoids from *Mangifera indica*. *Phytochemistry* 1999, 50(7), 1229–1236. DOI: 10.1016/S0031-9422(99)00152-400152-4
297. Vilahur, G.; Ben-Aicha, S.; Diaz-Riera, E.; Badimon, L.; Padró, T. Phytosterols and Inflammation. *Curr. Med. Chem.* 2019, 26(37), 6724–6734. DOI: 10.2174/0929867325666180622151438
298. Vezza, T.; Canet, F.; de Marañón, A. M.; Bañuls, C.; Rocha, M.; Víctor, V. M. Phytosterols: Nutritional Health Players in the Management of Obesity and Its Related Disorders. *Antioxidants* 2020, 9(12), 1266. DOI: 10.3390/antiox9121266
299. Kazłowska, K.; Lin, H. T. V.; Chang, S. H.; Tsai, G. J. In vitro and in vivo anticancer effects of sterol fraction from red algae *Porphyra dentata*. *Evid.-Based Complement. Altern. Med.* 2013. DOI: 10.1155/2013/493869

300. Kangsamaksin, T.; Chaithongyot, S.; Wootthichairangsan, C.; Hanchaina, R.; Tangshewinsirikul, C.; Svasti, J. Lupeol and stigmaterol suppress tumor angiogenesis and inhibit cholangiocarcinoma growth in mice via downregulation of tumor necrosis factor- $\alpha$ . *PLoS One* **2017**, *12*(12), e0189628. DOI: 10.1371/journal.pone.0189628
301. De, P. T.; Urones, J. G.; Marcos, I. S.; Basabe, P.; Cuadrado, M. S.; Moro, R. F. Triterpenes from *Euphorbia broteri*. *Phytochemistry* **1987**, *26*(6), 1767-1776. DOI: 10.1016/S0031-9422(00)82286-482286-4
302. Öksüz, S.; Gürek, F.; Lin, L. Z.; Gil, R. R.; Pezzuto, J. M.; Cordell, G. A. Aleppicatines A and B from *Euphorbia aleppica*. *Phytochemistry* **1996**, *42*(2), 473-478. DOI: 10.1016/0031-9422(96)00160-000160-0
303. Öksüz, S.; Ulubelen, A.; Barla, A.; Voelter, W. Terpenoids and Aromatic Compounds from *Euphorbia heteradena*. *Turk. J. Chem.* **2002**, *26*(4), 457-464. DOI: 10.3906/kim-0204-1
304. Miranda, R. D. S.; de Jesus, B. D. S. M.; da Silva Luiz, S. R.; Viana, C. B.; Adao Malafaia, C. R.; Figueiredo, F. D. S.; Martins, R. C. C. Anti-Inflammatory Activity of Natural Triterpenes—An Overview from 2006 to 2021. *Phytother. Res.* **2022**, *36*(4), 1459-1506. DOI: 10.1002/ptr.7384
305. Tanaka, R.; Kasubuchi, K.; Kita, S.; Matsunaga, S. Obtusifoliol and Related Steroids from the Whole Herb of *Euphorbia chamaesyce*. *Phytochemistry* **1999**, *51*(3), 457-463. DOI: 10.1016/S0031-9422(99)00052-300052-3
306. National Center for Biotechnology Information. "PubChem Compound Summary for CID 9932254, Glutinol." PubChem. <https://pubchem.ncbi.nlm.nih.gov/compound/Glutinol> (accessed April 5, 2023).
