

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

## Article

# Cryptic oral microbiota: What is its role as OSA-related periodontal pathogens?

Mayra A. Téllez Corral <sup>1,2,3\*</sup>, Eddy Herrera Daza <sup>4</sup>, Hayde K. Cuervo Jimenez<sup>2</sup>, María del Mar Bravo Becerra <sup>2</sup>, Jean Carlos Villamil <sup>1</sup>, Patricia Hidalgo Martinez <sup>5</sup>, Nelly S. Roa Molina <sup>1</sup>, Liliana Otero <sup>1</sup>, María E. Cortés <sup>3</sup>, Claudia M. Parra Giraldo <sup>2,6\*</sup>

- <sup>1</sup> Centro de Investigaciones Odontológicas, Facultad de Odontología, Pontificia Universidad Javeriana, Bogotá D.C., Colombia; [jean.villamil@javeriana.edu.co](mailto:jean.villamil@javeriana.edu.co) (J.C.V.); [nelly.roa@javeriana.edu.co](mailto:nelly.roa@javeriana.edu.co) (N.S.R.M.); [lotero@javeriana.edu.co](mailto:lotero@javeriana.edu.co) (L.O.)
  - <sup>2</sup> Unidad de Investigación en Proteómica y Miosis Humanas, Departamento de Microbiología, Facultad de Ciencias, Pontificia Universidad Javeriana, Bogotá D.C., Colombia; [hayde.cuervo@javeriana.edu.co](mailto:hayde.cuervo@javeriana.edu.co) (H.K.C.J.); [bravomaria@javeriana.edu.co](mailto:bravomaria@javeriana.edu.co) (MMB)
  - <sup>3</sup> Faculty of Dentistry and Innovation Technology Graduate Program, Universidade Federal de Minas Gerais, Belo Horizonte, Minas Gerais, Brazil; [mecortes@yahoo.com](mailto:mecortes@yahoo.com) (MEC)
  - <sup>4</sup> Departamento de Matemáticas, Facultad de Ciencias, Pontificia Universidad Javeriana, Bogotá D.C., Colombia; [eherrera@javeriana.edu.co](mailto:eherrera@javeriana.edu.co) (EHD)
  - <sup>5</sup> Sleep clinic, Hospital Universitario San Ignacio and Faculty of Medicine, Pontificia Universidad Javeriana, Bogotá D.C., Colombia; [phidalgo@husi.org.co](mailto:phidalgo@husi.org.co) (PHM)
  - <sup>6</sup> Departamento de Microbiología, Facultad de Farmacia, Universidad Complutense de Madrid., España
- \* Correspondence to: [tellezm@javeriana.edu.co](mailto:tellezm@javeriana.edu.co) (M.A.T.C.) and [claudia.parra@javeriana.edu.co](mailto:claudia.parra@javeriana.edu.co) (C.M.P.G.)

**Abstract:** Commonly the periodontitis has been linked to periodontopathogens categorized in Socransky's microbial complexes, however, there is a lack of knowledge regarding "other microorganisms" or "cryptic microorganisms", which are rarely thought of as significant oral pathogens and are neither previously categorized nor connected to illnesses in the oral cavity. This study hypothesized that these cryptic microorganisms could contribute to the modulation of oral microbiota present in health or disease (periodontitis and/or OSA patients). For this purpose, the presence and the correlation among these cultivable cryptic oral microorganisms were identified and their possible role in both conditions was determined. Data from oral samples of individuals with or without periodontitis and with or without OSA were obtained from a previous study. Demographic data, clinical oral characteristics, and genera and species of cultivable cryptic oral microorganisms identified by MALDI-TOF were recorded. The data of 75 participants were analyzed to determine the relative frequencies of cultivable cryptic microorganisms' genus and species, microbial clusters and correlations tests were performed. According to periodontal condition, Gingivitis - dental biofilm-induced in reduced periodontium and stage III periodontitis were found to have the highest diversity of cryptic microorganism species. Based on the experimental condition these findings showed that there are genera related to disease conditions and others related to healthy conditions, with species that could be related to different chronic diseases being highlighted as comorbidities periodontitis and OSA. The cryptic microorganisms within the oral microbiota of patients with periodontitis and OSA are present as potential pathogens, promoting the development of dysbiotic microbiota, and the occurrence of chronic diseases, which have been previously proposed to be common risk factors for periodontitis and OSA. Understanding the function of possible pathogens in the oral microbiota will take more research.

**Keywords:** periodontitis; obstructive sleep apnea; oral microbiota; pathogenic microbiota; chronic diseases; MALDI-TOF

## 1. Introduction

According to recent studies, patients with OSA have an increased risk of developing periodontitis [1,2]. Some hypotheses about the connection between periodontitis and OSA include a genetic predisposition, an inflammatory response that both disorders share, and a change in the oral microbiota [3].

Individuals with OSA have higher prevalence of increased periodontal parameters like Probing Depth (PD) and Clinical Attachment Level (CAL), as well as an Index of Apnea-Hypopnea (AHI) >15 events per hour, and they experience mouth breathing-related oral dryness. Also, these individuals have a microbiota characterized by an increase of Gram-negative bacteria, primarily periodontal pathogens [4,5]. It has been reported that the oral microbiota of patients with OSA is significantly different from that of individuals without OSA. The nasal microbiome of subjects with severe OSA was altered and enriched with *Streptococcus*, *Prevotella*, and *Veillonella*; Several common oral commensals (*Streptococcus*, *Rothia*, *Veillonella*, and *Fusobacterium*) correlated with apnea-hypopnea index [6]. Pyrosequencing was also utilized to detect bacteria associated with OSA and hypertension, revealing the presence of *Porphyromonas* spp. and *Aggregatibacter* sp. in both mild and moderate-severe OSA [7]; both genera are associated with the development of periodontitis.

