

Review

Not peer-reviewed version

Therapeutic Applications of Poly-miRNAs and miRNA Sponges

[Miguel Ibáñez-Hernández](#)*, [Cynthia Avendaño-Portugal](#), [Mariela Montaña-Samaniego](#),
[Raquel Guttman-Bazbaz](#), [Diana Marcela Bravo-Estupiñan](#)

Posted Date: 4 April 2025

doi: 10.20944/preprints202504.0362.v1

Keywords: miRNA; RNA-based therapy; biomedicine; miRNA sponge; poly-miRNA



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This open access article is published under a Creative Commons CC BY 4.0 license, which permit the free download, distribution, and reuse, provided that the author and preprint are cited in any reuse.

Review

Therapeutic Applications of poly-miRNAs and miRNA Sponges

Cynthia Avendaño-Portugal ¹, Mariela Montaña-Samaniego ^{1,2}, Raquel Guttman-Bazbaz ³,
Diana M. Bravo-Estupiñan ^{1,4} and Miguel Ibáñez-Hernández ^{1,*}

¹ Laboratorio de Terapia Génica, Departamento de Bioquímica, Escuela Nacional de Ciencias Biológicas, Instituto Politécnico Nacional, Carpio and Plan de Ayala, Col. Sto Tomás, Miguel Hidalgo, Mexico City 11340, Mexico.

² Laboratorio de Técnicas Fototérmicas, Departamento de Ciencias Básicas, Unidad Politécnica Interdisciplinaria de Biotecnología, Instituto Politécnico Nacional, Mexico City 07340, Mexico.

³ Facultad de Ciencias de la Salud, Universidad Anáhuac México, Av. Lomas Anáhuac 46, Col. Lomas Anáhuac, Huixquilucan, State of Mexico 52786, Mexico.

⁴ Laboratorio de Quimiosensibilidad Tumoral, Facultad de Microbiología, Universidad de Costa Rica, San Jose 11501-2060, Costa Rica.

* Correspondence: Author to whom correspondence should be addressed * mibanez@ipn.mx, mibanez_01@hotmail.com

Abstract: MicroRNAs (miRNAs) are small, non-coding RNA molecules that play crucial roles in regulating gene expression, and their dysregulation is implicated in various human diseases. Over the years, several research groups have identified miRNAs as promising therapeutic targets for intervention. Therapeutic strategies involve either the overexpression or knockdown of specific miRNAs. This review aims to provide a comprehensive overview of synthetic poly-miRNAs and miRNA sponges, highlighting their therapeutic applications. It begins with an introduction to miRNAs and their role in human diseases, followed by a detailed discussion on synthetic poly-miRNAs and miRNA sponges by exploring their application in cardiovascular, inflammatory, autoimmune, and metabolic disorders, as well as in cancer therapy. Additionally, strategies for targeted delivery, challenges, and limitations of these therapies are addressed.

Keywords: miRNA; RNA-based therapy; biomedicine; miRNA sponge; poly-miRNA

1. Introduction

MicroRNAs (miRNAs) are endogenous, small, conserved, and single-stranded non-coding RNA molecules of 21-25 nucleotides in length that play a crucial role in the post-transcriptional regulation of gene expression in eukaryotes. miRNAs bind to complementary sequences on their target messenger RNAs (mRNAs), which can lead to the degradation or translation inhibition of these mRNAs, thus regulating almost every important physiological process such as development, embryogenesis, proliferation, apoptosis, cell growth, viral defense, differentiation, and metabolism [1–3]. Polycistronic miRNAs (Poly-miRNAs) usually refer to the use of multiple miRNAs that are transcribed together as a single primary transcript (pri-miRNA) and are then processed into individual mature miRNAs [4–6]. Although these miRNAs are produced in a single transcript, they can target different mRNAs and act together in complex regulatory networks.

On the other hand, microRNA sponges (miRNA sponges) are a technology designed to bind and sequester miRNAs. These “sponges” contain multiple binding sites that are complementary to specific miRNAs (homo-sponges) or a group of miRNAs (hetero-sponges), acting as competitors for the binding of miRNAs and, therefore, inhibiting their function [7–9]. This technology allows the

derepression of miRNA target genes, which can be crucial in cases where miRNAs are overexpressed and contribute to the development and progression of disease. For instance, miRNAs have been associated with various pathological conditions, making them attractive targets for the treatment of cardiovascular, gastrointestinal, metabolic, neurodegenerative, and rheumatic diseases, as well as cancer and more. The ongoing development in this field is promising, with continuous improvements in the vehicles used for the delivery of poly-miRNAs and miRNA sponges, as well as the precision of targeting. In addition to their association with diseases, miRNAs have also been shown to play a role in the remodeling of the tumor microenvironment through their release in exosomes, mediated by cell-to-cell contact [10].

According to the NCBI database, in the past year, over 460,000 papers on miRNAs and their relation with multiple diseases have been published [11]. This growing body of evidence underscores the critical importance of investigating therapies targeting poly-miRNAs and miRNA sponges as a potential strategy for disease management and treatment. Hence, understanding the cooperative functions of miRNAs can offer a more comprehensive view of biological processes and diseases, leading to the identification of more precise therapeutic targets.

2. Overview of Synthetic Poly-miRNAs

Poly-miRNAs are more widely referenced in scientific literature as multi-miRNAs. Some similar concepts may be found in literature, including polycistronic miRNAs and miRNA clusters. Although closely related to poly-miRNAs, they cannot be employed as interchangeable terms. However, these concepts had, in a way, inspired the arrange of multiple miRNA precursors in tandem to synthetically build a single transcript coding for multiple miRNAs, as miRNA clusters have been found in several loci within the genome, showing a natural tendency of miRNAs to appear and function in clusters [12–14].

Poly-miRNAs replicate what has been found across genomes and may include multiple copies of identical miRNAs—forming a homo poly-miRNA—or different miRNAs—constituting a hetero poly-miRNA—(Figure 1) [14]. Thus, they enhance their effects by amplifying the expression of a single miRNA or, in a synergic manner, targeting multiple genes simultaneously. This makes it feasible to modulate multiple genes by engineering a single transcript [12–14].

It has been pointed out that single miRNAs effects are rarely biologically significant when compared to the linked effects within the whole miRNA network [12]; studying systematic miRNA deletions, conducted in *C. elegans*, and mice, has shown, reliably, that only less than 10% of miRNAs are individually essential for normal development or viability [15–17].

Moreover, to date, at least two clinical trials for therapeutic microRNAs -miR-34 in gastrointestinal cancers [18] and miR-16 in mesothelioma [19]- though never approved or even having entered phase III due to different concerns, showed data suggesting that the clinical impact may have been limited by the single miRNA modulation approach [12]. The work of Bhaskaran et al. confirms that microRNA clustering is a significantly more effective alternative due to a synergistic effect [12]. This approach has been applied to overexpress several inhibitory RNAs against cancer [20], hepatitis [21], influenza [22], and HIV [23]. Furthermore, it has been proved that it is achievable to transcribe multiple miRNA hairpins as a single transcript from the same Pol II promoter in mammalian cells (Figure 1). This confers an advantage since Pol II promoters are often stronger than others [24].

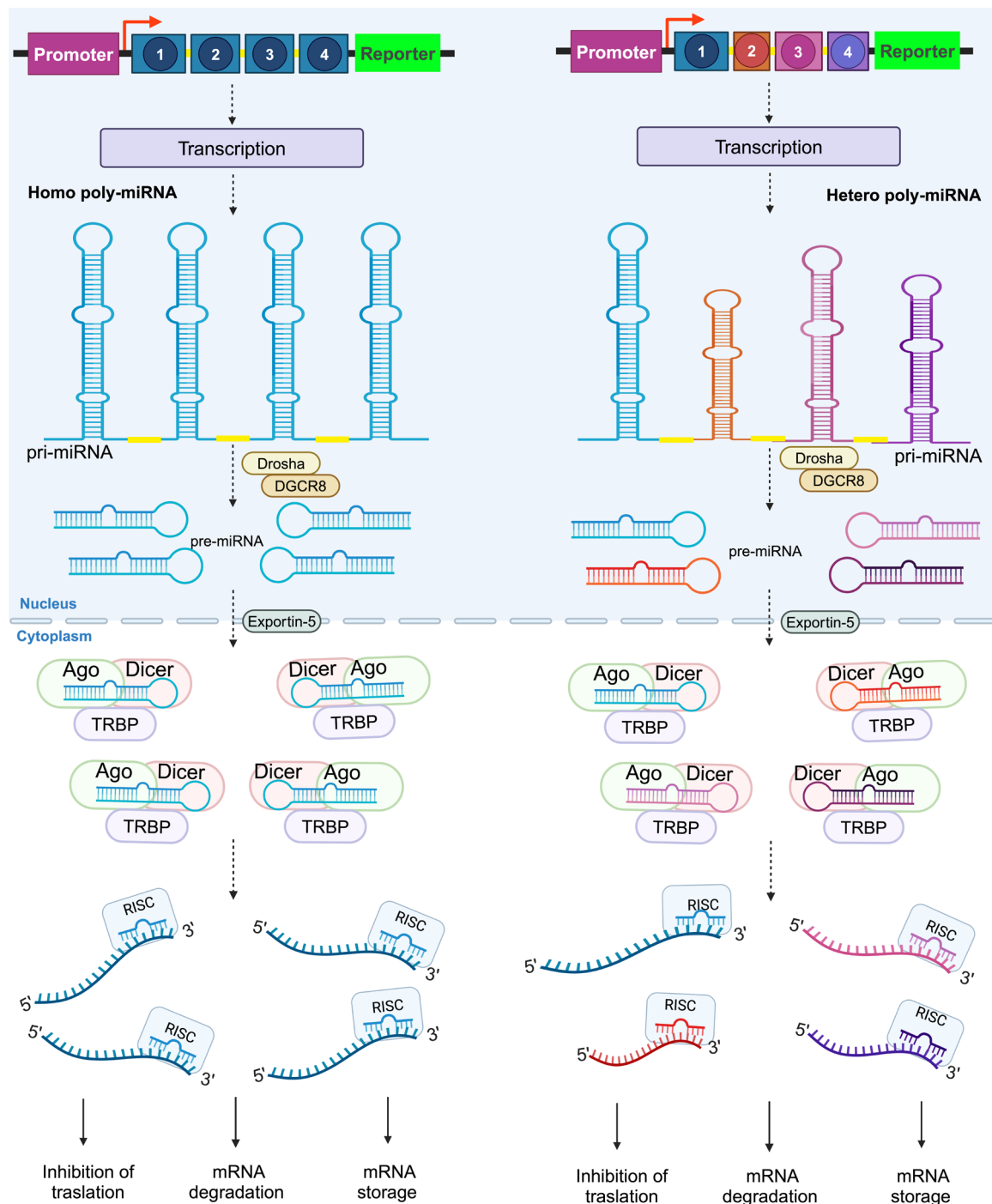


Figure 1. Schematic representation of poly-miRNA genetic constructs and their mode of action. The figure illustrates how poly-miRNA genes can be arranged within a genetic vector, driven by either a polymerase II or III promoter. On the left, an example of a homo poly-miRNA is shown, where identical miRNA sequences are repeated. On the right, a hetero poly-miRNA is depicted, where different miRNA sequences (represented in various colors) are arranged within the construct. After transcription, the poly-miRNA adopts a typical pri-miRNA stem-loop structure, with each stem-loop separated by a 'spacer sequence' (yellow). The pri-miRNA is then processed into mature miRNAs, which assemble with the RISC complex and bind to the 3' UTR of target mRNAs, leading to mRNA decay. Created in BioRender <https://BioRender.com/rnbsn8z>.

To achieve a potentially therapeutic poly-miRNA candidate, two main aspects should be considered: identifying the potential targets and constructing the backbone. Single miRNA prospect targets should be defined either by searching for known and fully characterized miRNAs in the

miRBase and miRTarBase repositories and the literature or by identifying new targets through miRNA overexpression assays [25–27].

Poly-miRNA backbones can be obtained through several methods, and it is critical as the integrity of the primary structures depends upon the backbone sequence. As more pri-miRNA or sequences are added to the poly-miRNA, obtaining a stable secondary structure and tertiary folding becomes more challenging. Moreover, to achieve the desired native assembly, maintaining the characteristic motifs is needed for proper recognition and processing into a mature and functional state [28]. Though mostly unraveled, those key features within the primary transcript are essential for miRNA microprocessor complex to discern between miRNAs and other hairpins [24,29,30], and it remains uncertain how many more are still yet to be uncovered or fully understood [24,30].

Given the challenge of designing de novo poly-miRNA sequences, and aiming to minimize the hurdles, time, and even molecular cloning requirements, strategies have been proposed to achieve a fully functioning poly-miRNA by modifying naturally occurring clustered-miRNA backbones [12,13,31,32].

Endogenous miRNA clusters have been explored as an innovative approach to designing customizable poly-miRNAs for therapeutic applications. One of the reported strategies consisted of using an endogenous miR 17-92 cluster that appears to be an efficient option since it allows for the highest number of miRNAs, six, within the shortest backbone, of about 800 bp [13]. Another approach leverages the pri-miR-155 backbone, which allows efficient maturation of several miRNA precursors with different stem sequences. The reported three-round cloning strategy used in this work, allows the construction of a poly-miRNA for up to 18 hairpins in a [24].

Despite promising preclinical results, miRNA therapeutics have faced a challenging path in the clinical field, since there are currently no miRNA-based drugs approved or even in clinical phase III [17,27,33–35]. The first in-human study for miRNA therapy was MRX34, a miR34a mimic, in 2013, halted after three years due to toxicity and safety issues [17,18,27].

Poly-miRNAs provide a particularly appealing approach towards complex diseases prophylaxis including cancer and other multifactorial disorders, as their pathogenesis is mainly driven by multigenic dysregulation [17,27,35] and poly-miRNAs allow to regulate multiple targets at once [12–14,24].

It is worth noting that, although poly-miRNAs are proven to work in a synergic manner, it has been suggested that this effect is more effectively achieved by expanding the targetome [13].

3. Overview of Synthetic miRNA Sponges

Naturally occurring non-coding RNAs (ncRNAs) function as regulators of endogenous miRNAs known as competing endogenous RNAs (ceRNAs). The most studied ceRNAs are long non-coding RNAs (lncRNAs) and circular RNAs (circRNAs), both of which act as natural miRNA sponges. However, if the sequence of a specific mature miRNA targeted for downregulation is known, a synthetic miRNA sponge can be designed [36].

Like lncRNAs, miRNA sponges act as competitive inhibitors, sequestering miRNAs away from their target mRNAs. This competitive inhibition leads to the derepression of mRNA targets, thereby altering protein synthesis and cellular function [36,37].

A hallmark feature of miRNA sponges is the presence of multiple miRNA binding sites (MBSs). The formation of the miRNA-miRNA sponge complex sequesters the miRNA, preventing it from binding to its endogenous mRNA targets. This sequestration effectively diminishes the miRNA's regulatory impact on gene expression, allowing the derepression of downstream genes (Figure 2) [7,38,39].