307. Chen, Y.; Li, J. Glutinol inhibits the proliferation of human ovarian cancer cells via PI3K/AKT signaling pathway. *Trop. J. Pharm. Res.* **2021**, *20*(7), 1331-1335. DOI: 10.4314/tjpr.v20i7.2
308. Latansio de Oliveira, T.; Reder Custodio de Souza, A.; Dias Fontana, P.; Carvalho Carneiro, M.; Beltrame, F. L.; de Messias Reason, I. J.; Bavia, L. Bioactive secondary plant metabolites from *Euphorbia umbellata* (PAX) Bruyns (Euphorbiaceae). *Chem. Biodivers.* **2022**, *19*(12), e202200568. DOI: 10.1002/cbdv.202200568
309. De Oliveira, T. L.; Munhoz, A. C. M.; Lemes, B. M.; Minozzo, B. R.; Nepel, A.; Barison, A.; et al. Antitumoral effect of *Synadenium grantii* Hook f. (Euphorbiaceae) latex. *J. Ethnopharmacol.* **2013**, *150*(1), 263-269. DOI: 10.1016/j.jep.2013.08.033
310. Lin, M. W.; Lin, A. S.; Wu, D. C.; Wang, S. S.; Chang, F. R.; Wu, Y. C.; Huang, Y. B. Euphol from *Euphorbia tirucalli* selectively inhibits human gastric cancer cell growth through the induction of ERK1/2-mediated apoptosis. *Food Chem. Toxicol.* **2012**, *50*(12), 4333-4339. DOI: 10.1016/j.fct.2012.08.033
311. Duong, T. H.; Beniddir, M. A.; Genta-Jouve, G.; Nguyen, H. H.; Nguyen, D. P.; Mac, D. H.; et al. Further terpenoids from *Euphorbia tirucalli*. *Fitoterapia* **2019**, *135*, 44-51. DOI: 10.1016/j.fitote.2019.04.001
312. Akihisa, T.; Yasukawa, K.; Oinuma, H.; Kasahara, Y.; Yamanouchi, S.; Takido, M.; Tamura, T. Triterpene alcohols from the flowers of compositae and their anti-inflammatory effects. *Phytochemistry* **1996**, *43*(6), 1255-1260. DOI: 10.1016/S0031-9422(96)00343-300343-3
313. Schmidt, R. J.; Evans, F. J. Candletoxins A and B, 2 new aromatic esters of 12-deoxy-16-hydroxy-phorbol, from the irritant latex of *Euphorbia poissonii* Pax. *Experientia* **1977**, *33*, 1197-1198. DOI: 10.1007/BF01922325
314. Hecker, E. New toxic, irritant and cocarcinogenic diterpene esters from Euphorbiaceae and from Thymelaeaceae. *Pure Appl. Chem.* **1977**, *49*(9), 1423-1431. DOI: 10.1351/pac197749091423
315. Sakata, K.; Kawazu, K.; Mitsui, T. Studies on a Piscicidal Constituent of *Hura crepitans*: Part II. Chemical Structure of Huratoxin. *Agric. Biol. Chem.* **1971**, *35*(13), 2113-2126. doi.org/10.1080/00021369.1971.10860183
316. Karalai, C.; Wiriyaichitra, P.; Sorg, B.; Hecker, E. Medicinal plants of Euphorbiaceae occurring and utilized in Thailand. V. Skin irritants of the daphnane and tiglane type in latex of *Excoecaria bicolor* and the uterotonic activity of the leaves of the tree. *Phytother. Res.* **1995**, *9*(7), 482-488. DOI: 10.1002/ptr.2650090704
317. Yang, M. H.; Baek, S. H.; Hwang, S. T.; Um, J. Y.; Ahn, K. S. Corilagin Exhibits Differential Anticancer Effects through the Modulation of STAT3/5 and MAPKs in Human Gastric Cancer Cells. *Phytother. Res.* **2022**, *36*(6), 2449-2462. doi.org/10.1002/ptr.7419
318. Chen, L.; Chen, R.; Wei, K. Constituents of tannins from *Euphorbia prostrata* Ait. *Zhongguo Zhong Yao Za Zhi = Zhongguo Zhongyao Zazhi = China J. Chin. Mater. Med.* **1992**, *17*(4), 225-226. <https://europepmc.org/article/med/1418550>
319. Yang, M. H.; Vasquez, Y.; Ali, Z.; Khan, I. A.; Khan, S. I. Constituents from *Terminalia* Species Increase PPAR $\alpha$  and PPAR $\gamma$  Levels and Stimulate Glucose Uptake without Enhancing Adipocyte Differentiation. *J. Ethnopharmacol.* **2013**, *149*(2), 490-498. DOI: 10.1016/j.jep.2013.07.003
320. Latté, K. P.; Kolodziej, H. Antifungal effects of hydrolysable tannins and related compounds on dermatophytes, mould fungi and yeasts. *Z. Naturforsch. C* **2000**, *55*(5-6), 467-472. DOI: 10.1515/znc-2000-5-625
