

1 Article

2 Evaluation of The Antifungal Activity of *Mentha x* 3 *piperita* (Lamiaceae) Of Pancalieri (Turin, Italy) 4 Essential Oil and Its Synergistic Interaction with 5 Azoles

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11 **Abstract:** The promising antimicrobial activity of essential oils (EOs) led researchers to use them in
12 combination with antimicrobial drugs in order to reduce drug toxicity, side effects, and resistance
13 with single agents. In Pancalieri (Turin, Italy), there is a local production of *Mentha x piperita*
14 worldwide known as “Mentha of Pancalieri”. The EO from this *Mentha* is considered as one of the
15 best peppermint EO in the world. In our research, we assessed the antifungal activity of “Mentha of
16 Pancalieri” EO either alone or in combination with azole drugs (fluconazole, itraconazole,
17 ketoconazole) against a wide panel of yeast and dermatophyte clinical isolates. The EO was
18 analyzed by GC-MS and its antifungal properties were evaluated by MIC/MFC parameters,
19 according to the CLSI guidelines, with some modifications. The interaction of peppermint EO with
20 azoles was evaluated through the checkerboard and isobologram methods. Results suggest this EO
21 exerts a fungicidal activity against yeasts, and a fungistatic activity against dermatophytes.
22 Interaction studies with azoles indicate mainly synergistic profiles between itraconazole and
23 peppermint EO vs. *Candida* spp., *Cryptococcus neoformans* and *Trichophyton mentagrophytes*.
24 Peppermint of Pancalieri EO may act as a potential antifungal agent and may serve as a natural
25 adjuvant for fungal infection treatment.

26 **Keywords:** essential oils; *Mentha x piperita*; “Mentha of Pancalieri”; azoles; antifungal activity; yeasts
27 and dermatophytes; synergism

29 1. Introduction

30 With the wide use of synthetic and semi-synthetic antimicrobial drugs, advantages and
31 disadvantages have been highlighted over the years, including the spread of drug-resistant
32 pathogens, and have focused research on natural products as useful antimicrobial tools. Currently,
33 there is evidence that EOs may exert remarkable biological activities against viruses, bacteria, fungi
34 and parasites. Several EOs are generally recognized as safe, do not accumulate in the liver or kidneys,
35 can stimulate the immune system, and cause no resistance, since microbes are unable to adapt to their
36 heterogeneous structure. *Mentha x piperita* L. (peppermint) EO is one of the most widely produced
37 and consumed EOs. Literature data have shown that peppermint EO and its main components
38 (menthol and menthone) display antimicrobial effects, but their mechanism of action is still not
39 elucidated [1-4]. Near Turin (Piedmont, Italy), in Pancalieri, there is a typical local production of *M.x*
40 *piperita* (Huds) var. OFFICINALIS (Sole), form RUBESCENS (Camus) (Lamiaceae), worldwide
41 known as “Mentha of Pancalieri” or “Mint Italo-Mitcham”, of which it is used or the green plant that
42 is dried for its conservation and used in herbal medicine, or the EO obtained from the steam current
43 distillation of the whole plant green grass (not shredded) [5]. The cultivation and distillation
44 processes of this plant boast a secular tradition. The EO from “Mentha of Pancalieri”, thanks to its

45 high quality and its peculiarities, is actually considered by experts to be as one of the best peppermint
 46 EO in the world. Due to the continuous increase of fungal infections and development of azole-
 47 resistant fungal strains, in our research, we assessed the antifungal activity of “*Mentha of Pancalieri*”
 48 EO against a wide panel of yeast and dermatophyte clinical isolates. The EO possible synergistic
 49 interaction with azoles, drugs used in fungal infections treatment, was also evaluated.
 50

51 2. Results

52 The phytochemical composition of “*Mentha of Pancalieri*” EO confirmed to be highly rich in
 53 oxygenated monoterpenes (menthol 41.7%; menthone 21.8%), in accord with data from the European
 54 Pharmacopoeia 8th Ed. (Table 1).

55 **Table 1.** Phytochemical composition of “*Mentha of Pancalieri*” essential oil in comparison with
 56 European Pharmacopoeia indication for *Mentha x piperita* (Lamiaceae) essential oil.