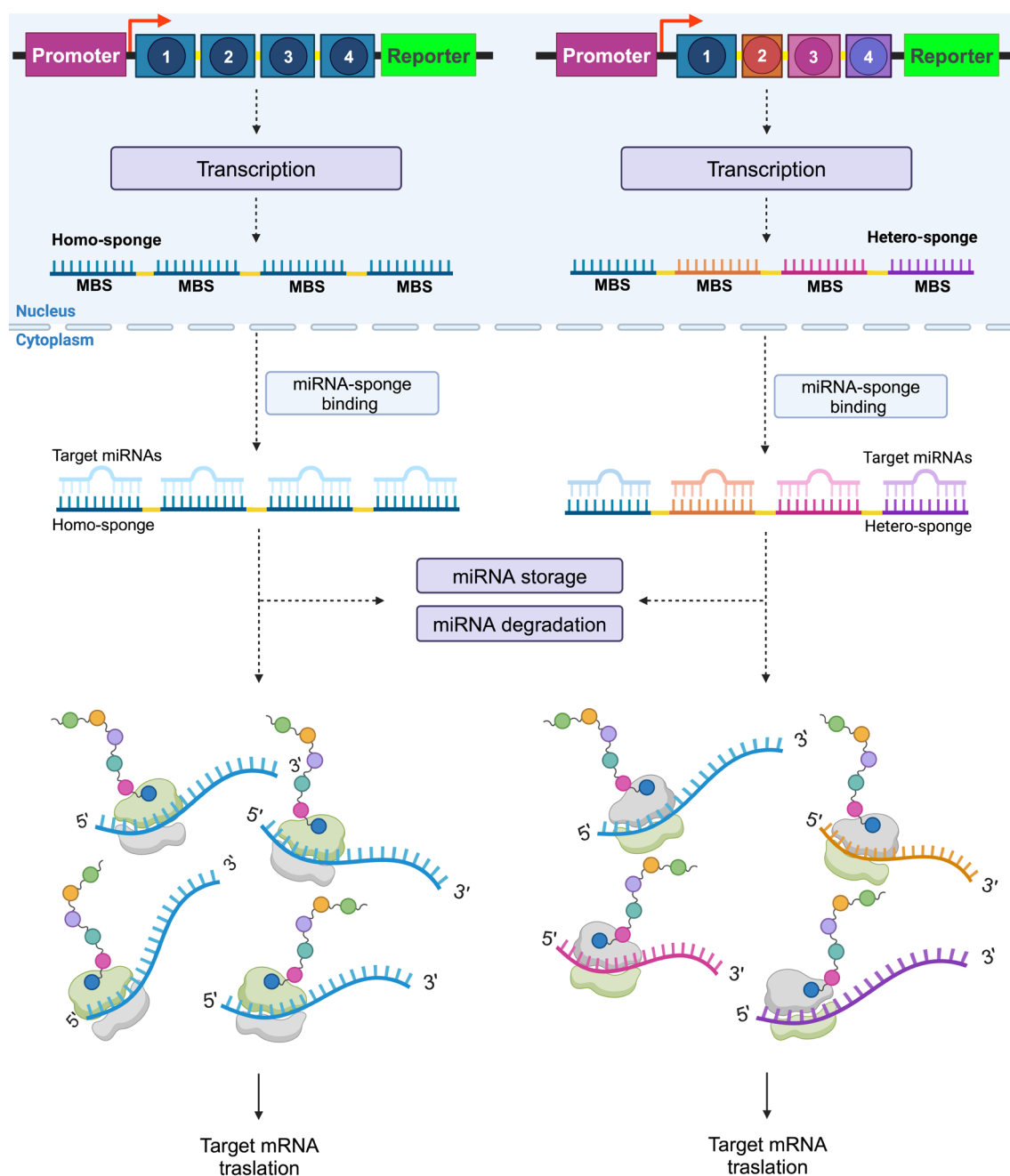


Figure 2. Schematic representation of synthetic miRNA sponges genetic constructs and their mechanism of action. The figure illustrates how miRNA sponge genes can be arranged within a genetic vector. On the left, an example of a homo sponge is shown, where identical MBSs are repeated. On the right, a hetero sponge is shown, where different MBSs (represented in various colors) are arranged within the construct; in both cases, the MBSs are separated by a spacer sequence (yellow). Once transcribed, the miRNA-sponge complex forms through complementary binding to the seed sequence of the miRNA, leading to mRNA derepression. Created in BioRender <https://BioRender.com/79wvrsy>.

The design of synthetic miRNA sponges is highly customizable. They can include MBSs that are either perfectly complementary to the mature miRNA of interest or contain bulged sections at the central position. It has been reported that perfectly base-paired miRNA sponges are vulnerable to cleavage by RNA-induced silencing complex (RISC) proteins, leading to rapid sponge degradation. In contrast, MBSs containing a 4-nucleotide bulge tend to be more stable [40,41]. Furthermore, non-perfect pairing sponges have better inhibition efficacy [37]. The MBSs can range from two to as many as the backbone and the structural properties of the transcript allow. Typically, they contain four to

ten MBSs, as increasing the number may have led to diminishing returns [42]. On the other hand, a research group managed to successfully construct sponges with 16 binding sites for one specific miRNA and others with 32 binding sites targeting two different miRNAs. In expression experiments, the sponges containing 16 binding sites for two different miRNAs (32 in total) showed enhanced miRNA downregulation [37]. The effectiveness of these designs depends on the level of upregulation of the miRNA in a given tissue or disease; also, if more miRNAs share the same seed sequence, the biological effects may be amplified.

MBSs are arranged in a repetitive sequence pattern and separated by a “spacer sequence” typically consisting of four to eight nucleotides. This characteristic is customizable depending on the desired structure [37,40]. Additionally, one of the greatest qualities of synthetic miRNA sponges is that they can be designed to inhibit entire miRNA clusters by incorporating multiple distinct MBSs within a single construct (Figure 2) [40,42].

Synthetic miRNA sponges are transcribed from strong promoters in DNA vectors in an mRNA-like manner when transcribed by RNA polymerase II, though polymerase III promoters can also be used [43]. Polymerase II promoters offer advantages, such as the capping and polyadenylation of the resulting transcript. Also, the sponge sequence can be placed in the 3' UTR of a reporter gene like EGFP. An additional advantage of polymerase II promoters for this purpose is that strong promoters like CMV, PGK, and EF1 α can be used, thus augmenting the expression of the transcript [40–42,44].

A key advantage of miRNA sponges over antisense oligonucleotides or antimiRs is their ability to interact with all members of a miRNA seed family. Since sponge-miRNA interaction relies on base-pairing within the seed region, the sponges are able to interact with all members of the miRNA seed family, whereas antisense oligonucleotides can only target one specific miRNA. Furthermore, many cell types, both in vitro and in vivo, exhibit resistance to oligonucleotide uptake. In contrast, for cell lines that are difficult to transfect or for in vivo applications, the sponge transgene can be introduced via a viral vector. By incorporating an open reading frame for a selectable marker or reporter gene within the vector, the selection, quantification, screening, fluorescence-activated cell sorting, or even laser capture microdissection of cells that strongly express the sponge can be facilitated. This enables the isolation of cells in which the miRNA family is significantly inhibited, allowing the detection of modifications in target gene expression [42].

Sponge stability and cellular localization can be optimized using chemical modifications and vector-based delivery systems [7,45,46]. Recent advancements in miRNA sponge technology have led to the development of inducible and tissue-specific sponges, which offer controlled and localized miRNA inhibition. These innovations minimize off-target effects and improve the therapeutic potential of miRNA sponges. Moreover, combination strategies integrating synthetic miRNAs and miRNA sponges have emerged, enabling the synergistic regulation of miRNA networks to achieve precise therapeutic outcomes [42].

Given their versatility, miRNA sponges have been widely explored for therapeutic applications in different diseases, particularly in conditions where specific miRNAs play a central role in disease progression. For instance, miR-21 sponges have been developed to counteract the overexpression of miR-21 in various cancers [47]. Similarly, sponges targeting miR-155 have shown potential in treating hematological malignancies, as miR-155 is implicated in the progression of leukemia and lymphoma [48].

In neurodegenerative diseases, miRNA sponges targeting miR-29 and miR-146a have been explored for Alzheimer's disease therapy. These miRNAs are known to regulate pathways involved in neuroinflammation and amyloid plaque formation. By inhibiting these miRNAs, synthetic sponges can potentially alleviate disease progression and neuronal damage [49].

In cardiovascular research, miR-92a sponges have been investigated for their role in promoting vascular repair and angiogenesis. Since miR-92a negatively regulates endothelial function, its inhibition via sponges can enhance post-injury tissue regeneration and improve recovery outcomes [50].

As research progresses, synthetic miRNA sponges continue to show promise as versatile tools for dissecting miRNA functions and developing novel miRNA-based therapies. Their ability to modulate miRNA activity with high specificity underscores their potential in personalized medicine and targeted gene regulation strategies. The integration of miRNA sponges with emerging gene editing and RNA delivery technologies may further expand their clinical applicability, paving the way for next-generation therapeutics.

4. Applications in Cardiovascular Diseases

Cardiovascular diseases (CVD) are the most common cause of death worldwide; their prevalence continues to increase despite significant advances in primary prevention and therapeutic strategies over recent years [51,52]. Atherosclerosis is a complex disease that may lead to ischemic heart disease, hypertension, aortic aneurysm, and stroke—the most important risk factor for CVD [9,53]. Therefore, it is essential to thoroughly investigate cardiovascular disease's molecular pathophysiology to discover new biomarkers that can improve early diagnosis and more efficient treatment for these diseases [2,3,54].

Several studies have reported the potential role of miRNAs in the cardiovascular system, such as in cardiac cell contractility and growth, angiogenesis, regulation of lipid metabolism, and plaque formation [55–57]. miRNA dysregulation has been observed in myocardial infarction [58,59], heart failure [60,61], and cardiac hypertrophy [51,62], in both, humans and animals, miRNAs are implicated in the development of atherosclerosis [3,63].

Numerous studies have examined the role of various miRNAs not only for diagnostic and prognostic purposes but also for treating several CVDs. For example, miR-33a/b is a proatherogenic miRNA that plays an important role in lipid metabolism and cholesterol homeostasis, impacting the progression of atherosclerosis [9,64]. Inhibiting the function of miR-33 is known to raise HDL levels and reduce atherogenic risk; however, long-term or generalized inhibition produced deleterious effects [65,66]. Therefore, the effect of decreasing miR-33 in specific tissues or organs has been studied either as basic research or as a therapeutic proposal using miRNA sponges to preserve its antiatherogenic function but omitting the well-known adverse effects [9,67].

Similarly, miR-122 has an important function in lipid metabolism and atherosclerosis development; however, it is also known to be elevated in heart failure patients [51,68,69]. Consequently, the use of miR-122 antagonists has been investigated. It was reported that the loss of miR-122 has anti-inflammatory effects in CVD and inhibits apoptosis of myocardial cells. In addition, miR-122 inhibition exerts antioxidant and anti-apoptotic effects on sepsis-induced myocardial injury [68,70]. It is important to notice that long-term depletion of miR-122 is associated with hepatocarcinogenesis, suggesting that complete depletion of miR-122 may have deleterious consequences in humans [71,72]. Therefore, additional research on heart-targeted inhibition is required.

miR-21 is one of the most studied miRNAs in CVD, given that it is the most prominent non-coding RNA associated with hypertension and atherosclerotic disease. miR-21 was upregulated in human atherosclerotic plaque and various heart diseases [73–75]. Consequently, inhibition of this miRNA—mainly by modified antisense oligonucleotides—has been evaluated and proven to be effective against cardiac fibrosis and dysfunction [76–80].

Previously it has been suggested that suppressing miR-214-3p could be a potential therapy for myocardial infarction, while other investigations provided evidence that miR-214-3p contributes to ferroptosis and its inhibition in vivo attenuates cardiac ferroptosis and malfunction induced by myocardial infarction [59,81]. In the same way, an adeno-associated virus serotype 9 (AAV9)-mediated delivery of anti-miR-214 was proven to restore cardiac function in hypertrophic mouse hearts [82]. Although miRNA sponges have been developed for the suppression and even targeted delivery of specific miRNAs, alternative approaches that leverage naturally occurring ncRNAs are also emerging, such as the use of lncRNAs to inhibit miR-214 and restore the levels of its target genes,

so it could be used to help alleviate osteoporosis [83,84], however, there have been no studies on miRNA sponges for CVD treatment.

Another miRNA of special interest is miR-34a, whose expression is elevated in mouse hearts with myocardial infarction and aging, as well as in human failing hearts. Inhibition of miR-34 has protective properties in mice after cardiac insult [85–88]. miR-34 has been successfully regulated in the heart using an adenoviral approach [89] and considering this, it was possible to generate a miR-34 sponge for the heart. Nevertheless, the sponge activity did not achieve significant inhibition of miR-34a using targeted approaches compared to previous studies employing locked nucleic acid inhibitors [1,85]. Moreover, chronic inhibition of miR-34 may not be optimal since it has been reported to induce tumorigenesis [90]. Therefore, the development of a more efficient therapeutic approach directly targeting the heart to inhibit miR-34 is necessary.

miR-155 is upregulated in patients or animal models with cardiac inflammation and atherosclerotic lesions. The levels of miR-155 in the blood of people who died due to heart conditions were about four times higher than those who did not have heart problems [91–93]. It is known that miR-155 inhibition ameliorates cardiac inflammation and that it attenuates atherosclerosis development and progression in mice [78,92,94]. It has been suggested that macrophage-specific miR-155 inhibition could be a feasible therapeutic strategy to decrease atherosclerosis-related inflammation [91]. Conversely, Chung and colleagues, based on miRNAs that are naturally expressed in clusters, designed a polycistronic expression vector for RNA interference based on BIC/miR-155, although, without any specific therapeutic purpose, demonstrated that this strategy can be used to express multiple miRNAs from a single transcription unit, in order to increase the inhibition of a target RNA [95].

The ability of miRNAs to regulate multiple pathways involved in disease progression positions them as attractive therapeutic targets. However, the translation to the clinic requires overcoming significant barriers in delivery, specificity, and safety. Continued research is crucial to developing the full potential of miRNA-based therapies in CVD.

5. Role in Metabolic Disorders

5.1. Role of miRNAs and Applications in Metabolic Disorders

Metabolic disorders, including obesity, diabetes, and dyslipidemia, are complex diseases influenced by genetic and environmental factors. Becoming a global health concern due to their increasing prevalence and association with serious complications such as CVD and cancer [53,96]. Traditional therapeutic approaches have limitations, prompting the search for novel strategies. Recent advances in the field of miRNAs highlight their crucial role in regulating metabolic processes and their potential as therapeutic targets [97].

5.2. Role in Obesity

Obesity is characterized by excessive fat accumulation and is a major risk factor for metabolic disorders, including diabetes and CVD [96,98]. Obesity leads to alterations in miRNA expression, and several miRNAs have been closely associated as regulators of adipogenesis and lipid metabolism, while also being dysregulated in obesity [99–102].

For example, long-term silencing and genetic knockout (KO) studies have shown that loss of miR-33 can lead to hepatic lipid accumulation, obesity, and metabolic dysfunction, particularly when combined with a high-fat diet (HFD). This is associated with increased food intake and impaired metabolic function in white adipose tissue and the liver [65,66]. Tissue-specific KO models have demonstrated that loss of miR-33 in the liver does not affect body weight but improves insulin sensitivity, glucose homeostasis, and reduces lipid accumulation in the liver, thereby protecting it from metabolic dysfunction-associated liver disease (MASLD) [63,103]. In another study, miR-33 was deleted in AgRP neurons, leading to metabolic dysregulation and obesity in mice [104]. miR-33b serum levels are potential biomarkers for obesity and hypercholesterolemia [98], as well as possible

treatments, underscoring the importance and need for more targeted miRNA treatments. Another well-established microRNA is miR-143 since it facilitates the differentiation of adipocytes [101,105–107] and its downregulation in cultured human pre-adipocytes led to the inhibition of adipocyte differentiation [108], whereas the KO in brown adipose tissue significantly enhanced thermogenesis while in white adipose tissue inhibited the process of adipogenesis [109]. In obese mice, circRNF111 functions as a sponge for miR-143, suggesting a protective role of circRNF111 in metabolic syndrome, which may serve as a promising therapeutic target for mitigating lipid accumulation [110].

On the other hand, overexpression of miR-21 in white adipose tissue has been linked to obesity, as it regulates adipogenic differentiation [111,112]. In 2021, researchers found that administering a miR-21 mimic to obese mice prevented weight gain induced by a high-fat diet [111], confirming the potential of the miR-21 mimic as a therapeutic option for obesity. However, the outcomes remain unclear since it has been previously demonstrated that long-term inhibition of miR-21 reduced body weight and adipocyte size, while also improving lipid homeostasis [113]. Recently, a miR-21 sponge construct released by exosomes was developed, which has been shown to have the potential to suppress miR-21 and upregulate miR-21 target genes [114].

Elevated levels of miR-126 were found in obese children [115], aligning with studies showing that miR-126b overexpression promotes lipid deposition and exacerbates obesity symptoms in mice, leading to higher adipose tissue weight and insulin resistance [116,117]. Targeting miR-126 with a sponge-based approach could offer a potential therapeutic strategy for pediatric obesity.

