

1 Article

2 Modeling Power Generation and Energy Efficiencies 3 in Air-Cathode Microbial Fuel Cells Based on Freter 4 Equations

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14 **Abstract:** The model proposed in this study was based on the assumption that the biomass attached
15 to the anode served as biocatalysts for MFC exoelectrogenesis, and this catalytic effect was
16 quantified by the exchange current density of anode. By modifying the Freter model and combining
17 it with the Butler-Volmer equation, this model could adequately describe the processes of electricity
18 generation, substrate utilization, and the suspended and attached biomass concentrations, at both
19 batch and continuous operating modes. MFC performance is affected by the operating variables
20 such as initial substrate concentration, external resistor, influent substrate concentration, and
21 dilution rate, and these variables were revealed to have complex interactions by data simulation.
22 The external power generation and energy efficiency were considered as indices for MFC
23 performance. The simulated results explained that an intermediate initial substrate concentration
24 (about 100 mg/L under this reactor configuration) needed to be chosen to achieve maximum overall
25 energy efficiency from substrate in the batch mode. An external resistor with the value about that
26 of the internal resistance boosted the power generation, and a resistor with several times of that of
27 the internal resistance achieved better overall energy efficiency. At continuous mode, dilution rate
28 significantly impacted the steady-state substrate concentration level (thus substrate removal
29 efficiency and rate), and attached biomass could be fully developed when the influent substrate
30 concentration was equal to or higher than 100 mg/L at any dilution rate of the tested range. Overall,
31 this relatively simple model provided a convenient way for evaluating and optimizing the
32 performance of MFC reactors by regulating operating parameters.

33 **Keywords:** exchange current; energy efficiency; heat generation; attached biomass; suspended
34 biomass; single chamber air-cathode

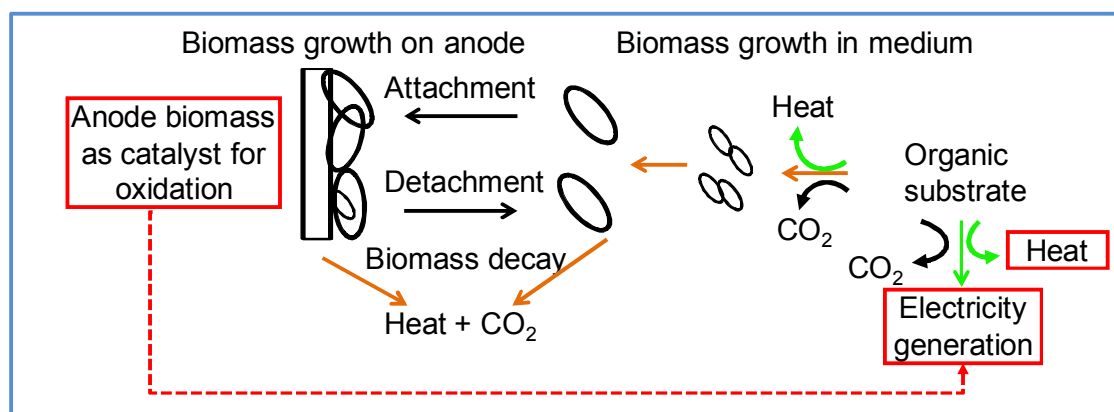
35 **Featured Application:** The established mathematical model for air-cathode microbial fuel cells can
36 be used to predict the electrical current and power generation as well as energy efficiencies at
37 varying operating conditions.
38

39 1. Introduction

40 A microbial fuel cell (MFC) is a device oxidizing organic substrates in the liquid environment
41 and releasing electrical energy to external loads. Anode attached biomass works as catalysts for
42 substrate oxidization [1]. The catalytic microorganisms at anodes, mostly enriched from mixed

43 microbial consortia, are capable of handling various organic substrates [2, 3], and can potentially be
 44 used in degrading the organic substrates in various types of wastewater.

45 Due to complicated interactions between various design and operating variables in this hybrid
 46 type of reactor, it is not easy to experimentally achieve optimal conditions for power and energy
 47 generation and efficient substrate removal. Mathematical models thus provide a simple approach to
 48 investigate the effects of different variables and optimize the MFC performances. The electrochemical
 49 reactions at anode are emphasized in MFC modeling [4, 5] since the anode reactions are features of
 50 MFC and MFC performance can partly be predicted from the growth status of anode attached
 51 bacteria [6]. The current MFC anode models are normally based on redox mediators supposed to
 52 exist in the medium. Research progress already reveals that the outer membrane-bound cytochromes
 53 and the nanowire of bacteria conduct electricity [1] so bacteria as a whole, which are directly attached
 54 to the anode surface, can be regarded as catalysts. It can thus be assumed that only the directly anode-
 55 attached bacteria, or bacteria of a monolayer [7], contribute to electricity generation. Besides, the
 56 suspended bacteria must also be included in the model because the attached and suspended bacterial
 57 populations reach a dynamic equilibrium between each other, and because both populations utilize
 58 substrate for growth. This inclusion is especially essential when studying substrate utilization
 59 between the populations. The Freter model, which originally describes the dynamics of the
 60 suspended bacteria and wall-attached bacteria in a bioreactor [8, 9], is herein adopted. The main
 61 reason favoring this model is that it emphasizes the formation of a bacterial monolayer on a wall
 62 structure which is similar to the process of anode-bacteria attachment. The Freter model is first
 63 revised to account for the feature of MFC where electricity generation consumes substrate (**Figure 1**)
 64 before using it to describe the MFC bacterial populations.
 65



66 **Figure 1.** A schematic of organic substrate and energy flow in a microbial fuel cell.
 67
 68

69 In this study, a dynamic model for MFC was developed based on microbial reaction,
 70 electrochemical reaction, and mass balances. Numerical simulation of the model was conducted to
 71 investigate the effect of varying operating parameters, including initial or influent substrate
 72 concentration, dilution rate, and external resistor, on the MFC microbial populations, substrate
 73 removal, power generation, and energy efficiency. Factors influencing the energy efficiency were
 74 especially emphasized in numerical simulation for both batch and continuous modes of operation.

75 2. Materials and Methods

76 2.1. MFC design and operation

77 Two identical single-chamber air-cathode MFC reactors, with a total volume of 155 mL, were
 78 fabricated from clear/extruded acrylic tubes (5 cm in internal diameter and 7.5 cm in length), covered
 79 with 0.25 cm thick plexiglass acrylic endplates at both ends. The anode was made from plain carbon
 80 cloth, while the air-cathode was made from 0.5 mg cm⁻² Pt-containing (10% in carbon black) carbon

81 cloth treated with Nafion, and with polytetrafluoroethylene (PTFE) as a gas diffusion backing layer
82 [10]. A 2.2 kΩ resistor was connected in the circuit as an external load.

83 Anaerobic sludge was obtained for MFC inoculation from a swine manure lagoon at the
84 University of Minnesota Southern Research and Outreach Center in Waseca, MN. The sludge was
85 sieved through a 1 mm mesh sieve to remove sand, gravels, and plant residues prior to use. Sodium
86 acetate was dissolved in 50 mM phosphate buffer solution (pH 7) to prepare a medium with 40 mM
87 acetate. Other nutrients also added to enhance and maintain the development of bacteria included (g
88 L⁻¹): 0.31 NH₄Cl, 0.13 KCl, and mineral and vitamin solutions.
89

90 2.1.1. Data acquisition

91 The voltage (U_{ext}) across the external resistor was measured by voltage probes and recorded by
92 a CR 1000 data logger (Campbell Scientific, Inc., Logan, UT). The anodic and cathodic potentials were
93 monitored using standard Ag/AgCl electrodes (MF-2072, BASi, Inc., West Lafayette, IN). The data
94 logger supporting software, LoggerNet Version 3.4, was used to monitor and collect the voltage data
95 of the MFC reactors.

