

Review

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Review

Odorant Receptors and Cancer

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Abstract: Odorant receptors (ORs) constitute the largest family of G protein-coupled receptors (GPCRs), with nearly 400 receptors identified in humans. The "omics" era has revealed an unexpected expression of ORs beyond olfactory tissues. For many decades these receptors were neglected from cancer research, largely due to the assumption that their expression in cancerous tissues was a background leakage, unrelated to conventional cancer pathways such as cell replication, differentiation, or DNA damage response. The Cancer Genome Atlas (TCGA) data shows, however, that OR expression profiles are specific to each tumor type. This evidence supports that ORs may be related to tumor malignancy. In this review, we explore the extranasal expression of ORs in cancer and discuss the potential implications of their presence in cancerous tissues.

Keywords: odorant receptor; extranasal; cancer; GPCR

1. Introduction

Emerging evidence has highlighted the expression and function of extranasal odorant receptors (ORs) in non-olfactory tissues, indicating that they may have different functions in these tissues. Although ORs outside the nose were first identified a few years after their discovery [1,2], their role in non-olfactory tissues was initially overlooked due to lack of indications for functional roles of these receptors outside the nasal cavity. Most extranasal ORs were unexpectedly discovered during investigations of differentially expressed genes across various non-olfactory tissues, including multiple types of cancer and their corresponding healthy tissue [3–5].

These extranasal ORs have been shown to play significant role in several physiological processes, such as: regulating blood pressure through renin secretion [6,7]; glucose and lipid metabolism regulation through a variety of metabolic events including insulin and glucagon secretion [8] and fatty acid oxidation [9]; maintenance of the oxygen homeostasis of breathing in the carotid body [10,11]; sperm chemotaxis [12,13] and many others, emphasizing the need for further research in this area. Additionally, studies have revealed the involvement of ORs in tumorigenesis and the progression of various cancers, including breast [4,14], skin [3], prostate [15,16], lung [17], colon [18] cancers among others. As a result, ORs have gained attention as potential diagnostic biomarkers and, more importantly, as promising drug targets [19].

2. Odorant Receptors

In 1991, Buck and Axel [20] identified the OR gene family in rats, which contains approximately 1000 genes in mice and 400 genes in humans [21,22]. They were awarded the Nobel Prize in Physiology or Medicine in 2004 for uncovering the molecular basis of smell.

The ORs are G-protein-coupled receptors (GPCRs) and are located in the cilia of olfactory sensory neurons (OSNs) within the olfactory mucosa. Only one OR gene type is expressed per olfactory neuron, following the "one neuron - one receptor" rule [23,24]. Binding of the odorant to its corresponding OR activates the odorant signaling pathway, leading to neuronal depolarization (Figure 1A).

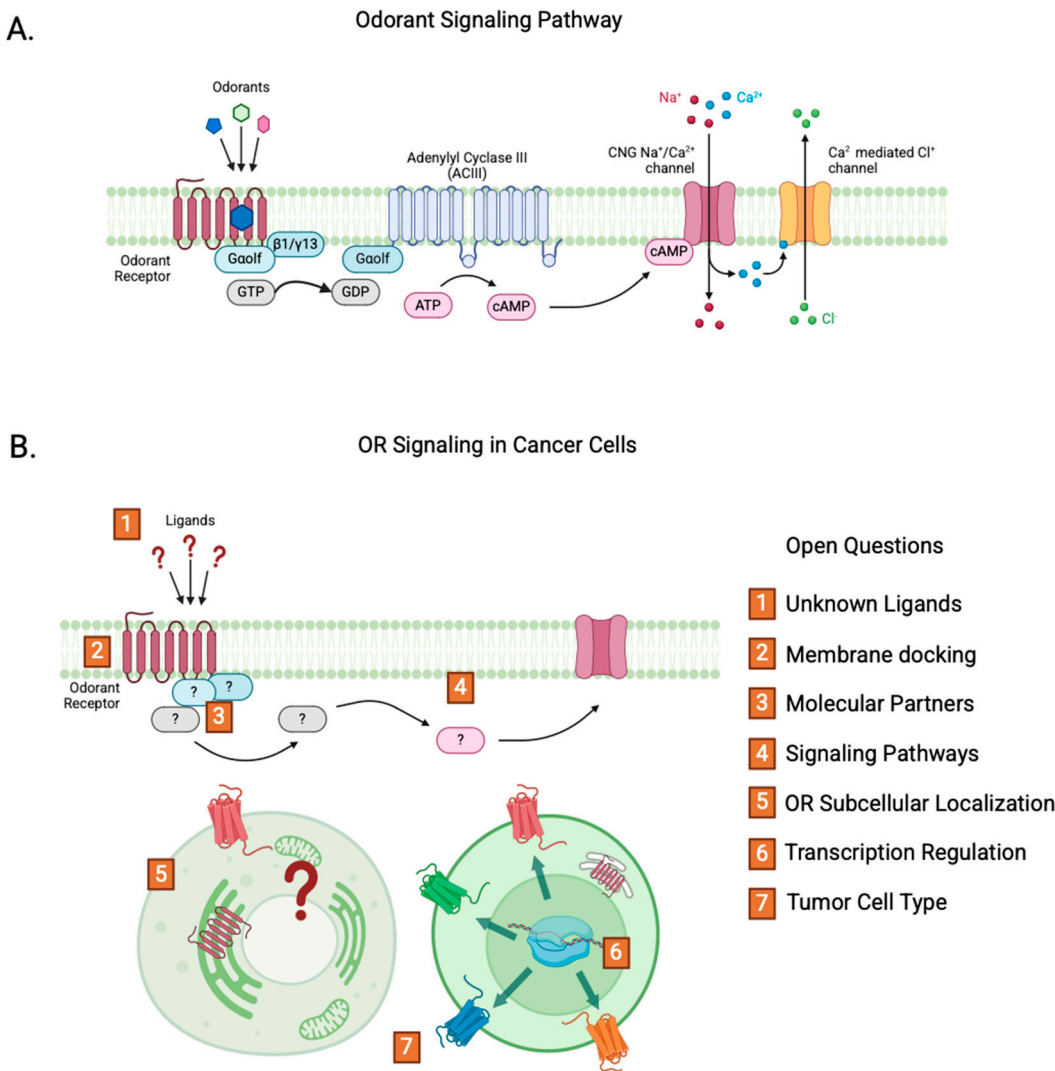


Figure 1. Schematic comparison between the OR signaling pathway in olfactory sensory neurons and in cancer cells. A. Odorant binding induces a conformational change of the OR structure leading to the dissociation of GDP from the Gαolf subunit and subsequent GTP binding to Gαolf. Gαolf-GTP activates adenylyl cyclase III (AC3), which increases the intracellular cAMP concentration. Consequently, cyclic nucleotide-gated (CNG) channels are opened and cation concentrations, including calcium ions (Ca²⁺), transiently increase within the OSN. This influx of cations depolarizes the ciliary membrane and activates calcium-activated chloride channels, enabling chloride ions to efflux from the cell and further amplify the membrane depolarization, thus enhancing the odorant signaling. B. Open questions regarding OR signaling in cancer cells: (1) while ORs in OSNs can be activated by a wide range of odorants with diverse chemical structures, the endogenous activators of ORs in cancer cells remain unidentified; (2) it is not clear if ORs are targeted to the cell membrane in cancer cells; (3) the specific G protein subunits involved in OR signaling in cancer cells remain unknown; (4) although *in vitro* studies suggest that ORs activate pathways such as ERK, MAPK, AKT, and PI3K, their precise signaling mechanisms in cancer cells and tissues *in vivo* are not understood.; (5) it is unclear whether ORs have specific intracellular functions or are localized within particular cellular organelles; (6) current *in vitro* cancer cell line data suggest that multiple ORs can be expressed within a single cancer cell, unlike the one neuron - one receptor rule in the olfactory system. The mechanisms governing OR gene transcriptional regulation in cancer cells remain to be determined; (7) some tumors exhibit significant cellular heterogeneity, but the expression patterns of ORs across different tumor cell subtypes are unclear. Most studies analyze bulk tumor samples, highlighting the need for single-cell approaches to resolve this complexity.

