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## 2 **Mechanical Energy before Chemical Energy at the** 3 **Origins of Life?**

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9 This paper is in honor of my distinguished colleague David Deamer and his 80<sup>th</sup> birthday.  
10 My UCSB ‘mica’ colleague Jacob Israelachvili introduced me to Deamer’s early work on  
11 encapsulating macromolecules in lipid vesicles [1] and the book Dave edited on Light  
12 Transducing Membranes from a joint United States-Australia conference held in Hawaii in  
13 1977.

14

15 **Abstract:** Forces and mechanical energy are prevalent in living cells. This may be  
16 because forces and mechanical energy preceded chemical energy at life’s origins.  
17 Mechanical energy is more readily available in non-living systems than the various  
18 other forms of energy used by living systems. Two possible prebiotic environments  
19 that might have provided mechanical energy are hot pools that experience wet/dry  
20 cycles and mica sheets as they move, open and shut, as heat pumps or in response  
21 to water movements.

22 **Keywords:** origin of life, origins of life, mechanical energy, work, entropic forces, mica, biotite,  
23 Muscovite, wet/dry cycles, clay

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### 25 **1. Introduction**

26 Forces and mechanical energy are prominent in living systems, at all size scales,  
27 from the molecular to the cellular and beyond [2-20]. Much of the cells’ chemical  
28 energy, such as ATP, is used to generate these forces. Perhaps mechanical energy  
29 in living cells is a remnant of mechanical energy that brought life into being, before  
30 chemical energy was readily available.

31 Mechanical forces shift reaction pathways [21]. Force lowers the transition  
32 states for reactions by tilting the energy landscape. Forces also give different  
33 reaction products than reactions without force [22].

34 Mechanical energy at life’s origins would resemble synthetic  
35 mechanochemistry, because there were no enzymes to carry out the (bio)chemical  
36 reactions. How feasible is synthetic mechanochemistry, in practice? Synthetic  
37 organic mechanochemistry has been used to produce many organic molecules,  
38 including pyrimidines [23], peptides, nucleosides, optically active products,

39 oxidations, reductions, condensations, nucleophilic reactions, and cascade reactions  
40 [24]. The industrial appeal of synthetic organic mechanochemistry is that it reduces  
41 the use of solvents.

42 Many forces pushed molecules around before 'biochemical' energy was  
43 available. These forces include hydration, dehydration, and surface forces, as well  
44 as entropic forces. Entropic forces "can have the counterintuitive effect of  
45 apparently introducing 'order'" [25]. The entropic forces are also known as  
46 'excluded-volume forces' or 'depletion interactions.' As water molecules become  
47 scarce, entropy increases when monomers join and free up a few water molecules  
48 that were constrained to linger near the ends of the monomers before their  
49 dimerization. The monomers experience an entropic force of attraction that brings  
50 them together.

51 These forces would generate mechanical energy (the product of force and  
52 distance), without chemical energy, at life's origins. In living systems, the  
53 motions and forces of enzymes need energy transduction from an energy source  
54 such as ATP. ATP and most of the energy sources now used by living systems  
55 were not available before at life's origins.

56 Mechanochemistry is a possible energy source for forming monomers as  
57 well as polymers. Some monomers, such as amino acids, may have been present on  
58 the early earth; but others, such as nucleotides were not present and were  
59 synthesized in some way/s [26].

60

## 61 **2. Materials and Methods**

62 Muscovite mica was obtained from New York Mica Co., New York, New York,  
63 and was scanned at 600 dpi with an HP Officejet 4635.

64

## 65 **3. Results and Discussion**

66 **Research on Previous 'Prebiotic' Polymerizations.** Proteins and  
67 nucleic acids are the main biopolymers involved in cell functions. Lipids are  
68 essential for cell functions but are not 'beads-on-a-string' polymers. Carbohydrates  
69 are also biopolymers but are used more for energy storage and cell structure. In most  
70 research investigating the prebiotic formation of proteins/peptides and nucleic  
71 acids/oligomers, the monomers are chemically activated, (e.g., [27-31]), which is not  
72 an ideal model for the origins of life.