Main components	<i>Mentha of Pancalieri</i> EO (area %)	Eur. Ph. 8th ed. (area %)
limonene	1.8	1-3.5
1,8-cineole	5.3	3.5-8
menthone	21.8¹	14-32
isomenthone	1.5	1.5-10
menthyl-acetate	4.8	2.8-10
isopulegole	0.16	max 0.2
menthol	41.7¹	30-55

57 ¹ Main components.

58 “*Mentha of Pancalieri*” EO was found to exert a good inhibitory activity against all tested fungal
 59 strains (Table 2), in comparison with azole drugs (Tables 3-5). Notably, this EO exerted the most
 60 remarkable antifungal activity against *Cryptococcus neoformans*, displaying the lowest Minimum
 61 inhibitory concentration (MIC) and Minimum fungicidal concentration (MFC) values (Table 2, range
 62 0.06-0.125 %, v/v). A higher antimicrobial activity was also detected against other non-*Candida* yeasts,
 63 such as *Saccharomyces cerevisiae*, and *Pichia carsonii* (MIC range 0.125-0.5 and 0.125-0.25 %, v/v,
 64 respectively). Based on MFC results, “*Mentha of Pancalieri*” EO was found to display a fungicidal
 65 activity against yeast cells (MIC=MFC), and a fungistatic activity against dermatophytes. In fact,
 66 MFCs for dermatophytes were found to be one or two concentrations higher than MICs.

67 **Table 2.** MIC¹ and MFC² ranges of “*Mentha of Pancalieri*” essential oil towards yeasts and
 68 dermatophytes clinical strains.

Species	Isolates (n.)	MIC range (%v/v)	MFC range (%v/v)
<i>Candida</i> spp.			
<i>Candida albicans</i>	ATCC 90028	0.5	1
<i>Candida albicans</i>	6	0.5-1	0.5-1
<i>Candida glabrata</i>	ATCC 90030	0.5	0.5
<i>Candida glabrata</i>	2	0.5	0.5-1
<i>Candida krusei</i>	1	0.25	>1
<i>Candida parapsilosis</i>	1	0.5	0.5
<i>Candida tropicalis</i>	1	1	1
<i>Candida valida</i>	2	0.25-0.5	0.25-0.5
<i>Candida lusitanae</i>	1	0.5	0.5
<i>Candida norvegensis</i>	2	0.25-0.5	0.25-0.5
non-<i>Candida</i> spp.			
<i>Cryptococcus neoformans</i>	7	0.06-0.125	0.06-0.125

<i>Saccharomyces cerevisiae</i>	4	0.125-0.5	0.25-1
<i>Pichia carsonii</i>	2	0.125-0.25	0.125-1
<i>Sporobolomyces salmonicolor</i>	1	0.5	1
<i>Kloeckera japonica</i>	1	0.5	0.5
Dermatophytes			
<i>Trichophyton mentagrophytes</i>	2	0.5	>1
<i>Microsporum canis</i>	2	0.125	>1
<i>Microsporum gypseum</i>	1	0.125	>1

69 ¹MIC = Minimum inhibitory concentration; ²MFC= Minimum fungicidal concentration

70 No significant differences in inhibitory and fungicidal concentrations were found between azole-
71 susceptible (S) and azole-resistant (R) strains (Tables 3, 4).

72 By checkerboard testing, binary combinations of FLC with “Mentha of Pancalieri” EO were found to
73 be additive (0.5<FICI≤1) against either azole-S *Candida albicans* or azole-S *C.glabrata* strains (Table 3).
74 Notably, when synergistic or additive effects were not observed, no antagonism was reported, since
75 binary mixtures of FLC/peppermint EO yielded indifferent effects on the azole-R *C.krusei* strain
76 (FICI=2). Notably, the combinatorial effects between ITZ and “Mentha of Pancalieri” EO were found
77 to be synergistic (FICI≤0.5) against all *Candida* spp. strains (Table 3).

