Herbicidal effects and cellular targets of aqueous extracts from young *Eucalyptus globulus* leaves - a novel post-fire management strategy of eucalyptus stands

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Abstract

Along with climate change, the native forest replacement by exotic species, such as *Eucalyptus globulus*, is contributing to a highly fire-prone environment. Since *E. globulus* detains several post-fire regeneration strategies, sustainable practices are needed to manage eucalyptus stands. Thus, eucalyptus allelopathic potential can be used for weed control. Therefore, this study aimed at assessing the herbicidal potential of post-fire regenerated *E. globulus* leaves in *Portulaca oleracea*, and unraveling the main physiological processes disturbed by biocide application. For this, an aqueous extract prepared with fresh leaves (FLE; 617 $g_{fresh\ weight}L^{-1}$) and other with oven-dried leaves (DLE; 250 $g_{dry\ weight}L^{-1}$) were prepared and foliar-sprayed twice-a-week at different dilutions in 7-days-old purslane plants. As positive control, glyphosate was used. After five weeks, results revealed that DLE at the highest dose detained the greatest herbicidal activity against *P. oleracea*. To understand how DLE impacted weed physiology, several biochemical and redox-related parameters were evaluated in purslane plants treated with

DLE highest dose. Results suggested an overproduction of hydrogen peroxide, causing severe oxidative damage in roots. Overall, this study showed that young *E. globulus* dried leaves had powerful herbicidal properties against *P. oleracea* and can represent a feasible approach for weed management, while reducing fire hazard.

Keywords: sustainable agriculture; allelopathy; biocidal potential; phytotoxic effects; weed management.

[1]**Abbreviations**

1. INTRODUCTION

Weeds are the main responsible for crop yield and quality losses, since both weeds and crops, compete for the same resources, such as light, water, minerals, and even space (Dadkhah and Asaadi, 2010). For this reason, modern agriculture is highly dependent on the use of synthetic herbicides, which can cost up to US$7 billion annually (Baucom, 2019). However, although these products are effective in eliminating weeds and avoiding major economic losses, their continued application has resulted in herbicide resistance acquisition by weed species (Baucom, 2019) and, also, in the contamination of several environmental matrices, such as air, soil and water (Geetha, 2019; Hernandez et al., 2013; Shahena et al., 2020), where they can negatively affect non-target organisms, including crops (Aktar et al., 2009; Soares et al., 2019b). In this way, new, eco-friendly, and economic solutions to combat weeds need to be urgently found.

Some plants have the ability to release compounds with a powerful biological activity into the environment, the allelochemicals, which prevent the growth and development of neighbouring plants (allelopathy) (Reigosa et al., 2000). This is the case of *Eucalyptus globulus* Labill. subsp. *globulus,* usually referred to as common eucalyptus or tasmanian blue gum, a hardwood tree native from southeast Australia, now widely distributed throughout the world

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[1] Abbreviations: AOX- antioxidant; CTL- control; DLE- dried leaf extract; DW- dry weight; FLE- fresh leaf extract; FW- fresh weight; GLY- glyphosate; IAS- invasive alien species; LP- lipid peroxidation; MDA- malondialdehyde; OECD- Organisation for Economic Co-operation and Development; ROS- reactive oxygen species; SD- standard deviation.

mainly due to its favourable wood characteristics for the paper industries (Águas et al., 2014). Although flowers, fruits, bark, and roots have allelochemicals in their composition, the recognised *E. globulus* allelopathic activity is mainly attributed to its leaves (Morsi and Abdelmigid, 2016). In fact, extracts prepared from leaves have been proven to be more effective in allelopathic inhibition than other organs (Morsi and Abdelmigid, 2016). Therefore, the allelopathic properties of *E. globulus* leaves can be used in a targeted way to suppress the growth of undesirable plant species, such as weeds (Puig et al., 2018).

Thus far, multiple studies were performed aiming at finding effective synthetic herbicide alternatives, taking advantage of the allelopathic properties of eucalyptus leaves (Aragão et al., 2015; El-Ghit and Hanan, 2016; Jaime and Ferrer, 2018). Most of these studies evaluated the herbicidal potential of concentrated essential oils from *Eucalyptus* species (Jaime and Ferrer, 2018; Rassaeifar et al., 2013). However, leaf essential oils are obtained mainly by hydrodistillation procedures, which require expensive and specialized equipment (Dilworth et al., 2017), making them dependent of industrial transformation. As so, aqueous extracts prepared with eucalyptus leaves can constitute affordable alternatives, allowing the obtention of not only essential oils but also polyphenolic compounds from the leaves, while ensuring easiness and reproducibility of the procedure. Nonetheless, there are still some gaps regarding the use of eucalyptus-based products as biocides.

Up to now, most of the works used extracts from *Eucalyptus citriodora* (Aragão et al., 2015; Benchaa et al., 2018; Chaturvedi et al., 2012; El-Rokiek and Eid, 2009) and *Eucalyptus camaldulensis* (Chaturvedi et al., 2012; Dadkhah, 2013; Verdeguer Sancho et al., 2018). Nevertheless, due to the favourable properties of its wood for the pulpwood industries, *E. globulus* is the most recurrent exotic species in several countries, such as Portugal (Águas et al., 2014). In this sense, the biocidal properties of this species have recently attracted more attention, with few studies evaluating the potential of concentrated essential oils (Jaime and Ferrer, 2018; Rassaeifar et al., 2013), leachates (Babu and Kandasamy, 1997; Padhy et al., 2000), and aqueous extracts (Ataollahi et al., 2014; Dejam et al., 2014; Morsi and Abdelmigid, 2016; Puig et al., 2018). However, concerning aqueous extracts, most of the studies conducted

so far evaluated the biocidal potential of *E. globulus* extracts in a pre-emergence context, concluding that they are capable of effectively impairing seed germination and seedling growth of *Solanum nigrum* L. (Ataollahi et al., 2014), *Solanum melongena* L. (Dejam et al., 2014), and *Hordeum vulgare* L. (Morsi and Abdelmigid, 2016), leaving much to clarify about the post-emergent biocidal properties of *E. globulus* aqueous extracts.

In addition, although some studies omit the age of the eucalyptus leaves utilized, only the mature leaves, collected from adult *E. globulus* trees, were used to prepare the aqueous extracts or leachates tested until now (Babu and Kandasamy, 1997; Bagavathy and Xavier, 2007; Puig et al., 2018). Thereby, the herbicidal potential of aqueous extracts prepared with young eucalyptus leaves constitutes a gap in knowledge and needs further investigation.

Due to their smaller heights, young eucalyptus trees or young resprouts tend to be more susceptible to herbivory than mature trees and, for this reason, leaves from young trees tend to have higher amounts of defensive specialized metabolites than those of adult trees (O'Reilly-Wapstra et al., 2007). The distinctive morphology between young and adult *E. globulus* leaves also points out to possible differences in their chemical composition: while leaves from young trees are blue-grey and have a thin and broad shape, mature leaves are typically dark green, thick and narrow (James and Bell, 2001).

Furthermore, the use of the leaf biomass from young eucalyptus trees, instead of mature ones, for the preparation of the biocides may represent a sustainable approach to control eucalyptus populations, outside plantations. Due to their fire-adaptation traits, when affected by a wildfire, *E. globulus* trees have a great ability to produce new shoots and thus, lead to the reestablishment of a vast leaf area, mainly by the resprouting from dormant buds located in aboveground parts of the trees, such as leaf axils, branches, and boles, or in the subterranean lignotubers or root collars (Burrows, 2002, 2013; Catry et al., 2013). This reproduction strategy contributes to a rapid dispersion of the species even over areas previously occupied by native trees (Catry et al., 2013), hampering post-fire management efforts in the urban-rural interface. As *E. globulus* is the most abundant species in multiple countries, outside its native geographic range, its elimination or replacement with native and less fire-prone species is very difficult due

to several political, economic, and ecological factors. Thereby, adding value to the leaf biomass of post-fire regenerated eucalyptus trees represents a way of controlling the dispersion of this exotic species, preventing the reestablishment of non-managed *Eucalyptus* stands, which, in turn, contributes to a better land-use planning. In fact, biological actions to eradicate, control or contain a population of certain invasive alien species (IAS) are included in the Article 19 from the European Union (EU) 1143/2014 for the management of widely spread IAS (*http://data.europa.eu/eli/reg/2014/1143/oj*).