96 2.2. Model description

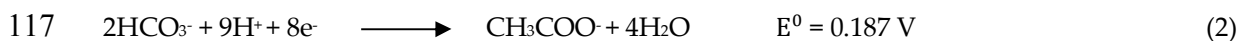
97 The mathematical model of microbial fuel cells to be developed simulated and predicted the
98 electrochemical performance in batch or continuous mode of operation. A few assumptions were
99 made to simplify the modeling of different MFC reactors. First, the anode-attached microbes were
100 assumed to be the catalyst for the exoelectrogenic process; second, the exchange current density, or
101 the catalytic effect, was proportional to the concentration of the attached biomass; third, the mass
102 transfer of acetate to anode and oxygen gas to cathode, assumed to be fast enough compared to the
103 electrochemical processes, was not a limiting factor and thus was neglected; and fourth, the
104 hydrolytic products of dead cells were assumed not providing a carbon source or energy to the
105 suspended or attached bacterial growth.

106 2.2.1. Electrode potentials

107 The electrode potential (against the standard hydrogen electrode, SHE or NHE) was
108 dependent on the standard electrode potential and the concentration of active species according to
109 the Nernst equation:

$$110 E^{o'} = E^0 + \frac{RT}{nF} \sum v_i \ln c_i \quad (1)$$

111 where $E^{o'}$ was the formal potential (pH- and concentration-adjusted reduction potential), E^0 was
112 the standard electrode potential, v_i was the stoichiometric number for species i (positive for
113 oxidized species, and negative for reduced species), and c_i was the concentration of the species i .
114 In the pH region of this study, the Mineql+ [11] simulation indicated that bicarbonate was the major
115 form in the carbonate equilibrium system. Thus, the anodic and cathodic reactions proceeded as
116 follows:



119 The anode and cathode electrode potentials ($E_A^{o'}$ and $E_C^{o'}$, respectively) based on the assumed
120 acetate and bicarbonate concentrations could be estimated as follows:

$$121 E_A^{o'} = E_A^0 + \frac{RT}{nF} \sum v_i \ln c_i = -0.335 \quad [\text{Ac}^-]=0.04 \text{ M}; [\text{HCO}_3^-]=10^{-4.5} \text{ M} \quad (4)$$

$$122 \quad E_C^{0'} = E_C^0 + \frac{RT}{nF} \sum v_j \ln c_j = 0.805 \quad P_{O_2}=0.2 \quad (5)$$

123 The experimentally measured cathodic potentials usually were much lower than the predicted
 124 values [12, 13], partly due to other reactions that might have happened on the cathode surface
 125 which resulted in a mixed cathodic potential of lower values. For example, hydrogen peroxide
 126 production could happen on cathode to deviate the cathode potential to a lower value (e.g., 0.36 V)
 127 [12]. In accordance with both the literature information and the cathode potential at open-circuit
 128 operation in this study, a cathode potential of 0.51 V (NHE), rather than 0.805 V (NHE), was
 129 adopted herein. The electromotive force (E_{emf}) was defined as the difference between the cathode
 130 and anode potentials:

$$131 \quad E_{emf} = E_C^{0'} - E_A^{0'} \quad (6)$$

133 2.2.2. Exchange current and overpotential

134 Overpotential is defined as the difference of an electrode potential from its formal potential in
 135 a fuel cell. The Butler-Volmer equation, neglecting the reverse cathodic reaction [14], was used to
 136 express the relationship between the exchange current and anode overpotential:

$$137 \quad I = j_{oA} A e^{\frac{\alpha_A F \eta_{act,A}}{RT}} \quad (7)$$

138 where I was the electric current, j_{oA} was the exchange current density of anode, α_A was the
 139 transfer coefficient of the anodic reaction, and $\eta_{act,A}$ was the anode overpotential. In a similar way,
 140 the cathode overpotential could be formulated with the reverse reaction neglected as follows:

$$141 \quad I = j_{oC} A e^{\frac{(\alpha_C - 1) F \eta_{act,C}}{RT}} \quad (8)$$

142 where j_{oC} was the exchange current density of cathode, α_C was the charge transfer coefficient of
 143 the cathode reaction, and $\eta_{act,C}$ was the cathode overpotential. The exchange current density of
 144 cathode was consistent during the MFC operation due to the unchanging amount of the catalyst (0.5
 145 mg cm⁻² of Pt), but that of anode was proportional to the increasing catalyst load, which was the
 146 attached biomass concentration here [15]:

$$147 \quad j_{oA} = j_{oAU} w \quad (9)$$

148 where j_{oAU} was the unit exchange current density of exoelectrogenic biomass, and w was the
 149 attached biomass concentration on anode.

151 2.2.3. Mass balances

152 A model that was proposed to describe the suspended and attached microbial formation [8, 9]
 153 in an aqueous environment was modified and adopted in this study. In this model, bacteria could
 154 divide, suspend and decay in the reactor medium, and could also attach to a surface with a
 155 dynamic equilibrium between attachment and detachment. Since bacterial growth and electricity
 156 generation consumed substrate when it was available, the three components of the system, the
 157 substrate concentration (S), the suspended biomass concentration (u), and the attached biomass
 158 concentration (w), which were dictated by mass balances, were expressed in three ordinary
 159 differential equations as follows:

$$160 \quad \frac{dS}{dt} = D(S_{in} - S) - \gamma^{-1}(u\mu_u + \delta w\mu_w) - r_e \quad (10)$$

$$161 \quad \frac{du}{dt} = u(\mu_u - D - k_u) + \beta\delta w + \delta w\mu_w(1 - G) - \alpha u(1 - W) \quad (11)$$

$$162 \quad \frac{dw}{dt} = w(\mu_w G - \beta - k_w) + \delta^{-1}\alpha u(1 - W) \quad (12)$$

163 where D was the dilution rate, S_{in} was the influent substrate concentration, γ was the biomass yield
 164 of substrate, μ_u was the suspended bacterial growth rate, μ_w was the attached bacterial growth
 165 rate, δ was the ratio of projected anode surface area to reactor volume, r_e was the substrate
 166 utilization rate by electricity and heat generation, k_u was the suspended bacterial death rate, k_w
 167 was the attached bacterial death rate, β was the detachment rate of bacteria from wall, α was the
 168 attachment rate of bacteria to anode surface, G was the fraction of daughter cells attached to
 169 anode, and W was the wall occupation fraction. Some of those parameters in the equations could
 170 be defined through some more fundamental design and operating parameters:

$$171 \quad D = Q/V \quad (13)$$

$$172 \quad \delta = A/V \quad (14)$$

$$173 \quad \mu_u = \frac{m_u S}{a_u + S} \quad (15)$$

$$174 \quad \mu_w = \frac{m_w S}{a_w + S} \quad (16)$$

$$175 \quad G = \frac{(1-W)}{(1.1-W)} \quad (17)$$

176 where Q was the dilution rate, V was the MFC reactor volume, A was the projected anode
 177 surface area, m_u was the maximum specific growth rate of suspended bacteria, a_u was the half
 178 saturation coefficient of suspended bacteria of the Monod kinetics, m_w was the maximum specific
 179 growth rate of attached bacteria, a_w was the half saturation coefficient of attached bacteria.

180 2.2.4. Ohm's law and Kirchhoff's voltage law

181 By applying Ohm's law, the electrical current of the circuit, the voltage drop (U_m) across the
 182 resistance (R_m) of electrolyte solution of MFC medium, and the voltage drop (U_c) across the contact
 183 resistance (R_c) of MFC could be obtained:

$$184 \quad I = U_{ext}/R_{ext} \quad (18)$$

$$185 \quad U_m = \frac{Id}{A\sigma} \quad (19)$$

$$186 \quad U_c = IR_c \quad (20)$$

187 where U_{ext} was the external voltage across the external resistor (R_{ext}), d was the distance between
 188 two electrodes, and σ was the conductivity of MFC medium. By combining all the polarization
 189 losses along the circuit of MFC, the voltage conservation according to Kirchhoff's voltage law could
 190 be denoted as follows:

$$191 \quad E_{emf} = E_C^{0'} - E_A^{0'} = U_{ext} + U_c + U_m + \eta_{act,A} - \eta_{act,C} \quad (21)$$

192 2.2.5. Power and Energy efficiencies

198 The power (P_{ext}) extracted by the external load of a resistor was the product of the external
199 voltage and current:

$$200$$

$$201 P_{\text{ext}} = U_{\text{ext}}I \quad (22)$$

202

203 The power extracted by the electrical circuit of MFC (P_{cell}), or termed as an MFC cell, was the
204 product of the electromotive force and current:

$$205 P_{\text{cell}} = E_{\text{emf}}I \quad (23)$$

206

207 The power lost through the polarization of the activation overpotentials of anode and cathode,
208 the contact resistance, and the medium resistance, was the power emitted in a form of heat:

$$209$$

$$210 P_{\text{heat}} = (U_c + U_m + \eta_{\text{act},A} - \eta_{\text{act},C})I \quad (24)$$

211

212 The rate of the substrate (acetate) loss through MFC cell (r_e) could be calculated as follows:

$$213$$

$$214 r_e = MP_{\text{cell}}/(\Delta H_c) \quad (25)$$

215

216 where ΔH_c was the heat of combustion of acetic acid, and M was the relative molecular weight of
217 acetic acid.

