

Research on cadmium adsorption-desorption dynamics of biochars

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Abstract: Biochar has high potential usage in retaining various contaminants, wastewater treatment, and water purification. In this study, three rice husk derived biochars with pyrolysis temperature 300, 400 and 500 °C, respectively, and pristine rice husk were used to remove cadmium from aqueous solution. The results showed that about 70% or more of Cd²⁺ adsorption occurred in the first 960 mins of adsorption kinetics. The Cd²⁺ adsorption capacity under equilibrium increased with increasing pyrolysis temperature, probably attributed to the increased specific surface area (SSA) under higher pyrolysis temperature noting that significant linear correlation occurred between Cd²⁺ adsorption capacity and SSA. The Cd²⁺ adsorption could be best fitted by pseudo-second order model relative to Elovich model and pseudo-first order model. The Cd²⁺ adsorption rates were higher in film diffusion stage, indicating that film diffusion stage was significant and fast in the early stage of Cd²⁺ adsorption. In contrast, Cd²⁺ adsorption by intra-particle diffusion accounted for 47.0%, 47.9% and 43.9% on average of the total Cd²⁺ adsorption, respectively, indicating that intra-particle diffusion of Cd²⁺ played a more predominant role in limiting Cd²⁺ adsorption rate. When reaching Cd²⁺ desorption equilibrium, removal ratio (RR) values were averaged 0.96, 0.91, and 0.90 under three initial concentrations. More than 90 percentage on average of Cd²⁺ was removed from aqueous solution by biochars and rice husk as well, thus biochars can be used to efficiently remove contaminants from aqueous environment. Cation exchange, electrostatic attraction, and the complexation with surface functional groups could be the main dominant mechanisms for Cd²⁺ adsorption-desorption on biochars.

Keywords Biochar; Cadmium; Pyrolysis temperature; Adsorption kinetics; Desorption

1 Introduction

Biochar has attracted global attention due to its high potential usage over the last decades such as wastewater treatment, water purification, retaining various contaminants, CO₂ capture, and soil amelioration to improve soil fertility and carbon sequestration, etc. (Ahmad et al. 2014; Leng et al. 2015; Tan et al. 2016; Oliveira et al. 2017; Hossain et al. 2020). Biochar is a carbon-rich material derived from a wide range of biomass or organic waste through the thermochemical route (Porpatham et al., 2012), and holds distinctive properties such as high carbon content, larger porosity, greater specific surface area (SSA), cation exchange capacity (CEC) and nutrient retention capacity, and stable structure (Sakhiya et al. 2020). Recent studies in literature showed that biochars had been used to remove organic contaminants such as insecticides (imidacloprid and clothianidin) from soils (Zhang et al. 2021), and herbicide (atrazine) and PAH (Phenanthrene) (Yang et al., 2020; Lian et al. 2019) from aqueous solution. Biochar shows strong alkalinity and is negatively charged in the surface, thereby can interact with metallic ions through ion exchange, electrostatic attraction (Jiang et al. 2012), and surface complexation with oxygen functional groups, precipitation, and metal- π electron interaction, in recent studies (Hossain et al. 2020).

Cadmium (Cd) is one kind of heavy metals that any organisms do not need due to its strong harmful effects on biochemical and physiological processes (Matei et al. 2015; Naushad et al. 2015). Cadmium, generally in the form of ions (Cd²⁺), was exposed in soil or aqueous environments through accumulation of garbage, sewage

irrigation and improper fertilization (Zhu et al. 2017). Cadmium showed very big mobility in soil-plant continuum, and did not leach with water easily (Bolan et al. 2014). Cadmium pollution has become a global environmental problem in recent years and show great potential harm to human body.

The Cd adsorption in soils could influence the mobilization, transformation and bioaccumulation of Cd, and decrease the bioavailability and environmental risks (Khan et al. 2018). Biochar containing functional groups can effectively reduce the effect of Cd stress by intensifying the adsorption of Cd in the soil, thereby significantly reducing the Cd content in plant (tobacco), and providing a theoretical basis and method to alleviate soil Cd pollution and enhance soil remediation (Ren et al. 2021). Biochar applied as an environmental friendly amendment in Cd-polluted soil could ameliorate the negative influences of Cd stress on plant (Khan et al. 2020b). and reduces Cd levels in the plant leaves due to the absorption properties of biochar, therefore the usage of biochar in contaminated soil could help reduce pollutions and decreases the human risk assessment (Mehdizadeh et al. 2021). Biochar could remove Cd from aqueous environment, e.g., MgCl₂ modified biochar had much higher Cd(II) adsorption of capability than that of pristine biochar due to its enhanced ion exchange ability (Zhang et al., 2021), and the H₃PO₄ modified biochar significantly increased the removal rate of Cd(II) from water (Li et al. 2020). In addition, (Khan et al. 2020a) presented a novel adsorbent with biochar modified with iron and MoS₂ which had higher surface area and stronger adsorption capability for Cd from aqueous solution than the unmodified biochar. The similar results had been

achieved by (Zhu et al. 2020), who showed that the adsorption capacity of surface aminated biochar and surface oxidized biochar for Cd was 23.54 and 19.04 mg g⁻¹, respectively, which was about three times higher than that of the pristine porous biochar. Much more researches including those described above indicated that biochars (both modified and unmodified) hold stronger adsorption ability for Cd, however, the adsorption-desorption dynamics, factors such as pyrolysis temperature, specific surface area (SSA) related to adsorption capacity, and sorption mechanisms need further study.

The present research was aimed to compare adsorption-desorption kinetic behavior of biochar obtained under three pyrolysis temperatures, examine the diversity for adsorption-desorption kinetics characteristics, and explore the adsorption mechanisms and more suitable ways to reduce the effectiveness of heavy metals such as cadmium.

2 Materials and methods

2.1 Materials

Biochar was made from rice husks. After drying at room temperature, rice husks were taken into the muffle furnace (Fisher Scientific, USA). At 3 different pyrolysis temperature (300°C, 400°C, 500°C), with residence time for 30 minutes, then carbonization process was completed.