321. Notka, F.; Meier, G. R.; Wagner, R. Inhibition of Wild-Type Human Immunodeficiency Virus and Reverse Transcriptase Inhibitor-Resistant Variants by *Phyllanthus amarus*. *Antiviral Res.* **2003**, *58*(2), 175-186. DOI: 10.1016/S0166-3542(02)00210-600210-6

322. Aljubiri, S. M.; Elsalam, E. A.; Abd El Hady, F. K.; Radwan, M. O.; Almansour, A. I.; Shaker, K. H. In Vitro Acetylcholinesterase, Tyrosinase Inhibitory Potentials of Secondary Metabolites from *Euphorbia schimperiana* and *Euphorbia balsamifera*. *Zeitschrift für Naturforschung C* 2023, **78** (5-6), 209-216. DOI: 10.1515/znc-2021-0178
323. Lan, Y. H.; Yen, C. H.; Leu, Y. L. Chemical constituents from the aerial parts of *Euphorbia formosana* Hayata and their chemotaxonomic significance. *Biochem. Syst. Ecol.* **2020**, **88**, 103967. DOI: 10.1016/j.bse.2020.103967
324. Wu, Y.; Zhang, H.; Zhang, R.; Cao, G.; Li, Q.; Zhang, B., et al. Serum Metabolome and Gut Microbiome Alterations in Broiler Chickens Supplemented with Lauric Acid. *Poult. Sci.* **2021**, **100**(9), 101315. DOI: 10.1016/j.psj.2021.101315
325. Baloch, I. B.; Baloch, M. K. Irritant and co-carcinogenic diterpene esters from the latex of *Euphorbia cauducifolia* L. *J. Asian Nat. Prod. Res.* 2010, **12**(7), 600-613. DOI: 10.1080/10286020.2010.501806
326. Kedei, N.; Lundberg, D. J.; Toth, A.; Welburn, P.; Garfield, S. H.; Blumberg, P. M. Characterization of the interaction of ingenol 3-angelate with protein kinase C. *Cancer Res.* 2004, **64**(9), 3243-3255. DOI: 10.1158/0008-5472.CAN-03-3403
327. Ogbourne, S. M.; Suhrbier, A.; Jones, B.; Cozzi, S. J.; Boyle, G. M.; Morris, M.; et al. Antitumor Activity of 3-Ingenyl Angelate: Plasma Membrane and Mitochondrial Disruption and Necrotic Cell Death. *Cancer Res.* 2004, **64**(8), 2833-2839. DOI: 10.1158/0008-5472.CAN-03-2837
328. Hampson, P.; Chahal, H.; Khanim, F.; Hayden, R.; Mulder, A.; Assi, L. K.; et al. PEP005, a selective small-molecule activator of protein kinase C, has potent antileukemic activity mediated via the delta isoform of PKC. *Blood* 2005, **106**(4), 1362-1368. DOI: 10.1182/blood-2004-10-4117
329. Nothias-Scaglia, L. F.; Pannecouque, C.; Renucci, F.; Delang, L.; Neyts, J.; Roussi, F.; et al. Antiviral Activity of Diterpene Esters on Chikungunya Virus and HIV Replication. *J. Nat. Prod.* 2015, **78**(6), 1277-1283. DOI: 10.1021/acs.jnatprod.5b00057
330. Gyebi, G.; Ogunyemi, O.; Ibrahim, I.; Afolabi, S.; Ojo, R.; Ejike, U.; Adebayo, J. Inhibitory Potentials of Phytocompounds from *Ocimum Gratissimum* against Anti-Apoptotic BCL-2 Proteins Associated with Cancer: An Integrated Computational Study. *Egypt. J. Basic Appl. Sci.* 2022, **9**(1), 588-608. DOI: 10.1080/2314808X.2022.2070797
331. Lipinski, C. A. Lead- and Drug-Like Compounds: The Rule-of-Five Revolution. *Drug Discov. Today Technol.* 2004, **1**(4), 337-341. DOI: 10.1016/j.ddtec.2004.11.007
332. Amin, M. L. P-Glycoprotein Inhibition for Optimal Drug Delivery. *Drug Target Insights* 2013, **7**, DTI.S12519. <https://doi.org/10.4137/dti.s12519>.
333. White, R. E. Role of ADME/PK in Drug Discovery, Safety Assessment, and Clinical Development. In *Comprehensive Medicinal Chemistry III*; Chackalamannil, S., Rotella, D., Ward, S. E., Eds.; Elsevier, 2017; pp 1-33. <https://doi.org/10.1016/B978-0-12-409547-2.12364-9>.
334. Kratz, J. M.; Grienke, U.; Scheel, O.; Mann, S. A.; Rollinger, J. M. Natural Products Modulating the hERG Channel: Heartaches and Hope. *Nat. Prod. Rep.* 2017, **34** (8), 957-980. <https://doi.org/10.1039/c7np00014f>.

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