With this in mind, this study aims to evaluate the post-emergent herbicidal potential of the leaves of young *E. globulus* trees that spontaneously resprout after a wildfire, in a common weed species, *Portulaca oleracea* L. (purslane) and to compare its effectiveness with synthetic herbicides, as well as to ascertain which biomolecules and cellular weed processes are affected by the biocide, in order to get an insight of its possible mode-of-action. Through the combination of physiological, histochemical and biochemical approaches, it was possible to conclude that, when dried, the leaves from post-fire regenerated *E. globulus* trees had the greatest herbicidal activity against purslane plants, mainly by inducing severe redox imbalances in their root system, which the antioxidant machinery was not able to overcome.

2. MATERIALS AND METHODS

2.1. Preparation of the aqueous extracts

Foliage from young *E. globulus* trees, recently regenerated after a wildfire, were collected in December 2019 in a forest area of Porto, Portugal (41.197288, -8.534506) burnt in the summer of 2019, according to locals. After manually detaching the leaves from the stems, a portion of the fresh leaves was oven-dried at 60 ºC until reaching a constant weight. Then, fresh and dried leaves were reduced to small fragments, in order to promote the release of the allelochemicals, and underwent an extraction in deionised water at 70 ºC, for 30 min, with regular agitation. The two obtained solutions were centrifuged twice (-4 ºC, 15 000 *g*), during 25 and 15 min, respectively. The final concentration of the aqueous extracts corresponded to the maximum

volume of leaves that could be fully submersed in 1 L of deionised water. Considering the moisture content of fresh leaves [59 ± 0.6 % (w/w)], two biocides were obtained: one was prepared with dried leaves for a final concentration, in a dry weight (dw) basis, of 250 g_{dw} L^{-1} and the other was made with fresh leaves at a final concentration, in a fresh weight (fw) basis, of 617 g_{fw} L^{-1}. Lastly, both aqueous extracts were filtered through nitrocellulose filters with 1.2 μm and stored at -80 ºC until use. In order to functionally characterize the obtained extracts, an exhaustive analysis of their phytochemical profile was obtained by gas chromatography with mass spectrometry detection (GC-MS) and high performance liquid chromatography with UV/Vis and tandem mass spectrometry detections (HPLC-UV/Vis-MS/MS) to identify volatile and polyphenolic compounds. Details on the obtained results can be found elsewhere (Pinto, 2020). No major differences were recorded between both extracts, but those prepared with dried leaves presented a much larger diversity of ellagitannins and greater amounts of galloyltannins, two groups of hydrolysable tannins with known allelopathic activities (Pinto, 2020; Pinto et al., in prep).

2.2. Preparation of the artificial soil

The substrate used in all experiments consisted in a standard artificial soil, manually prepared according to the guidelines of the Organisation for Economic Co-operation and Development (OECD) (OECD, 2006), composed of 70% (w/w) sand, 20% (w/w) kaolin, and 10% (w/w) peat [pH (KCl) 6.0 ± 0.5].

2.3. Test-species and growth conditions

Portulaca oleracea L. seeds were purchased from a local supplier and used as model weed species. Seeds were surface disinfected with 70% (v/v) ethanol for 10 min, followed by 20% (v/v) commercial bleach (5% active chlorine) for 10 min, and a series of successive washes in deionised water. Before sowing, visually damaged purslane seeds were discarded. The experiments were performed in plastic pots containing 150 g_{dw} of the artificial soil. In order to

ensure the maintenance of soil moisture, a cup with deionised water was placed under each pot. At the beginning of the assays, the cups were filled with 100 mL of Hoagland solution (HS) (Taiz et al., 2015), instead of deionised water, to guarantee nutrient availability. The communication between each plastic pot and the cup HS/deionised water was performed by a cotton rope inserted in a hole at the bottom of the pots. All experimental assays were performed in a growth chamber with controlled conditions [temperature: 25 ºC; photoperiod: 16 h/8 h light/dark; photosynthetically active radiation (PAR): 120 µmol m^{-2} s^{-1}].

2.4. Evaluation of the biocidal potential of the aqueous extracts in purslane plants - optimization of the most effective dose and extract (FLE or DLE)

To test the post-emergent herbicidal potential of the two aqueous extracts, as well as to determine the most effective formulation [fresh leaf extract (FLE) or dried leaf extract (DLE)] and concentration , FLE and DLE were foliar-applied (20 sprays each, corresponding to 15 mL of extract per pot), twice-a-week, at different concentrations [12.5, 25, 50, 75 and 100 % (v/v)], in 7-days-old purslane seedlings grown in OECD soil. The seedlings of the control (CTL) were sprayed, at the same rate, with deionised water. Glyphosate [RoundUp^{TM} UltraMax; 360 g L^{-1} glyphosate (GLY) as potassium salt] was used as a positive CTL to achieve a fair comparison between the biocides and commercially available options. Based on the manufacturer's recommendations, a single application equivalent to 10 L ha^{-1} was performed. Considering the area of the plastic pots (0.011 m^2), this application rate corresponded to 3.96 mg of GLY per pot/replicate. To ensure that the 15 mL of the pulverization carried the adequate amount of the active ingredient (4 mg), a solution 0.267 g L^{-1} of GLY was prepared. For each treatment, 4 experimental replicates were used, each one with 20 purslane seeds sown. The percentage of viable plants, as well as their growth and macroscopic phytotoxicity symptoms were weekly monitored throughout the assay. Upon 5 weeks of treatments, plants were collected, separated into shoots and roots, and used for the estimation of biometric parameters (shoot and root biometry and fresh biomass), and for histochemical assays.

2.4.1. Biometric evaluation

At the end the assay, and upon the subsequent separation of shoots from roots, the length and fresh weight of both organs were measured. The shoot length of purslane plants was measured from the shoot apex to the shoot-root transition. Regarding shoot and root length, each plant from each replicate was individually measured, while shoot and root fresh biomass was obtained by measuring together all the individuals from each replicate. To eliminate the interference of individuals' number, the shoot and root weight from each replicate was divided by the number of individuals of each replicate at the end of the assay.

2.4.2. Histochemical detection of cell death

The cellular viability was assessed based on the protocol of Romero-Puertas et al. (2004). For this purpose, purslane shoots (one per replicate) were immersed in a solution of 0.25% (w/v) Evans Blue for 5 h. Then, the plant material was transferred to boiling 96% (v/v) ethanol in order to remove the photosynthetic pigments. The appearance of blueish spots on leaf surface is indicative of unviable cells (cell death). The results were photographically recorded.

2.5. Assessment of the phytotoxicity induced by the application of the optimized aqueous extract in purslane plants

To understand the physiological processes affected by the aqueous extract application, 7-days-old purslane seedlings, grown in OECD soil, were foliar sprayed with DLE at 75% (v/v) and 100% (v/v) – the two most effective concentrations of the best biocide formulation. The CTL seedlings were sprayed with deionised water. For each treatment, 8 experimental replicates were considered. Five weeks upon the beginning of the treatments, purslane plants were collected, separated into roots and shoots, frozen in liquid nitrogen and stored at -80 ºC for further quantification of the levels of lipid peroxidation and H_2O_2 and of several biomolecules (sugars, proteins, amino acids, and photosynthetic pigments) as well as the activity of nitrogen metabolism-related enzymes (nitrate reductase and glutamine synthetase). For each

determination, samples from, at least, three experimental replicates from each experimental group [CTL, DLE at 75% (v/v) and 100% (v/v)] were used.

2.5.1. Lipid peroxidation (LP)

The spectrophotometric quantification of LP was performed by the quantification of malondialdehyde (MDA) levels, using samples of plant material homogenized in 0.1% (w/v) trichloroacetic acid (TCA) (Heath and Packer, 1968). After reading the absorbance (Abs) at 532 and 600 nm (unspecific turbidity) and considering an extinction coefficient of 155 mM cm^{-1}, MDA levels were calculated, and the results expressed in nmol g^{-1} fw.

2.5.2. Hydrogen peroxide (H_2O_2)

The determination of H_2O_2 levels was carried out based on the protocol of Alexieva et al. (2001), by following a spectrophotometric method in which H_2O_2 reacts with potassium iodide (KI; 1 M) for 1 h. After recording the Abs at 390 nm, the results were expressed as pmol g^{-1} fw, considering an extinction coefficient of 0.28 μM^{-1} cm^{-1}.