218 Besides the energy loss by heat generation in MFC, the chemical energy of substrate was also
219 lost by providing energy for the growth of the suspended and attached bacteria. The corresponding
220 powers for the suspended (P_u) and attached (P_w) bacteria could be calculated as follows:

$$221$$

$$222 P_u = \gamma^{-1}\Delta H_c u \mu_u V/M \quad (26)$$

$$223$$

$$224 P_w = \gamma^{-1}\Delta H_c \delta w \mu_w V/M \quad (27)$$

225

226 Although some studies [13] suggested that the energy efficiency was further reduced by the
227 more positive potential (close to 0 V, NHE) of redox intermediates than that of the substrate, this
228 energy loss was not considered because our experiment achieved anode potentials close to that of
229 the substrate. Therefore, the MFC cell energy efficiency (η_{cell}) and overall energy efficiency
230 (η_{overall}) were defined as follows:

$$231$$

$$232 \eta_{\text{cell}} = P_{\text{ext}}/P_{\text{cell}} \quad (28)$$

$$233$$

$$234 \eta_{\text{overall}} = P_{\text{ext}}/(P_{\text{cell}} + P_u + P_w) \quad (29)$$

235

236 Please note that in batch mode, the overall energy efficiency was calculated differently:

$$237$$

$$238 \eta_{\text{overall}} = \int_0^t P_{\text{ext}} dt / (\Delta H_c \Delta SV/M) \quad (30)$$

239

240 2.2.6. Model parameters

241 Some constants and parameters involved in microbial fuel cell processes described in
 242 Equations (from 1 to 30) were listed in Table 1. During modeling and simulation, units of those
 243 parameters and variables were not necessarily SI units, but some were adjusted to the suitable
 244 magnitudes. Microbial parameters of acetate utilizers in anaerobic condition were adopted from
 245 literature [16]. Some model parameters associated with electrochemical processes, including charge
 246 transfer coefficients (α_A and α_C) and exchange current densities (j_{oA} , j_{oAU} and j_{oC}) were estimated
 247 from the polarization dataset obtained from one of two MFC reactors in this experiment. The quasi-
 248 Newton approach was used to minimize the χ^2 test statistic as an error function:
 249

$$250 \chi^2 = \sum_{i=1}^n \frac{(y_i^{\text{cal}} - y_i^{\text{exp}})^2}{y_i^{\text{exp}}} \quad (31)$$

251
 252 where n was the number of data points, and y_i^{exp} was the i -th experimental value, and y_i^{cal} was
 253 the i -th calculated value. The polarization curve data of the other reactor was used for validation.
 254

255 Table 1. Constants, and design and operating parameters of a microbial fuel cell system

Parameter symbol	Description	Value	Units
Physical			
F	Faraday's constant	96485	C/mol
R	Ideal gas constant	8.314	J/mol/K
T	Room temperature	298	K
ΔH_c	Heat of combustion of acetic acid	-875000	J/mol
M	Relative molecular weight of acetic acid	60	g/mol
Electrochemical			
E_{A^0}	Formal reduction potential of anode	-0.335	V vs. NHE
E_{C^0}	Formal reduction potential of cathode	0.51	V vs. NHE
Reactor configuration			
A	Projected surface area of electrode	0.002	m ²
d	Distance between anode and cathode	0.075	m
V	Volume of MFC reactor	1.55×10 ⁻⁴	m ³
δ	Ratio of anode surface area to MFC volume	13	m ² /m ³
R_c	Contact resistance of MFC	20	Ω
Operating			
D	Dilution rate	0-2	d ⁻¹
S_{in}	Substrate concentration	10-2400	g/m ³
Q	Flow rate	0-7.8×10 ⁻⁵	m ³ /d
σ	Conductivity of MFC medium	1.1	S/m
R_{ext}	External resistor	2200	Ω
$[HCO_3^-]$	Bicarbonate concentration	1×10 ^{-4.5}	kmol/m ³
P_{O_2}	Partial oxygen pressure at cathode	0.2	atm
pH	-log ₁₀ of proton concentration	7	Dimensionless
Microbial			

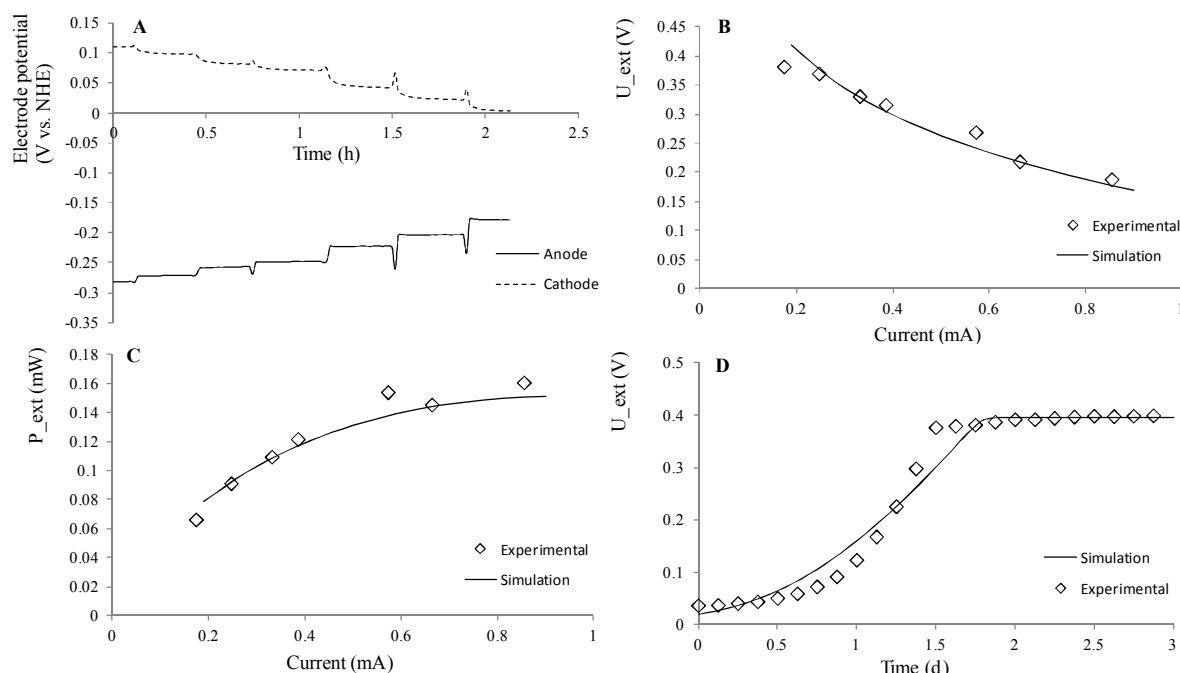
γ	Biomass yield from substrate	0.05	g/g
k_u	Death rate of suspended bacteria	0.02	d ⁻¹
k_w	Death rate of attached bacteria	0.02	d ⁻¹
α	Bacterial attachment rate	0.05	d ⁻¹
β	Bacterial detachment rate	0.05	d ⁻¹
m_u	Maximum specific growth rate of suspended bacteria	2.4	d ⁻¹
m_w	Maximum specific growth rate of attached bacteria	2.4	d ⁻¹
a_u	Half saturation coefficient of suspended bacteria	100	g/m ³
a_w	Half saturation coefficient of attached bacteria	100	g/m ³
w_{max}	Maximum bacterial attachment	0.33 ^a	g/m ²

256 ^a The maximum amount of bacteria attached on electrode was calculated from the bacteria
 257 dimensions observed in SEM images, and this value was close to the literature values of 0.47 to 0.52
 258 g/m² [7].