5.3. Role in Diabetes and Insulin Resistance

Insulin resistance (IR) is a precursor defect in the vast majority of patients with type 2 diabetes mellitus (T2DM). It is characterized by dysfunctional insulin action, leading to increased insulin levels to maintain blood glucose levels within a normal range. This condition can result in negative outcomes such as obesity, hypertension, and dyslipidemia among patients with T2DM [112,118].

miR-21 up-regulation is associated with an increase in insulin resistance [119], and its downregulation via knockdown plasmids or sponges, has been reported to be an effective therapy for diabetic nephropathy [120,121] in addition, a research group indicated that a miR-21 antagomir was able to lower blood glucose levels and improve insulin resistance in T2DM rats [118]. More recently, a study revealed an increased level of miR-21 expression in diabetic patients and that the circRNA circ_0000652 acts as a sponge for miR-21; hence, it could be a novel therapeutic target for treatment, including the prevention of T2DM [122].

In contrast, reduced miR-146a levels have been linked with insulin resistance and reduced glycemic control, consistent with reports of miR-146a downregulation in obese and T2DM patients [112,123,124]. Several investigations have shown that miR-146a can mitigate the progression of diabetes complications, including nephropathy, wound healing [125], neuropathy, and retinopathy. Furthermore, injection of miR-146a mimic has been confirmed to alleviate diabetes mellitus in animal models [123,126].

Other groups identified miR-690 as a key insulin-sensitizing miRNA that is highly expressed within exosomal-derived M2-polarized macrophages (M2 Exos). M2 Exos can improve insulin sensitivity in vivo [127,128]. Hence, if these results can be translated to humans, the miRNA contents of exosomes could potentially provide therapeutic options for treatment.

Another example of miRNA downregulation/silencing for insulin-resistant research is the silencing of miR-222, which reduces insulin-stimulated glucose uptake by ~40% [129]. Although several studies suggest that miR-222 sponges can effectively inhibit miR-222 activity, these have been tested for other diseases rather than diabetes [130,131].

miR-223 is associated with the regulation of glucose homeostasis and inflammatory response. Two independent groups demonstrated that miR-223 is overexpressed in adipose tissue from women with IR or obese subjects [132,133], and that the overexpression of miR-223 in human adipocytes inhibits glucose uptake stimulated by insulin, leading to impaired insulin signaling and GLUT4 trafficking [132,134]. Although miR-233 sponges have been evaluated, and a sponge-mediated

reduction level was achieved, these have not been specifically developed to evaluate their therapeutic potential in diabetes or IR [135,136]. In the same way, natural sponges such as lncRNAs and circRNAs can influence processes like kidney aging, cardiac hypertrophy, cancer progression, and liver fibrosis by modulating miR-223 activity [24,137–140], making them a promising strategy for the treatment of diabetes and IR.

5.4. Role in Dyslipidemia

Dyslipidemia is characterized by elevated lipid levels in the blood, is a common feature of metabolic disorders, and can lead to the development of obesity-associated diseases. Interestingly, IR is the primary catalyst for the development of dyslipidemia. miR-122, miR-27b, and miR-33 are involved in cholesterol and lipid metabolism but dysregulated in dyslipidemia. Since the inhibition of miR-122 has been shown to reduce plasma cholesterol and triglyceride levels [141,142], many circRNAs—such as circ_0005963, circRNA_002581, circ_0007142, circ_0011269, and circ_0006404—have been proven to serve as a sponge of miR-122 [139,143–145], hence, these miR-122 antagonists could be a promising strategy for the treatment of dyslipidemia and CVD, with respective precautions due to concerns regarding the inhibition of miR-122 expression, as mice with either entire body or liver-specific deficiencies develop steatohepatitis, fibrosis, and hepatocellular carcinoma [146].

On the other hand, miR-33 sponges or inhibitors can effectively regulate lipid metabolism by increasing HDL levels, offering potential therapeutic strategies for dyslipidemia [9,67,147]; however, it is clear that this inhibition must be controlled in a specific manner since genetic KO studies have shown that loss of miR-33 can lead to hepatic lipid accumulation and obesity [66].

miR-27b, in contrast, acts as a regulator in lipid metabolism, influencing dyslipidemia by modulating the expression of key lipid-metabolism genes and affecting lipid accumulation, adipogenesis, and cholesterol metabolism [148–150]. Besides, it has been demonstrated that the inhibition of miR-27b was effective for reducing fat and body weight, decreasing levels of total cholesterol, triglycerides, and LDL-Ch while increasing levels of HDL-Ch [150]. Recently, has been reported that statin-induced miR-33a and miR-27b up-regulation contributes to insulin resistance and metabolic dysfunction, potentially leading to T2DM [151], reiterating the importance of miR-27b and its proper regulation in health and disease. Ongoing research is essential to developing the full potential of miRNA-based therapies in metabolic disorders, as they require overcoming significant barriers in specificity and safety.

6. Applications in Inflammatory and Autoimmune Diseases

Inflammatory and autoimmune diseases are characterized by dysregulated immune responses, often involving aberrant miRNA expression patterns. miRNA-based therapies, including synthetic miRNA sponges, hold great promise in modulating immune responses and restoring immune homeostasis.

Several studies have highlighted the role of specific miRNAs in the pathogenesis of inflammatory diseases. For instance, miR-146a and miR-155 are key regulators of immune signaling pathways. While miR-146a functions as a negative regulator of inflammation by targeting TRAF6 and IRAK1, miR-155 promotes pro-inflammatory responses by modulating transcription factors such as NF- κ B [152,153]. Synthetic miRNA sponges targeting miR-155 have been investigated as potential treatments for autoimmune conditions such as multiple sclerosis and rheumatoid arthritis, aiming to dampen excessive inflammation [154].

In systemic lupus erythematosus (SLE), miR-21 is upregulated and contributes to disease progression by enhancing T-cell activation and reducing immune tolerance. A miR-21 sponge-based approach has been proposed to mitigate this hyperactivation and restore immune balance [155]. Similarly, miR-223, which is implicated in inflammatory bowel disease (IBD), is a promising target for sponge-based interventions to regulate intestinal immune responses [156].

Beyond direct miRNA inhibition, tissue-specific and inducible miRNA sponges are being developed to minimize off-target effects and improve therapeutic precision. These advanced sponges are particularly valuable in chronic inflammatory diseases, where long-term regulation of immune pathways is necessary.

Various synthetic miRNA sponges have been designed for different diseases, demonstrating significant therapeutic potential. For example, miR-21 sponges have shown promise in systemic lupus erythematosus (SLE) by sequestering miR-21 and restoring immune function, leading to reduced autoantibody production and alleviation of lupus symptoms [157]. Similarly, miR-155 sponges have been utilized in multiple sclerosis and rheumatoid arthritis to reduce inflammatory cytokine production and mitigate disease progression [154].

In the context of Neuroinflammatory Disorders, where miR-326 is often downregulated, synthetic miR-326 sponges have been used to fine-tune immune responses, balancing pro- and anti-inflammatory pathways and improving survival rates [158]. Cardiovascular disease has also been targeted using miRNA sponges, with miR-92a sponges developed to enhance vascular repair and angiogenesis, showing positive outcomes in models of myocardial infarction and stroke [159].

Other applications include antiviral therapy, such as miR-122 sponges that have been investigated for blocking hepatitis C virus (HCV) replication, resulting in reduced viral load and improved antiviral responses [160].

Neurodegenerative diseases such as Alzheimer's disease have been targeted with miR-29 sponges, which regulate pathways involved in neuroinflammation and amyloid plaque formation, leading to improvements in cognitive function [161]. Fibrotic diseases have seen potential therapeutic applications with miR-200 sponges, which reduce fibrosis progression in lung and liver fibrosis models [162].

Finally, in osteoarthritis, miR-16 sponges have been explored to regulate joint inflammation and cartilage degradation, offering a novel approach to improving joint health [163].

As miRNA sponge technologies continue to evolve, their integration with nanoparticle delivery systems and gene therapy approaches is expected to enhance their clinical viability. The potential to fine-tune immune responses through miRNA sponges positions them as an innovative and promising strategy in the treatment of inflammatory, autoimmune, and other chronic diseases.

7. Cancer Therapeutics

Despite significant progress in early detection and treatment strategies, cancer remains one of the leading causes of death worldwide —ranking second— and its incidence continues to rise. It is, therefore, imperative to continue searching for innovative treatments. In this context, it has been proven that miRNAs can repress hundreds of genes and regulate a wide range of cellular pathways, including those involved in cancer progression [164–166]. For instance, miRNAs can target the mRNA of tumor suppressor genes (oncomiRNAs) or oncogenes (tumor-suppressor miRNAs), thereby modulating processes such as metastasis, angiogenesis, epithelial-mesenchymal transition (EMT), migration, invasion, cell proliferation, immune escape, and chemoresistance [164,167]. By modulating the imbalance of miRNAs in cancer therapy, huge benefits can be achieved—from elucidating the heart of cancer pathogenesis to reducing metastasis and enhancing immune response [168–170]. Therefore, these therapies can function as an adjunct or even a replacement for conventional chemotherapy and radiotherapy [171,172].

For example, oncomiRNAs such as miR-21 have been reported to be upregulated in several cancer types, including melanoma, glioblastoma, colorectal, lung, stomach, prostate, pancreatic, and breast cancer. This upregulation promotes tumor growth, inhibition of cell death, and chemoresistance [173]. In 2015, an anti-miR-21 sponge with three binding sites encoded in plasmid DNA was delivered to MCF-7 breast cancer cells. A lower expression of miR-21 was observed, which resulted in G1-phase cell cycle arrest, activation of caspase-3 apoptosis pathway, and sensitization to doxorubicin and cisplatin [174]. Using a sponge targeting one oncomiRNA has proven to be a good strategy. However, based on these findings, researchers have explored sponges that simultaneously

target multiple oncomiRNAs. For instance, Jie et al. constructed sponges with multiple binding sites for two miRNAs. Concurrently, by developing a miR-21/miR-31 and miR-31/miR-155 sponges with 16 binding sites for each miRNA [37]. miR-31 is notably overexpressed in lung adenocarcinoma, squamous cell carcinoma, and large cell lung carcinoma, and this overexpression is associated with primary tumor growth and metastasis since it promotes activation of the RAS/MAPK pathway [175]. Conversely, miR-155 induces resistance to chemotherapeutics, and its downregulation in an orthotopic lung cancer model resensitizes tumors to chemotherapy. Also, miR-155 is linked to a downregulation of TP53, and its low expression is associated with shorter survival in lung cancer [176]. The sponges developed in that study, proved effective in lung cancer cell lines by inhibiting cell growth, with their effects being more pronounced than those observed with conventional miRNA inhibitors. In addition to these examples, the miR-221/222 cluster plays a crucial role in cancer progression, sharing the same seed sequence and targeting the mRNA of the cell cycle regulator p27^{kip1}, the overexpression of miR-221/222 promotes cell proliferation in thyroid papillary carcinoma, breast cancer, lung cancer, and hepatocellular carcinoma (HCC) [177]. As an alternative therapy for HCC—the second leading cause of cancer death in men [178]—genetically modified adeno-associated viral vectors (AAV) were developed to express a sponge with four binding sites for miR-221. When tested in HCC cell lines, the sponge repressed the expression and activity of miR-221, leading to increased apoptosis [179]. Moreover, as p27^{kip1} mRNA is not the only target of the miR-221/222 cluster, these oncomiRNAs have also been implicated in tamoxifen resistance in breast cancer [180]. Considering this, another group designed a miR-221/222 sponge to inhibit their activity in tamoxifen-resistant MCF-7 cells, observing that the sponge represses the miR-221/222 expression and, consequently, abrogates tamoxifen resistance by restoring ER expression [181].

On the other hand, tumor-suppressor miRNAs are often downregulated in cancer, resulting in the overexpression of genes involved in tumor growth, cell survival, and antiapoptotic proteins [182]. Restoring the levels of the type of miRNAs has demonstrated positive biological effects in cancerous tissues. For instance, miR-124 has been identified as a tumor suppressor across cancer, including glioblastoma [183,184]. Glioblastoma is known to be the most aggressive primary malignant brain tumor, with a median survival of less than 2 years [185]. In 2023, several tumor-suppressor miRNAs were identified on clinically relevant glioblastoma cell lines and proposed as candidates for miRNA-based therapy. As a result, a poly-miRNA to overexpress miR-124-2, miR-135a-2, and let-7i was constructed. Consequently, the study found that using the poly-miRNA was more effective against glioma than individually delivered miRNAs. The enhanced efficacy can be attributed to the high heterogeneity of glioblastoma tumors, where more than one molecular subtype may coexist; therefore, determining and overexpressing pan-subtype miRNAs can represent a better chance for therapeutic success [6]. In this context, miR-135a-2 regulates cell proliferation and invasion via MAPK and JAK/STAT3 pathways and inhibits EMT [186], while let-7i inhibits UDP-Galactose-4-epimerase, which is overexpressed in glioblastoma cells, promoting cell proliferation and migration [187].

Furthermore, poly-miRNAs represent a promising approach for cancer treatment but can also be integrated as tools to enhance other advanced therapies. For example, they may improve the development and efficacy of CAR T cell therapy, where T cells are genetically modified to express a chimeric antigen receptor that recognizes tumor antigens, and its recognition triggers the cascade of activation for the T cell [10,188]. CAR T cell therapy has been proven effective but has disadvantages, such as the population of CAR T cells decreasing over time. It has been observed that CAR T cells used for successful treatments have shown an increase in mitochondrial mass and fusion events and a reduction in anaerobic metabolism thus, promoting these events in CAR T cells using a cluster of miRNAs—such as poly-miRNAs—could improve the efficacy of CAR T cell therapy while offering a safe, non-antigenic alternative [10].

8. Targeted Delivery Approaches

Targeted delivery is still a major issue in gene therapy applications, as it is crucial to enhance specificity while minimizing the off-target effects [189]. Gene delivery strategies are classified into viral and non-viral vectors, both of which can be engineered to improve targeting efficiency.

Viral vectors, such as adeno-associated viruses (AAVs), lentiviruses, and retroviruses, exhibit superior delivery capacity compared to non-viral vectors [190]. However, achieving tissue-specific targeting requires advanced modifications. Strategies to enhance viral vector specificity include capsid engineering, ligand functionalization, and the use of cell-specific promoters. Capsid engineering modifies the aminoacidic sequence of the viral capsid to improve receptor affinity, while ligand functionalization involves attaching molecules such as antibodies, peptides, or aptamers to facilitate receptor-mediated endocytosis [191–193]. Nevertheless, the main challenge of using these vectors is biosafety in clinical translation, as their use may be highly immunogenic, which can impact the therapy's safety, durability, and efficacy [190]. Another major challenge to overcome is genotoxicity associated with some viral vectors, such as gammaretroviral vectors, which integrate into the cell host genome. This integration may be advantageous when stable, long-term expression is needed; however, it also comes with risks, as the inserted genes tend to integrate near transcriptional start sites and have an affinity for oncogenes [194].

On the other hand, non-viral vectors such as lipid nanoparticles (LNPs), liposomes, and exosomes offer an alternative with lower immunogenicity, cytotoxicity, and mutagenesis [195,196]. When delivering DNA or RNA, cationic lipids have proven to be the most effective [197]. Their targeting ability can be enhanced through surface modification, charge-based targeting, and microfluidic-based engineering. LNPs and liposomes can be conjugated with antibodies, aptamers, or peptides for site-specific delivery, while cationic lipids improve nucleic acid encapsulation and enhance cellular uptake, particularly for RNA-based therapies [198]. The disadvantages of non-viral vectors include high production costs and possible cytotoxic effects. Several research groups are still in the race to find more effective and secure strategies to overcome the first one, such as using microfluidic devices to produce nanovesicles for nucleic acid delivery. These devices create precise flow-controlled environments, making them ideal in terms of batch-to-batch reproducibility since they minimize the intermediate steps required in conventional method production [199].

Whether using viral or non-viral vectors, specificity can be achieved by using tissue-specific promoters, which are promoters that are only active in specific cell tissues. Since miRNA sponges and poly-miRs are coded in an mRNA-like manner, this approach may be more effective for targeted therapy since its use can restrict unwanted transgene expression [189,200]. Tissue-specific promoters have been used in several in vitro and in vivo studies. For example, the CC10 promoter restricts transgene expression to lung cells [201]; cytokeratin 18 and 19 promoters are specific to epithelial cells; the kallikrein promoter is specific for salivary glands [200]; and the PEPCK promoter is liver-specific [9], among others. One disadvantage of tissue-specific promoters is that many of them are relatively weak compared to viral promoters like CMV or SV40. However, several studies have developed methods to improve their activity without losing their specificity, such as creating chimeric promoters using parts of strong promoters, combining two specific promoters, or adding elements like TATA boxes [202,203].