The oral dryness affects bone remodeling triggered by hypoxia, with an increase in CO<sub>2</sub> levels [5], reducing the immune system's response to infections and allowing a higher diversity of microorganisms (bacteria and yeasts), that they are capable of generating dysbiotic polymicrobial communities. Recent studies have found that individuals with OSA and periodontitis had higher levels of periodontal pathogenic bacteria [8] associated to yeasts such *Candida* spp. [9]. Additionally, of cryptic microbiota which we describe as those microorganisms that are not often considered as significant oral pathogens and neither classified in the Socransky's microbial complexes [10], nor associated with specific pathologies in the oral cavity have been identified in periodontitis and OSA [9]. Although, these cryptic microorganisms could contribute to the periodontitis development in OSA patients. Therefore, the purpose of the present study was to analyze the presence of these cultivable cryptic oral microorganisms in individuals with periodontitis associated with OSA and to identify potential pathogens in both conditions.

## 2. Materials and Methods

### 2.1 Study Population

Demographic data, clinical oral characteristics and cultivable cryptic microorganisms identified by MALDI-TOF equipment (Microflex from<sup>®</sup> Burker Daltonik Inc), in oral samples of participants from a previous study were available for re-analysis for this study. As previously described [9], participants were recruited from the Sleep Clinic of the Hospital Universitario San Ignacio and the Sleep Clinic of the Faculty of Dentistry at the Pontificia Universidad Javeriana-PUJ, Colombia. Inclusionary criteria were set to select completed clinical oral data of participants with cryptic microorganisms identified in their oral samples. The participants that fulfilled the inclusion criteria were assigned to one of four groups according to the severity of their OSA and their periodontal diagnosis, as follows: Group 1 (G1) (H) healthy patients: non-periodontitis and non-OSA (n=20); Group 2 (G2) (P) periodontitis and non-OSA patients (n=13); Group 3 (G3) (OSA) OSA and non-periodontitis patients (n=18); and Group 4 (G4) (P-OSA) periodontitis and OSA patients (n=24). The data of these 75 participants were earlier related by Tellez Corral et al., 2022 [9] and in this study analyzed in relation with cryptic oral microorganisms.

### 2.2 Data register

The demographic data and clinical oral characteristics were recorded, including age, sex, and periodontal parameters: probing depth (PD), clinical attachment loss (CAL),

plaque index (PI), bleeding of probing (BOP) and missing teeth. Additionally, genus and species of cultivable cryptic oral microorganisms identified were recorded.

### 2.3 Statistical analysis

The first part of the present study consisted in performing a descriptive statistic. Two-way ANOVA with Tukey's multiple comparisons test were used to analyze the demographic data and periodontal parameters. The second part, tests were carried out to compare the cultivable cryptic oral microorganisms between each group, determining the relative frequencies of cryptic microorganisms' genus and species. The cluster analysis of cryptic microbial communities by group of patients was conducted using the Agglomerative Hierarchical Clustering (AHC) according to the frequency of the microorganisms. Principal Coordinates Analysis (PCoA) calculated by the relative abundance of microorganisms. Association tests were performed within each group using the Spearman r test ( $p$ -value  $<0.5$ ) to correlate periodontal parameters and the cultivable cryptic microorganisms. The software's GraphPad Prism 9.0.2 (GraphPad Software, California, USA) and XLSTAT statistical and data analysis solution (Addinsoft, New York, USA) were used.

## 3. Results

### 3.1 Clinical Data

The demographic variables and periodontal parameters of the study population are presented in Table 1. There was a higher percentage of men in Group 4 (P-OSA) than in the other groups. The teeth with periodontitis (%), the BOP (%) and the PI showed statistically significant differences between Group 2 (P) and Group 4 (P-OSA) vs. Group 1 (H) ( $p < 0.001$ ). As well, the PI showed statistically significant differences between Group 3 (OSA) ( $p < 0.001$ ) vs. Group 1 (H). Regarding the periodontal condition, the biofilm-induced gingivitis in a reduced periodontium was more prevalent in patients in Group 1 (H) and Group 3 (OSA) (56% and 79%, respectively). The Stage III periodontitis was more prevalent in patients in Group 2 (P) and Group 4 (P-OSA) (65% and 81%, respectively) (Figure 1).

**Table 1. Demographic variables and periodontal parameters of the group of patients**

Clinical variable	Group 1 (H) (n=20)	Group 2 (P) (n=13)	Group 3 (OSA) (n=18)	Group 4 (P-OSA) (n=24)
Age (years)	44.35 ± 14.24	40.69 ± 10.83	50.35 ± 13.09	49.33 ± 11.65
Gender (Males) (%)	32	41	37	69
Missing teeth	6.21 ± 4.42	5.69 ± 2.25	8.53 ± 6.53	7.08 ± 6.23
Teeth with periodontitis (%)	2.18 ± 3.99	47.91 ± 24.72*	1.37 ± 2.98»	39.16 ± 19.39*§
PD (mm)	1.81 ± 0.48	2.64 ± 0.45	2.36 ± 4.82	13.83 ± 9.95
Sites (%) PD ≥4 mm	0.29 ± 0.50	16.76 ± 10.62	2.01 ± 0.15	2.64 ± 0.41
CAL (mm)	1.33 ± 0.78	2.15 ± 1.01	1.52 ± 0.98	2.31 ± 1.14
BOP (%)	11.59 ± 11.46	49.19 ± 26.92*	24.02 ± 22.96»	48.67 ± 26.69*§
PI	19.30 ± 11.48	47.21 ± 26.28*	39.65 ± 21.43*	39.16 ± 19.39*