2.5.3. Total sugars, proteins, free amino acids, and proline

The levels of total sugars were quantified according to the anthrone-based method of Irigoyen et al. (1992), using frozen samples of shoots and roots. Upon reaction between the extract and anthrone at high temperature conditions (100 ºC; 10 min), the Abs of each sample was registered at 625 nm. Sugar levels were calculated by linear regression, using a curve prepared with glucose standards, and results were expressed in μg g^{-1} fw. The content of total soluble proteins was quantified according to the method of Bradford (1976), using frozen plant aliquots. The protein levels were expressed in mg g^{-1} fw and were calculated through a calibration curve, obtained by using different known bovine serum albumin concentrations. The levels of total free amino acids were quantified based on a ninhydrin-based protocol, as described by Lee and Takahashi (1966). A calibration curve with solutions of known concentration of cysteine was

prepared. The results were expressed in $\mu g\ g^{-1}$ fw. The quantification of proline was based on the spectrophotometric quantification of toluene-proline complex at 520 nm, following the method described by Bates et al. (1973), using ninhydrin. The proline content was expressed as $\mu g\ g^{-1}$ fw, after calculating its levels through linear regression with standards of proline.

2.5.4. Photosynthetic pigments

The quantification of total chlorophylls and carotenoids was performed based on Lichtenthaler (1987), using frozen shoot samples homogenized in 80% (v/v) acetone and quartz sand. The concentration of chlorophyll *a* (Chl *a*), chlorophyll *b* (Chl *b*), and carotenoids (Car) were calculated according to Lichtenthaler (1987) formulas and expressed in a fw basis.

2.5.5. Quantification of nitrogen metabolism-related enzymes

2.5.5.1. Nitrate reductase (NR; EC 1.6.6.1)

NR extraction from frozen plant samples was carried out on ice in 50 mM HEPES-KOH buffer (pH 7.8) with 1 mM phenylmethylsulfonyl fluoride (PMSF), 10 mM magnesium chloride ($MgCl_2$), and 2% (w/v) polyvinylpolypyrrolidone (PVPP), following the protocol of Kaiser and Brendle-Behnisch (1991). After a 25 min centrifugation at 4 ºC and 15 000 *g,* the supernatant (SN) was used for the quantification of proteins (as described in 2.5.3.) and NR activity. The quantification of NR activity was accomplished by monitoring the degradation of NADH and results were expressed as mmol $min^{-1}\ mg^{-1}$ of protein, using the extinction coefficient of NADH (6.22 $mM^{-1}\ cm^{-1}$).

2.5.5.2. Glutamine synthetase (GS; EC 6.3.1.2)

GS extraction and activity were done following the method of Shapiro and Stadtman (1970) by homogenizing the frozen plant material in an extraction buffer containing 25 mM Tris-HCl (pH 6.4), 10 mM $MgCl_2$, 1 mM dithiothreitol (DTT), 10% (v/v) glycerol, 0.05% (w/v) Triton-X, and 1% (w/v) PVPP. After centrifugation (20 min; 15 000 *g*; 4 ºC), the SNs were used to determine

the protein content (as described in 2.5.3.) and GS activity. The activity of GS was evaluated by the transferase assay and was expressed as nkat mg^{-1} of protein.

2.6. Statistics

Every experimental condition comprised, at least, three replicates ($n \geq 3$) and all results were expressed as mean ± standard deviation (SD). In order to test for significant differences between treatments, a one-way analysis of variance (ANOVA) was performed, followed by Dunnett's post-hoc test, whenever $p \leq 0.05$. The statistical analysis was carried out using GraphPad Prism® 7.0 software.

In order to test differences through time in the percentage of viable plants, a repeated-measures ANOVA was performed, defining the factors "within subjects" and "between subjects" as "weeks" and "concentration of the aqueous extract". Whenever significant differences were found ($p \leq 0.05$), by assuming the correction of Greenhouse-Geisser, a one-way ANOVA, followed by Dunnett's post-hoc test, was performed for each factor. This procedure was conducted in SPSS Statistics 22 (IBM SPSS®).

3.   RESULTS

3.1. Evaluation of the biocidal potential of the aqueous extracts in purslane plants - optimization of the most effective dose and extract (FLE or DLE)

The two aqueous extracts – FLE and DLE - were foliar-applied, individually and in increasing concentrations to seedlings of *P. oleracea*, in order to determine the most effective type of extract and the corresponding concentration. GLY was used as a positive CTL. After five weeks of treatments, the percentage of viable plants, macroscopic phytotoxicity symptoms, growth-related parameters, and cell death were analysed.

3.1.1. Percentage of viable plants over time

The percentage of viable plants was weekly monitored during the five weeks of treatments with
FLE (Figure 1a) or DLE (Figure 1b) (Table S1, Table S2 and Table S3). Concerning FLE, the
percentage of viable plants suffered a slight reduction of about 17% in the last week of the
experiment, when the plants were treated with FLE at 100% (v/v), compared to the CTL (Figure
1a). Regarding DLE, a decrease in the percentage of viable plants treated with the highest
concentration was observed from the third week until the end of the assay, following the same
trend of GLY, although not so evident (Figure 1b). In the last week of the assay, a reduction of
35% in the percentage of viable plants was recorded for the treatment to which DLE at 100%
(v/v) was applied to purslane plants, in comparison to the CTL plants. For the same week, the
single application of GLY induced a 75% decrease in this parameter, compared to the CTL.

Please insert Figure 1 here

3.1.2. Macroscopic phytotoxicity symptoms, biometric parameters and biomass production

As shown in Figure 2, the inhibitory effects were more pronounced with the application of DLE,
especially at 100% (v/v), which, in turn, were similar to those caused by GLY (positive CTL).
Furthermore, the plants treated with the highest concentration of DLE, only developed one new
leaf pair, with a lower leaf area, and a wilting appearance. Additionally, the application of FLE
and DLE at any of the tested concentrations induced leaf chlorosis.

Please insert Figure 2 here

Both aqueous extracts affected the growth of purslane plants, inducing different effects
depending on the applied concentration (Figure 2 and Table 1). The foliar spraying with low
doses [12.5, 25 and 50% (v/v)] of FLE enhanced shoot elongation by about 15%, 20% and 29%,
respectively, compared to the CTL. At higher concentrations [75 and 100% (v/v)], the shoot

length was only affected (decreased ca. 26%) when the extract at 100% (v/v) was applied. Likewise, the shoot biomass followed a similar trend: FLE at 25% (v/v) and 50% (v/v) induced an increase of 34% and 58%, respectively, while a reduction by about 32% was measured with the application of FLE at 100% (v/v) (Table 1). When it comes to roots, while the length was not significantly altered by the application of FLE at any of the tested concentrations, the fresh weight was significantly enhanced when *P. oleracea* plants were treated with FLE at 50% (v/v) and 75% (v/v), increasing 39% and 43%, respectively, and was impaired with the application of this extract at 100% (v/v), decreasing 31% (Table 1).

Regarding the treatment with DLE, the shoot length was enhanced by about 22% with the application of the extract at 50% (v/v) and was impaired, approximately by 17% and 36%, when the DLE was applied at 75% (v/v) and 100% (v/v), respectively (Table 1). With respect to the biomass of the aerial part, a significant reduction (ca. 52%) in response to DLE was found with the application of the extract at 75% (v/v), decreasing more than 70% with the spraying of the extract at 100% (v/v), compared to CTL (Table 1). This reduction in shoot biomass induced by the application of DLE at 100% (v/v) was comparable to the one obtained with the application of GLY, which reduced purslane plants' biomass in about 88% compared to the CTL (Table 1). In what concerns root length, DLE severely affected this endpoint, when it was applied at the two highest concentrations [75% (v/v) - decrease of 50%; 100% (v/v) - decrease of 65%, comparing to the CTL] (Table 1). Regarding root biomass, the foliar spraying of purslane plants with DLE at 75% (v/v) and 100% (v/v) caused a decrease in root biomass by 56% in both situations, in comparison to the CTL. The maximum inhibition rates for all evaluated endpoints (root length and biomass) were recorded for GLY-treated plants (Table 1). The statistical results of ANOVA analyses for shoot and root length and biomass are presented in Table S3.

Please insert Table 1 here

3.1.3. Cell viability

The histochemical detection of cell viability of *P. oleracea* plants treated with different concentrations of FLE and DLE, as well as with GLY, is represented in Figure 3 and Figure 4. In general, and compared to CTL, FLE only induced cell death when sprayed at 100% (v/v), where a pattern of cell death similar to the one obtained with GLY was found (Figure 3). In contrast, the application of DLE decreased cell viability in all tested concentrations, in a dose-dependent manner, with the highest concentration showing a similar cell death pattern to that of GLY (Figure 4).