In summary, while the targeted and effective delivery of genetic material remains the most significant challenge in gene therapy applications, numerous research groups continue exploring improved techniques from different perspectives—optimizing genetic constructs, modifying delivery vehicles, or refining extraction techniques. The continued acquisition of knowledge in this area is crucial for advancing the clinical application of these therapies.

9. Challenges and Limitations

Several challenges remain regarding using poly-miRNAs and miRNA sponges for therapeutic purposes. First, as stated, miRNAs are pleiotropic [13,17,204], capable of recognizing multiple targets,

therefore triggering different molecular mechanisms and pathways depending on the site or context [17]. In fact, several miRNAs have shown a potential dual effect as both oncomiRNAs and tumor-suppressors [205,206]. For example, miR-10b can act differently depending on cancer type [205], and the activity of miR-200c depends on the carcinogenic stage [206]. Although miRNA sponges could be significantly more specific when targeting microRNAs, the repression may work pleiotropically, with different effects in different cell types and cycle stages.

This concern could be addressed by targeting the effects of these constructed poly-miRNAs and miRNA sponges—whether through targeted delivery (including on-site administration, encapsulation in selective tropism-driven vectors and/or antibody-mediated targeting[19] [19,207]—or through targeted expression by using tissue or tumor-specific promoters to modulate transcription in the desired site, even when systemically administered [189,208].

On the other hand, the delivery of nucleic acids still represents a major obstacle for the development of therapeutic miRNAs, as it is imperative to overcome biological and pharmacological barriers—including stability in physiological environments, susceptibility to degradation, and the physical challenges of crossing biological membranes [206]. Therefore, effective delivery systems are crucial for translating miRNA-based therapies into clinical applications, whether using viral or non-viral vectors. In this regard, innovative package-free alternatives, such as ligand-conjugated miRNAs, are being explored [208]. Hence, for miRNA-based therapies to succeed in the clinic, delivery must be addressed so that miRNA molecules reach the desired cells, bypassing healthy cells and immune recognition [17].

Identifying potential targets remains challenging as the annotations on the MiRBase are often incomplete, which leads to poor concordance when comparing overexpression/inhibition experiments in vitro with the observations in vivo [27,209]. According to an editorial on Nature Biotechnology posted in November 2024, the main challenge holding back miRNAs from getting to market is the off-target effect [27]. In 2021, Zhang et al. evaluated 10 miRNA drug prospects (none of which reached phase III) to assess their miRNA targets and compared them to those of siRNA drug prospects. They found that for every miRNA, the target number ranged from tens to hundreds. In contrast, siRNA targets oscillated only between 1 and 3, thus shedding light on some of the hurdles that miRNA-based therapies have faced to become marketable or even go further in clinical trials. On a different assessment, the same group points out that the target number of each FDA-approved drug (since 1939) is no more than five [210], providing perspective on where miRNA molecules stand in the clinical field and the broader RNA-based therapy race.

Like any other miRNA-based technology, poly-miRNAs and miRNA sponges encounter hurdles in clinical phases, mostly due to toxicity and immunogenicity concerns. The first-ever phase 1 clinical trial for a miRNA-based drug prospect—a miR-34a mimic for tumor suppression in GI cancer—was terminated after three years due to severe immune-mediated toxicities and four deaths [18,27,205]. Moreover, the off-target particularity has also been related to neuro- and immunotoxicity [17,205,211].

As stated in sections 2 and 3, constructing a functional poly-miRNA or miRNA sponge sequence is difficult on its own. For poly-miRNAs, in silico analyses, folding predictions and processing might result differently than anticipated [13]. In addition, it has been suggested that as the number of microRNAs encoded within the mRNA-like transcript increases, the expression of mature microRNA decreases, possibly due to a reduced cleavage efficacy in the microprocessor complex [13].

Moreover, when considering miRNA-based solutions for therapeutic purposes, it is important to note how the pathogenesis occurred in the first place. As stated before, many complex diseases develop due to the altered regulation of microRNAs, which, among several factors, could be influenced by alterations in the biogenesis, processing, and cleavage of miRNAs. Consequently, this prevents therapeutic miRNAs from providing effective solutions [17,29,205,212–215]. Although most of this is in cancer, the same might hold true in every other disease, as it is widely accepted that miRNA dysregulation plays a significant role in pathogenesis and disease progression [17,35,205,212].

Therefore, it is imperative to continue the search for better bioinformatic tools and to conduct more research on specific miRNAs. This will enhance our understanding of the pathways in which they are involved and, in turn, enable the development of safer therapeutic strategies.

Author Contributions: Conceptualization, C.A.-P. and M.I.; methodology, C.A.-P.; writing—original draft preparation, C.A.-P., M.M.S., D.M.B.-E., R.G.-B.; writing—review and editing, C.A.-P., M.M.-S., D.M.B.-E., R.G.-B.; figure and table creation, C.A.-P.; supervision, M.I.; project administration, C.A.-P. All authors have read and approved the final manuscript.

Funding: This research was funded by the Instituto Politécnico Nacional (IPN), through Secretaría de Investigación y Posgrado (SIP), linked to project number 20251022.

Institutional Review Board Statement: Not applicable

Data Availability Statement: No new data were created or analyzed in this study. Data sharing is not applicable to this article.

Acknowledgments: Cynthia Avendaño-Portugal is a doctoral student supported by the Consejo Nacional de Humanidades, Ciencias y Tecnologías (CONAHCyT).

Conflicts of Interest: The authors declare no conflict of interest.

References

- Bernardo, B.C.; Gregorevic, P.; Ritchie, R.H.; McMullen, J.R. Generation of MicroRNA-34 Sponges and Tough Decoys for the Heart: Developments and Challenges. *Front. Pharmacol.* **2018**, *9*, 1090, doi:10.3389/fphar.2018.01090.
- Fu, Y.; Chen, J.; Huang, Z. Recent Progress in microRNA-Based Delivery Systems for the Treatment of Human Disease. *ExRNA* **2019**, *1*, 24, doi:10.1186/s41544-019-0024-y.
- Vishnoi, A.; Rani, S. miRNA Biogenesis and Regulation of Diseases: An Updated Overview. In *MicroRNA Profiling: Methods and Protocols*; Rani, S., Ed.; Springer US: New York, NY, 2023; pp. 1–12 ISBN 978-1-0716-2823-2.
- Castanotto, D.; Rossi, J.J. The Promises and Pitfalls of RNA-Interference-Based Therapeutics. *Nature* **2009**, *457*, 426–433, doi:10.1038/nature07758.
- Kotowska-Zimmer, A.; Pewinska, M.; Olejniczak, M. Artificial miRNAs as Therapeutic Tools: Challenges and Opportunities. *WIREs RNA* **2021**, *12*, e1640, doi:10.1002/wrna.1640.
- McDonald, M.F.; Hossain, A.; Momin, E.N.; Hasan, I.; Singh, S.; Adachi, S.; Gumin, J.; Ledbetter, D.; Yang, J.; Long, L.; et al. Tumor-Specific Polycistronic miRNA Delivered by Engineered Exosomes for the Treatment of Glioblastoma. *Neuro-Oncol.* **2023**, *26*, 236–250, doi:10.1093/neuonc/noad199.
- Kliver, J.; Gibcus, J.H.; Hettinga, C.; Adema, A.; Richter, M.K.S.; Halsema, N.; Slezak-Prochazka, I.; Ding, Y.; Kroesen, B.-J.; Berg, A. van den Rapid Generation of MicroRNA Sponges for MicroRNA Inhibition. *PLOS ONE* **2012**, *7*, e29275, doi:10.1371/journal.pone.0029275.
- Rama, A.R.; Perazzoli, G.; Cabeza, L.; Mesas, C.; Quiñonero, F.; García-Pinel, B.; Vélez, C. Novel MicroRNA Sponges to Specifically Modulate Gene Expression in Colon Cancer Cells. *Nucleic Acid Ther.* **2020**, *30*, 325–334, doi:10.1089/nat.2020.0861.
- Montaño-Samaniego, M.; Sánchez-Cedillo, J.; Lucas-González, A.; Bravo-Estupiñan, D.M.; Alarcón-Hernández, E.; Rivera-Gutiérrez, S.; Balderas-López, J.A.; Ibáñez-Hernández, M. Targeted Expression to Liver of an anti-miR-33 Sponge as a Gene Therapy Strategy against Hypercholesterolemia: In Vitro Study. *Curr. Issues Mol. Biol.* **2023**, *45*, 7043–7057, doi:10.3390/cimb45090445.
- Rad, S.M.A.H.; Halpin, J.C.; Tawinwung, S.; Suppipat, K.; Hirankarn, N.; McLellan, A.D. MicroRNA-Mediated Metabolic Reprogramming of Chimeric Antigen Receptor T Cells. *Immunol. Cell Biol.* **2022**, *100*, 424–439, doi:10.1111/imcb.12551.
- Sayers, E.W.; Beck, J.; Bolton, E.E.; Brister, J.R.; Chan, J.; Comeau, D.C.; Connor, R.; DiCuccio, M.; Farrell, C.M.; Feldgarden, M.; et al. Database Resources of the National Center for Biotechnology Information. *Nucleic Acids Res.* **2023**, *52*, D33–D43, doi:10.1093/nar/gkad1044.

12. Bhaskaran, V.; Nowicki, M.O.; Idriss, M.; Jimenez, M.A.; Lugli, G.; Hayes, J.L.; Mahmoud, A.B.; Zane, R.E.; Passaro, C.; Ligon, K.L.; et al. The Functional Synergism of microRNA Clustering Provides Therapeutically Relevant Epigenetic Interference in Glioblastoma. *Nat. Commun.* **2019**, *10*, 442, doi:10.1038/s41467-019-08390-z.
13. Bhaskaran, V.; Yao, Y.; Bei, F.; Peruzzi, P. Engineering, Delivery, and Biological Validation of Artificial microRNA Clusters for Gene Therapy Applications. *Nat. Protoc.* **2019**, *14*, 3538–3553, doi:10.1038/s41596-019-0241-8.
14. Sun, D.; Melegari, Margherita; Sridhar, Sunandini; Rogler, Charles E.; and Zhu, L. Multi-miRNA Hairpin Method That Improves Gene Knockdown Efficiency and Provides Linked Multi-Gene Knockdown. *BioTechniques* **2006**, *41*, 59–63, doi:10.2144/000112203.
15. Baer, C.; Claus, R.; Frenzel, L.P.; Zucknick, M.; Park, Y.J.; Gu, L.; Weichenhan, D.; Fischer, M.; Pallasch, C.P.; Herpel, E.; et al. Extensive Promoter DNA Hypermethylation and Hypomethylation Is Associated with Aberrant MicroRNA Expression in Chronic Lymphocytic Leukemia. *Cancer Res.* **2012**, *72*, 3775–3785, doi:10.1158/0008-5472.CAN-12-0803.
16. Lujambio, A.; Ropero, S.; Ballestar, E.; Fraga, M.F.; Cerrato, C.; Setién, F.; Casado, S.; Suarez-Gauthier, A.; Sanchez-Céspedes, M.; Gitt, A.; et al. Genetic Unmasking of an Epigenetically Silenced microRNA in Human Cancer Cells. *Cancer Res.* **2007**, *67*, 1424–1429, doi:10.1158/0008-5472.CAN-06-4218.
17. Seyhan, A.A. Trials and Tribulations of MicroRNA Therapeutics. *Int. J. Mol. Sci.* **2024**, *25*, 1–41, doi:10.3390/ijms25031469.
18. Beg, M.S.; Brenner, A.J.; Sachdev, J.; Borad, M.; Kang, Y.-K.; Stoudemire, J.; Smith, S.; Bader, A.G.; Kim, S.; Hong, D.S. Phase I Study of MRX34, a Liposomal miR-34a Mimic, Administered Twice Weekly in Patients with Advanced Solid Tumors. *Invest. New Drugs* **2017**, *35*, 180–188, doi:10.1007/s10637-016-0407-y.
19. van Zandwijk, N.; Pavlakis, N.; Kao, S.C.; Linton, A.; Boyer, M.J.; Clarke, S.; Huynh, Y.; Chrzanowska, A.; Fulham, M.J.; Bailey, D.L.; et al. Safety and Activity of microRNA-Loaded Minicells in Patients with Recurrent Malignant Pleural Mesothelioma: A First-in-Man, Phase 1, Open-Label, Dose-Escalation Study. *Lancet Oncol.* **2017**, *18*, 1386–1396, doi:10.1016/S1470-2045(17)30621-6.
20. Askou, A.L.; Aagaard, L.; Kostic, C.; Arsenijevic, Y.; Hollensen, A.K.; Bek, T.; Jensen, T.G.; Mikkelsen, J.G.; Corydon, T.J. Multigenic Lentiviral Vectors for Combined and Tissue-Specific Expression of miRNA- and Protein-Based Antiangiogenic Factors. *Mol. Ther. - Methods Clin. Dev.* **2015**, *2*, 14064, doi:10.1038/mtm.2014.64.
21. Yang, X.; Marcucci, K.; Anguela, X.; Couto, L.B. Preclinical Evaluation of An Anti-HCV miRNA Cluster for Treatment of HCV Infection. *Mol. Ther.* **2013**, *21*, 588–601, doi:10.1038/mt.2012.247.
22. Chen, S.C.-Y.; Stern, P.; Guo, Z.; Chen, J. Expression of Multiple Artificial MicroRNAs from a Chicken miRNA126-Based Lentiviral Vector. *PLoS ONE* **2011**, *6*, e22437, doi:10.1371/journal.pone.0022437.
23. Liu, Y.P.; Haasnoot, J.; ter Brake, O.; Berkhout, B.; Konstantinova, P. Inhibition of HIV-1 by Multiple siRNAs Expressed from a Single microRNA Polycistron. *Nucleic Acids Res.* **2008**, *36*, 2811–2824, doi:10.1093/nar/gkn109.
24. Wang, T.; Xie, Y.; Tan, A.; Li, S.; Xie, Z. Construction and Characterization of a Synthetic MicroRNA Cluster for Multiplex RNA Interference in Mammalian Cells. *ACS Synth. Biol.* **2016**, *5*, 1193–1200, doi:10.1021/acssynbio.5b00180.
25. Cui, S.; Yu, S.; Huang, H.-Y.; Lin, Y.-C.-D.; Huang, Y.; Zhang, B.; Xiao, J.; Zuo, H.; Wang, J.; Li, Z.; et al. miRTarBase 2025: Updates to the Collection of Experimentally Validated microRNA–Target Interactions. *Nucleic Acids Res.* **2025**, *53*, D147–D156, doi:10.1093/nar/gkae1072.
26. Huang, H.-Y.; Lin, Y.-C.-D.; Li, J.; Huang, K.-Y.; Shrestha, S.; Hong, H.-C.; Tang, Y.; Chen, Y.-G.; Jin, C.-N.; Yu, Y.; et al. miRTarBase 2020: Updates to the Experimentally Validated microRNA–Target Interaction Database. *Nucleic Acids Res.* **2020**, *48*, D148–D154, doi:10.1093/nar/gkz896.
27. What Will It Take to Get miRNA Therapies to Market? *Nat. Biotechnol.* **2024**, *42*, 1623–1624, doi:10.1038/s41587-024-02480-0.
28. Machowska, M.; Galka-Marciniak, P.; Kozłowski, P. Consequences of Genetic Variants in miRNA Genes. *Comput. Struct. Biotechnol. J.* **2022**, *20*, 6443–6457, doi:10.1016/j.csbj.2022.11.036.