The chemical reagent Cd(NO₃)₂·4H₂O (analytical grade) was resolved in a solution of 0.005 mol L⁻¹ CaCl₂·2H₂O, the initial Cd²⁺ concentrations were 0.5, 2.5

and 5 mg L^{-1} , respectively. The solution pH was adjusted to 6 using 0.1 mol L^{-1} NaOH or HCl solution.

2.2 Determination of biochar characters

Using 0.3 g biochar sample, the specific surface area (SSA) was measured by SSA and pore size analyzer (V-sorp4800b, Beijing, China). The energy spectrum analysis of biochar was measured by energy disperse spectroscopy (QUANTAX 50, Germany).

2.3 Adsorption kinetics

Firstly, 0.3g rice husk and biochar samples under three pyrolysis temperatures were put respectively into 100 ml centrifuge tube containing 0.005 mol L^{-1} $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$ solution with the initial Cd^{2+} concentrations as 0.5, 2.5 and 5 mg L^{-1} , respectively, shaken at stable temperature environment of $25 \text{ }^\circ\text{C}$, and taken out after 5min, 15min, 30min, 60min, 120min, 240min, 480min, 960min, 1920min, 3840min and 7680min, respectively. Three ml supernatant liquid was taken out each time, right after 20 min's (4000 r/min) centrifuge. Atomic absorption spectrophotometer (AA-7000, Japan) was used to measure Cd^{2+} concentration. After the adsorption kinetics experiment finished, 25 ml 0.01 mol/L $\text{Ca}(\text{NO}_3)_2$ solution was added into centrifuge tube which contained rice husk and biochar samples with adsorbed Cd^{2+} . and shaken under 25°C and took them out in 5min, 15min, 30min, 60min, 120min, 240min, 480min, 960min, 1920min, 3840min and 7680min, respectively. the samples were centrifuged for 20 min (4000 r/min) and poured out the supernatant fluid for

Cd^{2+} concentration determination. Then, 25 ml 0.01mol/L $\text{Ca}(\text{NO}_3)_2$ was put in centrifuge tube one more time to start another desorption cycle.

2.4 Data analysis

Three adsorption kinetics models were used to account for Cd^{2+} adsorption kinetics.

The pseudo-first order model was expressed as:

$$\ln(q_e - q_t) = \ln q_e - k_1 t \quad (1)$$

The pseudo-second order model was expressed as:

$$\frac{t}{q_t} = \frac{1}{k_2 q_e^2} + \frac{1}{q_e} t \quad (2)$$

where k_1 is the first order rate constant (h^{-1}), k_2 is the pseudo second-order rate constant ($\text{g mg}^{-1} \text{h}^{-1}$), q_e (mg g^{-1}) and q_t (mg g^{-1}) are the amounts of CIP adsorbed at equilibrium and at time t , respectively. The slope from the plot of $\ln(q_e - q_t)$ vs. t indicates the value of k_1 , and k_2 and q_e values were calculated from the slope and intercept of the t/q_t vs. t plot. t is the contact time (min).

The simplified Elovich model equation was expressed as:

$$q_t = \beta \ln(\alpha\beta) + \beta \ln t \quad (3)$$

where the constants α and β are obtained from the slope and intercept of the linear plot of q_t vs. $\ln t$.

The Weber-Morris intra-particle diffusion model equation was expressed as:

$$q_t = k_i t^{1/2} + C_i \quad (4)$$

where k_i is the intra-particle diffusion rate constant ($\text{mg g}^{-1} \text{h}^{-1/2}$) and C_i is a constant that indicates the thickness of the boundary layer.

The extent of desorption was described by removal ratio (RR) referring to Li et al. (2018) and modified, was calculated as:

$$RR = (q_{ads} - q_{des})/q_{ads} \quad (5)$$

where q_{ads} and q_{des} (mg kg^{-1}) represent the solid-phase concentrations Cd^{2+} adsorption and Cd^{2+} desorption, respectively.

The statistical analysis was conducted by SPSS 16.0. One-way analysis of variance (ANOVA; *F*-test) was conducted to compare the difference among treatments. A Pearson's bivariate linear correlation test was used for detecting the relationship among variables of interest.

3 Results and discussion

3.1 Specific surface area

The specific surface area (SSA) of biochars tended to increase linearly with the increase in pyrolysis temperatures (Table 1) ($r = 0.841$, $P = 0.159$). The significant differences ($P < 0.05$) in SSA were observed between higher temperature pyrolysed biochars, i.e., T-400 and T-500, and rice husks (T-0). The SSA of biochars with pyrolysis temperature 300 °C (T-300) increased by 26.5% as compared with rice husks (T-0), while the SSA of biochars with pyrolysis temperature 400 °C (T-400) and 500 °C (T-500) increased by 3.7 and 4.8 times as compared with rice husks (T-0). The high operational temperature during pyrolysis enhanced the surface area of biochar (Nguyen et al. 2010). The increase in SSA could enhance the adsorption

capacity of contaminants onto adsorbents (Li et al. 2014). In addition, the properties of biochar such as C content, C/H ratio, and the degree of aromatization increased with increasing pyrolysis temperature (Zhou et al. 2017), which may help increase the adsorption ability of biochar.