Odorants are small (200-300 Da) organic volatile molecules which exhibit diverse chemical structures. Despite the reduced number of functional ORs when compared to other mammals, it is estimated that humans can detect thousands of odorants. This is possible because one OR can recognize multiple odorants and one odorant is recognized by multiple ORs in a combinatorial code [25,26]. However, the majority of ORs are still orphan receptors whose ligand(s) are unknown. This is mostly due to the fact that functional assays are difficult to perform with these receptors. While the first OR activation assays were performed *in vivo*, via calcium imaging and single-cell RT PCR of OSNs of mice exposed to certain odorants [26], today the main activation assays are conducted using heterologous systems. In these experiments, cell lines such as HEK293T are transfected to express ORs which are then activated by odorants. The cells are co-transfected with Gαolf, as well as some type of reporter gene which generally relies on the increase of AMPc caused by the OR activation. However, OR expression in this system has proven itself challenging, as ORs are not always trafficked to the plasmatic membrane and can frequently be trapped in the endoplasmic reticulum [27]. The addition of chaperones, such as RTP1S, can increase the functional expression of some ORs, although other ORs have an increased expression in the absence of RTP1S [28,29]. A tag of rhodopsin, consisting of the first 20 amino acids of the protein, can also be inserted at the amino-terminal end of the OR to increase its trafficking to the membrane without compromising its function [30]. To date, only 83 ORs are fully deorphanized, with effective dose-response curves traced in different concentrations of odorants, as gathered in M2OR, a database for odorant and OR pairs that includes the results from most OR activation assays [31]. Another 253 ORs have been activated by specific odorants in primary and secondary screenings, although these results could not be reproduced in the attempts of generating a dose-response curve [31].

Recently, it was described that extranasal ORs can be activated not only by a variety of odorant like molecules, such as short-chain fatty acids (SCFAs) as acetate and propionate [32]; but also by different types of molecules like the peptide hormones asprosin and insulin peptide [33,34]. Asprosin has specifically activated the mouse OR olfr734 in an heterologous cell activation model based on HEK293T cells by measuring the activity of CRE-Luc, a reporter for evaluating GPCR-mediated cAMP signaling [33]. This unexpected activation of ORs by a peptide suggests that extranasal ORs may be also activated by ligands that are structurally different from odorants, and probably through a different mechanism, than the one that requires binding of the ligand to the odorant transmembrane binding pocket [35].

GPCRs are the largest family of drug targets in clinical trials, representing 34% of FDA-approved drugs [36]. There are ~800 human GPCRs, and half of them are ORs. The consistent expression of ORs in various types of cancer, detected through diverse OMICs approaches, has highlighted their potential as novel therapeutic targets, as indicated by the increasing number of publications on the subject in recent years (Figure 2). In this review, we focus on the available data regarding ORs and their potential involvement in cancer.

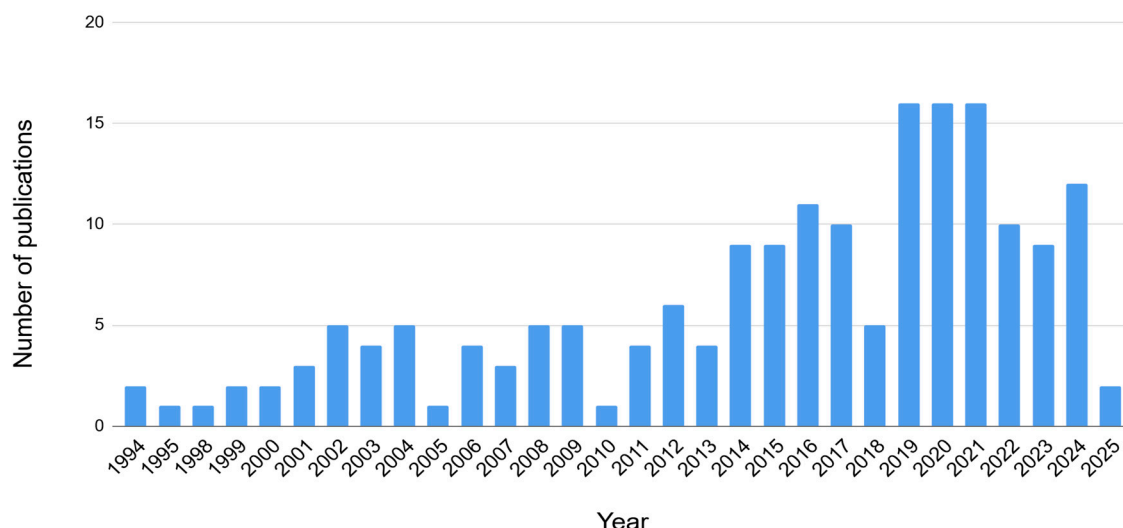


Figure 2. Emerging focus on olfactory receptors in cancer. The number of publications retrieved using the search query ["Olfactory receptor" + "cancer"] on the PubMed website as of February 14th, 2025.

2.1. ORs in Cancer

Curiously, although many cancer cell transcriptomes have ORs being expressed, they seldom express the same specific OR - on the contrary, it appears that each cancer has its own predominantly expressed OR. For example, a study investigated the expression of 301 OR genes in 968 cancer cell lines, and found that 332 of the tested cell lines expressed only one of the OR genes at levels considered to be above the background [37]. While some ORs had expression in cell lines of more than one type of cancer, such as OR4A467, OR1D5 and OR4C46, other ORs were expressed in a single tumor type, such as OR13A1, expressed in B cell malignancies and OR2C3, expressed in melanoma [37]. Most of the cell lines also expressed G protein alpha subunits and adenylyl cyclase III, indicating these receptors could in fact initiate a signaling pathway if present in the cell membrane [37].

The expression of ORs in cancer seems to surpass the "one neuron - one receptor" rule. Several breast cancer cell lines have 3 ORs being expressed at the same time, with HCC1396 having 112 OR genes expressed [4]. In the prostate, OR51E1 and OR51E2 are co-expressed in cell lines as well as patients, being super expressed in about 2/3 of tumour samples [38]. The same pattern can be seen in Acute Myeloid Leukemia (AML) patients, with 9 ORs expressed in a single patient [39]. Although the high number of ORs expressed could be due to different mutations leading to different profiles of cancerous cells in the same patient, the high percentage of cells expressing different ORs highly suggests that they are co-expressed in the same cell. This can be seen in AML, for example, with OR52H1 and OR52B6 being expressed in nearly 100% of The Cancer Genome Atlas (TCGA) patients' samples [39]. Altogether, that reinforces the possibility of a different mechanism of expression regulation in cancer cells, in comparison to the canonical monogenic expression of ORs in OSNs.