Please insert Figure 3 here

Please insert Figure 4 here

3.2. Assessment of the phytotoxicity induced by the application of the optimized aqueous extract on purslane plants

3.2.1. Redox status

As can be seen in Table 2, MDA levels in purslane shoots were not significantly altered when the two concentrations [75 and 100% (v/v)] of DLE were applied. In contrast, the content of MDA considerably increased, approximately by 89%, in the roots of plants treated with the highest concentration of the eucalyptus extract (Table 2), compared to the untreated situation.

Concerning H_2O_2, the levels of this reactive oxygen species (ROS) suffered a 3- and 5-fold increase with the spraying of the highest concentration [100% (v/v)] of DLE in shoots and roots, respectively, compared to CTL (Table 2). The ANOVA statistical results of MDA and H_2O_2 levels are denoted in Table S4.

Please insert Table 2 here

3.2.2. Total sugars, proteins, free amino acids and proline content

Regarding the total sugar levels, the treatment with the highest concentration of DLE increased this parameter by 3 times in the shoots and roots of purslane plants, compared to the CTL situation (Table 2 and Table S4).

Total soluble protein levels were substantially increased (about 89%) in the aerial part of purslane plants, with the application of DLE at 100% (v/v) (Table 2 and Table S4). In contrast, in the roots, the same concentration of DLE considerably decreased this parameter by about 50%, in relation to the CTL situation (Table 2 and Table S4).

As can be seen in Table 2 and Table S4, total free amino acid levels in *P. oleracea* shoots were reduced (26% over the CTL) only when the 75% (v/v) DLE was sprayed. Regarding the roots, the application of the highest concentration of the extract significantly increased, by approximately 59%, the levels of total free amino acids (Table 2 and Table S4). With regard to proline levels, there was a significant increase of about 37% in the shoots of purslane plants treated with DLE at the highest concentration, compared to the CTL situation (Table 2 and Table S4).

3.2.3. Levels of photosynthetic pigments

As shown in Table 2, total chlorophylls (*a* and *b*) were significantly impaired with the foliar spraying of the extract at 75% (v/v), decreasing the levels of these pigments by about 28%, compared to the untreated situation (Table S4). However, while the treatment with the highest concentration of DLE did not alter total chlorophyll levels, carotenoid production increased 19% when this dose was applied to purslane plants, compared to CTL (Table 2 and Table S4).

3.2.4. Nitrogen (N) metabolism

The impact of DLE application on N nutrition of purslane plants was evaluated through the quantification of two enzymes belonging to N metabolism, NR and GS, in the aerial part and in the roots of *P. oleracea* plants. As shown in Table 2 and Table S4, NR activity significantly

increased in the shoots of purslane plants treated with 75% (v/v) DLE, with an increase of 58% over the CTL, while in the roots, the NR activity did not change with the treatments (Table 2). With regard to GS, the results revealed that although the activity of this enzyme was not altered in the aerial part of the plants (Table 2 and Table S4), this enzyme was positively affected (67%) in the roots by the highest applied concentration of DLE, in relation to the CTL situation (Table 2 and Table S4).

4. DISCUSSION

The herbicidal potential of *E. globulus* leaf extracts has been explored by some authors (Ataollahi et al., 2014; Babu and Kandasamy, 1997; Dejam et al., 2014; Morsi and Abdelmigid, 2016; Padhy et al., 2000; Puig et al., 2018). However, most of these studies focused on the pre-emergent activity of *E. globulus* extracts, evaluating their effects only on seed germination and seedling growth. From all these studies, only Puig et al. (2018) evaluated the biocidal potential of an aqueous extract prepared with fresh leaves in a post-emergent context. In that study, the authors used leaves from mature trees to prepare the extract and concluded that it possessed a biocidal action when daily-applied, by foliar spraying or watering, for 3 weeks on *Lactuca sativa* L., an agronomically relevant species. In this sense, in the present study, the biocidal activity of the extracts prepared with the leaves obtained from young *E. globulus* trees was evaluated in a post-emergent context by foliar spraying, twice a week, 7-days-old purslane seedlings with several dilutions of FLE (617 g_{fw} L^{-1}) and DLE (250 g_{dw} L^{-1}).

4.1. DLE shows a high biocidal potential, impairing weed growth and compromising cell viability

The application of FLE at low concentrations, especially at 50% (v/v), presented curious fertilizing properties, enhancing plant growth, namely shoot elongation and biomass production. As expected, the treatment with 50% (v/v) FLE did not alter the percentage of viable plants over the five weeks of the experiment, and did not induce an increase of cell death in purslane plants.

Altogether, these results show that, at least up to 50% (v/v), the FLE did not have toxic effects on purslane plants. In fact, the allelopathic activity of eucalyptus leaves against other plants is only effective when the allelochemicals are present at certain concentrations (Dahiya et al., 2017), which were not achieved with the use of the diluted formulation.

Nevertheless, at higher concentrations [75 and 100% (v/v)], FLE has been shown to negatively affect plant growth and leaf area. Particularly when applied at its maximum dose (617 g_{fw} L^{-1}), FLE significantly reduced the shoot length and the fresh weight of both shoots and roots. In addition, although the application of FLE at the highest concentration has only induced a significant reduction in the percentage of viable plants after 5 weeks, purslane plants treated with this concentration presented a pattern of cell death very similar to that obtained for treatment with GLY, the most widely used chemical herbicide in a post-emergence context (Duke and Powles, 2008). Altogether these results evidence that the FLE at 100% (v/v) had an effective herbicidal activity on purslane plants, although with a slower action than the synthetic herbicide.

Regarding DLE, when applied at concentrations up to 50% (v/v), it did not induce relevant alterations in *P. oleracea* plants, as the FLE at the same percentages. Indeed, even though a significant increase in the shoot length was verified with the treatment of DLE at 50% (v/v), all the other endpoints remained unchanged. This may be due to the drying process of the leaves, which, on the one hand, may have induced the loss of volatile compounds and, on the other, may have caused the accumulation of greater amounts of compounds with allelopathic properties, such as hydrolysable tannins (Salminen, 2003).

In opposition, the treatment of purslane plants with the highest concentrations of DLE [75 and 100% (v/v)] caused a great reduction in most of the tested parameters, which evidences that DLE, at these concentrations, detained a strong and effective biocidal activity. As a matter of fact, the treatment with DLE at 75% (v/v) and 100% (v/v) markedly compromised cell viability in purslane leaves. Nevertheless, DLE at 100% (v/v) was the only one capable of effectively reducing the percentage of viable plants in the time frame of the experiment. Moreover, this concentration exhibited greater inhibitory effects on most of the growth-related parameters,

generally affecting the root system more than the aerial part of purslane plants, as will be discussed later. Accordingly, the treatment with DLE at 100% (v/v) severely impaired the growth of *P. oleracea* plants, compared to FLE at its maximum concentration.

Despite the observed differences between FLE and DLE, the application of both eucalyptus-based herbicides induced leaf chlorosis. The explanation for this observation may reside in the mode-of-action of these biocides. Plant-based herbicides, like eucalyptus extracts, can affect weed growth by disrupting nutrient uptake, photosynthetic process, and membrane permeability (Radhakrishnan et al., 2018). Briefly, the allelochemicals present in plant herbicides can alter the structure and function of cell membranes, decreasing the absorption of macro and micronutrients, such as magnesium (Mg), potassium (K), and iron (Fe) (Radhakrishnan et al., 2018). Therefore, the observed chlorosis in purslane leaves treated with FLE and DLE may result in the reduction of Mg, K, and Fe intracellular levels, whose presence is essential for the normal function of chloroplasts (Taiz et al., 2015). In fact, while Mg is present at the active centre of the chlorophyll molecule, Fe composes many cytochromes and nonheme iron proteins involved in the photosynthetic metabolism, and K is crucial for maintaining cell electroneutrality and participates in a series of photosynthetic reactions (Taiz et al., 2015). Accordingly, a reduction in the levels of total chlorophylls was detected in the plants treated with DLE at 75% (v/v). Although not statistically significant, probably due to the large decrease in the percentage of viable plants, the content of chlorophylls *a* and *b* in plants treated with the most effective concentration of DLE, presented a downward trend. Indeed, an analogous study in which an aqueous extract prepared with fresh *E. globulus* leaves was applied to lettuce plants reported that the application of the extract induced a reduction in the total chlorophyll content that was accompanied by a decline in chlorophyll *a* fluorescence (Puig et al., 2018). These results allowed the authors to conclude that the light-independent reactions of photosynthesis were directly affected by the application of the extract (Puig et al., 2018).