3.2 Adsorption kinetics modeling

The contact time is regarded as one of the important factors affecting the adsorption process (Li et al. 2018). Fig. 2 shows the dynamic adsorption process of Cd^{2+} onto rice husks (T-0) and biochars (T-300, T400, and T-500) under different initial Cd^{2+} concentrations. For biochar T-500 under 0.5 and 2.5 mg L^{-1} initial concentration, a sharp increase ($>$ or $\approx 80\%$) in Cd^{2+} adsorption took place within the first 240 mins (Fig.2 A, A1), which ran faster than rice husks and biochar T-300 and T-400. More than 90% Cd^{2+} adsorption onto biochar T-500 occurred within the first 480 mins under 0.5 mg L^{-1} initial concentration, however, this event lagged behind under 2.5 and 5 mg L^{-1} initial concentration relative to that under 0.5 mg L^{-1} initial concentration. Within the first 960 mins Cd^{2+} adsorption onto the adsorbents accounted for 79.3% on average (61-95%) of the total Cd^{2+} removal under 0.5 mg L^{-1} initial concentration (Fig.2 A1). Similarly, 73.7% on average (63-89%) and 69.7% on average (65-79%) of Cd^{2+} removal occurred within the first 960 mins under 2.5 and 5 mg L^{-1} initial concentration, respectively (Fig.2 B1, C1). This linear phase may be related to the time needed to saturate the external surface of the carbonaceous agglomerates (Ncibi and Sillanpää, 2015), and thereafter a transition phase was observed as the adsorption rate began to decrease attributable to the slow diffusion of

contaminants within the porous structure (960-7680 mins) (Li et al., 2018). Finally, the Cd^{2+} adsorption dynamic curve leveled off. The Cd^{2+} adsorption capacity under equilibrium increased with increasing pyrolysis temperature, which may be attributed to the increased SSA under higher pyrolysis temperature. A linear relationship was observed between Cd^{2+} adsorption capacity and SSA ($r = 0.958$, $P = 0.042$, under 0.5 mg L^{-1} initial concentration; $r = 0.997$, $P = 0.003$, under 2.5 mg L^{-1} initial concentration; $r = 0.936$, $P = 0.064$, under 5 mg L^{-1} initial concentration).

The adsorption kinetics could be fitted resorting to Lagergren pseudo-first order, pseudo-second order, and Elovich models in terms of their wide use (Konggidinata et al., 2017; Li et al., 2018). The calculated kinetic parameters for pseudo-first order (PFO), pseudo-second order (PSO), and Elovich models are shown in Table 2. For pseudo-first order model, no significant difference in the calculated rate constants of k_1 was found both under different temperatures and initial Cd^{2+} concentrations. In contrast, for pseudo-second order model the calculated rate constants of k_2 tended to decrease when the initial Cd^{2+} concentration increase from 0.5 to 5 mg L^{-1} for all adsorbents, indicating that Cd^{2+} removal efficiency depended on Cd^{2+} concentration within an aqueous environment, the Cd^{2+} removal was a slower process within higher Cd^{2+} concentration solution.

The Cd^{2+} adsorption could be best fitted by pseudo-second order rather than Elovich model and pseudo-first order model as shown by higher r^2 values (Table 2). The similar results were obtained by recent studies (Park et al., 2019; Penido et al., 2019; Khan et al. 2020a). The pseudo-second order model generally assumes that the

rate limiting step during the adsorption process is a chemical adsorption (Li et al. 2015b).

3.3 Rate controlling step analysis

The Weber-Morris intra-particle diffusion model suggested that film diffusion or intra-particle diffusion was a rate limiting step (Bedin et al., 2016; Konggudinata et al. 2017). Fig. 3 showed that the plots of Cd^{2+} adsorption on rice husks and three biochars against $t^{0.5}$ under different Cd^{2+} initial concentration were not straight lines which did not pass through the origin, indicating that intra-particle diffusion was not the sole rate-limiting step and more than one mechanism should account for the Cd^{2+} adsorption (Li et al. 2018) studied ciprofloxacin adsorption onto carbon nanotubes, and showed three stages of ciprofloxacin adsorption process in which film diffusion (external mass transfer) and intra-particle diffusion were distinct. In addition, (Parket al. 2019) suggested that the adsorption of Cd by pine tree residue biochars (PRBs) could be divided into two stages: rapid adsorption on the initial boundary layer and slow adsorption by intraparticle diffusion. The present study showed that Cd^{2+} adsorption rate (line slope in Fig. 4) of three stages decreased in the order: film diffusion > intraparticle diffusion > adsorption onto the active sites, significantly for Cd^{2+} adsorption within 2.5 and 5 mg L⁻¹ solution, indicating that film diffusion stage was significant and fast (within one hour) in the early stage of Cd^{2+} adsorption (Li et al., 2018). In this stage, Cd^{2+} adsorption accounted for 32.3, 25.8 and 25.8% on average of the total Cd^{2+} adsorption onto all adsorbents within 0.5, 2.5 and 5 mg L⁻¹ solution, respectively. In contrast, Cd^{2+} adsorption by intra-particle diffusion

accounted for 47.0%, 47.9% and 43.9% on average of the total Cd^{2+} adsorption, respectively, indicating that intra-particle diffusion of Cd^{2+} adsorption played a more predominant role in limiting Cd^{2+} adsorption rate on rice husk and biochars. Similarly, Zhu indicated that physicochemical adsorption and intraparticle diffusion dominated the Cd adsorption (Zhu et al. 2020), and surface functional groups were identified as key factors controlling adsorption. In the present research, the film and intra-particle diffusions could be limiting factors controlling Cd^{2+} adsorption rate, particularly for three biochars.

3.4 Desorption kinetics

Desorption kinetics of Cd^{2+} from rice husks and biochars are depicted in Fig. 5. For 0.5 mg L^{-1} initial Cd^{2+} concentration, Cd^{2+} desorption quickly reached equilibrium, 120 mins for rice husks, 60 mins for biochar T-300 and T-400, and 15 mins for biochar T-500. For 2.5 mg L^{-1} initial Cd^{2+} concentration, Cd^{2+} desorption reached equilibrium at 120 mins for rice husks, 60 mins for biochar T-300, and 1920 mins for biochar T-400 and T-500. For 5 mg L^{-1} initial Cd^{2+} concentration, Cd^{2+} desorption reached equilibrium at 30 mins for rice husks and biochar T-300, and 1920 mins for biochar T-400 and T-500.

Fig. 6 shows the removal ratio (RR) of Cd^{2+} on rice husks and biochars from aqueous solution. The RR stands for the Cd^{2+} fixation on adsorbents after desorption cycles. When the Cd^{2+} desorption equilibrium was reached, RR values were averaged 0.96 (0.93-0.98) for 0.5 mg L^{-1} initial concentration, 0.91 (0.85-0.96) for 2.5 mg L^{-1}

initial concentration, and 0.90 (0.85-0.94) for 5 mg L⁻¹ initial concentration. This indicated that more than 90 percentage of Cd²⁺ was removed from aqueous solution by rice rusk and biochars.