259 3. Results and Discussion

260 3.1. Parameter estimation

261 Two identically-made MFC reactors with respect to reactor configuration and electrode
 262 materials were inoculated with the same anaerobic sludge and operated in parallel. After MFC were
 263 fully inoculated and operated for 30 d, polarization curves of MFC was obtained by varying the
 264 external resistor from 2200 Ω to 220 Ω , with the electrode potentials recorded (Figure 2A). The
 265 resultant electrochemical parameters estimated by fitting polarization curves of electrodes were
 266 listed in Table 2. The simulation results based on the estimated parameters were in good agreement
 267 with the experimental data of both reactors, as shown in Figure 2B and C. The anodic exchange
 268 current density of 62.5 mA m⁻² was in the same order of a literature value of 29 mA m⁻² [17]. The
 269 cathodic exchange current density (0.975 mA m⁻²) observed in this study was higher than those of
 270 non-catalyzed and bio-catalyzed cathodes [18, 19] but lower than another Pt-based cathode [20]. To
 271 further validate the model, the external voltage (U_{ext}) during inoculation stage was simulated based
 272 on the listed parameters except the unit exchange current, which was fitted from the experimental
 273 data at the inoculation stage (0.129 mA mg⁻² biomass) and had a difference by 32% from 0.189 mA
 274 mg⁻¹ which was fitted from polarization curves. This difference could be a result of the change of
 275 the activity or catalytic effect of the attached biomass during the inoculation and 30 d operation.
 276 The simulated data and the experimental data of the inoculation stage generally showed good
 277 agreement to each other (Figure 2D).



278

279 Figure 2. Electrochemical parameters estimation and validation; (A) the electrode potential during
 280 polarization experiment; (B and C) model validation for experimental data from another
 281 identical MFC reactor (B for U_{ext} , and C for P_{ext}); and (D) comparison of the simulated
 282 and experimental U_{ext} during inoculation stage.

283

284

Table 2. Estimated electrochemical parameters for MFC in this study

Estimated parameters	Description	Value	Units
α_A	Charge transfer coefficient, anode	0.318	Dimensionless
α_C	Charge transfer coefficient, cathode	0.694	Dimensionless
j_{oA}	Exchange current density, anode	62.5	mA/m^2
j_{oAU}	Unit exchange current, anode	0.189	mA/mg
j_{oC}	Exchange current density, cathode	0.975	mA/m^2

285

286 3.2. Simulation of batch mode MFC

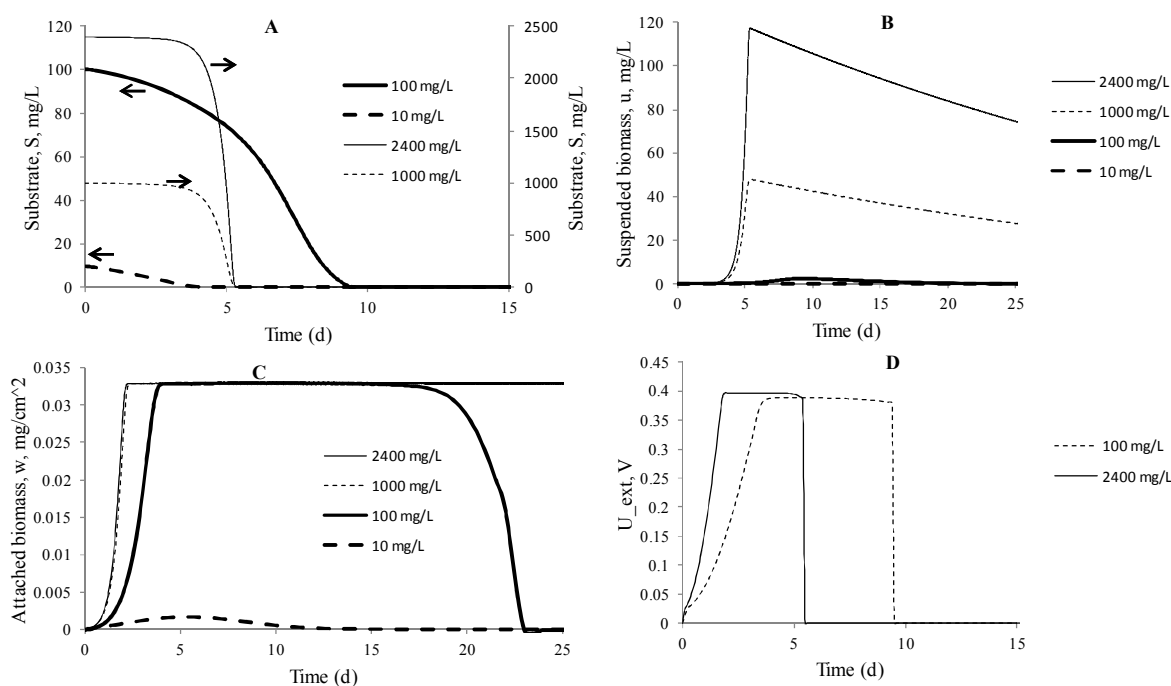
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288 3.2.1. Effect of initial substrate concentration

289 Simulation of the proposed model was performed through Matlab on a personal computer to
 290 illustrate the processes of substrate utilization, biomass concentration evolution, and voltage
 291 generation across an external resistor. Since most MFC studies in literature were carried out in
 292 batch mode, this simulation assumed the medium dilution rate $D=0$ with initial substrate (HAC)
 293 concentrations of 10, 100, 1000, and 2400 mg L^{-1} , consecutively.

294 The simulation results were shown in Figure 3. Initial substrate concentrations of 1000 and
 295 2400 mg L^{-1} did not show obvious decrease in the first 4 d (Figure 3A), but substrate was consumed
 296 dramatically after this period. MFCs started with the lower substrate concentrations behaved
 297 differently from the higher strength cases, as a continuous and gradual decrease was observed. The
 298 substrate in the case of 1000 mg L^{-1} lasted the longest, i.e., 9 d, which almost doubled that in the

299 higher strength cases. These results indicated that it was not always the case that more concentrated
 300 substrate could sustain the longest reaction period, and a moderate substrate concentration should
 301 be chosen for keeping MFC from starvation. Trends of substrate utilization were generally the work
 302 of suspended bacteria, which could accumulate to a high level and subsequently consumed
 303 substrate at a high rate that rapidly reduced substrate concentration (Figure 3B). For example, in
 304 the initial substrate concentrations of 100 and 2400 mg L⁻¹, the peak suspended biomass
 305 concentrations were 2.5 and 117 mg L⁻¹, respectively. The latter was 47 times higher than the former,
 306 which consumed substrate at a higher rate due to two reasons: the higher level of biomass
 307 concentration and the enhanced specific substrate utilization rate indicated by the Monod equation
 308 (Equations 15 & 16).

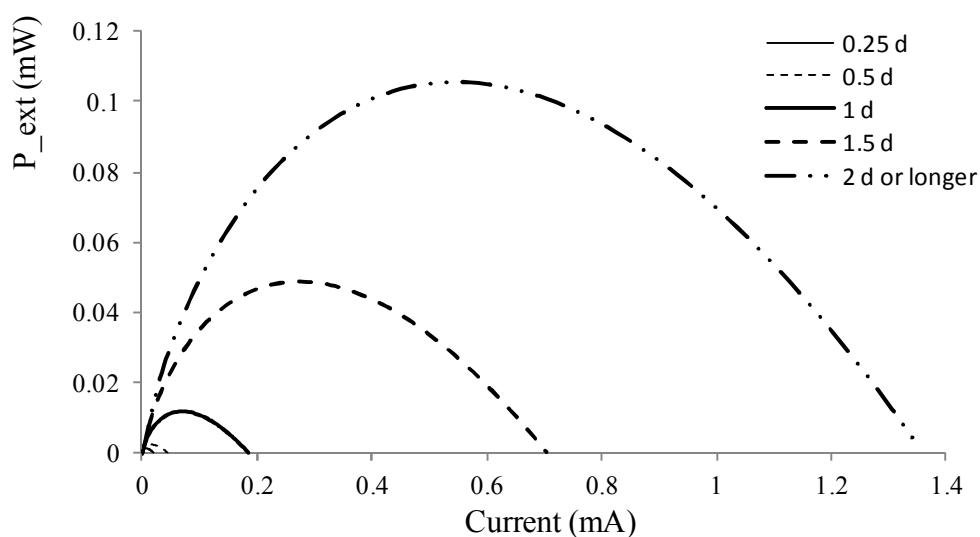


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 310 **Figure 3.** Simulated time-course profiles of process variables at different initial substrate
 311 concentration. (A) the substrate concentration; (B) suspended biomass concentration; (C) attached
 312 biomass concentration, and (D) the external voltage U_{ext} .