73 More relevant research uses unactivated amino acids and nucleotides.  
74 Unactivated amino acids polymerized into peptides when dried but hydrolyzed  
75 when wet [32], which was also not ideal for the origins of life. A newer better  
76 'prebiotic' way of forming peptides used hot/dry vs cool/wet cycles [33]. Another

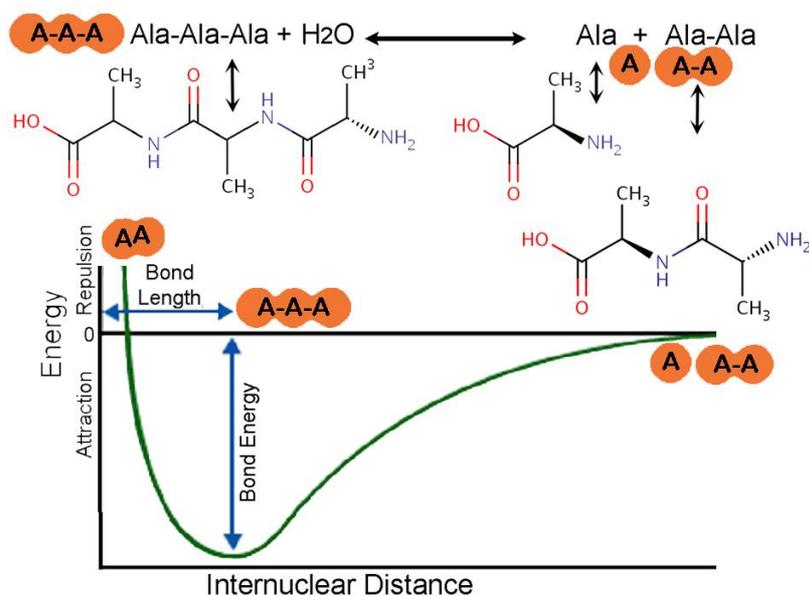
77 improvement was the use of both amino acids and hydroxy acids to form  
 78 depsipeptides, which have both amide (peptide) bonds and ester bonds. The ester  
 79 bonds form and break more easily than the peptide bonds. Initially, mostly ester  
 80 bonds formed. With cyclic wetting and drying, ester residues were replaced by  
 81 amino acid residues, leading to a hetero-polymer that was increasingly rich in amino  
 82 acids [33].

83 Clays have been used for many 'prebiotic' polymerization experiments,  
 84 catalyzing or supporting the formation of both peptides and oligonucleotides (e.g.,  
 85 [30, 31]). Clays are layered silicate minerals that swell when wet and shrink when  
 86 dry. Montmorillonite clays are best for these polymerizations. The anionic silicate  
 87 layers of Montmorillonite clays are held together by hydrated sodium (Na) ions.  
 88 The hydration of the Na ions causes the clay to shrink and swell in response to  
 89 drying and wetting.

90

91 **Two Embodiments of Mechanical Energy at Life's Origins.** Wet/dry  
 92 cycles and moving mica sheets are two sources of mechanical energy available for  
 93 powering the many types of chemical reactions that occurred as life was coming into  
 94 being. Mechanical energy might be capable of forming monomers and polymers of  
 95 prebiotic molecules. Polymer formation by mechanochemistry is diagrammed in  
 96 Fig.1 for the reaction of alanine + di-alanine to form tri-alanine.

97



98

99

100 **Figure 1.** Energy diagram of the way that mechanochemistry might polymerize  
 101 molecules, such as Alanine (A), shown here. (Top) tri-alanine, A-A-A, forming,  
 102 reversibly from alanine and di-alanine (A-A), with the release of a water molecule.

103 (Bottom) Force vs distance curve showing Attractive and Repulsive regimes as  
104 molecules are pushed closer together, to the bonding distance. Modified from [34].

105

106 “Fresh water” origins are assumed here, because low ionic-strength solutions  
107 are needed to form lipid membranes [35, 36]. (Deamer’s paper [35] has a wonderful  
108 analysis of lipids, membranes, and ‘informed guesses’ about their prebiotic  
109 assembly). There is also new evidence for origins in hot water on land [37-39] and  
110 evidence for origins in shallow ponds [40]. Whether life began in water on land or  
111 in water between mica sheets, the “fresh water” requirement holds for any origins  
112 scenario involving water-air interfaces. As Deamer points out, biochemists use  
113 dilute buffered aqueous solutions to do their biochemistry experiments, as opposed  
114 to ‘salt water’ [41].

115

#### 116 a. Wet/Dry Cycles

117 Cyclic wetting and drying on land occur in hot puddles where volcanoes were  
118 forming, such as the Kamchatka peninsula in Russia [42] or in active geothermal  
119 fields such as The Geysers in California [43]. Cyclic wetting and drying occurs on  
120 mineral or rock surfaces and has the advantage of concentrating prebiotic molecules  
121 during the drying phase, which overcomes the problems of dilution in Darwin’s  
122 “warm little ponds”. Prebiotic chemistry was tested in the Kamchatka peninsula, by  
123 pouring a sample of white powder into a hot clay-lined pool. The powder  
124 contained four amino acids and four chemical bases that compose naturally  
125 occurring nucleic acids, plus sodium phosphate, glycerol and a fatty acid. Foam and  
126 a white scum appeared quickly [37, 42].