Moreover, besides chlorosis, DLE at 100% (v/v) caused foliar wilting, a symptom exclusively found in this treatment. This macroscopic symptom is usually associated to a loss of membrane integrity (Radhakrishnan et al., 2018). Indeed, a study conducted with the purpose of

assessing the herbicidal potential of *E. citriodora* oil on *Phalaris minor* Retz. reported the occurrence of wilting and chlorotic leaves in the treated plants, which was the result of a high degree of ion leakage induced by the biocide application, causing the loss of membrane permeability and integrity (Batish et al., 2007). In accordance, the intracellular H_2O_2 levels were greatly increased in the shoots of the purslane plants treated with the highest concentration of DLE, as will be discussed later.

Overall, the herbicidal activity shown by DLE at 100% (v/v) was comparable to that of GLY although with an apparently slower action, but much faster and effective than that of FLE. In fact, the time lag to have an effect from DLE and FLE applications may depend mainly on the time needed to reach an effective dose, with the successive applications, rather than on the mode-of-action of the bioherbicides. Indeed, the reduction of the growth-related parameters and the pattern of cell death of the plants treated with DLE at 100% (v/v) were identical to the ones obtained when the synthetic herbicide was applied. In addition, the onset of the herbicidal effects of DLE was registered with only one week apart from that of GLY, as demonstrated by the results of the percentage of viable plants over the 5 weeks. Thus, from all the tested extracts and concentrations, it was possible to conclude that the DLE at 100% (v/v) had the greatest herbicidal activity against *P. oleracea* plants.

4.2. The biocidal potential of DLE is linked to an overproduction of H_2O_2, followed by a general deregulation of different cellular biomarkers

To be effective, herbicides must interfere with some weed physiological processes, disturbing certain cellular/metabolic processes, or inhibiting the biosynthesis of cellular/molecular components by blocking a specific step of a metabolic pathway (Sherwani et al., 2015). Consequently, irreparable damages and tissue injuries are induced in weeds, ultimately leading to cell death and thus, their elimination (Sherwani et al., 2015). With the purpose of identifying some of the physiological mechanisms potentially hindered by the application of DLE in

purslane plants, several endpoints related with many aspects of plant physiology were analysed in the weeds treated with the two most effective concentrations of DLE [75 and 100% (v/v)].

The induction of oxidative stress seems to be one of the commonest key factors behind herbicide-induced metabolic impairments (Caverzan et al., 2019). Often, an overproduction of ROS takes place, followed by the occurrence of oxidative damage, translated into the oxidation of the main biomolecules, from lipids and sugars, to proteins and nucleic acids (Kaur, 2019). Thus, the first step onto dissecting the effects of DLE in weed physiology was the evaluation of the redox homeostasis in both shoots and roots of the treated plants. The obtained results showed different responses for the shoots and roots of purslane plants exposed to the DLE treatment. In the shoots, the application of DLE at 100% (v/v) significantly increased the cell H_2O_2 levels. In fact, ROS overproduction generally occurs in response to adverse conditions, further leading to oxidative stress, when the antioxidant (AOX) system is inhibited and/or unable to eliminate the ROS in excess (Soares et al., 2019a). In this sense, the levels of MDA, a molecule resulting from the peroxidation of membrane lipids (Heath and Packer, 1968), which is commonly used as an indicator of oxidative stress occurrence, were also quantified. Unexpectedly, and contrasting to the ROS levels, the shoot MDA content was not altered in any treatment. Although it may seem a little surprising, this can be the result of the registered increase in the levels of proline and carotenoids in this plant organ. Indeed, besides their functions as osmoprotectant and accessory photosynthetic pigment, respectively, the ability of these AOX to scavenge ROS and to stabilize biological membranes is becoming widely recognized, being now considered as potent inhibitors of LP (Gill and Tuteja, 2010; Szabados and Savouré, 2010). Therefore, the unchanged MDA levels in shoots of purslane plants treated with DLE was most probably due to the increase in proline and carotenoid levels, along with the possible stimulation of other defense-related strategies. Accordingly, sugar and total soluble protein levels were enhanced in purslane plants exposed to DLE at 100% (v/v). Alongside with proline, sugars, such as glucose and sucrose, have important AOX roles either by directly reacting with ROS, forming less harmful products, or by inducing the transcription of stress-related genes, which results in the synthesis of important AOX players (Soares et al., 2019a). In

fact, the increase of the protein content observed when DLE at 100% (v/v) was applied can be indicative of the activation of gene expression, possibly resulting in the production of defense-related proteins. Indeed, it is known that under stress conditions, plants have altered gene expression, which results in the increase of the content of diverse proteins and metabolites (Ahmad et al., 2010). Altogether, the obtained results highlight the occurrence of several redox disorders in shoots of DLE-exposed purslane plants, favoring the accumulation of H_2O_2 and the synthesis of different AOX metabolites, which prevented LP and thus, the occurrence of a severe oxidative stress condition.

With regard to the roots, the same pattern of H_2O_2 accumulation as in the aerial part was found. In addition, and contrary to what was observed for shoots, the increase in H_2O_2 levels in the root may have exceeded the AOX system´s capacity to eliminate the excess of ROS, and induced the occurrence of oxidative stress, as demonstrated by the significant increase of MDA levels at the highest concentration. Thus, it appears that the significant increase of total sugars and free amino acids in the roots of purslane plants was not enough to counteract DLE-induced oxidative stress. A redox imbalance characterized by the overproduction of ROS can produce irreversible damages in plant cells, such as the oxidation of proteins, lipids, and even, nucleic acids (Soares et al., 2019a). Besides LP, the oxidation of proteins of purslane roots might have also occurred as a consequence of the oxidative stress imposed by the application of DLE at 100% (v/v), since protein levels significantly decreased in this organ. In fact, it has been reported that the application of plant-based extracts to weeds enhances the production of the superoxide radical ($O_2^{\cdot-}$) and H_2O_2, resulting in membrane and protein damages (Radhakrishnan et al., 2018). In addition, the application of these biocides induces an electrolytic leakage that, among others, release considerable amounts of proteases (Radhakrishnan et al., 2018). This protease release, along with the protein oxidation induced by the high concentrations of ROS, namely H_2O_2, might explain the great decrease in protein levels registered in *P. oleracea* roots.

In addition to oxidative stress, nutrient deficiencies or imbalances are one of the main causes that compromise plant growth (Rietra et al., 2017). Nitrogen is an essential macronutrient for plant growth and development since it is involved in the synthesis of multiple

organic compounds, such as amino acids and proteins (Taiz et al., 2015). Thus, disturbances in N nutrition are responsible for significant reductions in plant growth (Liu et al., 2017). The results of our study revealed that the foliar spraying of purslane plants with the highest concentration of DLE did not affect NO_3^- assimilation in both organs, since the activity of NR was not altered. Nevertheless, a great increase in the activity of GS in the roots of *P. oleracea* plants treated with the maximum concentration of DLE was found. As mentioned before, the GS product, glutamine, is an important precursor for the biosynthesis of other amino acids and chlorophylls, among others (Németh et al., 2018). In this sense, the increase in GS activity in the roots of the plants treated with DLE at 100% (v/v) might be responsible for the increase in the amino acid content registered in plants exposed to the same treatment, possibly in the attempt of restoring the cell protein levels that were substantially affected by the oxidative burst induced by DLE application.

Overall, the obtained results pointed out that the DLE at 100% (v/v) was able to induce severe phytotoxic effects in purslane plants, considerably impairing multiple weed physiological processes. Although the DLE at 100% (v/v) was applied by foliar spraying, the phytotoxic effects in the roots of purslane plants were more pronounced than in the shoots. This may have occurred because, after 20 sprays, the extract ended up accumulating in the soil, being absorbed by the plant´s root system. As a matter of fact, since the application of DLE at the maximum concentration induced great inhibitory effects in the growth and development of purslane plants, their root system was unable to fully develop, compromising the function of this organ. In these conditions, the allelochemicals can easily penetrate through the primary root structure (Radhakrishnan et al., 2018), causing more severe damage in the physiology of this organ than in the aerial part. In this way, the effects of DLE at 100% (v/v) in the shoots may have occurred due to the serious phytotoxic effects induced in the root system by the uptake of compounds with allelopathic activity and/or the interference of the allelochemicals with the absorption of nutrients by the roots, affecting several metabolic processes (Radhakrishnan et al., 2018).

5. CONCLUSIONS

The foliar spraying of aqueous extracts prepared with young leaves of *E. globulus*, especially when dried, revealed to have strong biocidal properties against *P. oleracea* weeds, mainly by severely disturbing the redox homeostasis of their root system. This represent the first study to unravel the herbicidal potential of the leaf biomass from young *E. globulus* trees, recently resprouted after a wildfire, which can contribute, on one hand, to decrease fire hazard by reducing the uncontrolled dispersion of this species, and on the other, to possibly increase crop production by eliminating weeds.