The OR gene family constitutes approximately 3% of the human genome, with around 400 functional and 600 OR pseudogenes [22,40,41]. Recent studies have highlighted the potential role of pseudogenes in various cellular processes, including tumorigenesis [17,42]. In our investigation on OR expression in AML [39], the most differentially expressed pseudogenes were OR7E128P and OR7E126P, both from the OR7E family. These findings align with the results of Flegel and colleagues who investigated the expression profile of ORs among 16 human tissues and found that 58% of the 62 ORs pseudogenes found expressed belong to the OR7E subfamily [43].

Both OR7E126P and OR7E128P genes are located within the 11q13.4 breakpoint interval on chromosome 11, home to the largest OR gene cluster, containing 43% of the OR genes in the human genome [22,44,45]. Chromosomal rearrangements involving chromosome 11, particularly in the

KMT2A gene, are common in acute leukemias [46]. While OR genes have been implicated in certain known translocations, such as t(4;11) [47], a recent study suggests that, despite their co-localization, ORs do not directly mediate structural rearrangements of chromosome 11. Instead, it is proposed that these alterations may arise from distinct DNA motifs and recombination mechanisms, whether based on homology or non-homology, which drive the chromosomal instability [48]. Other studies have identified the involvement of OR pseudogenes in lung squamous cell carcinoma and glioblastoma [17,49].

Current data suggest that ORs may play a role in four of the eight hallmarks of cancer [50]: proliferation; angiogenesis; invasion and metastasis; and cell death. Depending on the cancer type and context, ORs can either positively or negatively regulate these pathways. The signaling pathways activated by ORs in cancer differ from traditional olfactory signaling depending on the cell type. The most associated pathways include MAPK/ERK, PLC-IP3/Ca²⁺; PKA-CREB-HES1; PI3K-AKT; Gβ-γ-PI3Kγ-ARF1; MAPK p38 and SAPK/JNK [15,38,51–55].

In the next section, we will describe some ORs that were identified in different cancer types (Table 1). Some ORs exhibited a significant correlation between their expression and patient survival rates five years later, as shown in the overall survival rates (OS) column of Table 1. This association suggests that OR expression may have potential as a prognostic factor.

Table 1. List of cancer associated ORs.

Cancer	OR	OS*	Ref
Acute Myeloid Leukemia (AML)	OR2AE1, OR10A2, OR2G2	Lower	[39]
	OR5C1, OR1L6, OR10A4, OR13F1	Higher	[39]
	OR52B6, OR2L3, OR52H1, OR2L5, OR2AK2, OR13D1, OR9A4, OR52K2, OR52K1, OR2G3, OR2B2, OR10A5	-	[39]
Adrenocortical carcinoma (ACC)	OR7A5	Higher	[61]
	OR7A5	Higher	[61]
	OR2W3, OR2B6	Lower	[4]
	OR5B21	Higher	[57]
Breast cancer (BRCA)	OR6M1	-	[55]
	OR2T6	-	[14]
	OR51J1	-	[58]
	OR3A4	-	[59]
Chronic Myeloid Leukemia (CML)	OR2AT4	-	[52,60]
Colorectal Cancer (CRC)	OR3A4	-	[56]
Diffuse Large B-Cell Lymphoma (DLBCL)	OR3A4	-	[61]
Esophageal Cancer (EC)	OR3A4	-	[56]
Gallbladder Cancer (GBC)	OR3A4	-	[56]
Gastric Cancer (GC)	OR3A4	-	[56]

	OR7A5	Lower	[62]
Glioblastoma multiforme (GBM)	OR7E156P	-	[49]
	OR7D2, OR7E14P, OR4N2	-	[49]
Hepatocellular carcinoma	OR3A4	-	[63]
Kidney renal clear cell carcinoma (KIRC)	OR7A5	Lower	[62]
Kidney renal papillary cell carcinoma (KIRP)	OR7A5	Lower	[62]
Liver hepatocellular carcinoma (HCC)	OR7A5	Higher	[62]
	OR1A2	-	[51]
Low-Grade Glioma (LGG)	OR7A5	Lower	[62]
Lung adenocarcinoma (LUAD)	OR7A5	Lower	[62]
Lung squamous cell carcinoma (LUSC)	OR7A5	Higher	[62]
	OR51E1	-	[64]
	OR2C3	-	[37]
Malignant Melanoma (MM)	OR51E2	-	[3]
Non-small cell lung cancer (NSCLC)	OR2J3	-	[54]
Osteosarcoma	OR3A4	-	[65]
Ovarian Cancer	OR3A4	-	[66]
Ovarian Serous Cystadenocarcinoma (OSA)	OR7A5	Lower	[62]
	OR13C4	-	[67]
Pancreas (PDAC)	OR3A4	-	[56]
	OR51E1	-	[38,53]
Prostate (PCA)	OR51E2	-	[38]
Small intestine neuroendocrine carcinomas (SI-NEC)	OR51E1	-	[68]
Thymoma (THYM)	OR7A5	Higher	[62]
Thyroid cancer (THCA)	OR7A5	Higher	[62]
Urinary Bladder Cancer (BLCA)	OR10H1	-	[69]
Uterine Corpus Endometrial Carcinoma (UCEC)	OR7A5	Lower	[62]

*OS = Overall Survival in five years.

2.2.1. Prostate Cancer

Prostate cancer was one of the first to have an OR identified as up-regulated [70]. Initially thought to be a different type of GPCR, OR51E2 is also known as PSGR (prostate-specific G protein

coupled receptor). Besides OR51E2, another OR, the OR51E1, is also highly expressed in prostate cancer. Both ORs have an at least 10 times higher expression in 2 out of 3 prostate cancer tumors in comparison to a healthy prostate [38]. However, the super expression of OR51E1 alone leads to ERK1/2 phosphorylation and ultimately to cell cycle arrest via activation of p53 [38].

While OR51E1 is activated by longer aliphatic acids (3C-6C), OR51E2 is activated by shorter fatty acids, such as acetate and propionate [38]. OR51E2 is also activated by beta-ionone as well as by steroids such as ADT and 19OH AD, and inhibited by alpha-ionone [71]. Its activation was shown to induce the phosphorylation of MAPK p38 and SAPK/JNK, ultimately leading to a 50% reduction of cell proliferation [71,72]. Additionally, it has been shown in two prostate cancer cell lines, DU145 and LNCaP, that the activation of OR51E2 by beta-ionone was also able to lead to the activation of ERK1/2, via the $G\beta\gamma$ and PI3K γ signaling pathway. The pharmacological inhibition of $G\beta\gamma$ and PI3K γ signaling reduced metastasis, even with the activation of OR51E2, suggesting an important role of $G\beta\gamma$ and PI3K γ in mediating OR51E2 function in prostate cancer cells. [16].

2.2.2. Breast Cancer

In breast cancer, different odorants (citral, cyclovertal and citrathal R), both individually and in a mixture, were able to reduce the expression of the Ki67 proliferation marker in three cell lines, besides inducing the cells to undergo apoptosis through a higher expression of p53 [73]. Citral and cyclovertal were also able to activate signaling pathways related to cell growth and survival, by decreasing the expression of ERK1 and 2, and p38-MAPK [73].