78 **Table 3. MIC¹ and FICI² of “Mentha of Pancalieri” EO plus fluconazole or itraconazole against**
79 **azole-S/R *Candida* spp. strains**

Antifungals		<i>C. albicans</i> azole-S strain ¹	<i>C. glabrata</i> azole-S [*] /R ^{**} strain ¹	<i>C. krusei</i> azole-R strain ¹
<i>Mentha of Pancalieri</i> EO	MIC (% v/v) alone	0.5	0.5	0.25
Fluconazole	MIC (µg/ml) alone	0.12	4*	64
	FIC of EO	0.5	0.5	1
	FIC of FLC	0.12	0.12	1
	FICI	0.62	0.62	2
	Interpretation	ADD⁴	ADD	IND²
<i>Mentha of Pancalieri</i> EO	MIC (% v/v) alone	0.5	0.5	0.25
Itraconazole	MIC (µg/ml) alone	0.5	2**	2
	FIC of EO	0.12	0.25	0.25
	FIC of ITZ	0.12	0.25	0.25
	FICI	0.24	0.5	0.5
	Interpretation	SYN⁴	SYN	SYN

80 ¹MIC = Minimum inhibitory concentration; ²FICI= Fractional inhibitory concentration index

81 ³Breakpoints determining susceptibility/resistance to fluconazole (FLC) and itraconazole (ITZ) according to the CLSI
82 document: FLC S≤0.125 µg/ml and R≥4 µg/ml (*C.albicans*), FLC R≥64 µg/ml *C.glabrata*); *C.krusei* is basically resistant
83 to FLC; ITZ S≤0.125 µg/ml and R≥1 µg/ml (*C.krusei*, *C.glabrata*) [6,7]

84 ⁴ADD: additive; IND: indifferent; SYN: synergy.

85 Moreover, the isobologram profiles confirmed a synergistic associations between ITZ and “Mentha
86 of Pancalieri” EO (Figure 1 a,b,c).

