**Table S1.** The efficiency of each primer pair used for qRT-PCR analysis.

|  |  |  |
| --- | --- | --- |
| **Gene** | **Primer** | **PCR efficiency (*E*) 1** |
| SSUSC84\_RS00185 | Q0185F | 105% |
| Q0185R |
| SSUSC84\_RS00550 | Q0550F | 104% |
| Q0550R |
| SSUSC84\_RS01570 | Q1570F | 104% |
| Q1570R |
| SSUSC84\_RS03030 | Q3030F | 110% |
| Q3030R |
| SSUSC84\_RS03035 | Q3035F | 104% |
| Q3035R |
| SSUSC84\_RS03040 | Q3040F | 107% |
| Q3040R |
| SSUSC84\_RS03045 | Q3045R | 103% |
| Q3045F |
| SSUSC84\_RS03050 | Q3050F | 104% |
| Q3050R |
| SSUSC84\_RS06475 | Q6475F | 103% |
| Q6475R |
| SSUSC84\_RS07245 | Q7245F | 108% |
| Q7245R |
| 16s RNA | Q16S1 | 101% |
| Q16S2 |

1 The efficiency of each primer pair was determined using serially diluted genomic DNA, as previous described [1]. The efficiency (*E*) is defined as the fraction of target DNA molecules that are copied after one PCR cycle. *E* should be 100% when the number of target DNA molecules is perfectly doubled in every PCR cycle. Several factors, such as primer dimer, might result in *E* > 100%. According to the instructions of the NovoStart SYBR qPCR SuperMix Plus kit, 90% ≤ E ≤ 110% is acceptable for qRT-PCR analysis.

**Reference**

1. Svec, D.; Tichopad, A.; Novosadova, V.; Pfaffl, M.W.; Kubista, M. How good is a PCR efficiency estimate: Recommendations for precise and robust qPCR efficiency assessments. *Biomolecular detection and quantification* **2015**, *3*, 9-16, doi:10.1016/j.bdq.2015.01.005.