In the future, we intend to explore the biocidal potential of the leaves of young *E. globulus* trees in a pre-emergent scenario, and to assess their environmental safety, when applied in pre- and post-emergent contexts, in non-target organisms, such as crop species.

Supplementary Materials:

Table S1: Summary of repeated measures' ANOVA statistical data for the percentage of viable plants (% viable plants) treated with the extracts prepared with fresh leaves (FLE) and with dried leaves (DLE), **Table S2:** Summary of one-way ANOVA statistical data for the percentage of viable plants (% viable plants) for each tested concentration of the extracts prepared with fresh leaves (FLE) and with dried leaves (DLE), as well as glyphosate (GLY), over the exposure period (5 weeks), **Table S3:** Summary of ANOVA statistical data for purslane plants treated with increasing concentrations of the extracts prepared with fresh leaves (FLE) and with dried leaves (DLE), **Table S4:** Summary of ANOVA statistical data obtained for the shoots and roots of purslane plants treated with the extract prepared with dried leaves (DLE) at 75% (v/v) and 100% (v/v).

Author Contributions:

MP designed and performed the experiments, analyzed the results and wrote the manuscript. CS designed and carried out the experiments and contributed to data analysis and manuscript writing. MM performed the experiments and manuscript revision. BS carried out the

experiments and revised the manuscript. IV contributed to experimental design and manuscript revision. RP provided funding, conceptualization of the project, planned and recommended the appropriate statistical analysis and revised the manuscript. FF granted funding, conceptualization, and contributed to experimental design and manuscript revision.

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Conflicts of Interest:

The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

References

Águas, A., Ferreira, A., Maia, P., Fernandes, P.M., Roxo, L., Keizer, J., Silva, J.S., Rego, F.C., Moreira, F., 2014. Natural establishment of *Eucalyptus globulus* Labill. in burnt stands in Portugal. Forest Ecology and Management 323, 47-56.

Ahmad, P., Jaleel, C.A., Salem, M.A., Nabi, G., Sharma, S., 2010. Roles of enzymatic and nonenzymatic antioxidants in plants during abiotic stress. Critical Reviews in Biotechnology 30, 161-175.

Aktar, M.W., Sengupta, D., Chowdhury, A., 2009. Impact of pesticides use in agriculture: their benefits and hazards. Interdisciplinary Toxicology 2, 1-12.

Alexieva, V., Sergiev, I., Mapelli, S., Karanov, E., 2001. The effect of drought and ultraviolet radiation on growth and stress markers in pea and wheat. Plant, Cell and Environment 24, 1337-1344.

Aragão, F., Palmieri, M., Ferreira, A., Costa, A., Queiroz, V., Pinheiro, P., Andrade-Vieira, L., 2015. Phytotoxic and cytotoxic effects of *Eucalyptus* essential oil on lettuce (*Lactuca sativa* L.). Allelopathy Journal 35, 259-272.

Ataollahi, R., Dejam, M., Khaleghi, S.S., 2014. Phytotoxic effects of *Eucalyptus globulus* leaf extract on *Solanum nigrum*. South-Western Journal of Horticulture Biology and Environment 5, 43-53.

Babu, R.C., Kandasamy, O., 1997. Allelopathic effect of *Eucalyptus globulus* Labill. on *Cyperus rotundus* L. and *Cynodon dactylon* L. Pers. Journal of Agronomy and Crop Science 179, 123-126.

Bagavathy, S., Xavier, G., 2007. Effects of aqueous extract of *Eucalyptus globules* on germination and seedling growth of sorghum. Allelopathy 20, 395–402.

Bates, L.S., Waldren, R.P., Teare, I., 1973. Rapid determination of free proline for water-stress studies. Plant and Soil 39, 205-207.

Batish, D.R., Singh, H.P., Setia, N., Kohli, R.K., Kaur, S., Yadav, S.S., 2007. Alternative control of littleseed canary grass using eucalypt oil. Agronomy for Sustainable Development 27, 171-177.

Baucom, R.S., 2019. Evolutionary and ecological insights from herbicide-resistant weeds: what have we learned about plant adaptation, and what is left to uncover? New Phytologist 223, 68-82.

Benchaa, S., Hazzit, M., Abdelkrim, H., 2018. Allelopathic effect of *Eucalyptus citriodora* essential oil and its potential use as bioherbicide. Chemistry and Biodiversity 15, e1800202.

Bradford, M.M., 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Analytical Biochemistry 72, 248-254.

Burrows, G.E., 2002. Epicormic strand structure in *Angophora, Eucalyptus* and *Lophostemon* (Myrtaceae) – implications for fire resistance and recovery. New Phytologist 153, 111-131.

Burrows, G.E., 2013. Buds, bushfires and resprouting in the eucalypts. Australian Journal of Botany 61, 331-349.

Catry, F.X., Moreira, F., Tujeira, R., Silva, J.S., 2013. Post-fire survival and regeneration of *Eucalyptus globulus* in forest plantations in Portugal. Forest Ecology and Management 310, 194-203.

Caverzan, A., Piasecki, C., Chavarria, G., Stewart, C.N., Vargas, L., 2019. Defenses against ROS in crops and weeds: the effects of interference and herbicides. International Journal of Molecular Sciences 20, 1086.

Chaturvedi, S., Pandey, J., Dhyani, V., Guru, S., Kaushal, R., 2012. Phytotoxic potential of *Eucalyptus* leaf essential oil to control *Parthenium hysterophorus* L. Allelopathy Journal 29, 315-324.

Dadkhah, A., 2013. Allelopathic effect of sugar beet (*Beta vulgaris*) and eucalyptus (*Eucalyptus camaldulensis*) on seed germination and growth of *Portulaca oleracea*. Russian Agricultural Sciences 39, 117-123.

Dadkhah, A., Asaadi, A.M., 2010. Allelopathic effects of *Eucalyptus camaldulensis* on seed germination and growth seedlings of *Acroptilon repens*, *Plantago lanceolata* and *Portulaca oleracea*. Journal of Biological Sciences 5, 430-434.

Dahiya, S., Kumar, S., Khedwal, R.S., Jakhar, S., 2017. Allelopathy for sustainable weed management. Journal of Pharmacognosy and Phytochemistry 6, 832-837.

Dejam, M., Khaleghi, S.S., Ataollahi, R., 2014. Allelopathic effects of *Eucalyptus globulus* Labill. on seed germination and seedling growth of eggplant (*Solanum melongena* L.). International Journal of Farming and Allied Sciences 3, 81-86.

Dilworth, L., Riley, C., Stennett, D., 2017. Plant Constituents: Carbohydrates, Oils, Resins, Balsams, and Plant Hormones, Pharmacognosy. Elsevier, pp. 61-80.

Duke, S.O., Powles, S.B., 2008. Glyphosate: a once-in-a-century herbicide. Pest Management Science 64, 319-325.

El-Ghit, H.M.A., Hanan, M., 2016. Potent physiological allelopathic effect of eucalyptus leaf extract on *Malva parviflora* L (mallow) weed. Journal of Pharmaceutical, Chemical and Biological Sciences 3, 584-591.

El-Rokiek, K., Eid, R., 2009. Allelopathic effects of *Eucalyptus citriodora* on amaryllis and associated grassy weed. Planta Daninha 27, 887-899.

Geetha, A., 2019. Phytotoxicity due to fungicides and herbicides and its impact in crop physiological factors, Advances in Agriculture Sciences. AkiNik Publications, pp. 29-66.

Gill, S.S., Tuteja, N., 2010. Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. Plant Physiology and Biochemistry 48, 909-930.

Heath, R.L., Packer, L., 1968. Photoperoxidation in isolated chloroplasts: I. Kinetics and stoichiometry of fatty acid peroxidation. Archives of Biochemistry and Biophysics 125, 189-198.

Hernandez, A.F., Parron, T., Tsatsakis, A.M., Requena, M., Alarcon, R., Lopez-Guarnido, O., 2013. Toxic effects of pesticide mixtures at a molecular level: their relevance to human health. Toxicology 307, 136-145.

Irigoyen, J., Einerich, D., Sánchez-Díaz, M., 1992. Water stress induced changes in concentrations of proline and total soluble sugars in nodulated alfalfa (*Medicago sativa*) plants. Physiologia Plantarum 84, 55-60.

Jaime, M.D.I., Ferrer, M.A.B., 2018. Post-emergent herbicidal activity of *Eucalyptus globulus* Labill. essential oil. Nereis: Revista Iberoamericana Interdisciplinar de Métodos, Modelización y Simulación, 25-36.