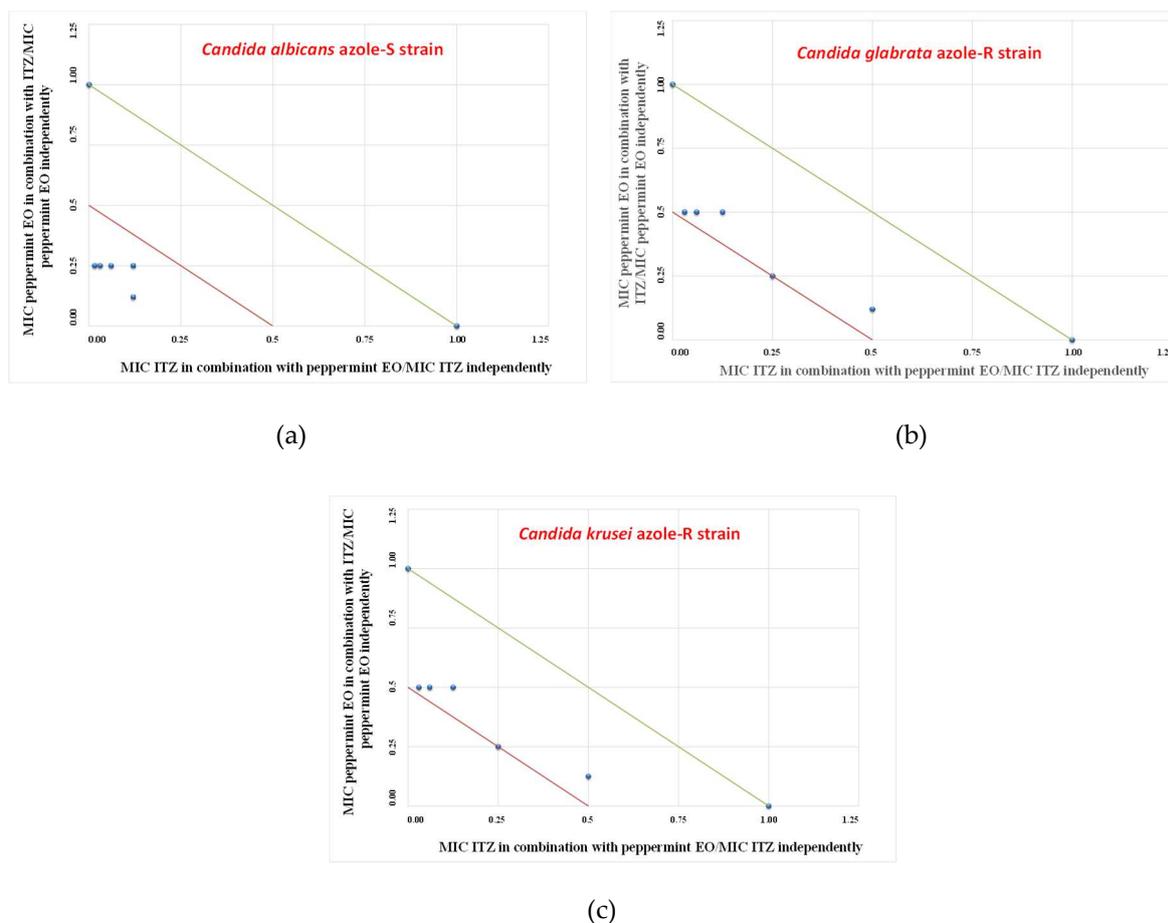


Figure 1. Isobolograms of synergistic interactions: plots of “*Mentha of Pancalieri*” EO and ITZ against *Candida* spp. (azole-susceptible/resistant strain). ITZ FIC data are drafted on x-axis, while EO FIC values are drafted on y-axis (a) Isobologram plot of EO and ITZ against *Candida albicans* azole-S strain; (b) Isobologram plot of EO and ITZ against *Candida glabrata* azole-R strain (c) Isobologram plot of EO and ITZ against *Candida krusei* azole-R strain.

In accord to the FIC indexes described for *C. neoformans* azole-S isolate, “*Mentha of Pancalieri*” EO was found to exert a synergistic profile (Table 4, $FICI \leq 0.5$), with a decrease of the MICs, as expressed by the isobologram in Figure 2. On the contrary, concerning *C. neoformans* azole-R strain, binary combination of ITZ/peppermint EO yielded additive effects ($FICI = 0.62$).

Table 4. MIC¹ and FIC² of “*Mentha of Pancalieri*” EO plus itraconazole against azole-S/R *Cryptococcus neoformans*.

Antifungals		<i>C. neoformans</i> azole-S strain	<i>C. neoformans</i> azole-R strain
<i>Mentha of Pancalieri</i> EO	MIC (% v/v) alone	0.06	0.06
Itraconazole	MIC (µg/ml) alone	0.5	2
	FIC of EO	0.12	0.5
	FIC of ITZ	0.25	0.12
	FICI	0.37	0.62
	Interpretation	SYN³	ADD

¹MIC = Minimum inhibitory concentration; ²FICI = Fractional inhibitory concentration index

³ADD: additive; SYN: synergy.

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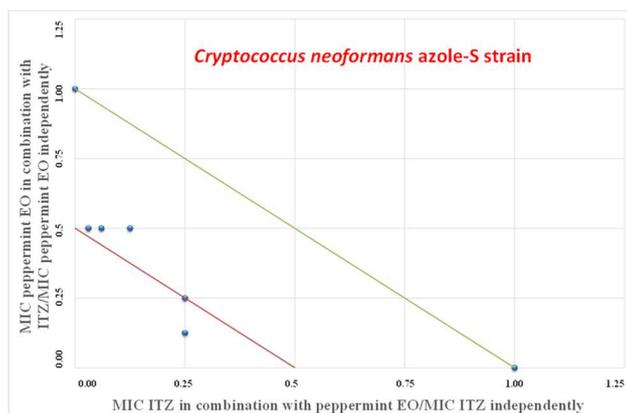
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Figure 2. Isobologram of synergistic interactions: plot of “Mentha of Pancalieri” EO and ITZ against *Cryptococcus neoformans* azole-S strain. ITZ FIC data are drafted on x-axis, while EO FIC values are drafted on y-axis.

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By checkerboard testing, binary combinations of ITZ/KTZ with “Mentha of Pancalieri” EO were found to be synergistic (Table 5, $FICI \leq 0.5$) against *Trichophyton mentagrophytes*, as represented by the corresponding isobologram in Figure 3 (a, b). Conversely, indifferent interactions of ITZ/KTZ and “Mentha of Pancalieri” EO binary mixtures were recorded against either *M.canis* or *M.gypseum* strains (Table 5, $1 < FICI < 4$).