313
 314 The attached biomass concentrations (Figure 3C) accumulated to their plateau values (0.033
 315 mg cm⁻²) for all initial substrate concentrations (in 2 d for 1000 and 2400 mg L⁻¹, and in 4 d for 100
 316 mg L⁻¹), except for the one with 10 mg L⁻¹ substrate, indicating that there might exist a lower limit
 317 for substrate concentration. Thus, a substrate concentration lower than 10 mg L⁻¹ should be avoided
 318 during inoculation stage. The higher the initial substrate concentration, the longer the attached
 319 biomass would sustain: both lasted more than 25 d for 1000 and 2400 mg L⁻¹, but only 23 d for 100
 320 mg L⁻¹ before the biomass disappeared. The electricity generation (Figure 3D) was simulated in a
 321 form of external voltage (U_{ext}) for the initial substrate concentrations of 100 and 2400 mg L⁻¹, because
 322 both of them fully developed the attached biomass, and were also the typical levels for the low and
 323 high substrate concentrations, respectively. During the first few days of anode inoculation, the
 324 growing voltage was caused by the growing attached biomass and eventually reached a plateau
 325 value of about 0.39 V. Since the substrate lasted a longer period in the 100 mg L⁻¹ MFC than in the

326 2400 mg L⁻¹ MFC, the electricity generation also lasted longer by 4 more days. Energy generated
 327 was 25.1 and 40 J for 2400 mg L⁻¹ MFC and for 100 mg L⁻¹ MFC, with overall energy efficiencies
 328 extracted from acetate ($\eta_{overall}$) of 0.46% and 17.7%, respectively. Therefore, based on electricity
 329 energy generation and overall energy efficiency, the MFC with 100 mg L⁻¹ of substrate substantially
 330 outperformed the one with 2400 mg L⁻¹. Controlling the initial substrate concentration was thus
 331 demonstrated as a way of regulating the suspended biomass concentration, and was helpful to
 332 harvest more energy out of the substrate for electricity.

333 The growing amount of the attached biomass not only increased the external voltage (U_{ext}), but
 334 also improved the external power, evidenced by the gradual expansion of the polarization curves
 335 against the time in Figure 4. These simulations were based on the high initial substrate
 336 concentration of 2400 mg L⁻¹ in order to avoid a noticeable substrate decrease in 2 d. The pattern of
 337 the curve expansion was similar to what was observed in literature [4, 6], illustrating the
 338 importance of the attached biomass on power generation.



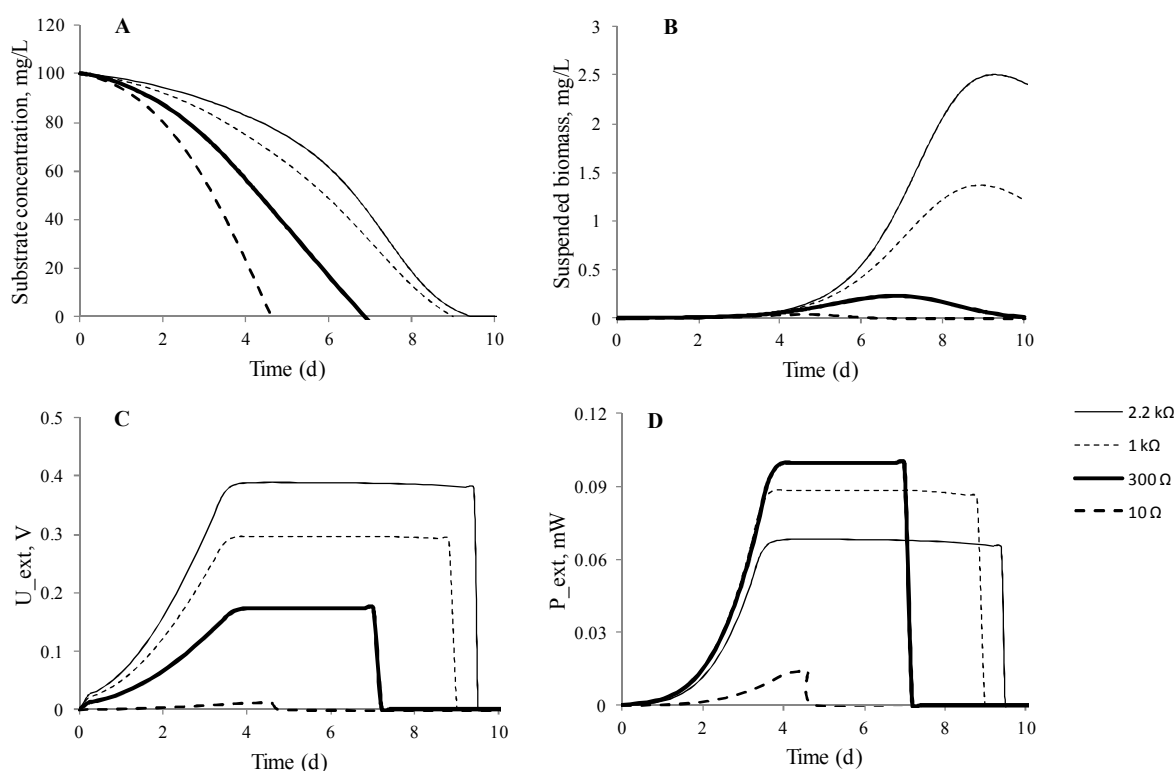
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 340 **Figure 4.** Simulated polarization curves at different moments of inoculation. Attached bacteria
 341 concentration reached a plateau value after about 2 d, and the polarization curves did not change
 342 much thereafter.

343 344 3.2.2. Effect of external resistor

345 From the comparisons of external voltage and overall energy efficiency, it could be seen that
 346 under this experimental condition, a 100 mg L⁻¹ of initial substrate worked best among the four
 347 tested substrate concentrations. The next step of this study simulated the impact of the external
 348 resistor on the batch mode MFC voltage and power generation as well as overall energy efficiency.
 349 In MFC study, the internal resistance (R_{in}) is used for indicating the overall resistance of a reactor,
 350 which is a useful index because varying an external resistor to the value of internal resistance
 351 generates the maximum external power density [21]. Depending on the reactor design and
 352 operation, the values of R_{in} might differ but usually was within a range of several hundreds of
 353 Ohms, which was also in accordance with the observations (332 Ω in the experimental study; and
 354 471 Ω in the simulation study) in MFC reactors in this study. In order to see the effect of external

355 resistors on MFC energy and power generation, a range of resistor covering the values of internal
 356 resistance was chosen for simulation, i.e., 10, 300, 1000, and 2200 Ω .

357 The simulation results were presented in Figure 5. The smaller the value of the external
 358 resistor, the more rapidly the substrate was consumed (Figure 5A), which was also observed in
 359 two-chamber MFC reactors for organic matter removal [22, 23]. The main reason was that the heat
 360 generation, which depicted the part of energy loss through overpotentials (Equation 24), was
 361 substantially increased with the smaller external resistor, which corresponded to a greater electrical
 362 current [24]. When the resistor reduced to 10 Ω , a major part of substrate was lost via heat so that
 363 there was not enough substrate to support suspended (Figure 5B) and attached microbial growth.
 364 On the other hand, the small resistor was beneficial for organic substrate removal because of its
 365 substrate utilization rate, and this value would be appropriate for treating high strength wastewater
 366 such as swine wastewater [25]. A greater external resistor harvested a higher external voltage
 367 (Figure 5C), and took a longer reaction period. The 10- Ω resistor caused an apparently impaired
 368 performance due to the failure of the microbial growth. Simulation of external power generation
 369 (Figure 5D) confirmed that 300 Ω , which was the closest among the three to the internal resistance
 370 observed in this study, generated the largest power. A similar trend was also observed in a
 371 previous experimental study [23]. However, this value of resistor sustained a shorter reaction time
 372 by 2 to 3 d compared to that of 1000 and 2200 Ω . Overall, the 1000 Ω resistor harvested the most
 373 energy from substrate, achieving overall energy efficiency of 20.7% (Table 3). Energy efficiency
 374 between 15% and 20% achieved in experimental studies was considered the best cases [26, 27],
 375 while a value of around 5% was more frequently observed for general cases [27, 28].