3.5 Sorption mechanisms

The present study indicated the biochars tested are strong and efficient adsorbents for Cd²⁺ removal from water solutions. The adsorption mechanisms include ion exchange, complexation, and electrostatic attraction, etc. Khan suggested that the adsorption process of Cd onto a surfaced modified biochar involved several mechanisms such as the complexation of Cd(II) with oxygen-based functional groups(Khan et al., 2020a), ion exchange, electrostatic attraction, Cd(II)- π interactions, metal-sulfur complexation, and inner-surface complexation. The interaction between carboxyl group and Cd dominated the adsorption performance of a surface oxidized biochar (OPBC), while the Cd²⁺- π interaction was weakened by increasing the π electron electrostatic potential of aromatic rings, Zhu showed in a recent research, and the lone pair electrons of the amino groups dominated the complexation of a surface aminated biochar (APBC) with Cd, and the π electron electrostatic potential was almost unaffected(Zhu et al. 2020). Park et al. (2019) demonstrated that the Cd adsorption on mixed pine tree residue biochars (M-PRB) was associated with functional groups such as C=C, COH and COOH, as shown by FTIR, and the exchangeable cations corresponds to 23.6% of the total adsorption amount. Zhang et al. (2019) indicated the Cd²⁺ adsorption on a rice straw-derived biochar was mainly the complexation between -NH₂ group on biochar surface and in

solution. Zhou et al. (2018) suggested that the cation- π interaction could act as the main mechanisms for Cd^{2+} adsorption on the biochar derived from tobacco stem at 700 °C for the residence time of 4–6 h. Chen et al. (2019) showed that electrostatic interaction, precipitation, and O–H bonds were the primary adsorption mechanisms for Cd^{2+} on a H_3PO_4 -modified biochar prepared using chicken feather. However, cation exchange and precipitation were considered as the dominant adsorption mechanisms of Cd^{2+} on rice straw biochars prepared at 400 °C and 700 °C, for Cd^{2+} and Ni^{2+} adsorption in the binary system (Deng et al. 2019). Chen et al. (2015) concluded that the main mechanism of the adsorption process of biochar derived from municipal sewage sludge for Cd^{2+} involves (1) surface precipitation by forming insoluble Cd^{2+} compounds in alkaline condition, and (2) ion exchange for Cd^{2+} with exchangeable cations in the biochar, such as Ca^{2+} . Microstructure characteristics and mechanism analysis further suggested that coprecipitation and surface complexation were the main mechanisms of Cd adsorption by pinecone biochar (Teng et al. 2020). In brief, cation exchange, electrostatic attraction, the complexation of Cd^{2+} with functional groups, surface precipitation, and Cd^{2+} - π interaction, etc., can account for the cadmium adsorption-desorption mechanisms on biochars derived from different sources.

4. Conclusions

The specific surface area (SSA) of biochars tended to increase linearly with the increase in pyrolysis temperatures. The significant differences ($P < 0.05$) in SSA were observed among the adsorbents tested. About 70% or more of Cd^{2+} adsorption

occurred in the first 960 mins. The Cd^{2+} adsorption capacity under equilibrium increased with increasing pyrolysis temperature, attributable to the increased SSA under higher pyrolysis temperature. Significant linear correlation was observed between Cd^{2+} adsorption capacity and SSA. The Cd^{2+} adsorption could be best fitted by pseudo-second order model relative to Elovich model and pseudo-first order model. The Cd^{2+} adsorption rate (line slope) defined by linear correlation in three stages decreased in the order: film diffusion > intraparticle diffusion > adsorption onto the active sites, significantly for Cd^{2+} adsorption within 2.5 and 5 mg L^{-1} solution, indicating that film diffusion stage was significant and fast (within one hour) in the early stage of Cd^{2+} adsorption. The Cd^{2+} adsorption by intra-particle diffusion accounted for more than 40% on average of the total Cd^{2+} adsorption, indicating that intra-particle diffusion of Cd^{2+} adsorption played a more predominant role in limiting Cd^{2+} adsorption rate. When reaching Cd^{2+} desorption equilibrium, removal ratio values were averaged more than 0.90 under three initial concentrations. More than 90 percentage on average of Cd^{2+} was removed from aqueous solution by biochars and rice rusk as well, thus biochars can be used to efficiently remove contaminants such as Cd^{2+} from aqueous environment. Cation exchange, electrostatic attraction, the complexation of Cd^{2+} with functional groups, surface precipitation, and Cd^{2+} - π interaction are the dominant mechanisms for Cd^{2+} adsorption-desorption on biochars.

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Data availability All data generated or analyzed during this study are included in this article.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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Table and figure legends

Table 1

Specific surface area (SSA) of rice husks (T-0) and biochars (T-300, T-400, T-500) under different pyrolysis temperature

Table 2.

Adsorption kinetics of Cd^{2+} onto rice husks (T-0) and biochars (T-300, T-400, T-500) with the initial concentrations as 0.5, 2.5, and 5 mg/L, respectively.

Table 3.

The fitted Weber-Morris intra-particle diffusion model

Fig 1. Biochar produced at pyrolysis temperatures as 300, 400, and 500 °C, respectively.

Fig. 2. Adsorption kinetics of Cd^{2+} on rice husk (T-0) and biochars with pyrolysis temperatures as 300 (T-300), 400 (T-400), 500 °C (T-500), respectively. A, B, C represent adsorption kinetics of Cd^{2+} from beginning to 7680 mins, while A1, B1, C1 represent adsorption kinetics of Cd^{2+} from beginning to 960 mins with initial concentrations as 0.5, 2.5, and 5 mg L⁻¹, respectively.

Fig. 3. Adsorption kinetics of Cd^{2+} on rice husk (T-0) and biochars with pyrolysis temperatures as 300 (T-300), 400 (T-400), 500 °C (T-500), respectively, depending upon $t^{0.5}$ (min^{0.5}). A, B, C represent adsorption kinetics of Cd^{2+} with initial

concentrations as 0.5, 2.5, and 5 mg L⁻¹, respectively.