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Table 5. MIC¹ and FICI² of “Mentha of Pancalieri” EO plus itraconazole or ketoconazole against dermatophytes

Antifungals		<i>Trichophyton mentagrophytes</i>	<i>Microsporium canis</i>	<i>Microsporium Gypseum</i>	
<i>Mentha of Pancalieri</i> EO	MIC (% v/v) alone	0.5	0.125	0.125	
	Itraconazole	MIC (µg/ml) alone	0.5	1	1
		FIC of EO	0.25	1	1
		FIC of ITZ	0.12	0.03	0.125
		FICI	0.37	1.03	1.125
Interpretation	SYN³	IND³	IND		
<i>Mentha of Pancalieri</i> EO	MIC (% v/v) alone	0.5	0.125	0.125	
	Ketoconazole	MIC (µg/ml) alone	2	4	2
		FIC of EO	0.25	1	1
		FIC of KTZ	0.12	0.015	0.015
		FICI	0.37	1.015	1.015
Interpretation	SYN	IND	IND		

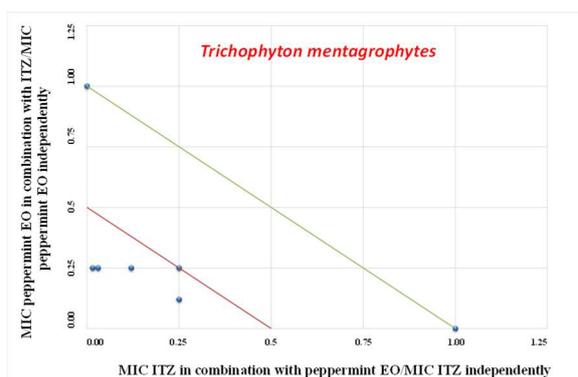
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¹MIC = Minimum inhibitory concentration; ²FICI= Fractional inhibitory concentration index

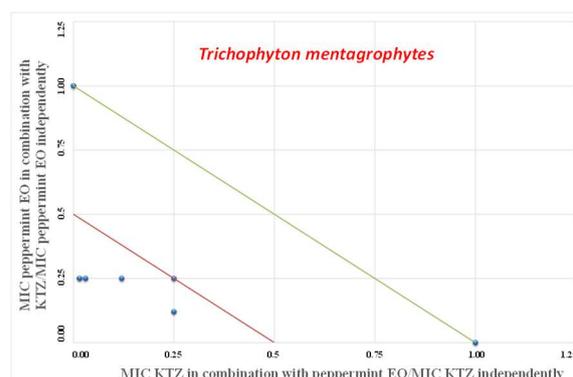
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³IND: indifferent; SYN: synergy.

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(a)

(b)

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Figure 3. Isobolograms of synergistic interactions: plots of “Mentha of Pancalieri” EO and ITZ or KTZ against *Trichophyton mentagrophytes*. ITZ/KTZ FIC data are drafted on x-axis, while EO FIC values are drafted on y-axis (a) Isobologram plot of EO and ITZ against *T.mentagrophytes*; (b) Isobologram plot of EO and KTZ against *T.mentagrophytes*.).

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3. Discussion

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Peppermint leaves (fresh and dried) and its EO are very well known herbal medicinal product, widely used since a long time, being a popular remedy inside and outside European countries, for its antispasmodic, choleric and carminative properties [8,9]. Some studies demonstrated that *Mentha x piperita* EO possesses antimicrobial properties and antioxidant activity [9]. In addition, *Mentha x piperita* EO is capable to inhibit the production of *Candida* and other microbial biofilms [10-12], reducing at subMIC concentrations in a dose-dependent manner the amount of exoproteins associated to *Staphylococcus aureus* virulence [13]. In this study, “Mentha of Pancalieri” EO exhibited a higher antimicrobial activity towards *C. neoformans*, *S.cerevisiae*, and *P.carsonii*, and displayed a good antimicrobial activity towards *C. glabrata* and *C. krusei*, species often resistant to conventional drugs. *Mentha x piperita* L. EO is able to carry out anticandidal activity with consequent cell “death”, by decreasing ergosterol amounts, producing intracellular acidification following PM-ATPase inhibition [4]. Recent studies have shown that *Mentha x piperita* L. EO inactivates *S. cerevisiae* that deteriorate fruit juices through the disorder of several physiological functions, i.e. enzymatic activity, efflux pump activity, early apoptosis [14]. “Mentha of Pancalieri” EO showed a higher activity against dermatophytes than azole drugs and displayed a fungistatic activity, in agreement with studies of Ibrahim *et al.* with *Mentha x piperita* L. EO [2].

The antimicrobial activity of “Mentha of Pancalieri” EO could be mainly related to menthol and/or menthone, the main compounds contained in the EO [4]. However, this activity may be due to a possible synergy between all the EO components [9, 10]. In fact, while our result about MICs for menthol (data not shown) indicated a better activity of this component against dermatophytes in comparison with EO *in toto*, MICs of menthone (data not shown) are twice higher those observed with the EO *in toto*.