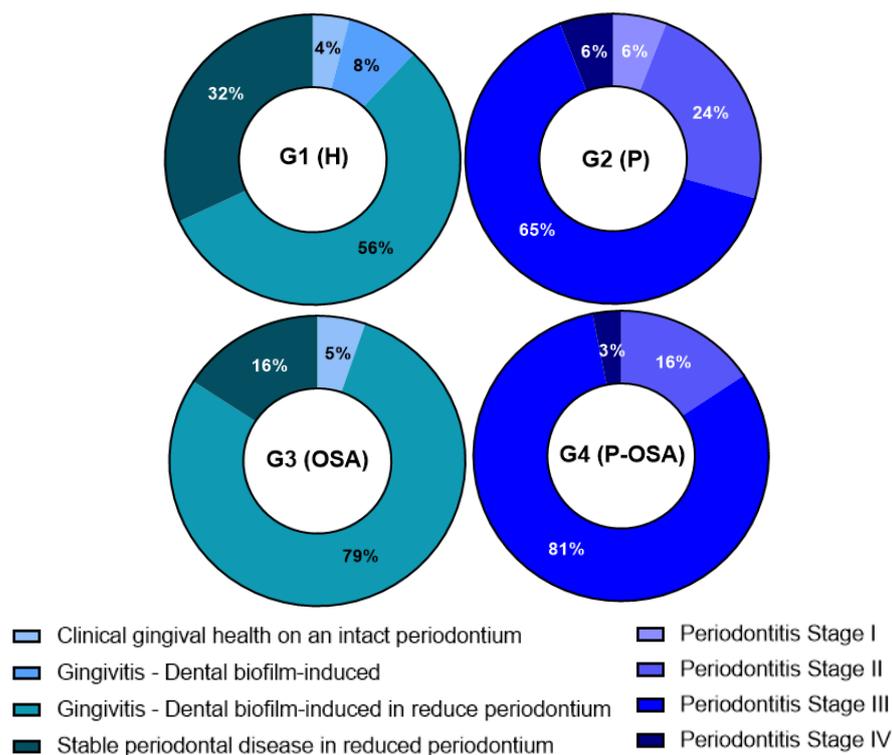
Values are given as mean ± standard deviation; Two-way ANOVA, Tukey's multiple comparisons test,  $p < 0.05$

PD: Probing depth; CAL: Clinical attachment loss; BOP: Bleeding of probing; PI: Plaque index

\* Significantly different to Group 1

» Significantly different to Group 2

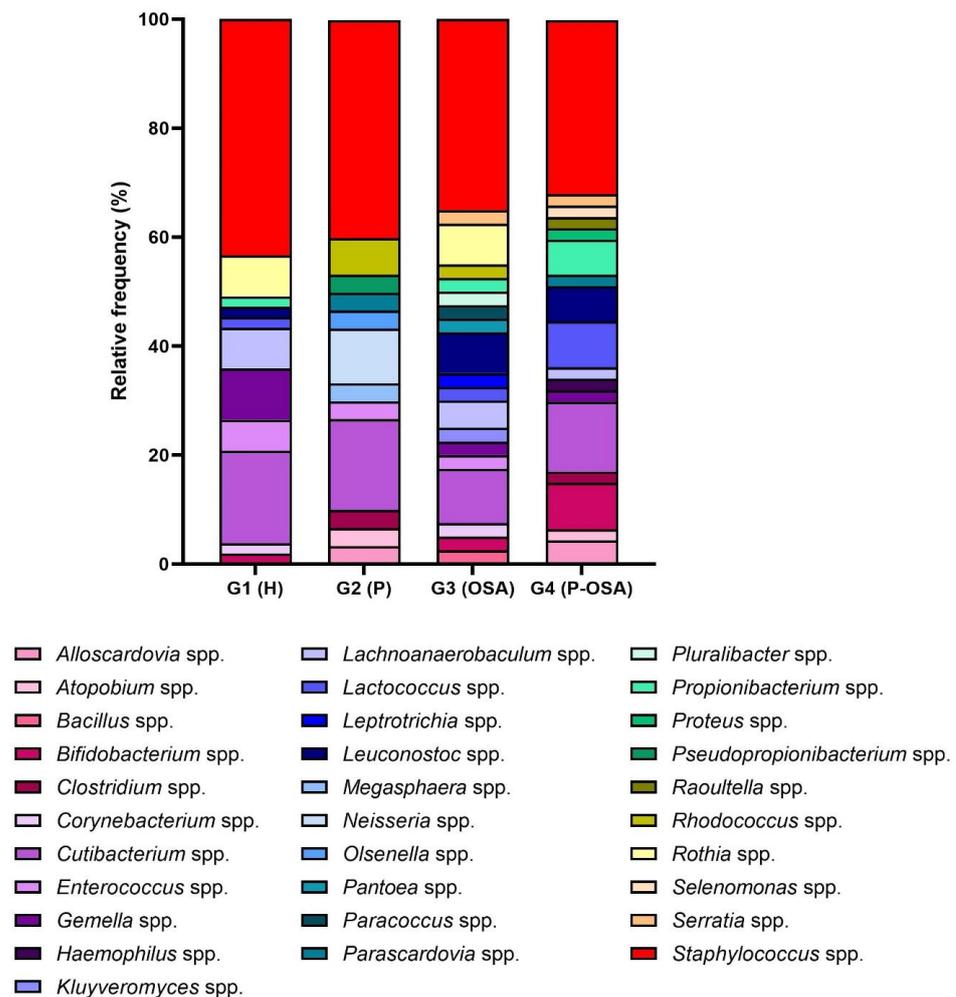
§ Significantly different to Group 3



**Figure 1.** Percentage of patients of each group according to periodontal condition (according to the new Classification of Periodontal and Peri-implant Diseases and Conditions by G. Caton et al., 2018). G1 (H): healthy patients, non-periodontitis and non-OSA (n=20); G2 (P) periodontitis and non-OSA patients (n=13); G3: (OSA) OSA and non-periodontitis patients (n=18); G4 (P-OSA) periodontitis and OSA patients (n=24).