29. Bofill-De Ros, X.; Kasprzak, W.K.; Bhandari, Y.; Fan, L.; Cavanaugh, Q.; Jiang, M.; Dai, L.; Yang, A.; Shao, T.-J.; Shapiro, B.A.; et al. Structural Differences between Pri-miRNA Paralogs Promote Alternative Drosha Cleavage and Expand Target Repertoires. *Cell Rep.* **2019**, *26*, 447–459.e4, doi:10.1016/j.celrep.2018.12.054.
30. Fang, W.; Bartel, D.P. The Menu of Features That Define Primary MicroRNAs and Enable De Novo Design of MicroRNA Genes. *Mol. Cell* **2015**, *60*, 131–145, doi:10.1016/j.molcel.2015.08.015.
31. Zhang, D.; Zhang, N.; Shen, W.; Li, J.-F. Engineered Artificial MicroRNA Precursors Facilitate Cloning and Gene Silencing in Arabidopsis and Rice. *Int. J. Mol. Sci.* **2019**, *20*, 5620, doi:10.3390/ijms20225620.
32. Zhang, N.; Zhang, D.; Chen, S.L.; Gong, B.-Q.; Guo, Y.; Xu, L.; Zhang, X.-N.; Li, J.-F. Engineering Artificial MicroRNAs for Multiplex Gene Silencing and Simplified Transgenic Screen. *Plant Physiol.* **2018**, *178*, 989–1001, doi:10.1104/pp.18.00828.
33. Chakraborty, C.; Sharma, A.R.; Sharma, G.; Lee, S.-S. Therapeutic Advances of miRNAs: A Preclinical and Clinical Update. *J. Adv. Res.* **2021**, *28*, 127–138, doi:10.1016/j.jare.2020.08.012.
34. Iacomino, G. miRNAs: The Road from Bench to Bedside. *Genes* **2023**, *14*, 314, doi:10.3390/genes14020314.
35. Kasina, V.; Wahane, A.; Liu, C.-H.; Yang, L.; Nieh, M.-P.; Slack, F.J.; Bahal, R. Next-Generation Poly-L-Histidine Formulations for miRNA Mimic Delivery. *Mol. Ther. - Methods Clin. Dev.* **2023**, *29*, 271–283, doi:https://doi.org/10.1016/j.omtm.2023.03.015.
36. Diener, C.; Keller, A.; Meese, E. The miRNA–Target Interactions: An Underestimated Intricacy. *Nucleic Acids Res.* **2024**, *52*, 1544–1557, doi:10.1093/nar/gkad1142.
37. Jie, J.; Liu, D.; Wang, Y.; Wu, Q.; Wu, T.; Fang, R. Generation of MiRNA Sponge Constructs Targeting Multiple MiRNAs. *J. Clin. Lab. Anal.* **2022**, *36*, e24527, doi:10.1002/jcla.24527.
38. Bak, R.O.; Mikkelsen, J.G. miRNA Sponges: Soaking up miRNAs for Regulation of Gene Expression. *Wiley Interdiscip. Rev. RNA* **2014**, *5*, 317–333, doi:10.1002/wrna.1213.
39. Tay, F.C.; Lim, J.K.; Zhu, H.; Hin, L.C.; Wang, S. Using Artificial microRNA Sponges to Achieve microRNA Loss-of-Function in Cancer Cells. *Adv. Drug Deliv. Rev.* **2015**, *81*, 117–127, doi:10.1016/j.addr.2014.05.010.
40. Barta, T.; Peskova, L.; Hampl, A. miRNAsong: A Web-Based Tool for Generation and Testing of miRNA Sponge Constructs in Silico. *Sci. Rep.* **2016**, *6*, 36625, doi:10.1038/srep36625.
41. Ortega, M.M.; Bouamar, H. Guidelines on Designing MicroRNA Sponges: From Construction to Stable Cell Line. In *MicroRNA Profiling: Methods and Protocols*; Rani, S., Ed.; Springer: New York, NY, 2017; pp. 221–233 ISBN 978-1-4939-6524-3.
42. Ebert, M.S.; Sharp, P.A. MicroRNA Sponges: Progress and Possibilities. *RNA* **2010**, *16*, 2043–2050, doi:10.1261/rna.2414110.
43. Tang, L.; Chen, H.-Y.; Hao, N.-B.; Tang, B.; Guo, H.; Yong, X.; Dong, H.; Yang, S.-M. microRNA Inhibitors: Natural and Artificial Sequestration of microRNA. *Cancer Lett.* **2017**, *407*, 139–147, doi:10.1016/j.canlet.2017.05.025.
44. Ebert, M.S.; Neilson, J.R.; Sharp, P.A. MicroRNA Sponges: Competitive Inhibitors of Small RNAs in Mammalian Cells. *Nat. Methods* **2007**, *4*, 721–726, doi:10.1038/nmeth1079.
45. Alkan, A.H.; Akgül, B. Endogenous miRNA Sponges. In *miRNomics: MicroRNA Biology and Computational Analysis*; Allmer, J., Yousef, M., Eds.; Springer US: New York, NY, 2022; pp. 91–104 ISBN 978-1-0716-1170-8.
46. Le, T.D.; Zhang, J.; Liu, L.; Li, J. Computational Methods for Identifying miRNA Sponge Interactions. *Brief. Bioinform.* **2017**, *18*, 577–590, doi:10.1093/bib/bbw042.
47. Mafi, A.; Rahmati, A.; Babaei Aghdam, Z.; Salami, R.; Salami, M.; Vakili, O.; Aghadavod, E. Recent Insights into the microRNA-Dependent Modulation of Gliomas from Pathogenesis to Diagnosis and Treatment. *Cell. Mol. Biol. Lett.* **2022**, *27*, 65, doi:10.1186/s11658-022-00354-4.
48. Witten, L.; Slack, F.J. miR-155 as a Novel Clinical Target for Hematological Malignancies. *Carcinogenesis* **2020**, *41*, 2–7, doi:10.1093/carcin/bgz183.
49. Gupta, P.; Bhattacharjee, S.; Sharma, A.R.; Sharma, G.; Lee, S.-S.; Chakraborty, C. miRNAs in Alzheimer Disease – A Therapeutic Perspective. *Curr. Alzheimer Res.* **2017**, *14*, 1198–1206, doi:10.2174/1567205014666170829101016.

50. Hinkel, R.; Penzkofer, D.; Zühlke, S.; Fischer, A.; Husada, W.; Xu, Q.-F.; Baloch, E.; van Rooij, E.; Zeiher, A.M.; Kupatt, C.; et al. Inhibition of microRNA-92a Protects against Ischemia/Reperfusion Injury in a Large-Animal Model. *Circulation* **2013**, *128*, 1066–1075, doi:10.1161/CIRCULATIONAHA.113.001904.
51. Cakmak, H.A.; Coskunpinar, E.; Ikitimur, B.; Barman, H.A.; Karadag, B.; Tiryakioglu, N.O.; Kahraman, K.; Vural, V.A. The Prognostic Value of Circulating microRNAs in Heart Failure: Preliminary Results from a Genome-Wide Expression Study. *J. Cardiovasc. Med. Hagerstown Md* **2015**, *16*, 431–437, doi:10.2459/JCM.0000000000000233.
52. WHO Cardiovascular diseases Available online: [https://www.who.int/es/news-room/fact-sheets/detail/cardiovascular-diseases-\(cvds\)](https://www.who.int/es/news-room/fact-sheets/detail/cardiovascular-diseases-(cvds)) (accessed on 4 August 2024).
53. WHO La OMS Revela Las Principales Causas de Muerte y Discapacidad En El Mundo: 2000-2019 Available online: <https://www.who.int/es/news/item/09-12-2020-who-reveals-leading-causes-of-death-and-disability-worldwide-2000-2019> (accessed on 20 July 2025).
54. Aghaei, S.M.; Hosseini, S.M. Inflammation-Related miRNAs in Obesity, CVD, and NAFLD. *Cytokine* **2024**, *182*, 156724, doi:10.1016/j.cyto.2024.156724.
55. Hata, A. Functions of MicroRNAs in Cardiovascular Biology and Disease. *Annu. Rev. Physiol.* **2013**, *75*, 69–93, doi:10.1146/annurev-physiol-030212-183737.
56. Sayed, D.; Abdellatif, M. MicroRNAs in Development and Disease. *Physiol. Rev.* **2011**, *91*, 827–887, doi:10.1152/physrev.00006.2010.
57. Wronska, A. The Role of microRNA in the Development, Diagnosis, and Treatment of Cardiovascular Disease: Recent Developments. *J. Pharmacol. Exp. Ther.* **2023**, *384*, 123–132, doi:10.1124/jpet.121.001152.
58. Dong, S.; Cheng, Y.; Yang, J.; Li, J.; Liu, X.; Wang, X.; Wang, D.; Krall, T.J.; Delphin, E.S.; Zhang, C. MicroRNA Expression Signature and the Role of MicroRNA-21 in the Early Phase of Acute Myocardial Infarction. *J. Biol. Chem.* **2009**, *284*, 29514–29525, doi:10.1074/jbc.M109.027896.
59. Liu, F.; Jiang, L.; Zhang, Y.; Xu, S.; Liu, S.; Ye, J.; Liu, P. Inhibition of miR-214-3p Attenuates Ferroptosis in Myocardial Infarction via Regulating ME2. *Biochem. Biophys. Res. Commun.* **2023**, *661*, 64–74, doi:10.1016/j.bbrc.2023.04.031.
60. Funahashi, H.; Izawa, H.; Hirashiki, A.; Cheng, X.W.; Inden, Y.; Nomura, M.; Murohara, T. Altered microRNA Expression Associated with Reduced Catecholamine Sensitivity in Patients with Chronic Heart Failure. *J. Cardiol.* **2011**, *57*, 338–344, doi:10.1016/j.jjcc.2011.01.009.
61. Shi, Y.; Zhang, Z.; Yin, Q.; Fu, C.; Barszcyk, A.; Zhang, X.; Wang, J.; Yang, D. Cardiac-specific Overexpression of miR-122 Induces Mitochondria-dependent Cardiomyocyte Apoptosis and Promotes Heart Failure by Inhibiting Hand2. *J. Cell. Mol. Med.* **2021**, *25*, 5326–5334, doi:10.1111/jcmm.16544.
62. Carè, A.; Catalucci, D.; Felicetti, F.; Bonci, D.; Addario, A.; Gallo, P.; Bang, M.-L.; Segnalini, P.; Gu, Y.; Dalton, N.D.; et al. MicroRNA-133 Controls Cardiac Hypertrophy. *Nat. Med.* **2007**, *13*, 613–618, doi:10.1038/nm1582.
63. Price, N.L.; Goedeke, L.; Suárez, Y.; Fernández-Hernando, C. miR-33 in Cardiometabolic Diseases: Lessons Learned from Novel Animal Models and Approaches. *EMBO Mol. Med.* **2021**, *13*, e12606, doi:10.15252/emmm.202012606.
64. Rotllan, N.; Ramírez, C.M.; Aryal, B.; Esau, C.C.; Fernández-Hernando, C. Therapeutic Silencing of MicroRNA-33 Inhibits the Progression of Atherosclerosis in Ldlr^{-/-} Mice—Brief Report. *Arterioscler. Thromb. Vasc. Biol.* **2013**, *33*, 1973–1977, doi:10.1161/ATVBAHA.113.301732.
65. Goedeke, L.; Salerno, A.; Ramírez, C.M.; Guo, L.; Allen, R.M.; Yin, X.; Langley, S.R.; Esau, C.; Wanschel, A.; Fisher, E.A.; et al. Long-term Therapeutic Silencing of miR-33 Increases Circulating Triglyceride Levels and Hepatic Lipid Accumulation in Mice. *EMBO Mol. Med.* **2014**, *6*, 1133–1141, doi:10.15252/emmm.201404046.
66. Price, N.L.; Singh, A.K.; Rotllan, N.; Goedeke, L.; Wing, A.; Canfrán-Duque, A.; Diaz-Ruiz, A.; Araldi, E.; Baldán, Á.; Camporez, J.-P.; et al. Genetic Ablation of miR-33 Increases Food Intake, Enhances Adipose Tissue Expansion, and Promotes Obesity and Insulin Resistance. *Cell Rep.* **2018**, *22*, 2133–2145, doi:10.1016/j.celrep.2018.01.074.

67. Price, N.L.; Zhang, X.; Fernández-Tussy, P.; Singh, A.K.; Burnap, S.A.; Rotllan, N.; Goedeke, L.; Sun, J.; Canfrán-Duque, A.; Aryal, B.; et al. Loss of Hepatic miR-33 Improves Metabolic Homeostasis and Liver Function without Altering Body Weight or Atherosclerosis. *Proc. Natl. Acad. Sci.* **2021**, *118*, e2006478118, doi:10.1073/pnas.2006478118.
68. Liu, Y.; Song, J.-W.; Lin, J.-Y.; Miao, R.; Zhong, J.-C. Roles of MicroRNA-122 in Cardiovascular Fibrosis and Related Diseases. *Cardiovasc. Toxicol.* **2020**, *20*, 463–473, doi:10.1007/s12012-020-09603-4.
69. Šatrauskienė, A.; Navickas, R.; Laucevičius, A.; Krilavičius, T.; Užupytė, R.; Zdanytė, M.; Ryliškytė, L.; Jucevičienė, A.; Holvoet, P. Mir-1, miR-122, miR-132, and miR-133 Are Related to Subclinical Aortic Atherosclerosis Associated with Metabolic Syndrome. *Int. J. Environ. Res. Public. Health* **2021**, *18*, 1483, doi:10.3390/ijerph18041483.
70. Song, W.; Zhang, Tiening; Yang, Ni; Zhang, Tao; Wen, Ri; and Liu, C. Inhibition of Micro RNA miR-122-5p Prevents Lipopolysaccharide-Induced Myocardial Injury by Inhibiting Oxidative Stress, Inflammation and Apoptosis via Targeting GIT1. *Bioengineered* **2021**, *12*, 1902–1915, doi:10.1080/21655979.2021.1926201.
71. Hsu, S.; Wang, B.; Kota, J.; Yu, J.; Costinean, S.; Kutay, H.; Yu, L.; Bai, S.; La Perle, K.; Chivukula, R.R.; et al. Essential Metabolic, Anti-Inflammatory, and Anti-Tumorigenic Functions of miR-122 in Liver. *J. Clin. Invest.* **2012**, *122*, 2871–2883, doi:10.1172/JCI63539.
72. Thakral, S.; Ghoshal, K. miR-122 Is a Unique Molecule with Great Potential in Diagnosis, Prognosis of Liver Disease, and Therapy Both as miRNA Mimic and Antimir. *Curr. Gene Ther.* **2015**, *15*, 142–150.
73. Eshraghi, R.; Rafiei, M.; Hadian Jazi, Z.; Shafie, D.; Raisi, A.; Mirzaei, H. MicroRNA-155 and Exosomal microRNA-155: Small Pieces in the Cardiovascular Diseases Puzzle. *Pathol. Res. Pract.* **2024**, *257*, 155274, doi:10.1016/j.prp.2024.155274.
74. Gangwar, R.S.; Rajagopalan, S.; Natarajan, R.; Deiuliis, J.A. Noncoding RNAs in Cardiovascular Disease: Pathological Relevance and Emerging Role as Biomarkers and Therapeutics. *Am. J. Hypertens.* **2018**, *31*, 150–165, doi:10.1093/ajh/hpx197.
75. Raitoharju, E.; Lyytikäinen, L.-P.; Levula, M.; Oksala, N.; Mennander, A.; Tarkka, M.; Klopp, N.; Illig, T.; Kähönen, M.; Karhunen, P.J.; et al. miR-21, miR-210, miR-34a, and miR-146a/b Are up-Regulated in Human Atherosclerotic Plaques in the Tampere Vascular Study. *Atherosclerosis* **2011**, *219*, 211–217, doi:10.1016/j.atherosclerosis.2011.07.020.
76. Holland, A.; Enrick, M.; Diaz, A.; Yin, L. Is miR-21 A Therapeutic Target in Cardiovascular Disease? *Int. J. Drug Discov. Pharmacol.* **2023**, *2*, 26–36, doi:10.53941/ijddp.0201003.
77. Huang, C.-K.; Bär, C.; Thum, T. miR-21, Mediator, and Potential Therapeutic Target in the Cardiorenal Syndrome. *Front. Pharmacol.* **2020**, *11*, doi:10.3389/fphar.2020.00726.
78. Laggerbauer, B.; Engelhardt, S. MicroRNAs as Therapeutic Targets in Cardiovascular Disease. *J. Clin. Invest.* **2022**, *132*, doi:10.1172/JCI159179.
79. Ramanujam, D.; Sassi, Y.; Laggerbauer, B.; Engelhardt, S. Viral Vector-Based Targeting of miR-21 in Cardiac Nonmyocyte Cells Reduces Pathologic Remodeling of the Heart. *Mol. Ther.* **2016**, *24*, 1939–1948, doi:10.1038/mt.2016.166.
80. Thum, T.; Gross, C.; Fiedler, J.; Fischer, T.; Kissler, S.; Bussen, M.; Galuppo, P.; Just, S.; Rottbauer, W.; Frantz, S.; et al. MicroRNA-21 Contributes to Myocardial Disease by Stimulating MAP Kinase Signalling in Fibroblasts. *Nature* **2008**, *456*, 980–984, doi:10.1038/nature07511.
81. Zhao, Y.; Ponnusamy, M.; Zhang, L.; Zhang, Y.; Liu, C.; Yu, W.; Wang, K.; Li, P. The Role of miR-214 in Cardiovascular Diseases. *Eur. J. Pharmacol.* **2017**, *816*, 138–145, doi:10.1016/j.ejphar.2017.08.009.
82. Duan, Q.; Yang, L.; Gong, W.; Chaugai, S.; Wang, F.; Chen, C.; Wang, P.; Zou, M.-H.; Wang, D.W. MicroRNA-214 Is Upregulated in Heart Failure Patients and Suppresses XBP1-Mediated Endothelial Cells Angiogenesis. *J. Cell. Physiol.* **2015**, *230*, 1964–1973, doi:10.1002/jcp.24942.
83. Feng, Y.; Wan, P.; Yin, L. Long Noncoding RNA X-Inactive Specific Transcript (XIST) Promotes Osteogenic Differentiation of Periodontal Ligament Stem Cells by Sponging MicroRNA-214-3p. *Med. Sci. Monit. Int. Med. J. Exp. Clin. Res.* **2020**, *26*, e918932-1-e918932-8, doi:10.12659/MSM.918932.
84. Huang, X.-Z.; Huang, J.; Li, W.-Z.; Wang, J.-J.; Song, D.-Y.; Ni, J.-D. LncRNA-MALAT1 Promotes Osteogenic Differentiation through Regulating ATF4 by Sponging miR-214: Implication of Steroid-Induced Avascular Necrosis of the Femoral Head. *Steroids* **2020**, *154*, 108533, doi:10.1016/j.steroids.2019.108533.