127 An amazing discovery from the Deamer lab is that unactivated mononucleotides  
128 will polymerize during wetting and drying in the presence of lipids [44, 45]. Lipids  
129 protect the oligonucleotides from the hydrolysis that occurs during drying without  
130 lipids.

131 This lipid-assisted origin of life is proposed for the ancient Dresser Formation  
132 in the Pilbara region of Australia. These rocks contain evidence of the earliest life  
133 on land, more than half a billion years earlier than previously believed [37, 46].  
134 Once an active geothermal field, the Dresser Formation now has 3.48 billion-year-  
135 old stromatolite fossils, which appear to have formed on land and not in oceans, as  
136 was previously believed [41].

137 As Damer describes the wet/dry cycles, there was a 3<sup>rd</sup> stage – a moist gel stage  
138 - in the cycles of vesicles forming and breaking [37, 39, 47]. Molecules reassorted  
139 and grew in complexity in the moist gel, climbing up an ‘evolutionary ladder’ and  
140 ‘booting up’ the functions of life, through ‘programs written in polymers.’ Doyle

141 uses similar language in describing the emerging complexity of life and its  
142 processes: “All life and advanced technologies rely on protocol-based  
143 architectures” that are ‘robust yet fragile’ [48].

144 A fractionation of organic molecules would occur during drying in rocky  
145 puddles, with the most lipid-rich material forming ‘bathtub rings’ [41] with  
146 divalent salts at the earliest stages of drying. With continued drying and  
147 concentration of salts, prebiotic lipid mixtures with less and less lipid would dry  
148 onto the rocky walls as the density of the salt solution increased, with a moist gel  
149 phase at the bottom that would be enriched in non-lipid molecules.

150 The situation is analogous to isolating lipoproteins from blood plasma, except  
151 that lipoproteins are isolated by increasing the salt concentration such that different  
152 lipoproteins float to the top. The most lipid-rich lipoproteins rise to the top of the  
153 initial solution, and successively more lipid-poor lipoproteins rise to the top as the  
154 solution density is adjusted with salt to densities of 1.02 g/mL for Low Density  
155 Lipoprotein (LDL) and 1.21 g/mL for High Density Lipoprotein [49]. A similar  
156 fractionation likely occurred in drying pools at life’s origins.

157 Theoretical analyses of the wet/dry cycles have provided additional insights:  
158 As polymers dry, polymerization rates increase, because diffusion distances are  
159 shorter. As polymers continue drying, polymerization rates decrease, because  
160 crowding decreases diffusion rates [50]. Polymerization can be explained  
161 thermodynamically by excluded volume effects and molecular crowding [51], where  
162 entropic forces would generate mechanical energy.

163

#### 164 **b.Moving Mica Sheets**

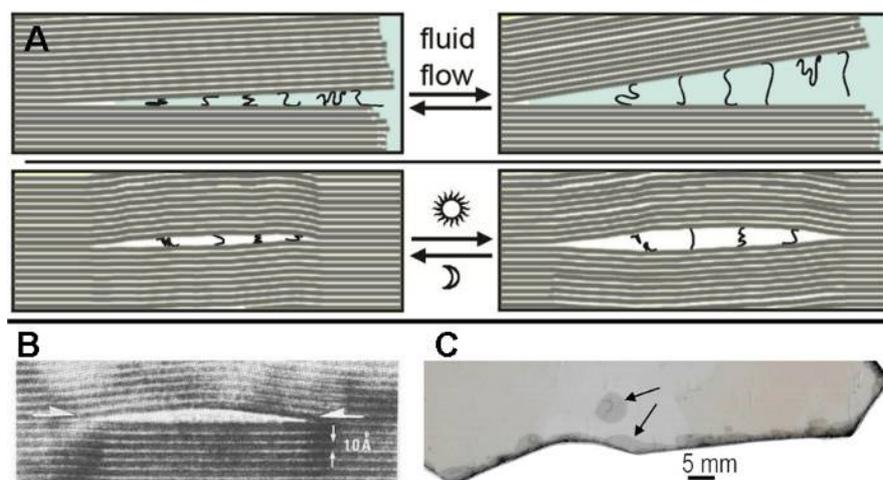
165 Mica is old – old enough to be the mineral from which life emerged [52]. Mica has a  
166 clay-like silicate structure with potassium (K) ions holding mica’s anionic sheets  
167 together. K ions are larger than Na ions, so there is no space for water molecules  
168 between unsplit mica sheets. Therefore, mica does not shrink and swell with  
169 drying and wetting, providing a more stable environment than clay particles.  
170 However, water seeps in at the edges of mica sheets, with cycles of heating and  
171 cooling (Fig. 2C) [53] and water can move farther in between the mica sheets,  
172 gradually, even to the point where the mica becomes ‘matted’, with large spaces  
173 between sheets.