376

377 **Figure 5.** Simulated batch performance of MFC with 100 mg L⁻¹ initial substrate at different
 378 external resistors. (A) substrate concentration; (B) suspended biomass concentration; (C) external
 379 voltage; and (D) power generation.

380

381 Table 3. Simulated peak power and energy recovery from an external resistor, and overall energy
 382 efficiency ($\eta_{overall}$) at different external resistors

External resistor	Peak power	Energy recovered	$\eta_{overall}$
10 Ω	0.0142	1.96 J	0.87%
300 Ω	0.0998 mW	37.1 J	16.4%
1000 Ω	0.0882 mW	47.3 J	20.7%
2200 Ω	0.0685 mW	40.0 J	17.7%

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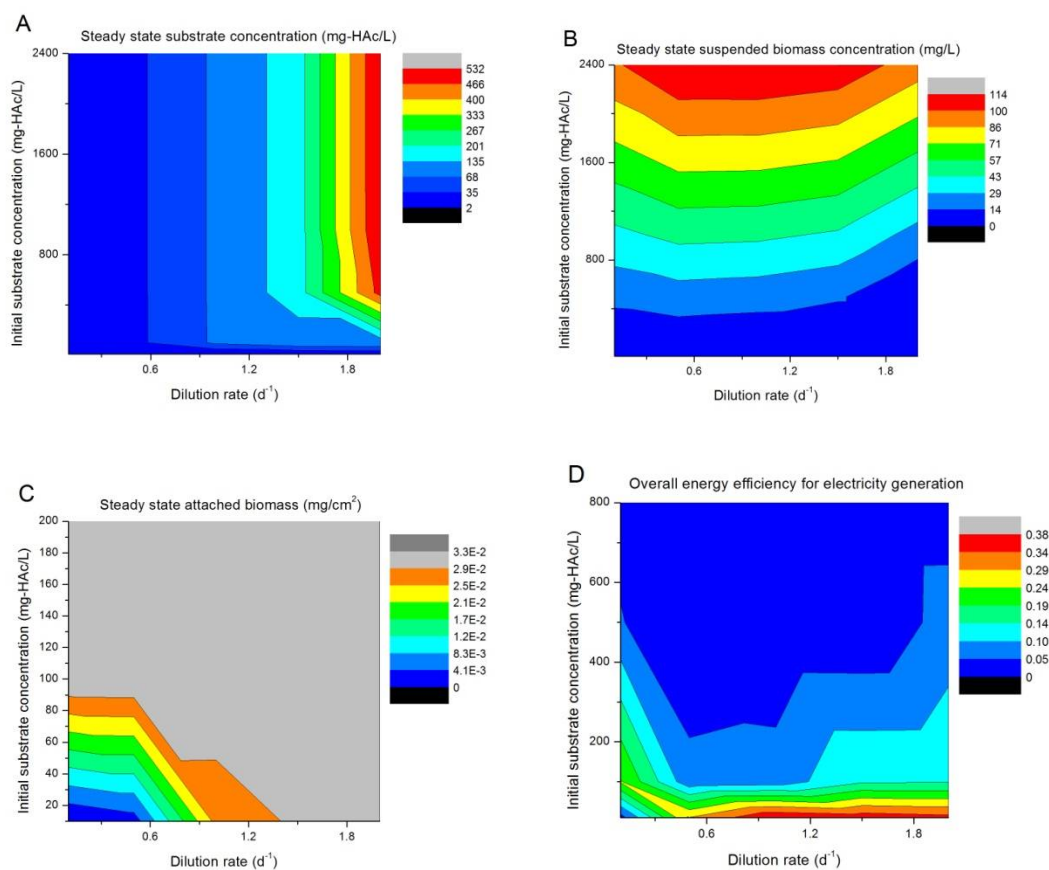
385 3.3. Simulation of continuous mode MFC

386 3.3.1. Effect of influent substrate concentration and dilution rate

387 When MFC reactors were operated in continuous mode, the operating variables that might
 388 have pronounced effects were dilution rate (D) and influent substrate concentration (S_{in}). These two
 389 variables controlled the substrate inflow rate and biomass washout rate, and therefore would
 390 eventually impact MFC power generation and energy efficiency at steady state. So in the
 391 simulation, the dilution rate was varied between 0.1 to 2 d⁻¹. The upper limit was chosen to be less
 392 than the maximum microbial specific growth rate (2.4 d⁻¹) to prevent biomass washout. The influent
 393 substrate (HAc) concentration was between 10 to 2400 mg L⁻¹. The results, including the substrate
 394 concentration, suspended biomass concentration, attached biomass concentration and overall
 395 energy efficiency, were plotted in Figure 6. Please note that the external power (P_{ext}) was not
 396 considered in the simulation of varying S_0 and D , because it was found that when assuming the
 397 attached biomass was fully grown at steady state, the external power was rather a function of the
 398 external resistor than a function of these two operating variables. It should be noted that little effort
 399 in MFC studies of varying these operating parameters was found in continuous mode, so literature
 400 data for comparison were not available.

401 At a given level of the influent substrate concentration, the steady state substrate concentration
 402 went up with the increasing dilution rate, basically due to the increased substrate inflow to the
 403 reactor (Figure 6A). When the influent substrate concentration was higher than 500 mg L⁻¹, the
 404 concentration did not have much impact on the steady state substrate concentration at a given
 405 dilution rate. But when the influent concentration was lower than 500 mg L⁻¹ and the dilution rate
 406 was greater than 1.5 d⁻¹, the steady state concentration increased along with the influent
 407 concentration. The suspended biomass concentration obviously increased with the influent
 408 substrate concentration in a linear manner at any dilution rate (Figure 6B). The dilution rate
 409 between 0.5 and 1 d⁻¹ resulted in a maximum suspended biomass at a given influent substrate
 410 concentration, because a lower rate caused substrate limitation and a higher rate could partially
 411 wash out suspended biomass. The attached biomass was more stable and could generally achieve
 412 its maximum in most cases (Figure 6C), except for the cases of S_{in} , which were much lower than
 413 100 mg L⁻¹ and were too low to provide enough carbon sources for supporting the bacterial growth
 414 and attachment. As to the overall energy efficiency, it could be concluded that a higher influent
 415 substrate concentration reduced the efficiency (Figure 6D). The only exception was the case with
 416 about 10 mg L⁻¹ of influent substrate and 0.1 d⁻¹ of dilution rate, under which condition the attached
 417 biofilm was not fully mature due to substrate deficiency so the MFC did not generate electricity.

418 Considering the tradeoff between higher electrical power generation (reflected by the attached
 419 biomass concentration, which was the catalyst) and acceptable overall energy efficiency, it was
 420 recommended to operate MFC with about 100 mg L^{-1} of influent substrate concentration.
 421 Meanwhile, at this substrate concentration, the choice of a lower dilution rate, e.g., 0.1 d^{-1} , would
 422 help achieve lower effluent substrate and better removal efficiency, and the choice of a larger
 423 dilution rate, e.g., 2 d^{-1} , might otherwise help generate more electricity energy.



424

425

426 **Figure 6.** Process variables and MFC performance predicted at steady state under different dilution
 427 rates and influent substrate concentrations. (A) steady state substrate concentration; (B) suspended
 428 biomass concentration; (C) attached biomass concentration, and (D) the overall energy efficiency
 429 η_{ext} .