85. Bernardo, B.C.; Ooi, J.Y.Y.; Matsumoto, A.; Tham, Y.K.; Singla, S.; Kiriazis, H.; Patterson, N.L.; Sadoshima, J.; Obad, S.; Lin, R.C.Y.; et al. Sex Differences in Response to miRNA-34a Therapy in Mouse Models of Cardiac Disease: Identification of Sex-, Disease- and Treatment-Regulated miRNAs. *J. Physiol.* **2016**, *594*, 5959–5974, doi:10.1113/JP272512.
86. Hua, C.-C.; Liu, X.-M.; Liang, L.-R.; Wang, L.-F.; Zhong, J.-C. Targeting the microRNA-34a as a Novel Therapeutic Strategy for Cardiovascular Diseases. *Front. Cardiovasc. Med.* **2022**, *8*, doi:10.3389/fcvm.2021.784044.
87. Huang, Y.; Qi, Yuan; Du, Jian-Qing; and Zhang, D. MicroRNA-34a Regulates Cardiac Fibrosis after Myocardial Infarction by Targeting Smad4. *Expert Opin. Ther. Targets* **2014**, *18*, 1355–1365, doi:10.1517/14728222.2014.961424.
88. Yang, Y.; Cheng, H.-W.; Qiu, Y.; Dupee, D.; Noonan, M.; Lin, Y.-D.; Fisch, S.; Unno, K.; Sereti, K.-I.; Liao, R. MicroRNA-34a Plays a Key Role in Cardiac Repair and Regeneration Following Myocardial Infarction. *Circ. Res.* **2015**, *117*, 450–459, doi:10.1161/CIRCRESAHA.117.305962.
89. Meloni, M.; Marchetti, M.; Garner, K.; Littlejohns, B.; Sala-Newby, G.; Xenophontos, N.; Floris, I.; Suleiman, M.-S.; Madeddu, P.; Caporali, A.; et al. Local Inhibition of MicroRNA-24 Improves Reparative Angiogenesis and Left Ventricle Remodeling and Function in Mice With Myocardial Infarction. *Mol. Ther.* **2013**, *21*, 1390–1402, doi:10.1038/mt.2013.89.
90. Misso, G.; Di Martino, M.T.; De Rosa, G.; Farooqi, A.A.; Lombardi, A.; Campani, V.; Zarone, M.R.; Gullà, A.; Tagliaferri, P.; Tassone, P.; et al. Mir-34: A New Weapon Against Cancer? *Mol. Ther. - Nucleic Acids* **2014**, *3*, e195, doi:10.1038/mtna.2014.47.
91. Bruen, R.; Fitzsimons, S.; Belton, O. miR-155 in the Resolution of Atherosclerosis. *Front. Pharmacol.* **2019**, *10*, 463, doi:10.3389/fphar.2019.00463.
92. Heymans, S.; Corsten, M.F.; Verhesen, W.; Carai, P.; van Leeuwen, R.E.W.; Custers, K.; Peters, T.; Hazebroek, M.; Stöger, L.; Wijnands, E.; et al. Macrophage microRNA-155 Promotes Cardiac Hypertrophy and Failure. *Circulation* **2013**, *128*, 1420–1432, doi:10.1161/CIRCULATIONAHA.112.001357.
93. Li, X.; Kong, D.; Chen, H.; Liu, S.; Hu, H.; Wu, T.; Wang, J.; Chen, W.; Ning, Y.; Li, Y.; et al. miR-155 Acts as an Anti-Inflammatory Factor in Atherosclerosis-Associated Foam Cell Formation by Repressing Calcium-Regulated Heat Stable Protein 1. *Sci. Rep.* **2016**, *6*, 21789, doi:10.1038/srep21789.
94. Yang, Y.; Yang, L.; Liang, X.; Zhu, G. MicroRNA-155 Promotes Atherosclerosis Inflammation via Targeting SOCS1. *Cell. Physiol. Biochem.* **2015**, *36*, 1371–1381, doi:10.1159/000430303.
95. Chung, K.-H.; Hart, C.C.; Al-Bassam, S.; Avery, A.; Taylor, J.; Patel, P.D.; Vojtek, A.B.; Turner, D.L. Polycistronic RNA Polymerase II Expression Vectors for RNA Interference Based on BIC/miR-155. *Nucleic Acids Res.* **2006**, *34*, e53, doi:10.1093/nar/gkl143.
96. WHO Obesity and overweight Available online: <https://www.who.int/es/news-room/fact-sheets/detail/obesity-and-overweight> (accessed on 10 August 2024).
97. Rahman, Md.A.; Islam, Md.M.; Ripon, Md.A.R.; Islam, Md.M.; Hossain, M.S. Regulatory Roles of MicroRNAs in the Pathogenesis of Metabolic Syndrome. *Mol. Biotechnol.* **2024**, *66*, 1599–1620, doi:10.1007/s12033-023-00805-z.
98. Masoumi-Ardakani, Y.; Eghbalian, M.; Fallah, H.; Jafari, A.; Shahouzehi, B. Exploring Serum miR-33b as a Novel Diagnostic Marker for Hypercholesterolemia and Obesity: Insights from a Pilot Case-Control Study. *BMC Endocr. Disord.* **2025**, *25*, 27, doi:10.1186/s12902-025-01849-9.
99. Elkhawaga, S.Y.; Ismail, A.; Elsakka, E.G.E.; Doghish, A.S.; Elkady, M.A.; El-Mahdy, H.A. miRNAs as Cornerstones in Adipogenesis and Obesity. *Life Sci.* **2023**, *315*, 121382, doi:10.1016/j.lfs.2023.121382.
100. Ibarra, P.E.; García-Solís, P.; Solís-Sáinz, J.C.; Cruz-Hernández, A. Expression of miRNA in Obesity and Insulin Resistance: A Review. *Endokrynol. Pol.* **2021**, *72*, 73–80, doi:10.5603/EP.a2021.0002.
101. Kilic, I.D.; Dodurga, Y.; Uludag, B.; Alihanoglu, Y.I.; Yildiz, B.S.; Enli, Y.; Secme, M.; Bostancı, H.E. microRNA -143 and -223 in Obesity. *Gene* **2015**, *560*, 140–142, doi:10.1016/j.gene.2015.01.048.
102. Silveira, A.; Gomes, J.; Roque, F.; Fernandes, T.; de Oliveira, E.M. MicroRNAs in Obesity-Associated Disorders: The Role of Exercise Training. *Obes. Facts* **2022**, *15*, 105–117, doi:10.1159/000517849.

103. Fernández-Tussy, P.; Cardelo, M.P.; Zhang, H.; Sun, J.; Price, N.L.; Boutagy, N.E.; Goedeke, L.; Cadena-Sandoval, M.; Xirouchaki, C.E.; Brown, W.; et al. miR-33 Deletion in Hepatocytes Attenuates MASLD-MASH-HCC Progression. *JCI Insight* **2024**, *9*, e168476, doi:10.1172/jci.insight.168476.
104. Price, N.L.; Fernández-Tussy, P.; Varela, L.; Cardelo, M.P.; Shanabrough, M.; Aryal, B.; de Cabo, R.; Suárez, Y.; Horvath, T.L.; Fernández-Hernando, C. microRNA-33 Controls Hunger Signaling in Hypothalamic AgRP Neurons. *Nat. Commun.* **2024**, *15*, 2131, doi:10.1038/s41467-024-46427-0.
105. Liu, J.; Wang, H.; Zeng, D.; Xiong, J.; Luo, J.; Chen, X.; Chen, T.; Xi, Q.; Sun, J.; Ren, X.; et al. The Novel Importance of miR-143 in Obesity Regulation. *Int. J. Obes.* **2023**, *47*, 100–108, doi:10.1038/s41366-022-01245-6.
106. Yi, C.; Xie, W.; Li, F.; Lv, Q.; He, J.; Wu, J.; Gu, D.; Xu, N.; Zhang, Y. MiR-143 Enhances Adipogenic Differentiation of 3T3-L1 Cells through Targeting the Coding Region of Mouse Pleiotrophin. *FEBS Lett.* **2011**, *585*, 3303–3309, doi:10.1016/j.febslet.2011.09.015.
107. Zarkesh, M.; Tabaei, K.; Akbarzadeh, M.; Daneshafrooz, A.; Zadeh-Vakili, A. Association of miR-34a and miR-143 Levels with PPAR γ Gene Expression in Adipose Tissues of Non-Diabetic Adults. *J. Physiol. Anthropol.* **2022**, *41*, 13, doi:10.1186/s40101-022-00286-0.
108. Esau, C.; Kang, X.; Peralta, E.; Hanson, E.; Marcusson, E.G.; Ravichandran, L.V.; Sun, Y.; Koo, S.; Perera, R.J.; Jain, R.; et al. MicroRNA-143 Regulates Adipocyte Differentiation. *J. Biol. Chem.* **2004**, *279*, 52361–52365, doi:10.1074/jbc.C400438200.
109. Liu, J.; Liu, J.; Zeng, D.; Wang, H.; Wang, Y.; Xiong, J.; Chen, X.; Luo, J.; Chen, T.; Xi, Q.; et al. miR-143-Null Is against Diet-Induced Obesity by Promoting BAT Thermogenesis and Inhibiting WAT Adipogenesis. *Int. J. Mol. Sci.* **2022**, *23*, 13058, doi:10.3390/ijms232113058.
110. Lin, X.; Du, Y.; Lu, W.; Gui, W.; Sun, S.; Zhu, Y.; Wang, G.; Eserberg, D.T.; Zheng, F.; Zhou, J.; et al. CircRNF111 Protects Against Insulin Resistance and Lipid Deposition via Regulating miR-143-3p/IGF2R Axis in Metabolic Syndrome. *Front. Cell Dev. Biol.* **2021**, *9*, doi:10.3389/fcell.2021.663148.
111. Lhamyani, S.; Gentile, A.-M.; Giráldez-Pérez, R.M.; Feijóo-Cuaresma, M.; Romero-Zerbo, S.Y.; Clemente-Postigo, M.; Zayed, H.; Oliva-Olivera, W.; Bermúdez-Silva, F.J.; Salas, J.; et al. miR-21 Mimic Blocks Obesity in Mice: A Novel Therapeutic Option. *Mol. Ther. - Nucleic Acids* **2021**, *26*, 401–416, doi:10.1016/j.omtn.2021.06.019.
112. Sekar, D.; Venugopal, B.; Sekar, P.; Ramalingam, K. Role of microRNA 21 in Diabetes and Associated/Related Diseases. *Gene* **2016**, *582*, 14–18, doi:10.1016/j.gene.2016.01.039.
113. Seeger, T.; Fischer, A.; Muhly-Reinholz, M.; Zeiher, A.M.; Dimmeler, S. Long-Term Inhibition of miR-21 Leads to Reduction of Obesity in Db/Db Mice. *Obesity* **2014**, *22*, 2352–2360, doi:10.1002/oby.20852.
114. Monfared, H.; Jahangard, Y.; Nikkhah, M.; Mirnajafi-Zadeh, J.; Mowla, S.J. Potential Therapeutic Effects of Exosomes Packed With a miR-21-Sponge Construct in a Rat Model of Glioblastoma. *Front. Oncol.* **2019**, *9*, 782, doi:10.3389/fonc.2019.00782.
115. Ma, F.; Cao, D.; Liu, Z.; Li, Y.; Ouyang, S.; Wu, J. Identification of Novel Circulating miRNAs Biomarkers for Healthy Obese and Lean Children. *BMC Endocr. Disord.* **2023**, *23*, 238, doi:10.1186/s12902-023-01498-w.
116. de Almeida-Faria, J.; Duque-Guimarães, D.E.; Ong, T.P.; Pantaleão, L.C.; Carpenter, A.A.; Loche, E.; Kusinski, L.C.; Ashmore, T.J.; Antrobus, R.; Bushell, M.; et al. Maternal Obesity during Pregnancy Leads to Adipose Tissue ER Stress in Mice via miR-126-Mediated Reduction in Lunapark. *Diabetologia* **2021**, *64*, 890–902, doi:10.1007/s00125-020-05357-4.
117. Shen, L.; He, J.; Zhao, Y.; Niu, L.; Chen, L.; Tang, G.; Jiang, Y.; Hao, X.; Bai, L.; Li, X.; et al. MicroRNA-126b-5p Exacerbates Development of Adipose Tissue and Diet-Induced Obesity. *Int. J. Mol. Sci.* **2021**, *22*, 10261, doi:10.3390/ijms221910261.
118. Wang, Y.; Yang, L.-Z.; Yang, D.-G.; Zhang, Q.-Y.; Deng, Z.-N.; Wang, K.; Mao, X.-J. MiR-21 Antagomir Improves Insulin Resistance and Lipid Metabolism Disorder in Streptozotocin-Induced Type 2 Diabetes Mellitus Rats. *Ann. Palliat. Med.* **2020**, *9*, 394–404, doi:10.21037/apm.2020.02.28.
119. Liu, R.; Liu, C.; He, X.; Sun, P.; Zhang, B.; Yang, H.; Shi, W.; Ruan, Q. MicroRNA-21 Promotes Pancreatic β Cell Function through Modulating Glucose Uptake. *Nat. Commun.* **2022**, *13*, 3545, doi:10.1038/s41467-022-31317-0.