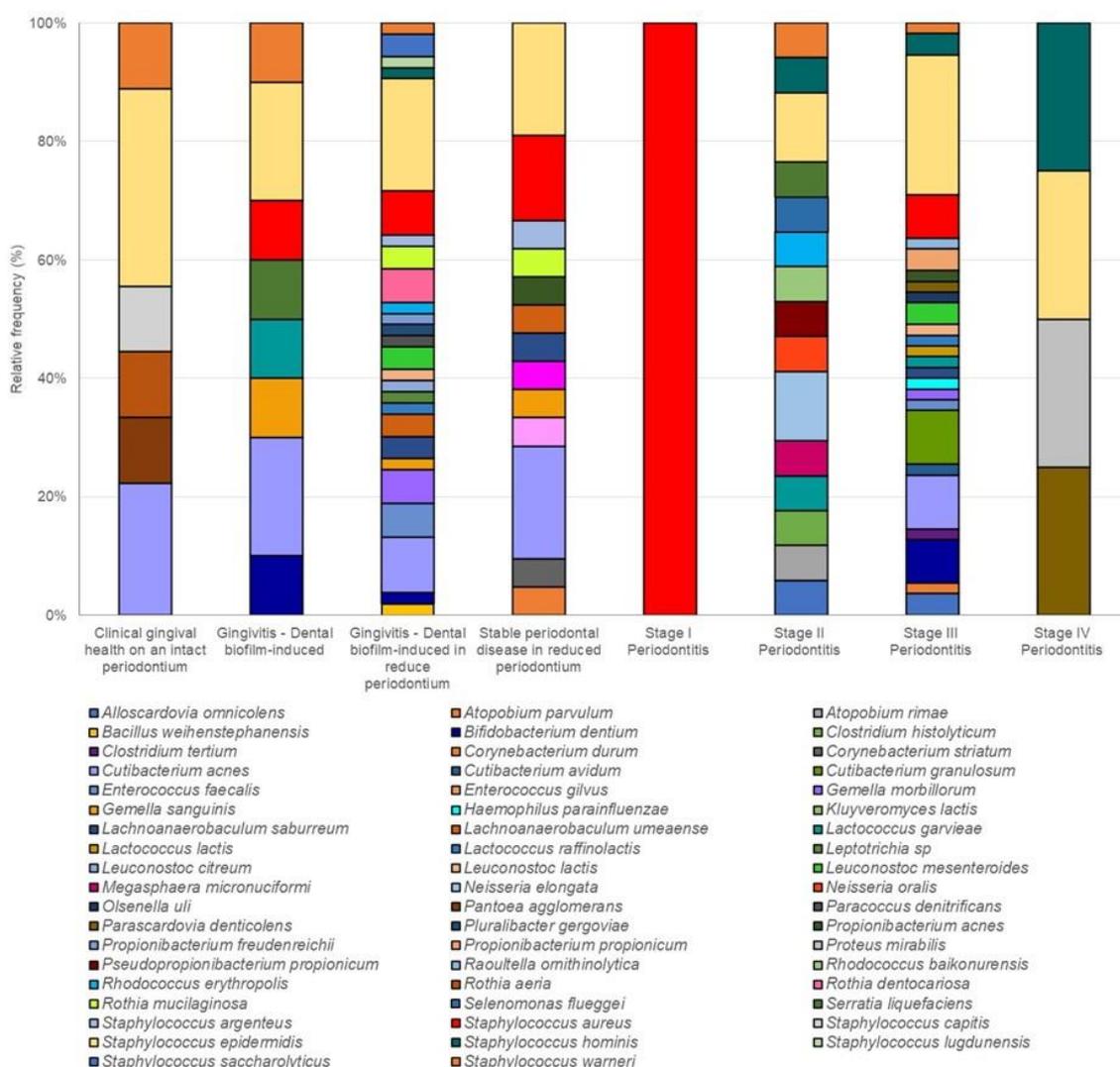
### 3.2 Microbiological data

According to the total number of microorganisms per genus identified by group, the percentages of the cryptic microorganisms for each group of patients were adjusted to percentages of relative frequency (Figure 2). Each patient group has a unique microbiological profile that is primarily made up of the genera *Staphylococcus* spp. and *Cutibacterium* spp.; the relative frequency of each genus ranged from 32-43% and 10-17%, respectively. Third-most frequent genera in each group stood out as *Gemella* spp. (9%) in G1 (H), *Neisseria* spp. (10%) in G2 (P), *Rothia* spp. and *Leuconostoc* spp. (7.5%) in G3 (OSA), and *Bifidobacterium* spp. and *Lactococcus* spp. (8.5%) in G4 (P-OSA). There was a decrease of the presence of four genus in G2 (P), G3 (OSA) and G4 (P-OSA) compared to G1 (H): *Staphylococcus* spp. decrease 3% in G2 (P), 8.4% in G3 (OSA) and 11.5% in G4 (P-OSA). *Enterococcus* spp. decrease 2.3% in G2 (P), 3.2% in G3 (OSA) and 5.7% in G4 (P-OSA). *Gemella* spp. decrease 9.4% in G2 (P), 6.9% in G3 (OSA) and 7.3% in G4 (P-OSA), and *Lachnoanaerobaculum* spp. decrease 7.6% in G2 (P), 2.6% in G3 (OSA) and 5.4% in G4 (P-OSA). Otherwise, there was an increase of the presence of five genus in G2 (P), G3 (OSA) and G4 (P-OSA) compared to G1 (H): *Alloscardovia* spp. increase 3.3% in G2 (P) and 4.3% in G4 (P-OSA), *Bifidobacterium* spp. increase 0.6% in G3 (OSA) and 6.6% in G4 (P-OSA). *Corynebacterium* spp. increase 0.6% in G3 (OSA). *Lactococcus* spp. increase 0.6% in G3 (OSA) and 6.6% in G4 (P-OSA). *Leuconostoc* spp. increase 5.6% in G3 (OSA) and 4.5% in G4 (P-OSA), and *Propionibacterium* spp. increase 0.6% in G3 (OSA) and 4.5% in G4 (P-OSA).