174 Mica’s mineral sheets move, open and shut, in response to water flow (Fig. 2A)  
175 and heating and cooling of the mica sheets (Fig. 2B). The movements of the mica  
176 sheets squeeze and stretch the molecules with enough force to make and break  
177 covalent bonds between them [53]. Spaces between mica sheets form cantilever-type

178 springs, capable of generating a vast array of different forces, depending on the area  
179 and thickness of the mica cantilever.

180 Longer DNA molecules bind more strongly to mica sheets than shorter DNA  
181 polymers, which are more likely to be washed away, as observed by atomic force  
182 microscopy (AFM) [54, 55]. This stronger binding favors the accumulation of longer  
183 nucleic-acid molecules on mica, thus accumulating nucleic acids long enough to  
184 carry 'enough' information.

185



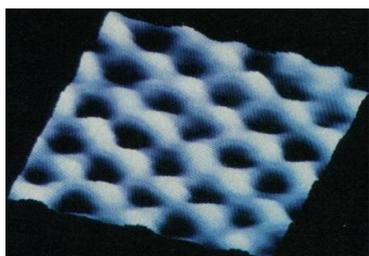
186 **Figure 2.** (A) Diagram of forces between biotite mica sheets, stretching and  
187 compressing polymers, due to: (Upper panels) water flow at the edges of the biotite  
188 sheets, and (Lower panels) heat pumps in a biotite bubble. (B) Biotite bubble  
189 imaged by HRTEM (high-resolution transmission electron microscopy). The  
190 thickness of a single biotite sheet is 1 nm (10 Angstroms) [56]. (C) Top view of a  
191 bubble in Muscovite mica (upper arrow) and of sheet separation at the edges of the  
192 mica sheet (bottom arrow). Bubbles are common even in 'high grade' micas. Biotite  
193 is now the preferred mica, for life's origins between mica sheets [57, 58]

194

195 Mica's anionic crystal lattice has a periodicity of 0.5 nm (Fig. 3), which is also the  
196 periodicity of phosphates on extended nucleic acids and sugar residues of  
197 carbohydrates. Amino acids in peptides have a smaller periodicity, such that a  
198 tripeptide has a length of ~ 1 nm. In mica, the 0.5-nm periodicity corresponds to the  
199 recessed hydroxyl groups in the mica surface, each of which carries  $\frac{1}{2}$  negative  
200 charge (i.e., either OH or O<sup>-</sup> groups).

201 DNA is an anionic polymer that interacts with inorganic cations in living cells.  
202 Perhaps life emerged from anionic mica sheets, where DNA also interacts with  
203 inorganic cations [55].

204



205  
206 **Figure 3.** Crystal lattice of Muscovite mica imaged by Atomic Force Microscopy  
207 (AFM), showing locations of recessed hydroxyl (OH) and ionized hydroxyl (O<sup>-</sup>)  
208 groups, as depressions (dark spots) on the mica surface. Image size is 2.6 nm x 2.6  
209 nm. Modified from [59].

210

211

212 Mica sheets might shelter emerging life without the need for membranes.  
213 Membranes are fragile. They leak, acquire and lose molecules, swell, and rupture. In  
214 living cells, membraneless organelles such as nucleoli contain RNA and protein.  
215 Although ribosomes are smaller than membraneless organelles, ribosomes have  
216 some of the most ancient RNAs and proteins. Ribosomes were present in the Last  
217 Universal Common Ancestor of life (LUCA) [60]. When life was coming into being,  
218 in the pre-LUCA stages, ribosomes and their precursors may have been the first  
219 ‘membraneless organelles’ [61].

220 Membranes and lipids are also compatible with mica surfaces, as seen by  
221 atomic force microscopy (AFM) of lipids on mica [62-64]. Therefore, mica could be  
222 the site for life’s origins, whether lipids were needed at the earliest stages of proto-  
223 life, or only at later stages in life’s emergence. Alternately, mica and hot pools might  
224 both be involved in the complex pathways to life’s origins.

225

#### 226 4. Conclusion

227 Mechanical energy in living systems provides energy in a form that is common  
228 in non-living systems. Wind, rain and waves do mechanical work on rocks, sand  
229 and water at all size scales from the molecular to the global. Entropic forces provide  
230 mechanical energy during drying or molecular crowding.

231 Mechanical work can be done without chemical energy, ion gradients, or proton  
232 gradients, which now provide energy for most of the processes in living systems.  
233 Ion gradients and proton gradients need an energy source to create the gradients,  
234 and they need a continuous supply of energy to maintain the gradients. Mechanical  
235 energy is a readily available energy source that may have brought life into being,  
236 and it is now found throughout living systems.

237

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244

245 **Conflicts of Interest:** The author declares no conflict of interest.

246

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