430

431 3.3.2. Effect of external resistor

432 The next simulation was implemented for changing electrical currents by varying R_{ext} at the
 433 operating conditions of S_{in} and D listed in Table 4. The relationship between R_{ext} and current I was
 434 depicted in the inset of Figure 7A. These results were useful for determining a suitable range of the
 435 external resistor for better substrate removal, power generation or overall energy recovery. As
 436 shown in Figure 7A and Table 4, all cases resulted in a maximum power generation ($P_{ext, max}$) of
 437 0.106 mW at R_{ext} of 352Ω , except for the case of the lowest substrate loading ($S_{in} = 100 \text{ mg L}^{-1}$ and D
 438 $= 0.1$), which had a highest power of 0.091 mW at R_{ext} of 1007Ω . For the lowest substrate loading, a
 439 further decrease of R_{ext} distributed more organic substrate utilization for heat generation and would
 440 finally lead to a system failure without electrical generation due to substrate limitation for biomass
 441 growth. On the contrary, all the MFCs with enough substrate generated the same external powers

442 at given R_{ext} . Figure 7B plotted the results of MFC cell efficiency (η_{cell} , defined in Equation 28),
 443 which showed that MFC cell efficiency decreased with the increasing circuit current (or the
 444 decreasing external resistor) because of the increased heat loss through the cell (Equation 24).
 445 Again, the curve of the lowest substrate loading differed from the others and was interrupted at R_{ext}
 446 less than 928 Ω due to insufficient substrate available for biomass growth.

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Table 4. Simulated MFC performance: substrate removal, power generation and overall energy efficiency

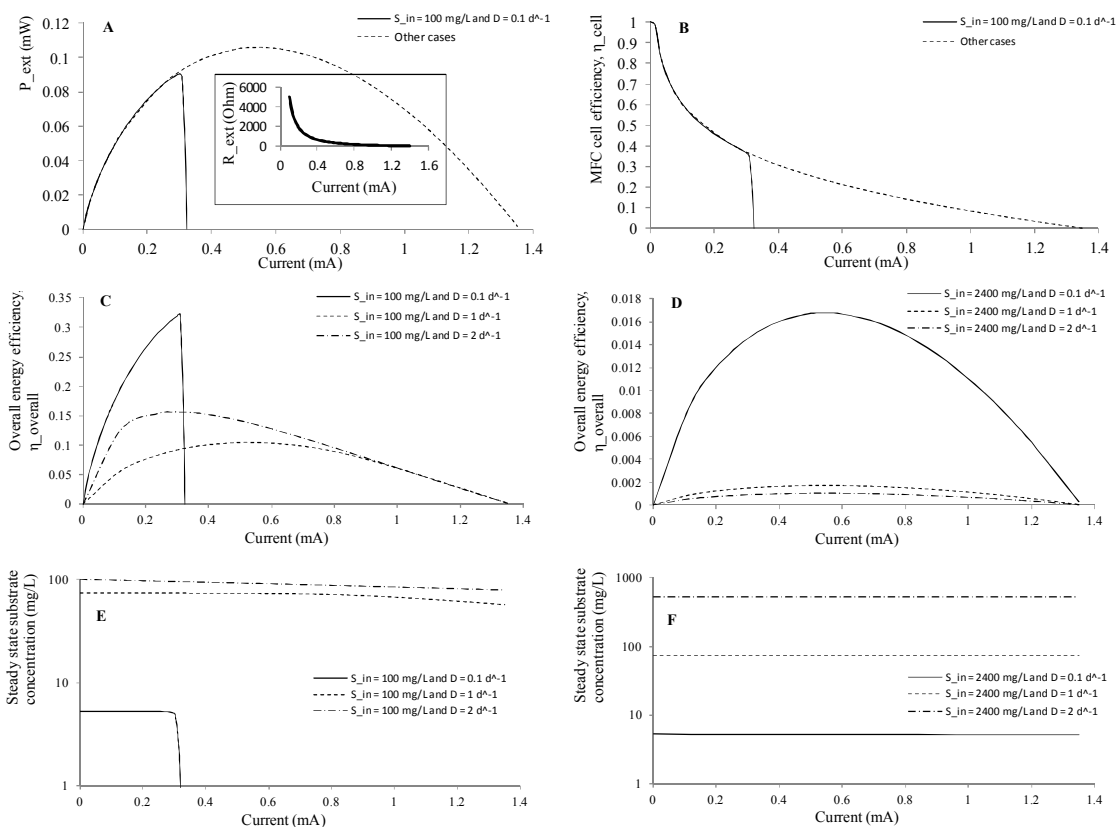
S_{in} mg/L	D d^{-1}	Steady state S mg/L	S removal %	$P_{ext, max}$ mW	$\eta_{overall, max}$	R_{in}^a Ω	$R_{ext, P}^b$ Ω	$R_{ext, \eta}^c$ Ω
100	0.1	0 - 5.26	> 94.7	0.091	0.322	471	1007	928
	1	57.6 - 73.5	26.5 - 42.4	0.106	0.105		352	405
2400	2	79 - 100	< 21.0	0.106	0.156	352	352	1017
	0.1	5.17 - 5.26	100	0.106	0.0167	352	352	352
2400	1	73.9	96.9	0.106	0.0017	352	352	352
	2	531	77.9	0.106	0.0011	352	352	352

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^a The internal resistance was estimated from the transiently simulated polarization curves in the electrical current range between 0.18mA to 0.82 mA.

^b This was the value of the external resistor where the $P_{ext, max}$ was achieved.

^c This was the value of the external resistor where the $\eta_{overall, max}$ was achieved.



456

457 **Figure 7.** Simulated MFC performance at steady state under different electrical currents (or external
 458 resistors), dilution rates and influent substrate concentrations. (A) steady state power density; (B)
 459 MFC cell energy efficiency; (C) overall energy efficiency at $S_{in}=100$ mg L⁻¹; and (D) $S_{in}=2400$ mg L⁻¹;
 460 (E) effluent substrate concentration at $S_{in}=100$ mg L⁻¹; and (F) $S_{in}=2400$ mg L⁻¹.

461

462 Figure 7C and D showed the overall energy efficiencies for the influent substrate
 463 concentrations of 100 and 2400 mg L⁻¹, respectively. For the lower substrate cases, the maximum
 464 overall energy efficiencies ($\eta_{overall, max}$) were between 0.105 and 0.322, and were achieved at different
 465 external resistors depending on dilution rates (928 Ω at $D=0.1$ d⁻¹; 405 Ω at $D=1$ d⁻¹; and 1017 Ω at
 466 $D=2$ d⁻¹). For the higher substrate cases, the maximum overall energy efficiencies were between
 467 0.0011 and 0.0167, and were achieved at $R_{ext} = 352$ Ω at which the $P_{ext, max}$ values were concurrently
 468 obtained. So the lower substrate concentration reactor had one or two orders of magnitude of
 469 higher overall efficiency than that of the higher substrate concentration. This substantial difference
 470 was a result of more proliferated biomass growth at higher substrate concentrations and the
 471 biomass in turn consumed more substrate. Please also note that the internal resistance R_{in} estimated
 472 by polarization curves was 471 Ω , and was independent of these operating parameters. The steady
 473 state substrate concentration (S) and substrate removal were almost staying unchanging at different
 474 electrical current (Figure 7E and F), but depended on the influent substrate concentration and
 475 dilution rate as discussed earlier. Removal efficiencies greater than 94.7% or about 100% were
 476 achieved at $S_{in} = 100$ mg L⁻¹ and $D = 0.1$ d⁻¹, and $S_{in} = 2400$ mg L⁻¹ and $D = 0.1$ d⁻¹, respectively.