**Figure 2.** Microbial profile: percentage of each patient group's relative frequency of the genus of cultivable cryptic microorganisms (other microorganisms). G1 (H): healthy patients; G2 (P) periodontitis patients; G3: (OSA) OSA patients; G4 (P-OSA) periodontitis and OSA patients.

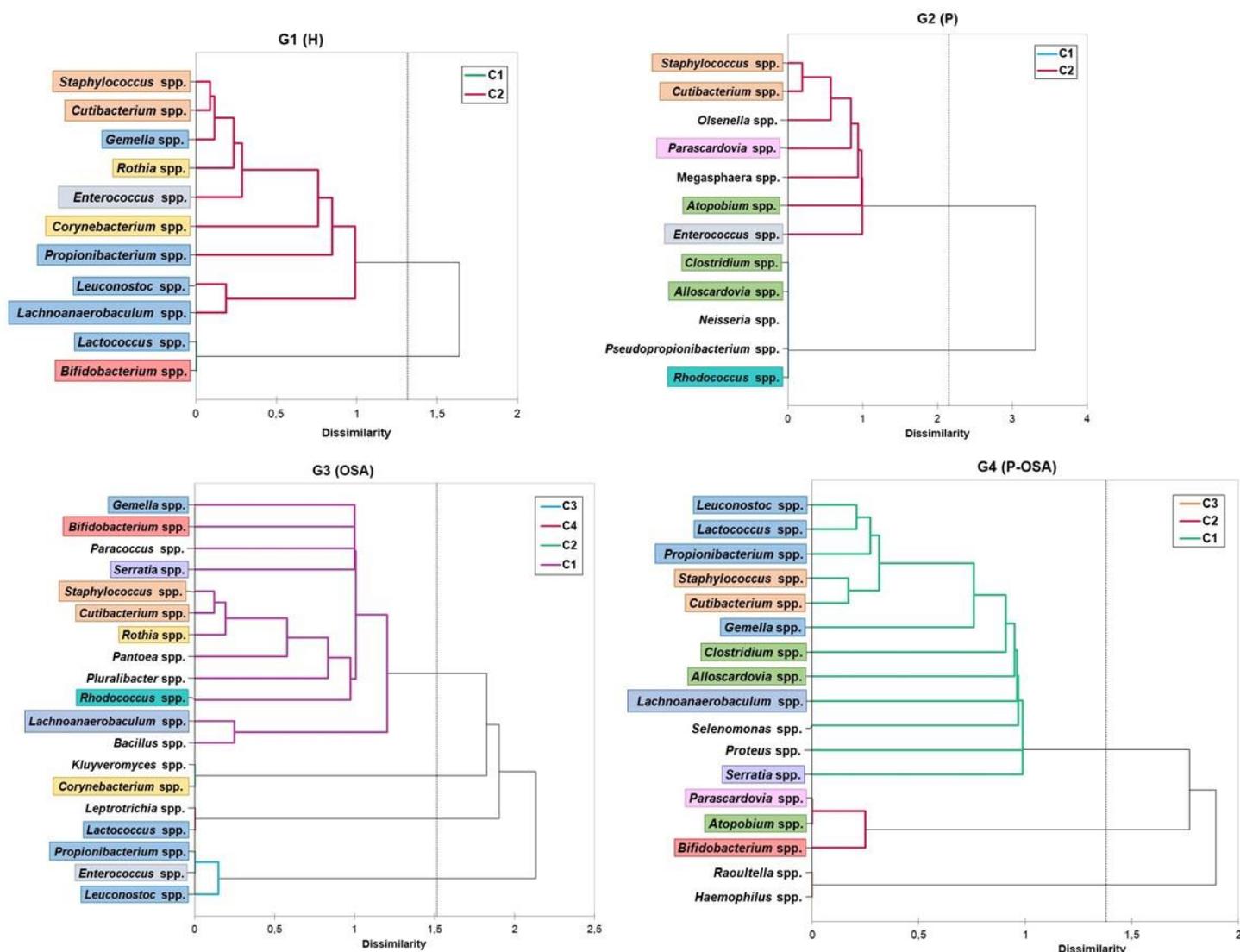
Some species were only identified in one of the four patient groups: (Table S1). Fifty-six species were found, distributed into thirty-one genera. The major diversity of species of cryptic microorganisms were identified in Gingivitis - dental biofilm-induced in reduced periodontium (26 species) and in stage III periodontitis (25 species), being *Staphylococcus epidermidis*, *Cutibacterium acnes* and *Bifidobacterium dentium* the most prevalent in both conditions (Figure 3).



**Figure 3.** Microbial profile: percentage of relative frequency of each species of cultivable oral cryptic microorganisms by periodontal condition: Healthy conditions: Clinical gingival health on an intact periodontium, Gingivitis - Dental biofilm-induced, Gingivitis - Dental biofilm-induced in reduce periodontium, Stable periodontal disease in reduced periodontium. Disease condition: Periodontitis stage I, II, III, IV.

The relative frequencies of microorganisms were analyzed using Agglomerative Hierarchical Clustering (AHC) in order to classify cryptic microorganisms based on the microbial community makeup by patient group. The results of this study are depicted as a dendrogram (Figure 4), which illustrates the degree to which the community makeup differs and how the cryptic microorganisms clustered for each patient group. The ensuing G1 (H) and G2 (P) dendrograms revealed two major well-defined clusters. In cluster 2, *Staphylococcus* spp., *Cutibacterium* spp., and *Enterococcus* spp. were shared by both groups. The G3

(OSA) displayed four clusters while G4 (P-OSA) displayed three clusters, with *Bifidobacterium* spp. constituting clusters 1 in G3 (OSA) and 2 in G4 (P-OSA).