Fig. 4. Adsorption kinetics of Cd²⁺ on rice rusk (T-0) and biochars with pyrolysis temperatures as 300 (T-300), 400 (T-400), 500 °C (T-500), respectively, depending upon t^{0.5} (min^{0.5}). A, B, C represent adsorption kinetics of Cd²⁺ with initial concentrations as 0.5, 2.5, and 5 mg L⁻¹, respectively. Correlation analysis between CIP adsorption and t^{0.5} (h^{0.5}) on the three stages, number 1, 2, and 3 represent stages of external mass transfer, intraparticle diffusion, and adsorption onto the active sites, respectively.

Fig. 5. Desorption kinetics of Cd²⁺ from rice rusk (T-0) and biochars with pyrolysis temperatures as 300 (T-300), 400 (T-400), 500 °C (T-500), respectively. A, B, C represent desorption of Cd²⁺ from beginning to 7680 mins, while A1, B1, C1 represent desorption of Cd²⁺ from beginning to 120 mins for 0.5 mg L⁻¹ initial concentration, and to 1920 mins for 2.5, and 5 mg L⁻¹, respectively.

Fig. 6. Removal rate of Cd²⁺ on rice rusk (T-0) and biochars with pyrolysis temperatures as 300 (T-300), 400 (T-400), 500 °C (T-500), respectively, from aqueous solution. A, B, C represent removal rate of Cd²⁺ from beginning to 7680 mins, while A1, B1, C1 represent removal rate of Cd²⁺ from beginning to 120 mins for 0.5 mg L⁻¹ initial concentration, and to 960 mins with as 2.5 and 5 mg L⁻¹, respectively.

Table 1

Specific surface area (SSA) of rice husks (T-0) and biochars (T-300, T-400, T-500) under different pyrolysis temperature

	T-0	T-300	T-400	T-500
Pyrolysis temperature (°C)	0	300	400	500
BET Specific surface area	9.17c	11.60c	52.42b	62.15a
Langmuir Specific surface area	16.71c	19.14c	68.24b	83.51a

The means in the same row with small letters differ significantly in the different treatments ($P < 0.05$).

Table 2.

Adsorption kinetics of Cd²⁺ onto rice husks (T-0) and biochars (T-300, T-400, T-500) with the initial concentrations as 0.5, 2.5, and 5 mg/L, respectively.

Initial concentration (mg/L)	pyrolysis temperature	Pseudo-first order kinetics			
		q_e	k_1	r^2_{adj}	P -value
0.5	T-0	58.8	0.0011	0.9158	<0.0001
	T-300	66.7	0.0008	0.9613	<0.0001
	T-400	61.5	0.0013	0.9466	<0.0001
	T-500	28.8	0.0011	0.9020	<0.0001
2.5	T-0	275.1	0.0010	0.9402	<0.0001
	T-300	286.6	0.0011	0.9338	<0.0001
	T-400	305.0	0.0010	0.9510	<0.0001
	T-500	188.0	0.0011	0.9484	<0.0001
5	T-0	488.3	0.0011	0.9493	<0.0001
	T-300	503.4	0.0012	0.9646	<0.0001
	T-400	532.0	0.0009	0.9568	<0.0001
	T-500	510.8	0.0009	0.9668	<0.0001
		Pseudo-second order kinetics			
		q_e	$k_2(\times 10^{-5})$	r^2_{adj}	P -value
0.5	T-0	58.8	5.08	0.9956	<0.0001
	T-300	69.9	2.51	0.9861	<0.0001
	T-400	84.0	8.21	0.9997	<0.0001
	T-500	83.3	29.1	0.9999	<0.0001
2.5	T-0	285.7	0.498	0.9789	<0.0001
	T-300	285.7	0.854	0.9960	<0.0001
	T-400	370.4	0.800	0.9972	<0.0001
	T-500	384.6	1.37	0.9997	<0.0001
5	T-0	526.3	0.602	0.9933	<0.0001

	T-300	555.6	0.392	0.9718	<0.0001
	T-400	625.0	0.352	0.9772	<0.0001
	T-500	714.3	0.537	0.9961	<0.0001
Elovich model					
		α	β	r^2_{adj}	P -value
0.5	T-0	0.012	8.41	0.9316	<0.0001
	T-300	0.007	9.63	0.8827	<0.0001
	T-400	0.017	12.3	0.9632	<0.0001
	T-500	0.365	9.18	0.8914	<0.0001
2.5	T-0	0.002	40.0	0.9533	<0.0001
	T-300	0.002	41.3	0.9263	<0.0001
	T-400	0.002	53.8	0.9559	<0.0001
	T-500	0.012	51.9	0.8692	<0.0001
5	T-0	0.002	73.4	0.9858	<0.0001
	T-300	0.002	75.4	0.9814	<0.0001
	T-400	0.001	84.9	0.9826	<0.0001
	T-500	0.001	104.2	0.9778	<0.0001

T-0, T-300, T-400, and T-500 represent rice rusk and the biochars pyrolysed with temperatures as 300, 400, 500 °C, respectively. The calculation methods of r^2_{adj} followed $r^2_{\text{adj}} = 1 - (1-r^2)(N-2)/(N-P-2)$, N : the number of experimental data points; P : number of parameters in the model.

Table 3.