The promising antimicrobial activity of EOs, has led researchers to use them in combination with available antimicrobial drugs, to assess their possible clinical use. Drug combinations represent a promising strategy for the development of new antifungal strategies, in order to overcome drug

163 resistance, limit the side effect and toxicity of drugs and increase therapeutic efficacy [10,15]. In this
164 research, we assessed the combined effect of “Mentha of Pancalieri” EO and three antifungal azoles,
165 such as FLC, ITZ, and KTZ. Our data showed a synergistic (against *Candida* spp., *C.neoformans*, and
166 *Trichophyton mentagrophytes*) interaction between ITZ and “Mentha of Pancalieri” EO, with a
167 noteworthy reduction in the concentration of the azoles at the MIC. However, our FICI result about
168 menthol and menthone in combination with ITZ or KTZ (data not shown) indicated only an additive
169 activity of these components against dermatophytes in comparison with EO *in toto*, suggesting once
170 again that the activity of an EO is due to the phytoextract that expresses its effectiveness.

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172 4. Materials and Methods

173 4.1 Essential oil.

174 Commercial “Mentha of Pancalieri” EO, obtained from the fresh leaves of *M.x piperita* (Huds) var.
175 OFFICINALIS (Sole), form RUBESCENS (Camus) (Lamiaceae) by steam distillation, was purchased
176 from Erbe Aromatiche Essenzialmenta, Pancalieri (Turin, Italy). “Mentha of Pancalieri” EO
177 composition was analyzed by GC-MS at Drug Science and Technology Department (University of
178 Turin, Italy) with a Supelcowax capillary column (Supelco, Bellefonte, PA) as previously described
179 [16]. For susceptibility testing, EO was dissolved in ethanol (1:2.5), and diluted (1:20) up to 2% (v/v)
180 in RPMI-1640 medium with L-glutamine and without sodium bicarbonate (Sigma-Aldrich, Rome,
181 Italy), as previously described [17]. Then, 0.165 M morpholinepropanesulfonic acid (MOPS) (Sigma-
182 Aldrich) and 0.2% glucose were added to EO solution. Final pH was 7.0. The EO was protected from
183 light and humidity and maintained at 4 °C just before use [17].

184 4.2 Antifungal agents.

185 Fluconazole (FLC; F8929), itraconazole (ITZ; I6657), and ketoconazole (KTZ; K1003) powders
186 (≥98% purity by HPLC) were achieved from Sigma-Aldrich. FLC solutions were prepared in sterile
187 distilled water, whereas ITZ and KTZ solutions were prepared in 100% DMSO (Sigma-Aldrich), and
188 stored at -20°C just before use [15].

189 4.3 Fungal strains.

190 16 *Candida* spp., (*C.albicans*, *C.glabrata*, *C.krusei*, *C.parapsilosis*, *C.tropicalis*, *C.valida*, *C.lusitaniae*,
191 *C.norvegensis*), 15 non-*Candida* spp. clinical strains (*Cryptococcus neoformans*, *Saccharomyces cerevisiae*,
192 *Kloekera japonica*, *Pichia carsonii*, *Sporobolomyces salmonicolor*), and 5 dermatophyte clinical isolates
193 (*Trichophyton mentagrophytes*, *Microsporum canis*, *M.gypseum*) were tested (Table 2). *C.albicans* ATCC
194 90028 and *C.glabrata* ATCC 90030 were also included.

195 Yeasts identification was determined by the API ID32C systems (BioMérieux, Rome, Italy). Then,
196 strains were maintained at -80 C in Microbanks™ (Pro-Lab Diagnostics, Neston, UK), and cultured
197 twice on Sabouraud dextrose agar (SDA, Oxoid, Milan, Italy) at 35°C for 72h before the assays.
198 Dermatophyte identification was carried out by macroscopic and microscopic observation of the
199 colonies and reproductive structures after 15 days of incubation on Mycobiotic agar at 25°C (Merck,
200 KGAA, Germany) [17,18]. Molds were stored in SDA at 4°C until use. For susceptibility testing non-
201 germinated conidial suspensions were prepared as previously described [17].

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203 4.4 In vitro antifungal susceptibility assays.