**Figure 4.** Microbial profiles dendrogram: clustering of oral cryptic microorganisms based on Euclidean distance dissimilarity matrix and agglomeration method of Ward (Agglomerative Hierarchical Clustering-AHC). The microorganisms present in two or more patient groups are indicated in color. Those without color were only identified in a group. C: cluster; G1 (H): healthy patients; G2 (P) periodontitis patients; G3: (OSA) OSA patients; G4 (P-OSA) periodontitis and OSA patients.

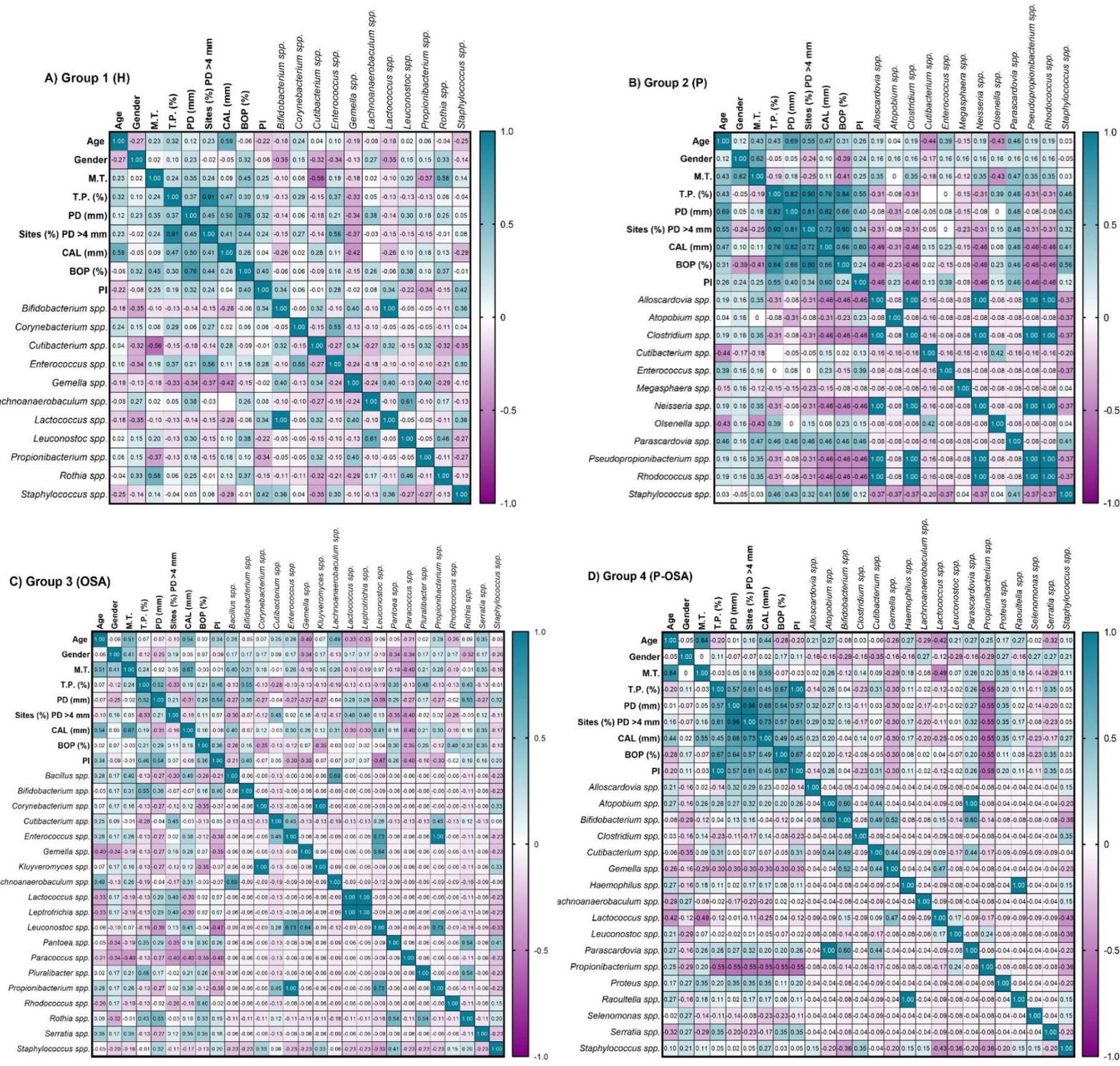
In order to observe the distributions of the cryptic microbiota in each category, PCoA was applied to the relative abundances in each group of patients (Figures S1-S4). The G1 scattering showed that component 1 was defined by *Bifidobacterium* spp., *Lactococcus* spp. and *Gemella* spp. and *Enterococcus* spp. and component 2 was defined by *Corynebacterium* spp. However, when applying marimax rotation this component is redefined with *Lachnoanaerobaculum* spp. and *Leuconostoc* spp., being the distribution for men and women associated with component 1. The G2 scattering showed that *Alloscardovia* spp., *Clostridium* spp., *Neisseria* spp. and *Pseudopropionibacterium* spp. highly defined the first component and the component is mainly defined by *Staphylococcus* spp. and this was maintained in rotation, being directly associated with those of stage II periodontitis. The G3 scattering showed that component 1 was defined by *Enterococcus* spp., *Leuconostoc* spp. and *Propionibacterium* spp., being highly associated with moderate OSA and inversely with severe OSA and

women. Component 2 was mainly defined by *Staphylococcus* spp. and this is maintained in rotation, being inversely associated with men. The G4 scattering showed that *Haemophilus* spp., *Staphylococcus* spp., and *Raoultella* spp. defined component 2 with an associative pattern for men and women while *Bifidobacterium* spp., *Atopobium* spp., and *Parascardovia* spp. defined component 1 with an association trend with severe OSA.