Three ORs have been associated with different subtypes of breast cancer [4,57,73]. OR2W3 is correlated with stage IV breast cancer, which is slightly more prevalent in the triple negative subtype and in the basal-like subtype, known to be more aggressive. This receptor was highly correlated with genes of tumor invasion, such as CTSV and MMP11 [4]. Moreover, OR2W3 has been associated with shorter survival, of only 35% after 150 months against 85% after 150 months in patients with a normal expression of the OR. Therefore, this OR could be used as a prognostic marker for the subtypes where it is present [4].

OR5B21, on the other hand, is highly expressed in breast cancer metastasis, especially in the brain but also in bones and lungs [57]. In assays where OR5B21 was knocked down, the cells showed less invasion and migration capacity, through the inhibition of STAT3 and NF- κ B p65 phosphorylation. Although proliferation was not altered, there was a reduction in metastasis and an overall better survival rate [57]. Finally, OR2B6 has been correlated to the breast cancer of subtype Luminal A, a less aggressive subtype [4]. Still, it is correlated to cell proliferation markers, such as MKI67, MYBL2 and CCNB1 [4].

Lastly, OR6M1, expressed in MCF-7 cells, a human cancer cell line derived from estrogen receptor (ER)-positive breast cancer, was shown to be activated by antroquinone in concentrations of 50 and 100 μ M, ultimately leading to a decrease in cell viability as well as cell death [55].

2.2.3. Leukemia

Different types of leukemia have exhibited altered expression of ORs. In chronic myeloid leukemia (CML), OR2AT4 was found to be highly expressed in the K562 cell line, and its activation by sandalore was able to induce apoptosis and differentiation, as well as leading to a reduction of proliferation [52]. Another study with the same cell line showed that the same OR can be activated by (-)-epigallocatechin gallate (EGCG), leading to a reduction in cell viability as well as extrinsic apoptosis via Caspase-3 and 8 [60]. A decrease in phosphorylation of proteins related to cell proliferation was also shown following the activation, especially in AKT, p38-MAPK and STAT5. The same assay was conducted in a cell line derived from an acute lymphoblastic leukemia (ALL) patient, MOLT-4, which does not express OR2AT4, and none of the previous mentioned results were obtained, indicating these results are OR dependent [60]. Other non-olfactory GPCRs have been described to be correlated to subtypes of AML, and it is possible that some ORs could also be

associated with different AML subtypes, which were probably filtered out from this study due to their relative low expression levels [74].

Analysis of the TCGA cohort brought up 19 ORs enriched in AML in comparison to healthy blood samples, with 16 of them being validated in an independent AML cohort, the BEAT AML cohort [39]. Of these, OR52B6, OR52H1, OR2AK2, OR13D1 and OR52K2 were found in 90% of the samples. Given the different subtypes of AML, these ORs could be valuable markers or even drug targets for the disease, independently of the subtype, since these ORs also did not show a considerable expression in any of the healthy tissue samples available on Genotype-Tissue Expression Project (GTEx). Moreover, these ORs seem to be AML specific, since they do not show significant expression in other TCGA cancer samples. Besides, 3 of the 16 identified ORs (OR2G2, OR2AE1 and OR10A2) were correlated with worse survival rates [39].

2.2.4. Melanoma

In melanoma cell lines, OR2C3 stands out by having a higher expression among all other ORs in 8 out of 52 of the tested cell lines [37]. The same OR can be found expressed in melanoma samples from patients derived from TCGA, although it is expressed in only 14 out of the 474 samples. Still, none of the healthy tissues available on GTEx had such high expression of this gene [37].

Another OR, OR51E2, was fourfold upregulated in metastatic melanoma cells, but not on other dermal tissue cells around the cancer [3]. *In vitro* assays showed an OR51E2-dependent increase in intracellular Ca^{2+} in melanoma primary cell cultures in presence of β -ionone. This led to a dose-dependent decrease in cell number through apoptosis, besides inhibition of cell migration [3].

2.2.5. Gliomas

In glioblastoma (GBM), more than 9 ORs have been reported as having differential expression when compared to healthy tissues [49]. ORs such as OR2B6 and OR51E1, had a higher expression in GBM samples when compared to low-grade glioma (LGG) [49]. OR7D2 was expressed in all malignant stages of GBM, regardless of sample heterogeneity. OR4N2 was more frequent in malign cells, and is likely involved in the tumoral plasticity mechanism, promoting GBM adaptation and resistance. OR51E1 was highly expressed in pericytes in the tumor microenvironment, likely related to angiogenesis and contributing to the tumor microenvironment (TME) remodeling. OR2B11 was also found expressed in the TME, but specifically associated with TME macrophages, as well as in the mesenchymal subtype of GBM. It is believed that OR2B11 has a role in the modulation of an immunosuppressant TME, via NF- κ B/TGF- β signaling, which could lead to immunotherapy resistance. Finally, OR2L13 was found expressed predominantly in oligodendrocytes and neurons, and could contribute to tumor recurrence and resistance. Nonetheless, two OR pseudogenes also had significant expression in GBM. OR7E156P had been associated with miR-143/HIF1A, known to promote tumor growth and invasion, thus favoring GBM progression. OR7E14P, on the other hand, was enriched in MES-like GBM state and could be related to the subtype aggressiveness as well as tumor resistance [49].

In another study, a few more ORs were found to be upregulated both in LGG and GBM, including OR2A9P, another pseudogene [62]. OR7A5 was linked to a worse prognostic and lower overall survival in five years, being considered as an oncogene and involved in the initiation and progression of the disease with an increase in *in vitro* cell proliferation depending on the expression of the receptor. The OR was found to modulate lipids metabolism possibly through pathways such as cAMP/HSL, cAMP/CREB and cAMP/MAPK, which could affect the malignant progression of the tumour [62].

2.2.5. Gastric Cancer

Gastric cancer is the third most frequent cancer to lead to the death of patients, because its diagnosis often happens in advanced stages of the disease leading to poor prognosis [56]. Although

not functionally expressed, lncRNA OR3A4 was found to affect gastric cancer [56]. The lncRNA OR was super expressed in primary tissues as well as metastasis tissues and in the blood of patients. Its presence in the circulating blood poses the lncRNA OR as an interesting potential biomarker to the disease, with its detection reaching 86.94% sensitivity and 91.27% specificity in patients, which could enhance early diagnosis of the cancer and, thus, better prognosis. High levels of lncRNA OR3A4 were related to shorter OS, shorter interval before relapse and, most importantly, highly associated with both metastasis and angiogenesis. *In vitro*, lncRNA OR3A4 super expression led to significant increase of both proliferation and cell invasion and migration, while its silencing reduced all of them [56].

Curiously, the lncRNA OR3A4 is expressed in other types of cancer, such as in breast cancer, ovarian cancer, hepatocellular carcinoma, colorectal cancer, non-small cell lung cancer, osteosarcoma and in diffuse large B-cell lymphoma (DLBCL) [56,59,61,63,65,66]. In DLBCL patients, the FOXM1 induced the upregulation of lncRNA OR3A4 and patients with high lncRNA OR3A4 expression presented poor prognosis. Moreover, the same study showed that the knockdown of OR3A4 suppressed cell proliferation and promoted cell apoptosis in DLBCL by inactivating the Wnt/ β -catenin signaling pathway [61].