The fitted Weber-Morris intra-particle diffusion model

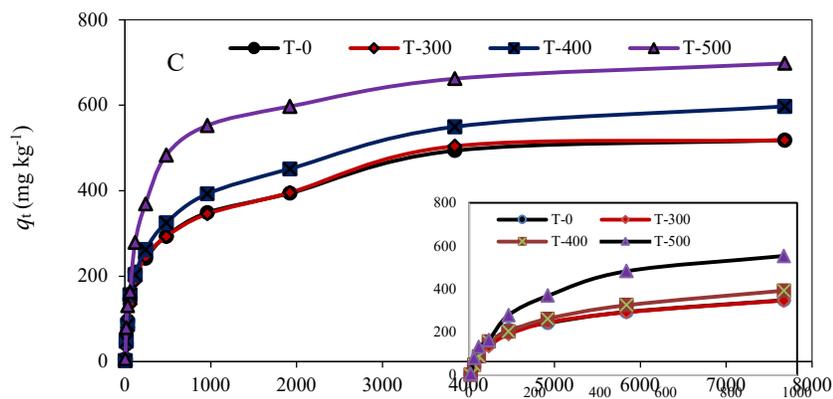
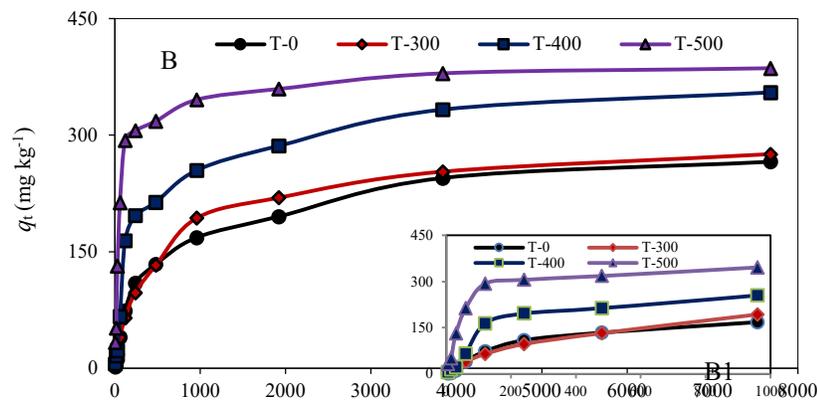
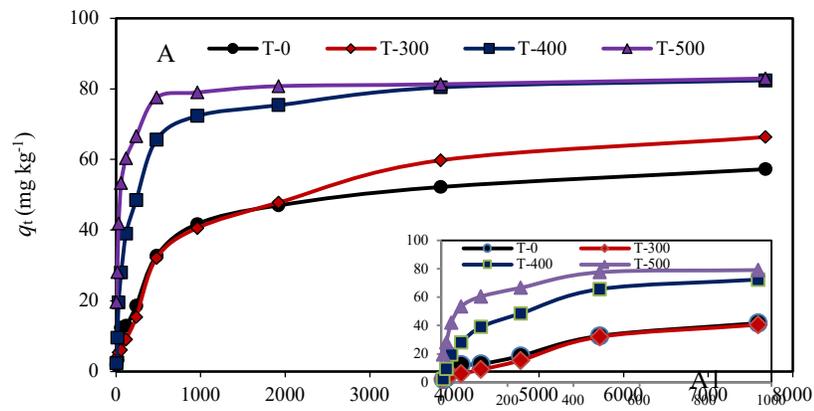
Initial concentration (mg/L)	pyrolysis temperatures	Intra-particle diffusion model			
		K_i	C_i	r^2_{adj}	P -value
0.5	T-0	0.688	8.097	0.8222	0.000
	T-300	0.834	4.139	0.8952	0.000

	T-400	0.901	23.64	0.6210	0.001
	T-500	0.613	44.78	0.4279	0.009
2.5	T-0	3.287	25.98	0.8497	0.000
	T-300	3.427	28.86	0.8514	0.000
	T-400	4.161	63.51	0.7249	0.000
	T-500	3.458	163.5	0.4277	0.009
5	T-0	5.848	98.21	0.8129	0.000
	T-300	6.028	89.42	0.8159	0.000
	T-400	6.852	97.47	0.8383	0.000
	T-500	7.958	154.3	0.7177	0.000

T-0, T-300, T-400, and T-500 represent rice rusk and the biochars pyrolysed with temperatures as 300, 400, 500 °C, respectively. The calculation methods of r_{adj}^2 followed $r_{\text{adj}}^2 = 1 - (1-r^2)(N-2)/(N-P-2)$, N : the number of experimental data points; P : number of parameters in the model.



Fig 1. Biochar produced at pyrolysis temperatures as 300, 400, and 500 °C, respectively.



C1

t (min)

Fig. 2. Adsorption kinetics of Cd^{2+} on rice rusk (T-0) and biochars with pyrolysis temperatures as 300 (T-300), 400 (T-400), 500 °C (T-500), respectively. A, B, C represent adsorption kinetics of Cd^{2+} from beginning to 7680 mins, while A1, B1, C1 represent adsorption kinetics of Cd^{2+} from beginning to 960 mins with initial concentrations as 0.5, 2.5, and 5 mg L^{-1} , respectively.