### 3.3 Association between periodontal parameters and the cryptic oral microorganisms

This correlation between periodontal parameters and the genera of species of cryptic microorganisms present in the four groups of patients evaluated was found by the analysis of multicomponent matrices. The association can be positive (+) or negative (-) according to the Spearman correlation range ( $r_s$ ). The  $r_s$  values over zero indicate a positive correlation in cyan tones, whereas  $r_s$  values below zero indicate a negative correlation in purple tones. (Figure 5 - Table S2).

In G1 (H) there was a positive, statistically significant correlation between the missing teeth and *Rothia* spp. ( $r_s=0.58$ ,  $p=0.004$ ), PD (mm) and *Lachnoanaerobaculum* spp. ( $r_s=0.38$ ,  $p=0.047$ ), Sites (%) PD >4 mm and *Enterococcus* spp. ( $r_s=0.56$ ,  $p=0.006$ ), BOP (%) and *Leuconostoc* spp. ( $r_s=0.38$ ,  $p=0.048$ ), and PI and *Staphylococcus* spp. ( $r_s=0.42$ ,  $p=0.032$ ), whereas *Cutibacterium* spp. and the missing teeth, and CAL and *Gemella* spp. showed a negative statistical significant correlation ( $r_s=-0.56$ ,  $p=0.005$  and  $r_s=-0.42$ ,  $p=0.032$ , respectively) (Figure 5A). In G2 (P), there was a positive correlation with statistical significance between *Staphylococcus* spp. and Teeth with periodontitis (%) ( $r_s=0.46$ ,  $p=0.056$ ), as with BOP (%) ( $r_s=0.56$ ,  $p=0.026$ ) (Figure 5B). In G3 (OSA), there was a positive correlation with statistical significance between age and *Lachnoanaerobaculum* spp. ( $r_s=0.49$ ,  $p=0.018$ ), Missing teeth and *Bacillus* spp. ( $r_s=0.40$ ,  $p=0.05$ ), Teeth with periodontitis (%) and *Bifidobacterium* spp. ( $r_s=0.55$ ,  $p=0.01$ ), as with *Pluralibacter* spp. and *Rothia* spp. ( $r_s=0.48$ ,  $p=0.022$ ;  $r_s=0.43$ ,  $p=0.036$ , respectively). Also, PD (mm) and *Rothia* spp. ( $r_s=0.36$ ,  $p=0.011$ ), Sites (%) PD >4 mm and *Cutibacterium* spp. ( $r_s=0.45$ ,  $p=0.03$ ). Whereas there was a negative statistical significance between *Leuconostoc* spp. and PD (mm) ( $r_s=-0.39$ ,  $p=0.046$ ), as PI ( $r_s=-0.47$ ,  $p=0.023$ ) (Figure 5C). In G4 (P-OSA), *Proteus* spp. was correlated positively with the missing teeth, PD (mm) and Sites (%) PD >4 mm ( $r_s=0.56$ ,  $p=0.026$ ). *Serratia* spp. was correlated positively with teeth with periodontitis (%), BOP (%) and PI ( $r_s=0.35$ ,  $p=0.048$ ). Whereas *Lactococcus* spp. was correlated negatively with the missing teeth ( $r_s=-0.49$ ,  $p=0.008$ ) and *Propionibacterium* spp. with all the periodontal parameters ( $r_s=-0.55$ ,  $p=0.003$ ) (Figure 5D).



**Figure 5.** Multicomponent matrix for the correlation of periodontal parameters and oral cryptic microorganisms present in each of the groups of patients evaluated: A) G1 (H), n=20; B) G2 (P), n=13; C) G3 (OSA), n=18; D) G4 (P-OSA), n=24, using the Spearman's rank correlation coefficient  $r_s > 0.30$ ,  $p < 0.05$ . The association can be positive (+) or negative (-) according to the Spearman correlation range ( $r_s$ ). The  $r_s$  values over zero indicate a positive correlation in cyan tones, whereas  $r_s$  values below zero indicate a negative correlation in purple tones. M.T., Missing teeth; T.P., Teeth with periodontitis; PD: probing depth; CAL: clinical attachment loss; BOP: bleeding of probing; PI: plaque index.

#### 4. Discussion

This study is the first to analyze the presence of microorganisms found in oral samples that are not often linked to oral diseases such as periodontitis related to OSA. A previous research [9] demonstrated that there was a greater diversity of microorganisms in oral samples from individuals with periodontitis and OSA, and showed the association between both diseases by sharing risk factors such as comorbidities and presence of the bacteria of the orange and red complexes, associated with *Candida albicans*. Additionally, this study identified cultivable cryptic microorganisms in healthy individuals (G1), individuals with periodontitis (G2), individuals with OSA (G3) and individuals with periodontitis and with OSA (G4), that we describe as those microorganisms that are not often considered as significant oral pathogens and neither classified in the Socransky's microbial complexes [10], nor associated with specific pathologies in the oral cavity. Although, they could be cryptic microorganisms that contribute to the periodontitis development in OSA patients and could be associated to other chronic pathologies. It is crucial to clarify the role and modulation of this diverse group of microorganisms [11–18], which goes beyond those first identified as periodontal pathogens, in both disease and pathogenicity, taking into consideration the effectiveness of current identification technologies. It is important to consider that cryptic microorganisms may contribute to health and the emergence of periodontal disease or may not. To clarify the role of these microorganisms, the purpose of the current study was to analyze their presence in health or disease (periodontitis and/or OSA) and to identify potential pathogens.

The periodontal parameters BOP (%) and PI were highest in all of groups of patients, compare to patients of G1, highlighting that the patients of G3 had an increased PI with statistically significant difference ( $p=0.001$ ). This evidence suggests that OSA might favor the oral biofilm formation, and is consistent with previous reports [19,20]. Also, other study demonstrated that periodontal parameters as PD (mm) and CAL were higher in patients with OSA [4], supporting the idea that OSA pathophysiology, that include hypoxia, hypercapnia, and oral dryness, can be contributing to develop periodontitis.