2.3. ORs and Tumor Microenvironment

OR2B11, OR52K1 and OR3A2 were found not only in GBM but in its TME, with a possible role in the promotion of tumorigenicity [49]. Besides, OR2B11 was also found in macrophages of the TME associated with the tumor, and could have an important role in the modulation of the TME to sustain tumor growth. OR2B11 has also been related as having a high expression in macrophages in bone marrow and microglia. In the mesenchymal subtype of GBM, it is believed that OR2B11 could promote mesenchymal transition, contributing to an aggressive TME and therefore associated with a lower survival rate and decreased response to immunotherapy [49].

In mice, Olfr78 was expressed in macrophages derived from the blood marrow [75]. Curiously, this OR could be activated by the lactate found in TME because of the Warburg effect. Tumor associated macrophages (TAMs) can be categorized in two subtypes: M1, antitumoral and M2, tumoral. The activation of Olfr78 by lactate seems to induce the transformation of TAM into an M2 phenotype. In the absence of Olfr78, this effect was not seen in the presence of lactate. This could explain why lower expression of the human Olfr78 counterpart OR51E2, in cancers such as breast, glioblastoma and lung adenocarcinoma, is correlated with longer survival rates [75].

2.4. ORs and Angiogenesis

Angiogenesis is essential, especially to solid tumors, for the oxygen transportation during cancer development. OR51E1 has been found in endothelial cells as well as cells in the pericyte, besides being correlated with genes of vascular remodeling and angiogenesis, and could have a vascular function [49]. In gastric cancer, OR3A4 was found to modulate the expression of pro-angiogenesis factors such as VEGF-C and MMP9, with its super expression increasing significantly the expression of both genes while its silencing led to a reduction of their expression. Also, the same OR had the capacity of regulating the activity of other genes involved in angiogenesis, such as *NTN4* [56].

3. Perspectives

Despite the potential of ORs as therapeutic targets for various cancers, several challenges explain why, although 40% of all drugs target GPCRs, none currently target ORs (Figure 1B).

A primary challenge is the lack of specific agonists for most ORs, which remain "orphan" receptors. Outside the olfactory system, endogenous ligands with different structures when compared to that of odorants, may also bind to and activate the extranasal ORs. As described above, hormone peptides, which are larger in size than odorants, have also been described as being able to activate ORs [33,34]. However, the previously mentioned difficulty of achieving heterologous

expression of these receptors limits the identification of potential ligands. Additionally, for a ligand to be used as an activator for an OR in cancer, it would have to be specific to that given OR. In the olfactory system, odorants are known for being able to activate more than one OR [26]. Given the expression of several ORs throughout the human body, a non-specific ligand could trigger unforeseen effects outside the cancer cells.

While ORs are expressed in certain cancers, the absence of viable antibodies complicates the validation of their functional presence in the membrane — in OSNs and in *in vitro* heterologous systems, this process is dependent on specific chaperone proteins. Still, it is uncertain whether the functional expression is necessary for ORs to have a role in cancer, as shown from the example described in the gastric cancer session of this review, where a superexpression of the lncRNA OR3A4 leads to increase in cell proliferation, migration and invasion. Additionally, certain OR pseudogenes may play a role in lung cancer and other malignancies [17,42].

Despite these hurdles, the abnormal and tumor-specific expression of extranasal ORs offers a valuable tool for precise cancer diagnosis, prognosis, and patient stratification. Targeted expression of these receptors could also assist in measuring minimal residual disease (MRD) in patients undergoing treatment, since many cancers have no molecular marker to perform the molecular follow up. Furthermore, this specificity has been exploited for therapeutic purposes, such as using chimeric antigen receptor (CAR)-expressing T cells to target OR2H1 expressing tumor cells both *in vitro* and *in vivo* in mice [76]. While CAR T cell therapy has shown relative success in treating hematological malignancies, it has not yet proven effective for solid tumors. The recombinant OR2H1 IgG generated in this study specifically detected the OR2H1 protein in 60 human lung cancers, 40 ovarian carcinomas, and 73 cholangiocarcinomas, offering a promising new therapeutic approach for epithelial cancers [76].

In conclusion, advancing our understanding of extranasal ORs in both healthy tissues and tumors is crucial for enhancing cancer treatment strategies and modulating cellular processes to better control cancer progression.

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Abbreviations

The following abbreviations are used in this manuscript:

OR	Odorant receptor
GPCR	G protein coupled receptor
OSN	Olfactory sensory neuron
AML	Acute Myeloid Leukemia
TME	Tumor microenvironment
GBM	Glioblastoma
DLBCL	Diffuse large B-cell lymphoma
TAM	Tumor associated macrophages
LGG	Low-grade glioma
OS	Overall Survival