James, S.A., Bell, D.T., 2001. Leaf morphological and anatomical characteristics of heteroblastic *Eucalyptus globulus* ssp. *globulus* (Myrtaceae). Australian Journal of Botany 49, 259-269.

Kaiser, W.M., Brendle-Behnisch, E., 1991. Rapid modulation of spinach leaf nitrate reductase activity by photosynthesis: I. Modulation *in vivo* by CO_2 availability. Plant Physiology 96, 363-367.

Kaur, G., 2019. Herbicides and its role in induction of oxidative stress- a review. International Journal of Environment, Agriculture and Biotechnology 4, 995-1004.

Lee, Y.P., Takahashi, T., 1966. An improved colorimetric determination of amino acids with the use of ninhydrin. Analytical Biochemistry 14, 71-77.

Lichtenthaler, H.K., 1987. Chlorophylls and carotenoids: pigments of photosynthetic biomembranes, Methods in Enzymology. Elsevier, pp. 350-382.

Liu, M., Sun, J., Li, Y., Xiao, Y., 2017. Nitrogen fertilizer enhances growth and nutrient uptake of *Medicago sativa* inoculated with *Glomus tortuosum* grown in Cd-contaminated acidic soil. Chemosphere 167, 204-211.

Morsi, M.M., Abdelmigid, H.M., 2016. Allelopathic activity of *Eucalyptus globulus* leaf aqueous extract on *Hordeum vulgare* growth and cytogenetic behaviour. Australian Journal of Crop Science 10, 1551-1556.

Németh, E., Nagy, Z., Pécsváradi, A., 2018. Chloroplast glutamine synthetase, the key regulator of nitrogen metabolism in wheat, performs its role by fine regulation of enzyme activity via negative cooperativity of its subunits. Frontiers in Plant Science 9.

O'Reilly-Wapstra, J.M., Humphreys, J.R., Potts, B.M., 2007. Stability of genetic-based defensive chemistry across life stages in a *Eucalyptus* species. Journal of Chemical Ecology 33, 1876-1884.

OECD, 2006. Terrestrial plant test: seedling emergence and seedling growth test. OECD-Organization for Economic Cooperation and Development, Paris 208, 6.

Padhy, B., Patnaik, P., Tripathy, A., 2000. Allelopathic potential of *Eucalyptus* leaf litter leachates on germination and seedling growth of fingermillet. Allelopathy Journal 7, 69-78.

Pinto, M., 2020. Assessing the effectiveness and environmental safety of young eucalyptus leaf biomass as a biocide. MSc Dissertation. Biology Department. Faculty of Sciences of the University of Porto.

Puig, C.G., Reigosa, M.J., Valentao, P., Andrade, P.B., Pedrol, N., 2018. Unravelling the bioherbicide potential of *Eucalyptus globulus* Labill: Biochemistry and effects of its aqueous extract. PLoS One 13, e0192872.

Radhakrishnan, R., Alqarawi, A.A., Abd_Allah, E.F., 2018. Bioherbicides: Current knowledge on weed control mechanism. Ecotoxicology and Environmental Safety 158, 131-138.

Rassaeifar, M., Hosseini, N., Asl, N.H.H., Zandi, P., Aghdam, A.M., 2013. Allelopathic effect of *Eucalyptus globulus* essential oil on seed germination and seedling establishment of *Amaranthus blitoides* and *Cyndon dactylon*. Trakia Journal of Sciences 11, 73-81.

Reigosa, M., Gonzalezy, L., Souto, X., Pastoriza, J., 2000. Allelopathy in forest ecosystems, Allelopathy in Ecological Agriculture and Forestry. Springer, pp. 183-193.

Rietra, R.P.J.J., Heinen, M., Dimkpa, C.O., Bindraban, P.S., 2017. Effects of nutrient antagonism and synergism on yield and fertilizer use efficiency. Communications in Soil Science and Plant Analysis 48, 1895-1920.

Romero-Puertas, M., Rodríguez-Serrano, M., Corpas, F., Gomez, M.d., Del Rio, L., Sandalio, L., 2004. Cadmium-induced subcellular accumulation of $O_2^{\cdot-}$ and H_2O_2 in pea leaves. Plant, Cell and Environment 27, 1122-1134.

Salminen, J.-P., 2003. Effects of sample drying and storage, and choice of extraction solvent and analysis method on the yield of birch leaf hydrolyzable tannins. Journal of Chemical Ecology 29, 1289-1305.

Shahena, S., Rajan, M., Chandran, V., Mathew, L., 2020. Controlled release herbicides and allelopathy as sustainable alternatives in crop production, Controlled Release of Pesticides for Sustainable Agriculture. Springer, pp. 237-252.

Shapiro, B.M., Stadtman, E., 1970. Glutamine synthetase (*Escherichia coli*), Methods in Enzymology. Elsevier, pp. 910-922.

Sherwani, S.I., Arif, I.A., Khan, H.A., 2015. Modes of action of different classes of herbicides, in: Price, A., Kelton, J., Sarunaite, L. (Eds.), Herbicides, Physiology of Action, and Safety. IntechOpen, pp. 165-186.

Soares, C., Carvalho, M.E., Azevedo, R.A., Fidalgo, F., 2019a. Plants facing oxidative challenges—A little help from the antioxidant networks. Environmental and Experimental Botany 161, 4-25.

Soares, C., Pereira, R., Spormann, S., Fidalgo, F., 2019b. Is soil contamination by a glyphosate commercial formulation truly harmless to non-target plants? - Evaluation of oxidative damage and antioxidant responses in tomato. Environmental Pollution 247, 256-265.

Szabados, L., Savouré, A., 2010. Proline: a multifunctional amino acid. Trends in Plant Science 15, 89-97.

Taiz, L., Zeiger, E., Møller, I.M., Murphy, A., 2015. Plant physiology and development, Twelfth edition ed, USA.

Verdeguer Sancho, M.M., Blazquez, M., Boira Tortajada, H., 2018. Phytotoxic potential of *Lantana camara, Eucalyptus camaldulensis, Eriocephalus africanus, Cistus ladanifer* and *Artemisia gallica* aqueous extracts to control weeds. Journal of Allelochemical Interactions 4, 17-26.

Figures and tables' captions

Figure 1. Percentage of viable plants over the five weeks of treatments with deionised water (CTL), glyphosate (GLY), and different concentrations [12.5, 25, 50, 75 and 100% (v/v)] of the aqueous extract prepared with fresh leaves (FLE; 617 g_{fw} L^{-1}) (**a**) and with dried leaves (DLE; 250 g_{dw} L^{-1}) (**b**). Results are expressed as mean ± SD (n ≥ 3). º refers to statistically significant differences ($p \leq 0.05$) between treatments and the control (CTL), for the last week; * refers to statistically significant differences ($p \leq 0.05$) between weeks, for each treatment and the CTL.

Figure 2. Macroscopic phytotoxicity symptoms of purslane plants after five weeks of treatments with deionised water (CTL), glyphosate (GLY), and different concentrations [12.5, 25, 50, 75 and 100% (v/v)] of the aqueous extract prepared with fresh leaves (FLE; 617 g_{fw} L^{-1}) and with dried leaves (DLE; 250 g_{dw} L^{-1}).

Figure 3. Cell death staining in shoots of purslane plants treated for five weeks with deionised water (CTL), glyphosate (GLY), and different concentrations [12.5, 25, 50, 75 and 100% (v/v)] of the aqueous extract prepared with fresh leaves (FLE; 617 g_{fw} L^{-1}).

Figure 4. Cell death staining in shoots of purslane plants treated for five weeks with deionised water (CTL), glyphosate (GLY), and different concentrations [12.5, 25, 50, 75 and 100% (v/v)] of the aqueous extract prepared with dried leaves (DLE; 250 g_{dw} L^{-1}).

Table 1. Shoot and root length and fresh weight of the purslane plants treated for five weeks with deionised water (CTL), glyphosate (GLY), and different concentrations [12.5, 25, 50, 75

and 100% (v/v)] of the aqueous extract prepared with fresh leaves (FLE; 617 g_{fw} L^{-1}) and with dried leaves (DLE; 250 g_{dw} L^{-1}). Results are expressed as mean ± SD (n ≥ 3). * above bars denotes significant differences ($p \leq 0.05$) from the CTL situation.