204 Fungal strains were assayed for susceptibility to “Mentha of Pancalieri” EO, and to azoles (FLC,
205 ITZ, KTZ), by a broth microdilution method, in accordance to CLSI guidelines (CLSI M27-A3 and
206 M27-S4) [19, 20] for yeasts, and CLSI M38-A2 [21] for dermatophytes, with some modifications for
207 the EO [15,17]. Tween 80 (Sigma-Aldrich) (final concentration 0.001%, v/v) was employed to enhance
208 EO solubility, without precluding fungal proliferation. Blastoconidia yeast suspensions and
209 dermatophyte conidial suspensions were prepared to yield final inocula of ~1.5×10³ CFU/ml, and
210 ~1.5×10⁴ CFU/ml, respectively [15, 17]. Microdilution plates were set up, as previously described

211 [15,17] and yeast strains were incubated at 35°C for 24h, whereas dermatophytes were incubated at
212 30°C for 7 days. MICs of FLC, ITZ, and KTZ were determined visually as the lowest concentration of
213 drug that produced complete inhibition (dermatophytes) or a significant reduction ($\geq 50\%$ inhibition,
214 yeasts) of growth, in comparison with control. MICs of EO were considered the lowest concentration
215 with no visible growth. MFC was determined by inoculating 10 μ l from wells not turbid on SDA agar
216 plates incubated for 3 (yeasts) and 4 (dermatophytes) days at 30°C or at 35°C (*Cryptococcus* sp). MFC
217 was defined as the lowest concentration resulting in no growth on subculture from the MICs [15,17].

218 *4.5 Checkerboard assays and evaluation of the fractional inhibitory concentration index (FICI).*

219 A two-dimensional checkerboard with serial twofold dilutions below the MIC to 2xMIC of each
220 compound was performed in a 96-well microtiter plate, as previously described [15, 22].
221 Microorganism suspensions were joined to each well containing mixtures of azole drug/EO. FICI
222 values were calculated according to the following formula [22]: $FICI = \frac{FICa + FICb}{MICa}$ in
223 combination/MICa tested alone + MICb in combination/MICb tested alone; where MICa and MICb
224 are the MICs of azole and of EO used alone. Synergy and antagonism were defined by FICI values of
225 ≤ 0.5 and > 4 , respectively. A FICI value between 0.5 and 1.0 was interpreted as additive, whereas a
226 value between 1.0 and 4.0 was interpreted as indifferent [22].

227 *4.6. Isobolograms.*

228 Data of the checkerboard assays were expressed graphically by isobolograms, as previously
229 described [15,22]. FIC values of azole drug were reported on y-axis, whereas FIC values of
230 peppermint EO on x-axis. The straight line that joins the intercept points is the line of additivity
231 (FICI=1). Below this line we can find the additive area ($0.5 < FICI \leq 1$) and synergistic (FICI ≤ 0.5) effects,
232 respectively. FICI values above of the straight line corresponded to indifferent ($1 < FICI < 4$) or
233 antagonistic (FICI > 4) interactions [22].

234 5. Conclusions

235 The EO of “*Mentha of Pancalieri*” contains mainly menthol and menthone, in according with other
236 countries and European Pharmacopeia guidelines. Our data are difficult to compare with literature
237 data because of different methodology used in experimental procedures and different *Mentha x*
238 *piperita* L. EOs composition. For example, the EO acquired from the dried leaves of *Mentha x piperita*
239 in Brasil contains 51% of linalool, as the main compound [23], probably due to different cultivation
240 conditions. It is very important that mint cultivation and EO production be strictly controlled,
241 because it is recognized that the chemical composition of mint EOs can be influenced by many factors,
242 such as, humidity, nutrients, temperature, etc.,. From this point of view, “*Mentha of Pancalieri*” can
243 guarantee controlled composition, and consequently high quality that make it an effective product
244 for its exploitation.

245 “*Mentha of Pancalieri*” EO may act as a potential antifungal agent and may serve as a natural
246 adjuvant for fungal infection treatment. Further researches are necessary to confirm these data.

247 **Author Contributions:** Conceptualization, V.T. and D.S.; methodology, N.M. and J.R.; investigation, V.T.; data
248 curation, N.M and J.R.; writing—original draft preparation, V.T. and D.S.; writing—review and editing, V.T.

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253 **Conflicts of Interest:** The authors declare no conflict of interest.

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330 **Sample Availability:** Samples of the compounds Mentha of Pancalieri EO, fluconazole, itraconazole,
331 ketoconazole, and fungal clinical strains (yeasts and dermatophytes) are available from the authors.