References

1. Parmentier, M., Libert, F., Schurmans, S., Schiffmann, S., Lefort, A., Eggerickx, D., Ledent, C., Mollereau, C., Gerard, C., Perret, J., et al. Expression of members of the putative olfactory receptor gene family in mammalian germ cells. *Nature* 1992, 355, 453-455, doi:10.1038/355453a0.
2. Spehr, M., Gisselmann, G., Poplawski, A., Riffell, J. A., Wetzell, C. H., Zimmer, R. K. and Hatt, H. Identification of a testicular odorant receptor mediating human sperm chemotaxis. *Science* 2003, 299, 2054-2058, doi:10.1126/science.1080376.
3. Gelis, L., Jovancevic, N., Bechara, F. G., Neuhaus, E. M. and Hatt, H. Functional expression of olfactory receptors in human primary melanoma and melanoma metastasis. *Exp Dermatol* 2017, 26, 569-576, doi:10.1111/exd.13316.
4. Masjedi, S., Zwiebel, L. J. and Giorgio, T. D. Olfactory receptor gene abundance in invasive breast carcinoma. *Sci Rep* 2019, 9, 13736, doi:10.1038/s41598-019-50085-4.
5. Orecchioni, M., Kobiyama, K., Winkels, H., Ghosheh, Y., McArdle, S., Mikulski, Z., Kiesses, W. B., Fan, Z., Wen, L., Jung, Y., et al. Olfactory receptor 2 in vascular macrophages drives atherosclerosis by NLRP3-dependent IL-1 production. *Science* 2022, 375, 214-221, doi:10.1126/science.abg3067.
6. Poll, B. G., Xu, J., Gupta, K., Shubitowski, T. B. and Pluznick, J. L. Olfactory receptor 78 modulates renin but not baseline blood pressure. *Physiol Rep* 2021, 9, e15017, doi:10.14814/phy2.15017.
7. Xu, J., Choi, R., Gupta, K., Warren, H. R., Santhanam, L. and Pluznick, J. L. An evolutionarily conserved olfactory receptor is required for sex differences in blood pressure. *Sci Adv* 2024, 10, eadk1487, doi:10.1126/sciadv.adk1487.
8. Yang, Z., Cheng, J., Shang, P., Sun, J. P. and Yu, X. Emerging roles of olfactory receptors in glucose metabolism. *Trends Cell Biol* 2023, 33, 463-476, doi:10.1016/j.tcb.2022.09.005.
9. Tong, T., Ryu, S. E., Min, Y., de March, C. A., Bushdid, C., Golebiowski, J., Moon, C. and Park, T. Olfactory receptor 10J5 responding to alpha-cedrene regulates hepatic steatosis via the cAMP-PKA pathway. *Sci Rep* 2017, 7, 9471, doi:10.1038/s41598-017-10379-x.
10. Chang, A. J., Ortega, F. E., Riegler, J., Madison, D. V. and Krasnow, M. A. Oxygen regulation of breathing through an olfactory receptor activated by lactate. *Nature* 2015, 527, 240-244, doi:10.1038/nature15721.
11. Colinas, O., Mombaerts, P., Lopez-Barneo, J. and Ortega-Saenz, P. Carotid Body Function in Tyrosine Hydroxylase Conditional Olfr78 Knockout Mice. *Function (Oxf)* 2024, 5, zqae010, doi:10.1093/function/zqae010.
12. Fukuda, N., Yomogida, K., Okabe, M. and Touhara, K. Functional characterization of a mouse testicular olfactory receptor and its role in chemosensing and in regulation of sperm motility. *J Cell Sci* 2004, 117, 5835-5845, doi:10.1242/jcs.01507.
13. Flegel, C., Vogel, F., Hofreuter, A., Schreiner, B. S., Osthold, S., Veitinger, S., Becker, C., Brockmeyer, N. H., Muschol, M., Wennemuth, G., et al. Characterization of the Olfactory Receptors Expressed in Human Spermatozoa. *Front Mol Biosci* 2015, 2, 73, doi:10.3389/fmolb.2015.00073.
14. Li, M., Wang, X., Ma, R. R., Shi, D. B., Wang, Y. W., Li, X. M., He, J. Y., Wang, J. and Gao, P. The Olfactory Receptor Family 2, Subfamily T, Member 6 (OR2T6) Is Involved in Breast Cancer Progression via Initiating Epithelial-Mesenchymal Transition and MAPK/ERK Pathway. *Front Oncol* 2019, 9, 1210, doi:10.3389/fonc.2019.01210.
15. Rodriguez, M., Siwko, S., Zeng, L., Li, J., Yi, Z. and Liu, M. Prostate-specific G-protein-coupled receptor collaborates with loss of PTEN to promote prostate cancer progression. *Oncogene* 2016, 35, 1153-1162, doi:10.1038/onc.2015.170.
16. Xu, X., Khater, M. and Wu, G. The olfactory receptor OR51E2 activates ERK1/2 through the Golgi-localized Gbetagamma-PI3Kgamma-ARF1 pathway in prostate cancer cells. *Front Pharmacol* 2022, 13, 1009380, doi:10.3389/fphar.2022.1009380.
17. Zhao, Y. Q., Zhang, H. H., Wu, J., Li, L., Li, J., Zhong, H., Jin, Y., Lei, T. Y., Zhao, X. Y., Xu, B., et al. Prediction of Tumor Microenvironment Characteristics and Treatment Response in Lung Squamous Cell Carcinoma by Pseudogene OR7E47P-related Immune Genes. *Curr Med Sci* 2023, 43, 1133-1150, doi:10.1007/s11596-023-2798-2.

18. Weber, L., Al-Refae, K., Ebbert, J., Jagers, P., Altmüller, J., Becker, C., Hahn, S., Gisselmann, G. and Hatt, H. Activation of odorant receptor in colorectal cancer cells leads to inhibition of cell proliferation and apoptosis. *PLoS One* 2017, 12, e0172491, doi:10.1371/journal.pone.0172491.
19. Naressi, R. G., Schechtman, D. and Malnic, B. Odorant receptors as potential drug targets. *Trends Pharmacol Sci* 2023, 44, 11-14, doi:10.1016/j.tips.2022.08.003.
20. Buck, L. and Axel, R. A novel multigene family may encode odorant receptors: a molecular basis for odor recognition. *Cell* 1991, 65, 175-187, doi:10.1016/0092-8674(91)90418-x.
21. Zhang, X. and Firestein, S. The olfactory receptor gene superfamily of the mouse. *Nat Neurosci* 2002, 5, 124-133, doi:10.1038/nn800.
22. Malnic, B., Godfrey, P. A. and Buck, L. B. The human olfactory receptor gene family. *Proc Natl Acad Sci U S A* 2004, 101, 2584-2589, doi:10.1073/pnas.0307882100.
23. Monahan, K. and Lomvardas, S. Monoallelic expression of olfactory receptors. *Annu Rev Cell Dev Biol* 2015, 31, 721-740, doi:10.1146/annurev-cellbio-100814-125308.
24. Nagai, M. H., Armelin-Correa, L. M. and Malnic, B. Monogenic and Monoallelic Expression of Odorant Receptors. *Mol Pharmacol* 2016, 90, 633-639, doi:10.1124/mol.116.104745.
25. Kurian, S. M., Naressi, R. G., Manoel, D., Barwich, A. S., Malnic, B. and Saraiva, L. R. Odor coding in the mammalian olfactory epithelium. *Cell Tissue Res* 2021, 383, 445-456, doi:10.1007/s00441-020-03327-1.
26. Malnic, B., Hirono, J., Sato, T. and Buck, L. B. Combinatorial receptor codes for odors. *Cell* 1999, 96, 713-723, doi:10.1016/s0092-8674(00)80581-4.
27. Gimelbrant, A. A., Haley, S. L. and McClintock, T. S. Olfactory receptor trafficking involves conserved regulatory steps. *J Biol Chem* 2001, 276, 7285-7290, doi:10.1074/jbc.M005433200.
28. Saito, H., Kubota, M., Roberts, R. W., Chi, Q. and Matsunami, H. RTP family members induce functional expression of mammalian odorant receptors. *Cell* 2004, 119, 679-691, doi:10.1016/j.cell.2004.11.021.
29. Sharma, R., Ishimaru, Y., Davison, I., Ikegami, K., Chien, M. S., You, H., Chi, Q., Kubota, M., Yohda, M., Ehlers, M., et al. Olfactory receptor accessory proteins play crucial roles in receptor function and gene choice. *Elife* 2017, 6, doi:10.7554/eLife.21895.
30. Zhuang, H. and Matsunami, H. Synergism of accessory factors in functional expression of mammalian odorant receptors. *J Biol Chem* 2007, 282, 15284-15293, doi:10.1074/jbc.M700386200.
31. Lalis, M., Hladis, M., Khalil, S. A., Briand, L., Fiorucci, S. and Topin, J. M2OR: a database of olfactory receptor-odorant pairs for understanding the molecular mechanisms of olfaction. *Nucleic Acids Res* 2024, 52, D1370-D1379, doi:10.1093/nar/gkad886.
32. Fleischer, J., Bumbalo, R., Bautze, V., Strotmann, J. and Breer, H. Expression of odorant receptor Olfr78 in enteroendocrine cells of the colon. *Cell Tissue Res* 2015, 361, 697-710, doi:10.1007/s00441-015-2165-0.
33. Li, E., Shan, H., Chen, L., Long, A., Zhang, Y., Liu, Y., Jia, L., Wei, F., Han, J., Li, T., et al. OLFR734 Mediates Glucose Metabolism as a Receptor of Asprosin. *Cell Metab* 2019, 30, 319-328 e318, doi:10.1016/j.cmet.2019.05.022.
34. Cheng, J., Yang, Z., Ge, X. Y., Gao, M. X., Meng, R., Xu, X., Zhang, Y. Q., Li, R. Z., Lin, J. Y., Tian, Z. M., et al. Autonomous sensing of the insulin peptide by an olfactory G protein-coupled receptor modulates glucose metabolism. *Cell Metab* 2022, 34, 240-255 e210, doi:10.1016/j.cmet.2021.12.022.
35. Billesbølle, C. B., de March, C. A., van der Velden, W. J. C., Ma, N., Tewari, J., Del Torrent, C. L., Li, L., Faust, B., Vaidehi, N., Matsunami, H., et al. Structural basis of odorant recognition by a human odorant receptor. *Nature* 2023, 615, 742-749, doi:10.1038/s41586-023-05798-y.
36. Hauser, A. S., Attwood, M. M., Rask-Andersen, M., Schiøth, H. B. and Gloriam, D. E. Trends in GPCR drug discovery: new agents, targets and indications. *Nat Rev Drug Discov* 2017, 16, 829-842, doi:10.1038/nrd.2017.178.
37. Ranzani, M., Iyer, V., Ibarra-Soria, X., Del Castillo Velasco-Herrera, M., Garnett, M., Logan, D. and Adams, D. J. Revisiting olfactory receptors as putative drivers of cancer. *Wellcome Open Res* 2017, 2, 9, doi:10.12688/wellcomeopenres.10646.1.
38. Pronin, A. and Slepak, V. Ectopically expressed olfactory receptors OR51E1 and OR51E2 suppress proliferation and promote cell death in a prostate cancer cell line. *J Biol Chem* 2021, 296, 100475, doi:10.1016/j.jbc.2021.100475.