Table 2. Levels of MDA, H_2O_2, total sugars, total free amino acids, proline, total soluble proteins, photosynthetic pigments (chlorophylls and carotenoids) and of the activity of NR and GS in the shoots and roots of purslane plants treated for five weeks with deionised water (CTL), 75% (v/v) and 100% (v/v) of the aqueous extract prepared with dried leaves (DLE; 250 g_{dw} L^{-1}). Results are expressed as mean ± SD (n ≥ 3). * above bars denotes significant differences ($p \leq$ 0.05) from the CTL situation.

Figure 1

(a)   (b)
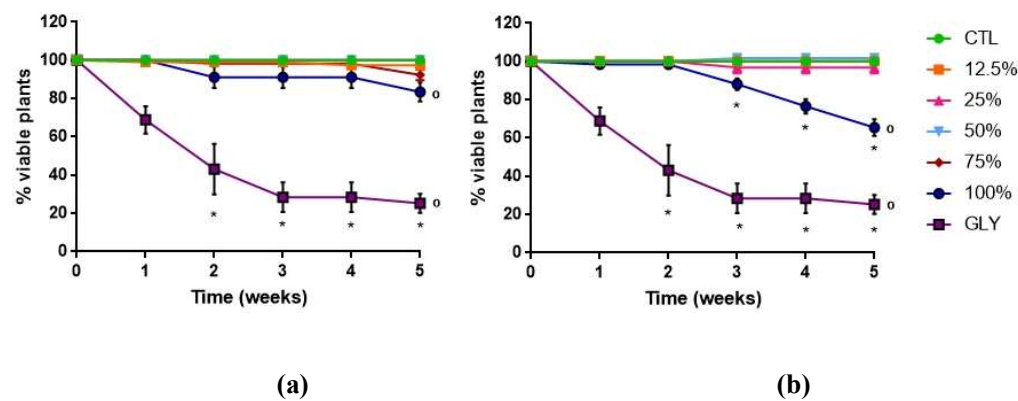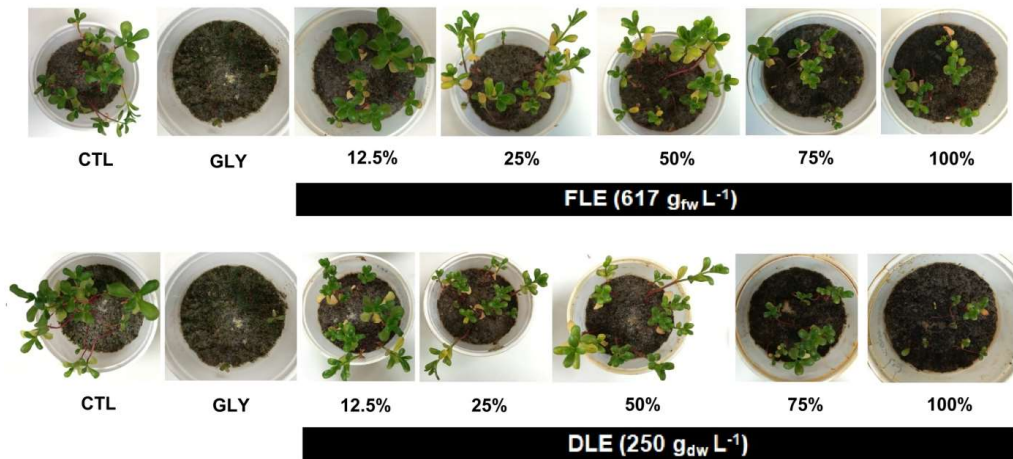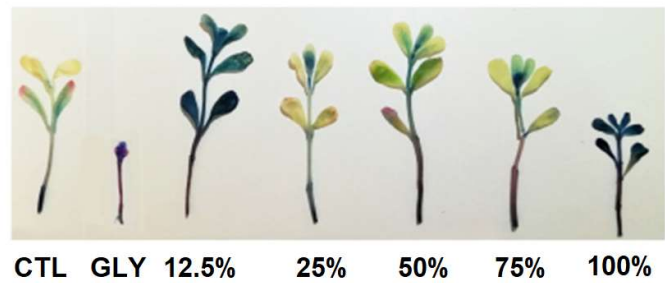
Figure 2

Figure 3

Figure 4
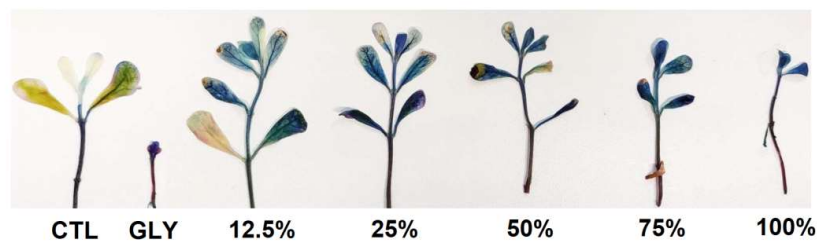
Table 1

Endpoint Treatment		Length (cm)		Fresh weight (g)	
		Shoots	**Roots**	**Shoots**	**Roots**
FLE	**CTL**	7.37 ± 0.30	6.51 ± 0.22	0.26 ± 0.014	0.020 ± 0.0034
	12.5%	8.47 ± 0.13*	6.13 ± 0.56	0.25 ± 0.026	0.019 ± 0.0028
	25%	8.85 ± 0.45*	5.92 ± 0.78	0.34 ± 0.047*	0.015 ± 0.00023
	50%	9.52 ± 0.44*	7.27 ± 0.15	0.41 ± 0.0069*	0.028 ± 0.0016*
	75%	6.88 ± 0.09	6.43 ± 0.59	0.28 ± 0.0039	0.029 ± 0.0011*
	100%	5.47 ± 0.44*	6.12 ± 0.42	0.18 ± 0.035*	0.014 ± 0.0047*
DLE	**CTL**	7.31 ± 0.73	7.11 ± 1.01	0.27 ± 0.024	0.025 ± 0.0046
	12.5%	7.72 ± 0.52	8.02 ± 0.49	0.25 ± 0.010	0.020 ± 0.0027
	25%	7.33 ± 0.47	7.20 ± 0.59	0.25 ± 0.043	0.022 ± 0.0047
	50%	8.94 ± 0.77*	6.83 ± 0.15	0.26 ± 0.025	0.027 ± 0.0031
	75%	6.10 ± 0.051*	3.58 ± 0.43*	0.13 ± 0.013*	0.011 ± 0.0017*
	100%	4.65 ± 0.60*	2.46 ± 0.46*	0.079 ± 0.017*	0.011 ± 0.0001*
GLY		2.65 ± 0.11*	0.79 ± 0.35*	0.037 ± 0.0074*	0.003 ± 0.00088*

Table 2

Treatment / Endpoint	CTL		DLE			
			75% (v/v)		100% (v/v)	
	Shoots	Roots	Shoots	Roots	Shoots	Roots
MDA (nmol g^{-1} fw)	23.3 ± 3.4	3.24 ± 1.1	18.5 ± 3.7	1.53 ± 0.51	19.6 ± 1.1	6.11 ± 1.3*
H_2O_2 (pmol g^{-1} fw)	13.9 ± 1.3	6.79 ± 0.81	21.8 ± 4.0	10.7 ± 2.5	43.5 ± 9.9*	34.4 ± 3.3*
Total sugars (µg g^{-1} fw)	1.32 ± 0.16	4.73 ± 2.1	2.00 ± 0.01	7.37 ± 0.61	3.96 ± 0.81*	12.2 ± 1.7*
Total free amino acids (µg g^{-1} fw)	262 ± 11	343 ± 36	195 ± 14*	381 ± 30	247 ± 2.1	544 ± 83*
Proline (µg g^{-1} fw)	37.1 ± 2.4	-	48.3 ± 6.6	-	59.0 ± 15*	-
Total soluble proteins (mg g^{-1} fw)	1.21 ± 0.14	1.23 ± 0.12	1.40 ± 0.049	1.27 ± 0.27	2.29 ± 0.35*	0.61 ± 0.099*
Total chlorophylls (mg g^{-1} fw)	0.315 ± 0.013	-	0.205 ± 0.05*	-	0.278 ± 0.033	-
Carotenoids (mg g^{-1} fw)	0.0697 ± 0.0012	-	0.066 ± 0.0066	-	0.082 ± 002*	-
NR activity (mmol min^{-1} mg^{-1} protein)	110 ± 12	96.4 ± 15	175 ± 7.3*	111 ± 12	92.7 ± 7.2	108 ± 6.1
GS activity (nkat mg^{-1} protein)	4.71 ± 1.2	49.2 ± 7.2	6.88 ± 0.77	58.7 ± 1.6	7.21 ± 3.9	82.1 ± 6.2*