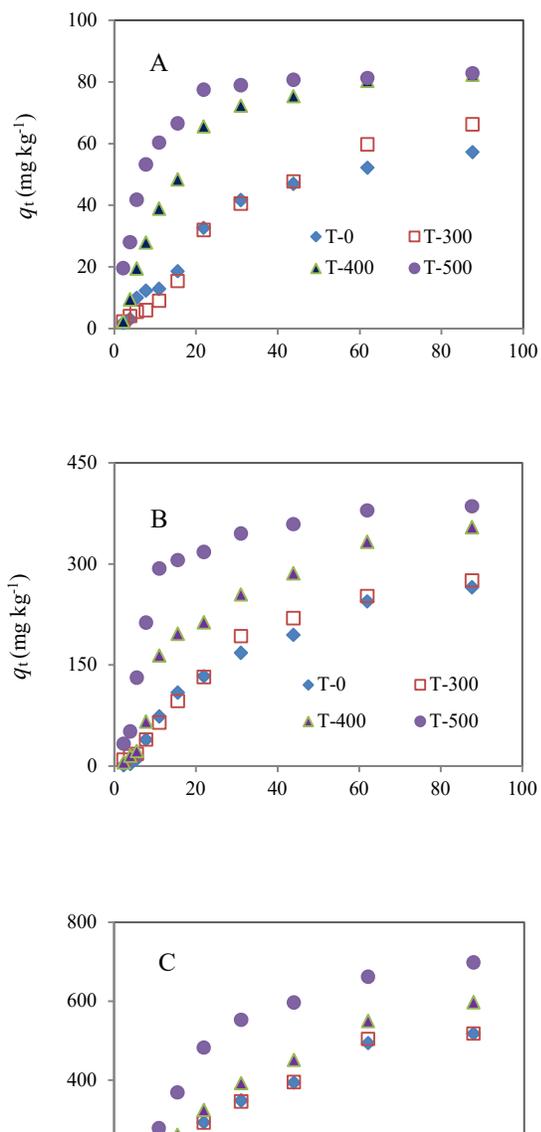
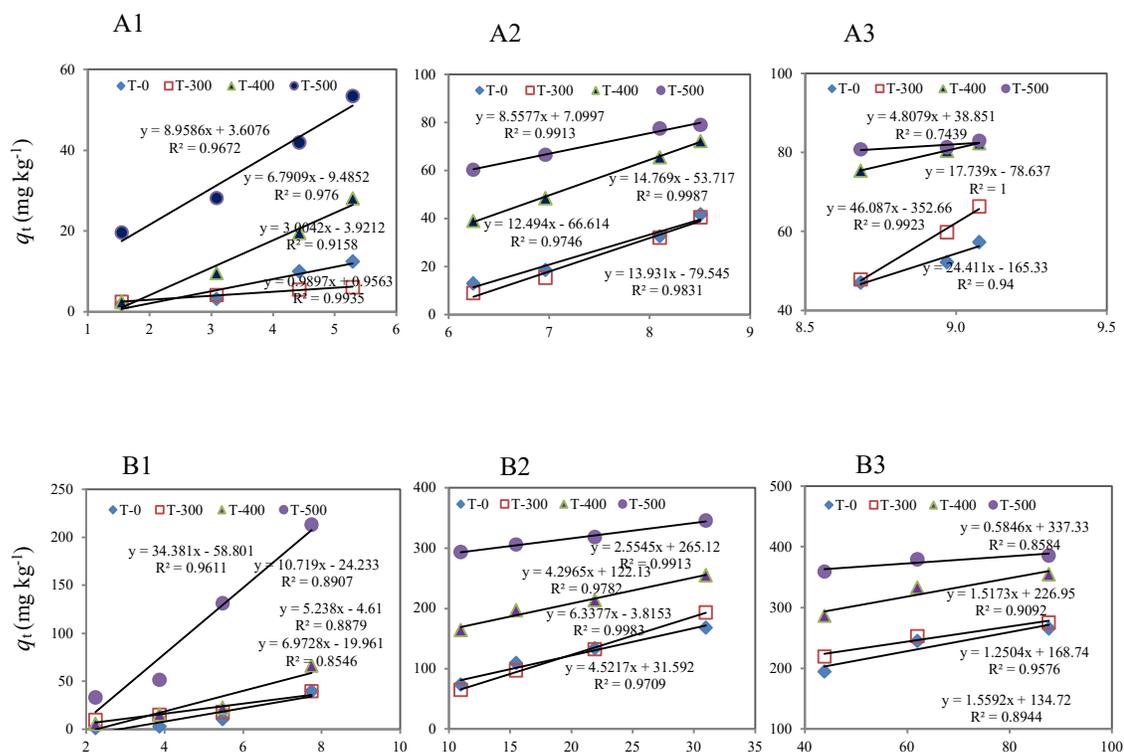




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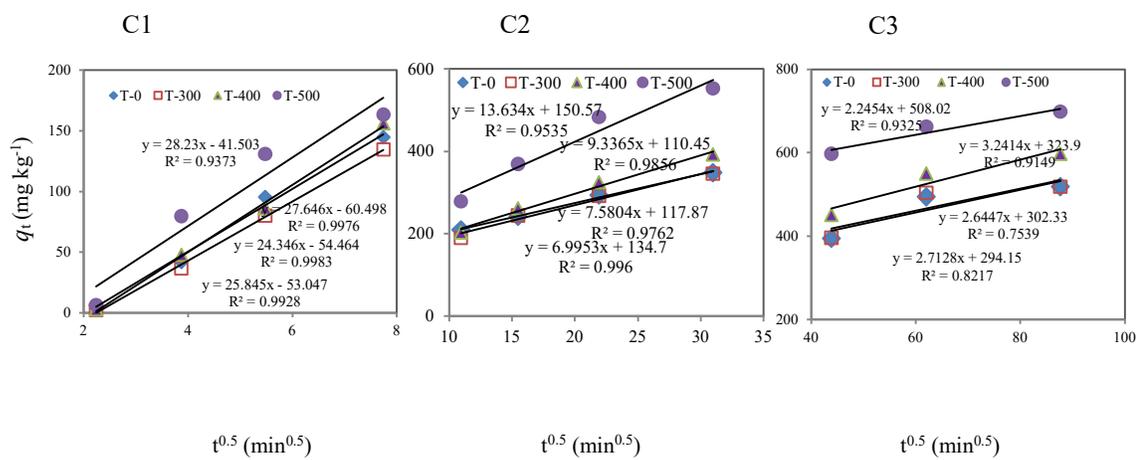
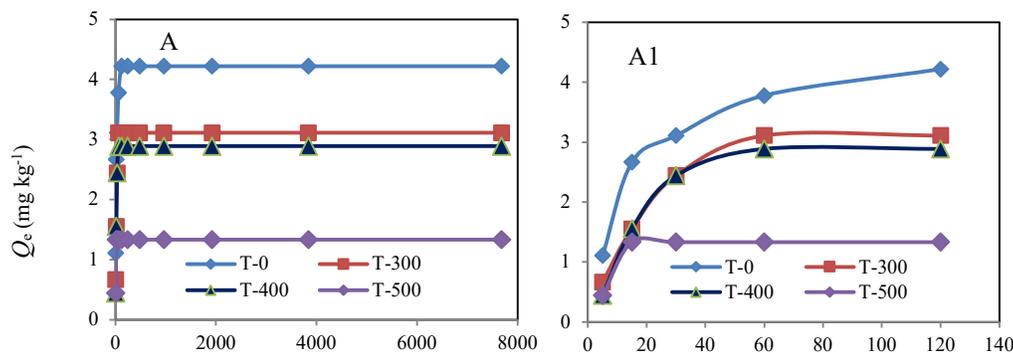