39. Guardia, G. D. A., Naressi, R. G., Buzzato, V. C., da Costa, J. B., Zalcborg, I., Ramires, J., Malnic, B., Gutiyama, L. M. and Galante, P. A. F. Acute Myeloid Leukemia Expresses a Specific Group of Olfactory Receptors. *Cancers (Basel)* 2023, 15, doi:10.3390/cancers15123073.
40. Glusman, G., Yanai, I., Rubin, I. and Lancet, D. The complete human olfactory subgenome. *Genome Res* 2001, 11, 685-702, doi:10.1101/gr.171001.
41. Niimura, Y. and Nei, M. Evolution of olfactory receptor genes in the human genome. *Proc Natl Acad Sci U S A* 2003, 100, 12235-12240, doi:10.1073/pnas.1635157100.
42. Nakamura-Garcia, A. K. and Espinal-Enriquez, J. Pseudogenes in Cancer: State of the Art. *Cancers (Basel)* 2023, 15, doi:10.3390/cancers15164024.
43. Flegel, C., Manteniotis, S., Osthold, S., Hatt, H. and Gisselmann, G. Expression profile of ectopic olfactory receptors determined by deep sequencing. *PLoS One* 2013, 8, e55368, doi:10.1371/journal.pone.0055368.
44. Buettner, J. A., Glusman, G., Ben-Arie, N., Ramos, P., Lancet, D. and Evans, G. A. Organization and evolution of olfactory receptor genes on human chromosome 11. *Genomics* 1998, 53, 56-68, doi:10.1006/geno.1998.5422.
45. Olender, T., Feldmesser, E., Atarot, T., Eisenstein, M. and Lancet, D. The olfactory receptor universe--from whole genome analysis to structure and evolution. *Genet Mol Res* 2004, 3, 545-553.
46. Meyer, C., Larghero, P., Almeida Lopes, B., Burmeister, T., Groger, D., Sutton, R., Venn, N. C., Cazzaniga, G., Corral Abascal, L., Tsaur, G., et al. The KMT2A recombinoome of acute leukemias in 2023. *Leukemia* 2023, 37, 988-1005, doi:10.1038/s41375-023-01877-1.
47. Ou, Z., Stankiewicz, P., Xia, Z., Breman, A. M., Dawson, B., Wiszniewska, J., Szafranski, P., Cooper, M. L., Rao, M., Shao, L., et al. Observation and prediction of recurrent human translocations mediated by NAHR between nonhomologous chromosomes. *Genome Res* 2011, 21, 33-46, doi:10.1101/gr.111609.110.
48. Redaelli, S., Grati, F. R., Tritto, V., Giannuzzi, G., Recalcatti, M. P., Sala, E., Villa, N., Crosti, F., Roversi, G., Malvestiti, F., et al. Olfactory receptor genes and chromosome 11 structural aberrations: Players or spectators? *HGG Adv* 2024, 5, 100261, doi:10.1016/j.xhgg.2023.100261.
49. Cho, H. J., Yeo, D. J., Yang, H. and Koo, J. Comprehensive Transcriptomic Analysis Reveals Cell-Type-Specific Roles of Human Odorant Receptors in Glioblastoma and the Tumor Microenvironment. *Int J Mol Sci* 2024, 25, doi:10.3390/ijms252413382.
50. Hanahan, D. Hallmarks of Cancer: New Dimensions. *Cancer Discov* 2022, 12, 31-46, doi:10.1158/2159-8290.CD-21-1059.
51. Massberg, D., Simon, A., Haussinger, D., Keitel, V., Gisselmann, G., Conrad, H. and Hatt, H. Monoterpene (-)-citronellal affects hepatocarcinoma cell signaling via an olfactory receptor. *Arch Biochem Biophys* 2015, 566, 100-109, doi:10.1016/j.abb.2014.12.004.
52. Manteniotis, S., Wojcik, S., Brauhoff, P., Mollmann, M., Petersen, L., Gothert, J. R., Schmiegel, W., Duhrsen, U., Gisselmann, G. and Hatt, H. Functional characterization of the ectopically expressed olfactory receptor 2AT4 in human myelogenous leukemia. *Cell Death Discov* 2016, 2, 15070, doi:10.1038/cddiscovery.2015.70.
53. Massberg, D., Jovancevic, N., Offermann, A., Simon, A., Baniahmad, A., Perner, S., Pungsrinont, T., Luko, K., Philippou, S., Ubrig, B., et al. The activation of OR51E1 causes growth suppression of human prostate cancer cells. *Oncotarget* 2016, 7, 48231-48249, doi:10.18632/oncotarget.10197.
54. Kalbe, B., Schulz, V. M., Schlimm, M., Philippou, S., Jovancevic, N., Jansen, F., Scholz, P., Lubbert, H., Jarocki, M., Faissner, A., et al. Helional-induced activation of human olfactory receptor 2J3 promotes apoptosis and inhibits proliferation in a non-small-cell lung cancer cell line. *Eur J Cell Biol* 2017, 96, 34-46, doi:10.1016/j.ejcb.2016.11.004.
55. Choi, Y. R., Shim, J., Park, J. H., Kim, Y. S. and Kim, M. J. Discovery of Orphan Olfactory Receptor 6M1 as a New Anticancer Target in MCF-7 Cells by a Combination of Surface Plasmon Resonance-Based and Cell-Based Systems. *Sensors (Basel)* 2021, 21, doi:10.3390/s21103468.
56. Guo, X., Yang, Z., Zhi, Q., Wang, D., Guo, L., Li, G., Miao, R., Shi, Y. and Kuang, Y. Long noncoding RNA OR3A4 promotes metastasis and tumorigenicity in gastric cancer. *Oncotarget* 2016, 7, 30276-30294, doi:10.18632/oncotarget.7217.