120. Dey, N.; Das, F.; Mariappan, M.M.; Mandal, C.C.; Ghosh-Choudhury, N.; Kasinath, B.S.; Choudhury, G.G. MicroRNA-21 Orchestrates High Glucose-Induced Signals to TOR Complex 1, Resulting in Renal Cell Pathology in Diabetes. *J. Biol. Chem.* **2011**, *286*, 25586–25603, doi:10.1074/jbc.M110.208066.
121. Zhong, X.; Chung, A.C.K.; Chen, H.Y.; Dong, Y.; Meng, X.M.; Li, R.; Yang, W.; Hou, F.F.; Lan, H.Y. miR-21 Is a Key Therapeutic Target for Renal Injury in a Mouse Model of Type 2 Diabetes. *Diabetologia* **2013**, *56*, 663–674, doi:10.1007/s00125-012-2804-x.
122. Mostafa, A.; Abusree Ahmed, A.; Hassanien, R.T.M.; Mahfouz, H.; Salah, M.; Amr, H.M.; Fahim, S.A. Emerging Role of *Hsa_circ_0000652*, *Hsa-miR-21*, *SMAD2*, and *Foxo1* in Type 2 Diabetes Mellitus Pathogenesis. *Hum. Gene* **2025**, *43*, 201386, doi:10.1016/j.humgen.2025.201386.
123. Ghaffari, M.; Razi, S.; Zalpoor, H.; Nabi-Afjadi, M.; Mohebichamkhorami, F.; Zali, H. Association of MicroRNA-146a with Type 1 and 2 Diabetes and Their Related Complications. *J. Diabetes Res.* **2023**, *2023*, 2587104, doi:10.1155/2023/2587104.
124. Runtsch, M.C.; Nelson, M.C.; Lee, S.-H.; Voth, W.; Alexander, M.; Hu, R.; Wallace, J.; Petersen, C.; Panic, V.; Villanueva, C.J.; et al. Anti-Inflammatory microRNA-146a Protects Mice from Diet-Induced Metabolic Disease. *PLoS Genet.* **2019**, *15*, e1007970, doi:10.1371/journal.pgen.1007970.
125. Peng, X.; He, F.; Mao, Y.; Lin, Y.; Fang, J.; Chen, Y.; Sun, Z.; Zhuo, Y.; Jiang, J. miR-146a Promotes M2 Macrophage Polarization and Accelerates Diabetic Wound Healing by Inhibiting the TLR4/NF- κ B Axis. **2022**, doi:10.1530/JME-21-0019.
126. Roos, J.; Dahlhaus, M.; Funcke, J.-B.; Kustermann, M.; Strauss, G.; Halbgebauer, D.; Boldrin, E.; Holzmann, K.; Möller, P.; Trojanowski, B.M.; et al. miR-146a Regulates Insulin Sensitivity via NPR3. *Cell. Mol. Life Sci. CMLS* **2021**, *78*, 2987–3003, doi:10.1007/s00018-020-03699-1.
127. Rohm, T.V.; Castellani Gomes Dos Reis, F.; Isaac, R.; Murphy, C.; Cunha e Rocha, K.; Bandyopadhyay, G.; Gao, H.; Libster, A.M.; Zapata, R.C.; Lee, Y.S.; et al. Adipose Tissue Macrophages Secrete Small Extracellular Vesicles That Mediate Rosiglitazone-Induced Insulin Sensitization. *Nat. Metab.* **2024**, *6*, 880–898, doi:10.1038/s42255-024-01023-w.
128. Ying, W.; Gao, H.; Dos Reis, F.C.G.; Bandyopadhyay, G.; Ofrecio, J.M.; Luo, Z.; Ji, Y.; Jin, Z.; Ly, C.; Olefsky, J.M. MiR-690, an Exosomal-Derived miRNA from M2-Polarized Macrophages, Improves Insulin Sensitivity in Obese Mice. *Cell Metab.* **2021**, *33*, 781–790.e5, doi:10.1016/j.cmet.2020.12.019.
129. Shi, Z.; Zhao, C.; Guo, X.; Ding, H.; Cui, Y.; Shen, R.; Liu, J. Differential Expression of MicroRNAs in Omental Adipose Tissue From Gestational Diabetes Mellitus Subjects Reveals miR-222 as a Regulator of ER α Expression in Estrogen-Induced Insulin Resistance. *Endocrinology* **2014**, *155*, 1982–1990, doi:10.1210/en.2013-2046.
130. Raimondi, G.; Gea-Sorlí, S.; Otero-Mateo, M.; Fillat, C. Inhibition of miR-222 by Oncolytic Adenovirus-Encoded miRNA Sponges Promotes Viral Oncolysis and Elicits Antitumor Effects in Pancreatic Cancer Models. *Cancers* **2021**, *13*, 3233, doi:10.3390/cancers13133233.
131. Zhou, L.; Jiang, F.; Chen, X.; Liu, Z.; Ouyang, Y.; Zhao, W.; Yu, D. Downregulation of miR-221/222 by a microRNA Sponge Promotes Apoptosis in Oral Squamous Cell Carcinoma Cells through Upregulation of PTEN. *Oncol. Lett.* **2016**, *12*, 4419–4426, doi:10.3892/ol.2016.5250.
132. Chuang, T.-Y.; Wu, H.-L.; Chen, C.-C.; Gamboa, G.M.; Layman, L.C.; Diamond, M.P.; Azziz, R.; Chen, Y.-H. MicroRNA-223 Expression Is Upregulated in Insulin Resistant Human Adipose Tissue. *J. Diabetes Res.* **2015**, *2015*, 943659, doi:10.1155/2015/943659.
133. Deiluiis, J.A.; Syed, R.; Duggineni, D.; Rutsky, J.; Rengasamy, P.; Zhang, J.; Huang, K.; Needleman, B.; Mikami, D.; Perry, K.; et al. Visceral Adipose MicroRNA 223 Is Upregulated in Human and Murine Obesity and Modulates the Inflammatory Phenotype of Macrophages. *PloS One* **2016**, *11*, e0165962, doi:10.1371/journal.pone.0165962.
134. Sánchez-Ceinos, J.; Rangel-Zuñiga, O.A.; Clemente-Postigo, M.; Podadera-Herreros, A.; Camargo, A.; Alcalá-Díaz, J.F.; Guzmán-Ruiz, R.; López-Miranda, J.; Malagón, M.M. miR-223-3p as a Potential Biomarker and Player for Adipose Tissue Dysfunction Preceding Type 2 Diabetes Onset. *Mol. Ther. - Nucleic Acids* **2021**, *23*, 1035–1052, doi:10.1016/j.omtn.2021.01.014.

135. Huerta-Zavala, M.L.; Lopez-Castillejos, E.S.; Requenez-Contreras, J.L.; Granados-Riveron, J.T.; Aquino-Jarquín, G. A Single miRNA and miRNA Sponge Expression System for Efficient Modulation of miR-223 Availability in Mammalian Cells. *J. Gene Med.* **2019**, *21*, e3100, doi:10.1002/jgm.3100.
136. M'baya-Moutoula, E.; Marchand, A.; Six, I.; Bahrar, N.; Celic, T.; Mougenot, N.; Maitrias, P.; Massy, Z.A.; Lompré, A.-M.; Metzinger, L.; et al. Inhibition of miR-223 Expression Using a Sponge Strategy Decreases Restenosis in Rat Injured Carotids. *Curr. Vasc. Pharmacol.* **2020**, *18*, 507–516, doi:10.2174/1570161117666190705141152.
137. Gao, F.; Wei, L.; Li, J.; Zeng, L.; Wang, M.; Lan, P.; Liang, S.; Huang, X.; Chen, L.; Jiang, H. Circular RNA CircVmn2r1 Acts as a miR-223-3p Sponge to Promote Kidney Aging by Regulating NLRP3 Expression in Mice. *J. Gerontol. Ser. A* **2024**, *79*, glae067, doi:10.1093/gerona/glae067.
138. Hou, Y.; Sun, J.; Huang, J.; Yao, F.; Chen, X.; Zhu, B.; Zhao, D. Circular RNA circRNA_0000094 Sponges microRNA-223-3p and up-Regulate F-Box and WD Repeat Domain Containing 7 to Restrain T Cell Acute Lymphoblastic Leukemia Progression. *Hum. Cell* **2021**, *34*, 977–989, doi:10.1007/s13577-021-00504-4.
139. Ji, D.; Chen, G.-F.; Wang, J.-C.; Ji, S.-H.; Wu, X.-W.; Lu, X.-J.; Chen, J.-L.; Li, J.-T. Hsa_circ_0070963 Inhibits Liver Fibrosis via Regulation of miR-223-3p and LEMD3. *Aging* **2020**, *12*, 1643–1655, doi:10.18632/aging.102705.
140. Wu, H.; Dai, Y.; Zhang, D.; Zhang, X.; He, Z.; Xie, X.; Cai, C. LINC00961 Inhibits the Migration and Invasion of Colon Cancer Cells by Sponging miR-223-3p and Targeting SOX11. *Cancer Med.* **2020**, *9*, 2514–2523, doi:10.1002/cam4.2850.
141. Chen, W.; Xu, J.; Wu, Y.; Liang, B.; Yan, M.; Sun, C.; Wang, D.; Hu, X.; Liu, L.; Hu, W.; et al. The Potential Role and Mechanism of circRNA/miRNA Axis in Cholesterol Synthesis. *Int. J. Biol. Sci.* **2023**, *19*, 2879–2896, doi:10.7150/ijbs.84994.
142. Mirzaei, R.; Karampoor, S.; Korotkova, N.L. The Emerging Role of miRNA-122 in Infectious Diseases: Mechanisms and Potential Biomarkers. *Pathol. Res. Pract.* **2023**, *249*, 154725, doi:10.1016/j.prp.2023.154725.
143. Wang, X.; Zhang, H.; Yang, H.; Bai, M.; Ning, T.; Deng, T.; Liu, R.; Fan, Q.; Zhu, K.; Li, J.; et al. Exosome-delivered circRNA Promotes Glycolysis to Induce Chemoresistance through the miR-122-PKM2 Axis in Colorectal Cancer. *Mol. Oncol.* **2020**, *14*, 539–555, doi:10.1002/1878-0261.12629.
144. Xu, Y.; Kong, S.; Qin, S.; Shen, X.; Ju, S. Exosomal circRNAs: Sorting Mechanisms, Roles and Clinical Applications in Tumors. *Front. Cell Dev. Biol.* **2020**, *8*, 581558, doi:10.3389/fcell.2020.581558.
145. Yang, F.; Chen, Y.; Luo, L.; Nong, S.; Li, T. circFOXO3 Induced by KLF16 Modulates Clear Cell Renal Cell Carcinoma Growth and Natural Killer Cell Cytotoxic Activity through Sponging miR-29a-3p and miR-122-5p. *Dis. Markers* **2022**, *2022*, 6062236, doi:10.1155/2022/6062236.
146. Fernández-Tussy, P.; Ruz-Maldonado, I.; Fernández-Hernando, C. MicroRNAs and Circular RNAs in Lipoprotein Metabolism. *Curr. Atheroscler. Rep.* **2021**, *23*, 33, doi:10.1007/s11883-021-00934-3.
147. Ortega, R.; Liu, B.; Persaud, S.J. Effects of miR-33 Deficiency on Metabolic and Cardiovascular Diseases: Implications for Therapeutic Intervention. *Int. J. Mol. Sci.* **2023**, *24*, 10777, doi:10.3390/ijms241310777.
148. Goedeke, L.; Rotllán, N.; Ramírez, C.M.; Aranda, J.F.; Canfrán-Duque, A.; Araldi, E.; Fernández-Hernando, A.; Langhi, C.; de Cabo, R.; Baldán, Á.; et al. miR-27b Inhibits LDLR and ABCA1 Expression but Does Not Influence Plasma and Hepatic Lipid Levels in Mice. *Atherosclerosis* **2015**, *243*, 499–509, doi:10.1016/j.atherosclerosis.2015.09.033.
149. Vickers, K.C.; Shoucri, B.M.; Levin, M.G.; Wu, H.; Pearson, D.S.; Osei-Hwedie, D.; Collins, F.S.; Remaley, A.T.; Sethupathy, P. MicroRNA-27b Is a Regulatory Hub in Lipid Metabolism and Is Altered in Dyslipidemia. *Hepatology* **2013**, *57*, 533–542, doi:10.1002/hep.25846.
150. Wang, X.; Lu, Y.; Zhu, L.; Zhang, H.; Feng, L. Inhibition of miR-27b Regulates Lipid Metabolism in Skeletal Muscle of Obese Rats During Hypoxic Exercise by Increasing PPAR γ Expression. *Front. Physiol.* **2020**, *11*, 1090, doi:10.3389/fphys.2020.01090.
151. Galicia, U.; Benito-Vicente, A.; Jebari-Benslaïman, S.; Belloso-Urbe, K.; Larrea, A.; José, A.S.; Plagaro, C.M. Statin-Induced miR-33a and miR-27b up-Regulation Contributes to the Development of New-Onset Type 2 Diabetes. *Atherosclerosis* **2023**, *379*, S30, doi:10.1016/j.atherosclerosis.2023.06.766.
152. Testa, U.; Pelosi, E.; Castelli, G.; Labbaye, C. miR-146 and miR-155: Two Key Modulators of Immune Response and Tumor Development. *Non-Coding RNA* **2017**, *3*, 22, doi:10.3390/ncrna3030022.

153. Zhou, C.; Zhao, L.; Wang, K.; Qi, Q.; Wang, M.; Yang, L.; Sun, P.; Mu, H. MicroRNA-146a Inhibits NF- κ B Activation and pro-Inflammatory Cytokine Production by Regulating IRAK1 Expression in THP-1 Cells. *Exp. Ther. Med.* **2019**, *18*, 3078–3084, doi:10.3892/etm.2019.7881.
154. Maciak, K.; Dziedzic, A.; Miller, E.; Saluk-Bijak, J. miR-155 as an Important Regulator of Multiple Sclerosis Pathogenesis. A Review. *Int. J. Mol. Sci.* **2021**, *22*, 4332, doi:10.3390/ijms22094332.
155. Garchow, B.; Kiriakidou, M. MicroRNA-21 Deficiency Protects from Lupus-like Autoimmunity in the Chronic Graft-Versus-Host Disease Model of Systemic Lupus Erythematosus. *Clin. Immunol. Orlando Fla* **2016**, *162*, 100–106, doi:10.1016/j.clim.2015.11.010.
156. Neudecker, V.; Haneklaus, M.; Jensen, O.; Khailova, L.; Masterson, J.C.; Tye, H.; Biette, K.; Jedlicka, P.; Brodsky, K.S.; Gerich, M.E.; et al. Myeloid-Derived miR-223 Regulates Intestinal Inflammation via Repression of the NLRP3 Inflammasome. *J. Exp. Med.* **2017**, *214*, 1737–1752, doi:10.1084/jem.20160462.
157. Kourti, M.; Sokratous, M.; Katsiari, C.G. Regulation of microRNA in Systemic Lupus Erythematosus: The Role of miR-21 and miR-210. *Mediterr. J. Rheumatol.* **2020**, *31*, 71–74, doi:10.31138/mjr.31.1.71.
158. Gaudet, A.D.; Fonken, L.K.; Watkins, L.R.; Nelson, R.J.; Popovich, P.G. MicroRNAs: Roles in Regulating Neuroinflammation. *The Neuroscientist* **2018**, *24*, 221–245, doi:10.1177/1073858417721150.
159. Kablak-Ziemicka, A.; Badacz, R.; Okarski, M.; Wawak, M.; Przewłocki, T.; Podolec, J. Cardiac microRNAs: Diagnostic and Therapeutic Potential. *Arch. Med. Sci. AMS* **2023**, *19*, 1360–1381, doi:10.5114/aoms/169775.
160. Panigrahi, M.; Palmer, M.A.; Wilson, J.A. MicroRNA-122 Regulation of HCV Infections: Insights from Studies of miR-122-Independent Replication. *Pathogens* **2022**, *11*, 1005, doi:10.3390/pathogens11091005.
161. Abidin, S.Z.; Mat Pauzi, N.A.; Mansor, N.I.; Mohd Isa, N.I.; Hamid, A.A. A New Perspective on Alzheimer's Disease: microRNAs and Circular RNAs. *Front. Genet.* **2023**, *14*, 1231486, doi:10.3389/fgene.2023.1231486.
162. Li, H.; Liu, T.; Yang, Y.; Cho, W.C.; Flynn, R.J.; Harandi, M.F.; Song, H.; Luo, X.; Zheng, Y. Interplays of Liver Fibrosis-Associated microRNAs: Molecular Mechanisms and Implications in Diagnosis and Therapy. *Genes Dis.* **2023**, *10*, 1457–1469, doi:10.1016/j.gendis.2022.08.013.
163. Felekis, K.; Pieri, M.; Papanephytous, C. Exploring the Feasibility of Circulating miRNAs as Diagnostic and Prognostic Biomarkers in Osteoarthritis: Challenges and Opportunities. *Int. J. Mol. Sci.* **2023**, *24*, 13144, doi:10.3390/ijms241713144.
164. Bhatia, A.; Upadhyay, A.K.; Sharma, S. miRNAs Are Now Starring in “No Time to Die: Overcoming the Chemoresistance in Cancer.” *IUBMB Life* **2023**, *75*, 238–256, doi:10.1002/iub.2652.
165. Mehterov, N. Role of MicroRNAs in Cancer Development and Treatment. *Int. J. Mol. Sci.* **2023**, *24*, 11058, doi:10.3390/ijms241311058.
166. Romano, G.; Acunzo, M.; Nana-Sinkam, P. microRNAs as Novel Therapeutics in Cancer. *Cancers* **2021**, *13*, 1526, doi:10.3390/cancers13071526.
167. Sell, M.C.; Ramlogan-Steel, C.A.; Steel, J.C.; Dhungel, B.P. MicroRNAs in Cancer Metastasis: Biological and Therapeutic Implications. *Expert Rev. Mol. Med.* **2023**, *25*, e14, doi:10.1017/erm.2023.7.
168. Arghiani, N.; Shah, K. Modulating microRNAs in Cancer: Next-Generation Therapies. *Cancer Biol. Med.* **2022**, *19*, 289–304, doi:10.20892/j.issn.2095-3941.2021.0294.
169. Chauhan, N.; Jaggi, M.; Chauhan, S.C.; Yallapu, M.M. COVID-19: Fighting the Invisible Enemy with microRNA. *Expert Rev. Anti Infect. Ther.* **2021**, *19*, 137–145, doi:10.1080/14787210.2020.1812385.
170. He, B.; Zhao, Z.; Cai, Q.; Zhang, Y.; Zhang, P.; Shi, S.; Xie, H.; Peng, X.; Yin, W.; Tao, Y.; et al. miRNA-Based Biomarkers, Therapies, and Resistance in Cancer. *Int. J. Biol. Sci.* **2020**, *16*, 2628–2647, doi:10.7150/ijbs.47203.
171. Bayraktar, E.; Bayraktar, R.; Oztatlici, H.; Lopez-Berestein, G.; Amero, P.; Rodriguez-Aguayo, C. Targeting miRNAs and Other Non-Coding RNAs as a Therapeutic Approach: An Update. *Non-Coding RNA* **2023**, *9*, 27, doi:10.3390/ncrna9020027.
172. Milán Rois, P.; Latorre, A.; Rodriguez Diaz, C.; Del Moral, Á.; Somoza, Á. Reprogramming Cells for Synergistic Combination Therapy with Nanotherapeutics against Uveal Melanoma. *Biomimetics* **2018**, *3*, 28, doi:10.3390/biomimetics3040028.
173. Rhim, J.; Baek, W.; Seo, Y.; Kim, J.H. From Molecular Mechanisms to Therapeutics: Understanding MicroRNA-21 in Cancer. *Cells* **2022**, *11*, 2791, doi:10.3390/cells11182791.