477

478 4. Conclusions

479 The model proposed in this study was based on the assumption that the anode attached bacterial
 480 monolayer served as biocatalysts for MFC exoelectrogenesis. By modifying the Freter model and
 481 combining it with the Butler-Volmer equation, this model could adequately describe the processes
 482 of electricity generation, substrate utilization, and suspended and attached biomass growth, in both
 483 batch and continuous operating mode. MFC performances were impacted by various operating
 484 variables such as initial substrate concentration, external resistor, influent substrate concentration,
 485 and dilution rate, and their interactions, which were revealed by data simulation to be complicated.
 486 The simulation results explained that in batch mode, an intermediate initial substrate concentration
 487 (S_0 about 100 mg L⁻¹ at this reactor configuration) was appropriate to achieve maximum overall
 488 energy efficiency. With the S_0 of 100 mg L⁻¹, an external resistor (about 300 Ω) with the value of
 489 around the internal resistance (R_{in} of about 471 Ω) could boost the power generation, and a resistor
 490 of about two times (about 1000 Ω) of that value achieved better overall energy efficiency. However,
 491 a small external resistor should be adopted for MFC reactors aiming to rapidly remove organic
 492 substrates. In continuous mode and at $R_{ext}=2200$ Ω , the anode-attached biomass could fully grow
 493 when the influent substrate concentration was equal to or higher than 100 mg L⁻¹ at any dilution
 494 rates within the tested range. The maximum external power of 0.106 mW could be achieved at $R_{ext}=$
 495 352 Ω when there was enough substrate to sustain the power generation. An influent substrate
 496 concentration of 100 mg L⁻¹ achieved the overall energy efficiency greater than 10%. The high
 497 substrate concentration, e.g., 2400 mg L⁻¹, encouraged substantial biomass growth and thus reduced
 498 the overall energy efficiency to less than 2%. For the low influent substrate concentration (100 mg L⁻
 499 ¹), the maximum power generation and overall energy efficiency could not be concurrently

500 achieved, and operating MFC at an external resistor R_{ext} close to the internal resistance (R_{in}) or two
501 times of R_{in} was recommended to obtain optimal values for both indices. Overall, this relative
502 simple model provided a convenient way for evaluating and optimizing performance of MFC
503 reactors by regulating operating parameters.
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505 References

- 506 1. Logan, B. E., Exoelectrogenic bacteria that power microbial fuel cells. *Nat Rev Microbiol*
507 **2009**, 7, (5), 375-381.
- 508 2. Ren, Z.; Ward, T. E.; Regan, J. M., Electricity production from cellulose in a microbial fuel
509 cell using a defined binary culture. *Environ Sci Technol* **2007**, 41, (13), 4781-4786.
- 510 3. Rezaei, F.; Xing, D.; Wagner, R.; Regan, J. M.; Richard, T. L.; Logan, B. E., Simultaneous
511 cellulose degradation and electricity production by *Enterobacter cloacae* in a microbial fuel
512 cell. *Appl Environ Microb* **2009**, 75, (11), 3673-3678.
- 513 4. Picioreanu, C.; Head, I. M.; Katuri, K. P.; van Loosdrecht, M. C. M.; Scott, K., A
514 computational model for biofilm-based microbial fuel cells. *Water Res* **2007**, 41, (13), 2921-
515 2940.
- 516 5. Hamelers, H. V. M.; Ter Heijne, A.; Stein, N.; Rozendal, R. A.; Buisman, C. J. N., Butler-
517 Volmer-Monod model for describing bio-anode polarization curves. *Bioresource Technol*
518 **2011**, 102, (1), 381-387.
- 519 6. Ren, Z.; Ramasamy, R. P.; Cloud-Owen, S. R.; Yan, H.; Mench, M. M.; Regan, J. M., Time-
520 course correlation of biofilm properties and electrochemical performance in single-chamber
521 microbial fuel cells. *Bioresource Technol* **2011**, 102, (1), 416-421.
- 522 7. Bond, D. R.; Lovley, D. R., Electricity production by *Geobacter sulfurreducens* attached to
523 electrodes. *Appl Environ Microb* **2003**, 69, (3), 1548-1555.
- 524 8. Freter, R.; Brickner, H.; Fekete, J.; Vickerman, M. M.; Carey, K. E., Survival and
525 implantation of *Escherichia coli* in the intestinal tract. *Infection and Immunity* **1983**, 39, (2),
526 686-703.
- 527 9. Mašić, A.; Eberl, H. J., Persistence in a Single Species CSTR Model with Suspended Floccs
528 and Wall Attached Biofilms. *Bulletin of mathematical biology* **2012**, 1-26.
- 529 10. Liu, H.; Logan, B. E., Electricity generation using an air-cathode single chamber microbial
530 fuel cell in the presence and absence of a proton exchange membrane. *Environ Sci Technol*
531 **2004**, 38, (14), 4040-4046.
- 532 11. Schecher, W.; McAvoy, D., MINEQL+ Version 4.5. *Environ. Res. Software, Hallowell, ME*
533 **1998**.
- 534 12. Logan, B. E., *Microbial Fuel Cells*. 1st ed.; Wiley-Interscience: Hoboken, New Jersey, 2008.
- 535 13. Schröder, U., Anodic electron transfer mechanisms in microbial fuel cells and their energy
536 efficiency. *Physical Chemistry Chemical Physics* **2007**, 9, (21), 2619-2629.
- 537 14. Zeng, Y.; Choo, Y. F.; Kim, B. H.; Wu, P., Modelling and simulation of two-chamber
538 microbial fuel cell. *J Power Sources* **2010**, 195, (1), 79-89.
- 539 15. Neyerlin, K.; Gu, W.; Jorne, J.; Gasteiger, H. A., Study of the exchange current density for
540 the hydrogen oxidation and evolution reactions. *J Electrochem Soc* **2007**, 154, (7), B631-B635.

- 541 16. Batstone, D. J.; Keller, J.; Angelidaki, I.; Kalyuzhnyi, S.; Pavlostathis, S.; Rozzi, A.; Sanders,
542 W.; Siegrist, H.; Vavilin, V., The IWA Anaerobic Digestion Model No 1(ADM 1). *Water*
543 *Science & Technology* **2002**, 45, (10), 65-73.
- 544 17. Liu, J.; Lowy, D.; Baumann, R.; Tender, L., Influence of anode pretreatment on its microbial
545 colonization. *Journal of applied microbiology* **2007**, 102, (1), 177-183.
- 546 18. Freguia, S.; Rabaey, K.; Yuan, Z.; Keller, J., Non-catalyzed cathodic oxygen reduction at
547 graphite granules in microbial fuel cells. *Electrochim Acta* **2007**, 53, (2), 598-603.
- 548 19. Freguia, S.; Rabaey, K.; Yuan, Z.; Keller, J., Sequential anode-cathode configuration
549 improves cathodic oxygen reduction and effluent quality of microbial fuel cells. *Water Res*
550 **2008**, 42, (6-7), 1387-1396.
- 551 20. Wang, B.; Han, J.-I., A single chamber stackable microbial fuel cell with air cathode.
552 *Biotechnol Lett* **2009**, 31, (3), 387-393.
- 553 21. Fan, Y.; Sharbrough, E.; Liu, H., Quantification of the internal resistance distribution of
554 microbial fuel cells. *Environ Sci Technol* **2008**, 42, (21), 8101-8107.
- 555 22. Jia, Y. H.; Tran, H. T.; Kim, D. H.; Oh, S. J.; Park, D. H.; Zhang, R. H.; Ahn, D. H.,
556 Simultaneous organics removal and bio-electrochemical denitrification in microbial fuel
557 cells. *Bioprocess and biosystems engineering* **2008**, 31, (4), 315-321.
- 558 23. Katuri, K. P.; Scott, K.; Head, I. M.; Picioreanu, C.; Curtis, T. P., Microbial fuel cells meet
559 with external resistance. *Bioresource Technol* **2011**, 102, (3), 2758-2766.
- 560 24. Mench, M. M., *Fuel Cell Engines*. Wiley Online Library: 2008.
- 561 25. Wu, X.; Yao, W.; Zhu, J., Effect of pH on continuous biohydrogen production from liquid
562 swine manure with glucose supplement using an anaerobic sequencing batch reactor. *Int J*
563 *Hydrogen Energ* **2010**, 35, (13), 6592-6599.
- 564 26. Jung, S.; Regan, J. M., Comparison of anode bacterial communities and performance in
565 microbial fuel cells with different electron donors. *Appl Microbiol Biot* **2007**, 77, (2), 393-402.
- 566 27. Liu, H.; Cheng, S.; Logan, B. E., Power generation in fed-batch microbial fuel cells as a
567 function of ionic strength, temperature, and reactor configuration. *Environ Sci Technol* **2005**,
568 39, (14), 5488-5493.
- 569 28. He, Z.; Wagner, N.; Minteer, S. D.; Angenent, L. T., An upflow microbial fuel cell with an
570 interior cathode: assessment of the internal resistance by impedance spectroscopy. *Environ*
571 *Sci Technol* **2006**, 40, (17), 5212-5217.

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