57. Asadi, M., Ahmadi, N., Ahmadvand, S., Jafari, A. A., Safaei, A., Erfani, N. and Ramezani, A. Investigation of olfactory receptor family 51 subfamily j member 1 (OR51J1) gene susceptibility as a potential breast cancer-associated biomarker. *PLoS One* 2021, 16, e0246752, doi:10.1371/journal.pone.0246752.
58. Liu, G., Hu, X. and Zhou, G. Long non-coding RNA OR3A4 promotes proliferation and migration in breast cancer. *Biomed Pharmacother* 2017, 96, 426-433, doi:10.1016/j.biopha.2017.10.011.
59. Choi, Y. R., Na, H. J., Lee, J. A., Kim, Y., Kim, Y. S. and Kim, M. J. Discovery of (-)-epigallocatechin gallate, a novel olfactory receptor 2AT4 agonist that regulates proliferation and apoptosis in leukemia cells. *Heliyon* 2024, 10, e30298, doi:10.1016/j.heliyon.2024.e30298.
60. Meng, H., Zhao, B. and Wang, Y. FOXM1-induced upregulation of lncRNA OR3A4 promotes the progression of diffuse large B-cell lymphoma via Wnt/beta-catenin signaling pathway. *Exp Mol Pathol* 2020, 115, 104451, doi:10.1016/j.yexmp.2020.104451.
61. Bao, Y., Tang, Z., Chen, R., Yu, X. and Qi, X. Pan-cancer analysis identifies olfactory receptor family 7 subfamily A member 5 as a potential biomarker for glioma. *PeerJ* 2024, 12, e17631, doi:10.7717/peerj.17631.
62. Li, W., Fu, Q., Man, W., Guo, H. and Yang, P. LncRNA OR3A4 participates in the angiogenesis of hepatocellular carcinoma through modulating AGGF1/akt/mTOR pathway. *Eur J Pharmacol* 2019, 849, 106-114, doi:10.1016/j.ejphar.2019.01.049.
63. Giandomenico, V., Cui, T., Grimelius, L., Oberg, K., Pelosi, G. and Tsolakis, A. V. Olfactory receptor 51E1 as a novel target for diagnosis in somatostatin receptor-negative lung carcinoids. *J Mol Endocrinol* 2013, 51, 277-286, doi:10.1530/JME-13-0144.
64. Wang, X., Chen, K. and Zhao, Z. LncRNA OR3A4 Regulated the Growth of Osteosarcoma Cells by Modulating the miR-1207-5p/G6PD Signaling. *Onco Targets Ther* 2020, 13, 3117-3128, doi:10.2147/OTT.S234514.
65. Guo, F., Du, J., Liu, L., Gou, Y., Zhang, M., Sun, W., Yu, H. and Fu, X. LncRNA OR3A4 Promotes the Proliferation and Metastasis of Ovarian Cancer Through KLF6 Pathway. *Front Pharmacol* 2021, 12, 727876, doi:10.3389/fphar.2021.727876.
66. Wei, P., Tang, H. and Li, D. Insights into pancreatic cancer etiology from pathway analysis of genome-wide association study data. *PLoS One* 2012, 7, e46887, doi:10.1371/journal.pone.0046887.
67. Cui, T., Tsolakis, A. V., Li, S. C., Cunningham, J. L., Lind, T., Oberg, K. and Giandomenico, V. Olfactory receptor 51E1 protein as a potential novel tissue biomarker for small intestine neuroendocrine carcinomas. *Eur J Endocrinol* 2013, 168, 253-261, doi:10.1530/EJE-12-0814.
68. Weber, L., Schulz, W. A., Philippou, S., Eckardt, J., Ubrig, B., Hoffmann, M. J., Tannapfel, A., Kalbe, B., Gisselmann, G. and Hatt, H. Characterization of the Olfactory Receptor OR10H1 in Human Urinary Bladder Cancer. *Front Physiol* 2018, 9, 456, doi:10.3389/fphys.2018.00456.
69. Xu, L. L., Stackhouse, B. G., Florence, K., Zhang, W., Shanmugam, N., Sesterhenn, I. A., Zou, Z., Srikantan, V., Augustus, M., Roschke, V., et al. PSGR, a novel prostate-specific gene with homology to a G protein-coupled receptor, is overexpressed in prostate cancer. *Cancer Res* 2000, 60, 6568-6572.
70. Neuhaus, E. M., Zhang, W., Gelis, L., Deng, Y., Noldus, J. and Hatt, H. Activation of an olfactory receptor inhibits proliferation of prostate cancer cells. *J Biol Chem* 2009, 284, 16218-16225, doi:10.1074/jbc.M109.012096.
71. Abaffy, T., Bain, J. R., Muehlbauer, M. J., Spasojevic, I., Lodha, S., Bruguera, E., O'Neal, S. K., Kim, S. Y. and Matsunami, H. A Testosterone Metabolite 19-Hydroxyandrostenedione Induces Neuroendocrine Trans-Differentiation of Prostate Cancer Cells via an Ectopic Olfactory Receptor. *Front Oncol* 2018, 8, 162, doi:10.3389/fonc.2018.00162.
72. Klauser, A. L., Hirschfeld, M., Ritter, A., Rucker, G., Jager, M., Gundarova, J., Weiss, D., Juhasz-Boss, I., Berner, K., Erbes, T., et al. Anticarcinogenic Effects of Odorant Substances Citral, Citrathal R and Cyclovertal on Breast Cancer in vitro. *Breast Cancer (Dove Med Press)* 2021, 13, 659-673, doi:10.2147/BCTT.S322619.
73. Li, M., Schweiger, M. W., Ryan, D. J., Nakano, I., Carvalho, L. A. and Tannous, B. A. Olfactory receptor 5B21 drives breast cancer metastasis. *iScience* 2021, 24, 103519, doi:10.1016/j.isci.2021.103519.

74. Maiga, A., Lemieux, S., Pabst, C., Lavallee, V. P., Bouvier, M., Sauvageau, G. and Hebert, J. Transcriptome analysis of G protein-coupled receptors in distinct genetic subgroups of acute myeloid leukemia: identification of potential disease-specific targets. *Blood Cancer J* 2016, 6, e431, doi:10.1038/bcj.2016.36.
75. Vadevoo, S. M. P., Gunassekaran, G. R., Lee, C., Lee, N., Lee, J., Chae, S., Park, J. Y., Koo, J. and Lee, B. The macrophage odorant receptor Olfr78 mediates the lactate-induced M2 phenotype of tumor-associated macrophages. *Proc Natl Acad Sci U S A* 2021, 118, doi:10.1073/pnas.2102434118.
76. Martin, A. L., Anadon, C. M., Biswas, S., Mine, J. A., Handley, K. F., Payne, K. K., Mandal, G., Chaurio, R. A., Powers, J. J., Sprenger, K. B., et al. Olfactory Receptor OR2H1 Is an Effective Target for CAR T Cells in Human Epithelial Tumors. *Mol Cancer Ther* 2022, 21, 1184-1194, doi:10.1158/1535-7163.MCT-21-0872.

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