174. Gao, S.; Tian, H.; Guo, Y.; Li, Y.; Guo, Z.; Zhu, X.; Chen, X. miRNA Oligonucleotide and Sponge for miRNA-21 Inhibition Mediated by PEI-PLL in Breast Cancer Therapy. *Acta Biomater.* **2015**, *25*, 184–193, doi:10.1016/j.actbio.2015.07.020.
175. Davenport, M.L.; Echols, J.B.; Silva, A.D.; Anderson, J.C.; Owens, P.; Yates, C.; Wei, Q.; Harada, S.; Hurst, D.R.; Edmonds, M.D. miR-31 Displays Subtype Specificity in Lung Cancer. *Cancer Res.* **2021**, *81*, 1942–1953, doi:10.1158/0008-5472.CAN-20-2769.
176. Van Roosbroeck, K.; Fanini, F.; Setoyama, T.; Ivan, C.; Rodriguez-Aguayo, C.; Fuentes-Mattei, E.; Xiao, L.; Vannini, I.; Redis, R.S.; D'Abundo, L.; et al. Combining Anti-Mir-155 with Chemotherapy for the Treatment of Lung Cancers. *Clin. Cancer Res.* **2017**, *23*, 2891–2904, doi:10.1158/1078-0432.CCR-16-1025.
177. Garofalo, M.; Quintavalle, C.; Romano, G.; Croce, C.M.; Condorelli, G. miR221/222 in Cancer: Their Role in Tumor Progression and Response to Therapy. *Curr. Mol. Med.* **2012**, *12*, 27–33.
178. Asafo-Agyei, K.O.; Samant, H. Hepatocellular Carcinoma. In *StatPearls*; StatPearls Publishing: Treasure Island (FL), 2025.
179. Moshiri, F.; Callegari, E.; D'Abundo, L.; Corrà, F.; Lupini, L.; Sabbioni, S.; Negrini, M. Inhibiting the Oncogenic Mir-221 by microRNA Sponge: Toward microRNA-Based Therapeutics for Hepatocellular Carcinoma. *Gastroenterol. Hepatol. Bed Bench* **2014**, *7*, 43–54.
180. Miller, T.E.; Ghoshal, K.; Ramaswamy, B.; Roy, S.; Datta, J.; Shapiro, C.L.; Jacob, S.; Majumder, S. MicroRNA-221/222 Confers Tamoxifen Resistance in Breast Cancer by Targeting p27Kip1. *J. Biol. Chem.* **2008**, *283*, 29897–29903, doi:10.1074/jbc.M804612200.
181. Ouyang, Y.X.; Feng, J.; Wang, Z.; Zhang, G.J.; Chen, M. miR-221/222 Sponge Abrogates Tamoxifen Resistance in ER-Positive Breast Cancer Cells through Restoring the Expression of ER α . *Mol. Biomed.* **2021**, *2*, 20, doi:10.1186/s43556-021-00045-0.
182. Smolarz, B.; Durczyński, A.; Romanowicz, H.; Szyłło, K.; Hogendorf, P. miRNAs in Cancer (Review of Literature). *Int. J. Mol. Sci.* **2022**, *23*, 2805, doi:10.3390/ijms23052805.
183. Liu, Y.; Yang, Y.; Wang, X.; Yin, S.; Liang, B.; Zhang, Y.; Fan, M.; Fu, Z.; Shen, C.; Han, Y.; et al. Function of microRNA-124 in the Pathogenesis of Cancer (Review). *Int. J. Oncol.* **2024**, *64*, 1–21, doi:10.3892/ijo.2023.5594.
184. Lv, Z.; Yang, L. miR-124 Inhibits the Growth of Glioblastoma through the Downregulation of SOS1. *Mol. Med. Rep.* **2013**, *8*, 345–349, doi:10.3892/mmr.2013.1561.
185. Tan, A.C.; Ashley, D.M.; López, G.Y.; Malinzak, M.; Friedman, H.S.; Khasraw, M. Management of Glioblastoma: State of the Art and Future Directions. *CA. Cancer J. Clin.* **2020**, *70*, 299–312, doi:10.3322/caac.21613.
186. Cao, Z.; Qiu, J.; Yang, G.; Liu, Y.; Luo, W.; You, L.; Zheng, L.; Zhang, T. MiR-135a Biogenesis and Regulation in Malignancy: A New Hope for Cancer Research and Therapy. *Cancer Biol. Med.* **2020**, *17*, 569–582, doi:10.20892/j.issn.2095-3941.2020.0033.
187. Sun, X.; Xue, H.; Xiong, Y.; Yu, R.; Gao, X.; Qian, M.; Wang, S.; Wang, H.; Xu, J.; Chen, Z.; et al. GALE Promotes the Proliferation and Migration of Glioblastoma Cells and Is Regulated by miR-Let-7i-5p. *Cancer Manag. Res.* **2019**, *11*, 10539–10554, doi:10.2147/CMAR.S221585.
188. Nair, R.; Westin, J. CAR T-Cells. In *Immunotherapy*; Naing, A., Hajjar, J., Eds.; Springer International Publishing: Cham, 2020; pp. 215–233 ISBN 978-3-030-41008-7.
189. Montaña-Samaniego, M.; Bravo-Estupiñan, D.M.; Méndez-Guerrero, O.; Alarcón-Hernández, E.; Ibáñez-Hernández, M. Strategies for Targeting Gene Therapy in Cancer Cells With Tumor-Specific Promoters. *Front. Oncol.* **2020**, *10*, 605380, doi:10.3389/fonc.2020.605380.
190. Wang, J.-H.; Gessler, D.J.; Zhan, W.; Gallagher, T.L.; Gao, G. Adeno-Associated Virus as a Delivery Vector for Gene Therapy of Human Diseases. *Signal Transduct. Target. Ther.* **2024**, *9*, 1–33, doi:10.1038/s41392-024-01780-w.
191. Eid, F.-E.; Chen, A.T.; Chan, K.Y.; Huang, Q.; Zheng, Q.; Tobey, I.G.; Pacouret, S.; Brauer, P.P.; Keyes, C.; Powell, M.; et al. Systematic Multi-Trait AAV Capsid Engineering for Efficient Gene Delivery. *Nat. Commun.* **2024**, *15*, 6602, doi:10.1038/s41467-024-50555-y.
192. Puzzo, F.; Zhang, C.; Powell Gray, B.; Zhang, F.; Sullenger, B.A.; Kay, M.A. Aptamer-Programmable Adeno-Associated Viral Vectors as a Novel Platform for Cell-Specific Gene Transfer. *Mol. Ther. Nucleic Acids* **2023**, *31*, 383–397, doi:10.1016/j.omtn.2023.01.007.

193. Santiago-Ortiz, J.L.; Schaffer, D.V. Adeno-Associated Virus (AAV) Vectors in Cancer Gene Therapy. *J. Control. Release Off. J. Control. Release Soc.* **2016**, *240*, 287–301, doi:10.1016/j.jconrel.2016.01.001.
194. Bulcha, J.T.; Wang, Y.; Ma, H.; Tai, P.W.L.; Gao, G. Viral Vector Platforms within the Gene Therapy Landscape. *Signal Transduct. Target. Ther.* **2021**, *6*, 1–24, doi:10.1038/s41392-021-00487-6.
195. Vyas, K.; Patel, M.M. Insights on Drug and Gene Delivery Systems in Liver Fibrosis. *Asian J. Pharm. Sci.* **2023**, *18*, 100779, doi:10.1016/j.ajps.2023.100779.
196. Bravo-Estupiñan, D.M.; Montaña-Samaniego, M.; Mora-Rodríguez, R.A.; Ibáñez-Hernández, M. Cationic Lipid Derived from a Basic Amino Acid: Design and Synthesis. *Appl. Sci.* **2024**, *14*, 10892, doi:10.3390/app142310892.
197. Wang, C.; Pan, C.; Yong, H.; Wang, F.; Bo, T.; Zhao, Y.; Ma, B.; He, W.; Li, M. Emerging Non-Viral Vectors for Gene Delivery. *J. Nanobiotechnology* **2023**, *21*, 272, doi:10.1186/s12951-023-02044-5.
198. Wei, P.-S.; Thota, N.; John, G.; Chang, E.; Lee, S.; Wang, Y.; Ma, Z.; Tsai, Y.-H.; Mei, K.-C. Enhancing RNA-Lipid Nanoparticle Delivery: Organ- and Cell-Specificity and Barcoding Strategies. *J. Controlled Release* **2024**, *375*, 366–388, doi:10.1016/j.jconrel.2024.08.030.
199. Li, X.; Qin, Z.; Wang, S.; Zhang, L.; Jiang, X. Microfluidics-Assembled Nanovesicles for Nucleic Acid Delivery. *Acc. Chem. Res.* **2025**, *58*, 570–582, doi:10.1021/acs.accounts.4c00738.
200. Zheng, C.; Baum, B.J. Evaluation of Promoters for Use in Tissue-Specific Gene Delivery. *Methods Mol. Biol. Clifton NJ* **2008**, *434*, 205–219, doi:10.1007/978-1-60327-248-3_13.
201. Meuwissen, R.; Berns, A. Mouse Models for Human Lung Cancer. *Genes Dev.* **2005**, *19*, 643–664, doi:10.1101/gad.1284505.
202. Nakamura, S.; Watanabe, S.; Ohtsuka, M.; Maehara, T.; Ishihara, M.; Yokomine, T.; Sato, M. Cre-loxP System as a Versatile Tool for Conferring Increased Levels of Tissue-Specific Gene Expression from a Weak Promoter. *Mol. Reprod. Dev.* **2008**, *75*, 1085–1093, doi:10.1002/mrd.20847.
203. Shepelev, M.V.; Kopantzev, E.P.; Vinogradova, T.V.; Sverdlov, E.D.; Korobko, I.V. hTERT and BIRC5 Gene Promoters for Cancer Gene Therapy: A Comparative Study. *Oncol. Lett.* **2016**, *12*, 1204–1210, doi:10.3892/ol.2016.4718.
204. Plotnikova, O.; Baranova, A.; Skoblov, M. Comprehensive Analysis of Human microRNA-mRNA Interactome. *Front. Genet.* **2019**, *10*, 933, doi:10.3389/fgene.2019.00933.
205. Menon, A.; Abd-Aziz, N.; Khalid, K.; Poh, C.L.; Naidu, R. miRNA: A Promising Therapeutic Target in Cancer. *Int. J. Mol. Sci.* **2022**, *23*, 11502, doi:10.3390/ijms231911502.
206. Toden, S.; Zumwalt, T.J.; Goel, A. Non-Coding RNAs and Potential Therapeutic Targeting in Cancer. *Biochim. Biophys. Acta Rev. Cancer* **2021**, *1875*, 188491, doi:10.1016/j.bbcan.2020.188491.
207. Samad, A.F.A.; Kamaroddin, M.F. Innovative Approaches in Transforming microRNAs into Therapeutic Tools. *Wiley Interdiscip. Rev. RNA* **2023**, *14*, e1768, doi:10.1002/wrna.1768.
208. Orellana, E.A.; Abdelal, A.M.; Rangasamy, L.; Tenneti, S.; Myoung, S.; Low, P.S.; Kasinski, A.L. Enhancing MicroRNA Activity through Increased Endosomal Release Mediated by Nigericin. *Mol. Ther. Nucleic Acids* **2019**, *16*, 505–518, doi:10.1016/j.omtn.2019.04.003.
209. Fromm, B.; Zhong, X.; Tarbier, M.; Friedländer, M.R.; Hackenberg, M. The Limits of Human microRNA Annotation Have Been Met. *RNA N. Y. N* **2022**, *28*, 781–785, doi:10.1261/rna.079098.122.
210. Zhang, S.; Cheng, Z.; Wang, Y.; Han, T. The Risks of miRNA Therapeutics: In a Drug Target Perspective. *Drug Des. Devel. Ther.* **2021**, *15*, 721–733, doi:10.2147/DDDT.S288859.
211. Chen, Y.; Gao, D.-Y.; Huang, L. In Vivo Delivery of miRNAs for Cancer Therapy: Challenges and Strategies. *Adv. Drug Deliv. Rev.* **2015**, *81*, 128–141, doi:10.1016/j.addr.2014.05.009.
212. Ali Syeda, Z.; Langden, S.S.S.; Munkhzul, C.; Lee, M.; Song, S.J. Regulatory Mechanism of MicroRNA Expression in Cancer. *Int. J. Mol. Sci.* **2020**, *21*, 1723, doi:10.3390/ijms21051723.
213. Bortoletto, A.S.; Parchem, R.J. KRAS Hijacks the miRNA Regulatory Pathway in Cancer. *Cancer Res.* **2023**, *83*, 1563–1572, doi:10.1158/0008-5472.CAN-23-0296.

214. Fu, Z.; Wang, L.; Li, S.; Chen, F.; Au-Yeung, K.K.-W.; Shi, C. MicroRNA as an Important Target for Anticancer Drug Development. *Front. Pharmacol.* **2021**, *12*, 736323, doi:10.3389/fphar.2021.736323.
215. Ricarte-Filho, J.C.; Casado-Medrano, V.; Reichenberger, E.; Spangler, Z.; Scheerer, M.; Isaza, A.; Baran, J.; Patel, T.; MacFarland, S.P.; Brodeur, G.M.; et al. DICER1 RNase IIIb Domain Mutations Trigger Widespread miRNA Dysregulation and MAPK Activation in Pediatric Thyroid Cancer. *Front. Endocrinol.* **2023**, *14*, 1083382, doi:10.3389/fendo.2023.1083382.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.