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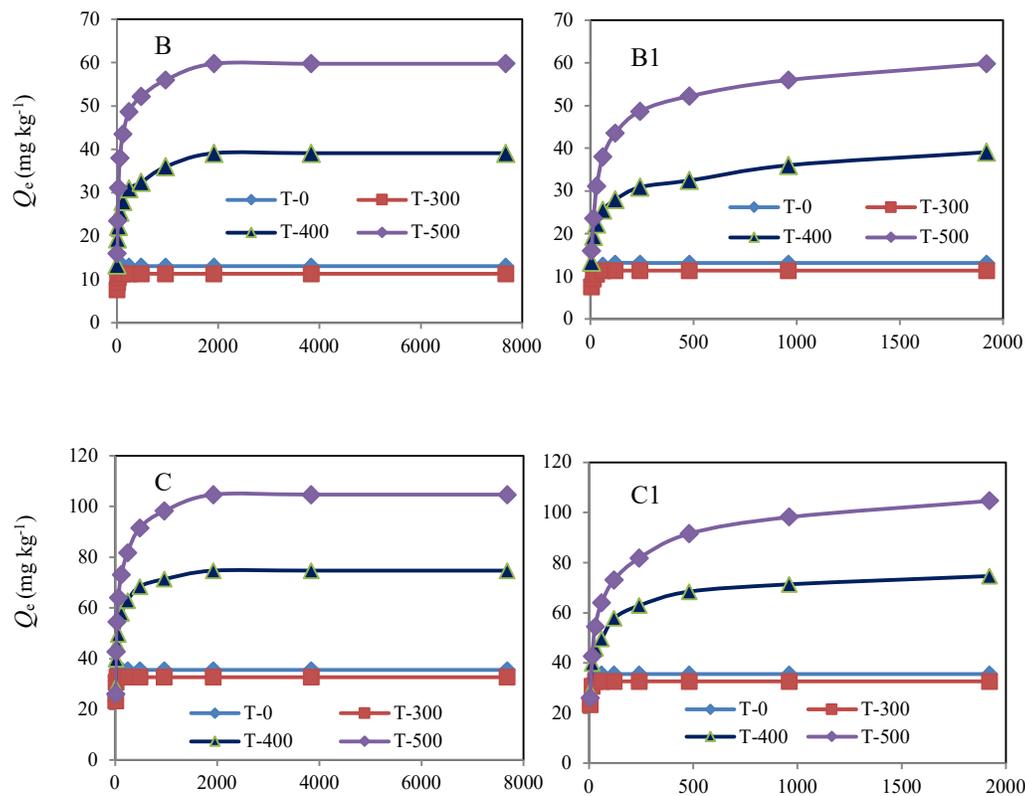
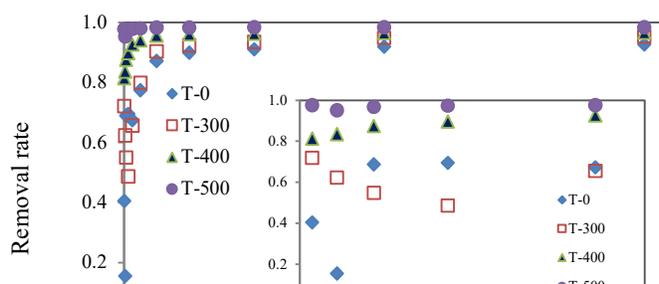


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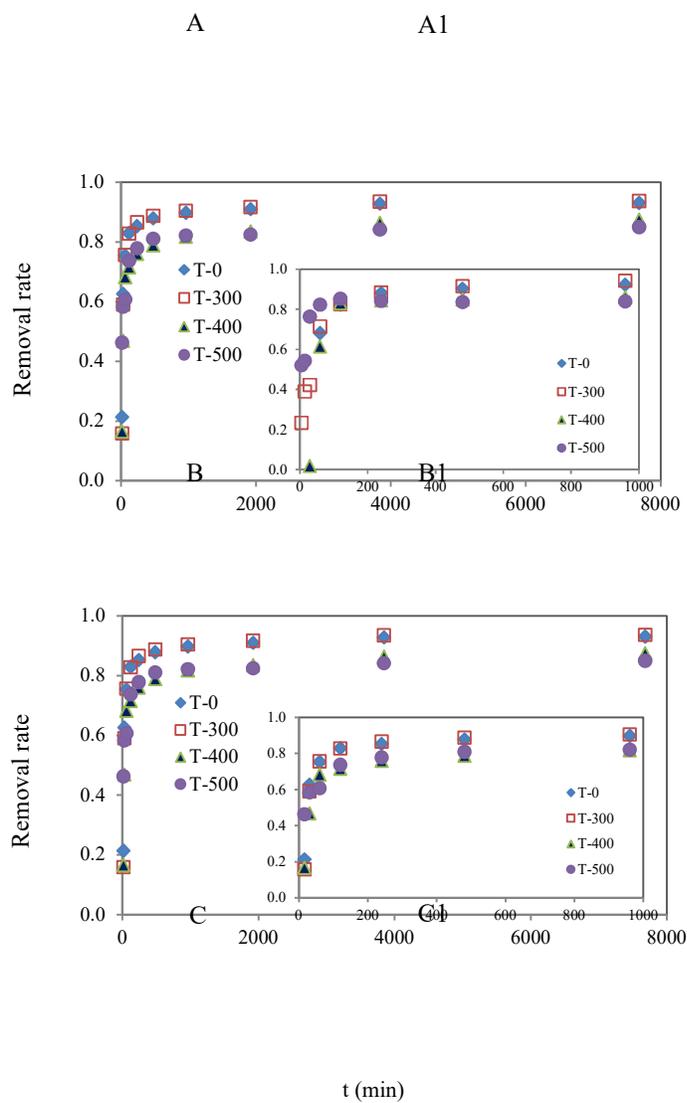


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