According to the new Classification of Periodontal and Peri-implant Diseases and Conditions [21], the periodontal condition of the patients of each group was determined. The Gingivitis -Dental biofilm-induced in reduced periodontium was more frequent in G1 and G3 (56% and 79%, respectively). Meanwhile, Periodontitis Stage III was more frequent in G2 and G4 (65% and 85%, respectively). These results suggest that OSA is a factor that increase the risk of periodontitis and it is crucial to comprehend that OSA individuals should undergo periodic periodontal screenings. The percentage of relative frequency of each species of cultivable oral cryptic microorganisms showed that Gingivitis -Dental biofilm-induced in reduced periodontium as a healthy condition and the stage III of periodontitis as disease condition, had the major diversity of species. There was evidence that the presence and absence of certain species vary depending on health and disease conditions, as a possible modulation between the cryptic microorganisms.

According to cluster conformation, periodontitis and OSA affect the diversity and distribution of cryptic microorganisms and unique genera in each group formed majority clusters; In G2 (P), *Olsenella* spp. and *Megasphaera* spp.: both genera have been reported as periodontal pathogens. *M. micronuciformis* has been isolated from women suffering from preterm birth [22,23]; In G3 (OSA), *Paracoccus* spp., *Pantoea* spp., *Pluralibacter* spp. and *Bacillus* spp.: These microorganisms are related to biofilm formation, abscess, bacteremia, pneumonia, urinary tract infection, septic arthritis, osteomyelitis, peritonitis, choledocholithiasis, dacryocystitis, and endophthalmitis [24]. *Bacillus* sp. have pathogenic potential by the production of enterotoxins [12,25,26]; and in G4 (P-OSA), *Selenomonas* spp. and *Proteus* spp.: they are related to pathogenesis of periodontal disease [27] and Catheter-associated urinary tract infections (CAUTIs) [28], respectively. Furthermore, other microorganisms were found in each group, but in small clusters: *Neisseria* spp. (G2), this genus is related to endocarditis and osteomyelitis [29], *N. oralis* was identified in systemic infection

and cystitis in a diabetic adult [30]. *Leptotrichia* sp. (G3) has been related to periodontal disease and abscesses of the oral cavity, endocarditis and septicemia [31], and *Kluyveromyces* sp. (G3), a yeast producer of pGK1 killer toxin [32]. *Raoultella* spp. (G4) has been related to infected root canals and urinary, gastrointestinal, hepatobiliary, osteoarticular infections [33], and *Haemophilus* spp. (G4) has been related to endocarditis, meningitis, pneumonia, otitis media, sinusitis and epiglottitis [34]. The PCoA identified that *Alloscordavia* spp., *Clostridium* spp., *Neisseria* spp. and *Staphylococcus* spp. were associated to Periodontitis stage III in G2 (P). *Alloscordavia* spp. has been related to dental caries [11] and *Clostridium* spp. is related to gas gangrene, bacteremia, meningitis, septic arthritis, enterocolitis, spontaneous bacterial peritonitis, post-traumatic brain abscess, and pneumonia [13,14]. *Enterococcus* spp., *Leuconostoc* spp. and *Propionibacterium* spp. were associated to Moderate OSA in G3 (OSA). *Enterococcus* spp., is related to endodontic disease, bacteremia, endocarditis, urinary tract infections, diabetic foot ulcers, and cholecystitis [17,18], and *Leuconostoc* spp. is related to bacterial meningitis and bacteremia [35–37]. These results suggest that severe conditions of periodontitis and OSA may harbor microorganisms that favor the development of systemic infectious disease.

Regarding the cryptic microorganisms identified (Table S3) [15,16,38–44], *Staphylococcus* spp. and *Cutibacterium* spp. were the genera more common in all patient groups. However, it was clear that their abundance was higher in G1 compared to the other patients' groups. *Staphylococcus* spp. is frequent in skin and soft tissue infections, bloodstream infections, endocarditis, osteomyelitis, lung infection, suppurative diseases, pneumonia, prosthetic joint infections and toxic shock syndrome [45–53]. *S. aureus* stands out in this genus, considered an opportunistic pathogen that is a part of the skin and nasal microbiota. It has been related to infectious disease of the oral cavity, as periodontitis [22], coinciding with the positive correlation found between *Staphylococcus* spp. and TP(%) and with BOP(%) in G2, and an in vitro study determined that *S. aureus* has the ability to bind to periodontal pathogenic bacteria, as *F. nucleatum* and *P. gingivalis*; supporting the idea that *S. aureus* can become part of the complex oral microbiota and contribute to the development of oral infections [54].

The presence of *S. epidermidis* and *S. hominis* are all associated with skin conditions like atopic dermatitis or psoriasis, bloodstream infections including endocarditis and peritonitis, osteomyelitis, and infections of the bones and joints [48,49]. Both bacteria had been isolated from subgingival samples of healthy and periodontitis patients, without significant differences between both conditions [55], also supported by our results. The current study found that patients with periodontitis and OSA had lower levels of *Staphylococcus* spp. A highly diverse of microbial community that influences the microenvironment and controls this bacterium's growth may be responsible for this bacterium's decline. Furthermore, a study found that *Staphylococcus* spp. increased during treatment with continuous positive airway pressure (CPAP), the primary therapy for OSA patients [56]. This finding might explain the increase of *Staphylococcus* spp. in healthy patients due to the restored microenvironment that favors its growth.

*Gemella* spp. is mainly linked to poor dental health, dental manipulation or surgery, colorectal disease or procedures, steroid therapy, diabetes mellitus, or hepatocellular dysfunction [57,58]. Moreover, *Gemella* spp. is also linked to septic arthritis and oral abscesses, which can result in serious endovascular infections like endocarditis and pericarditis [59]. CPAP-using OSA patients were shown to have lower levels of this bacterium [56]. In contrast, the healthy patients (G1) in the current study had a higher prevalence of these bacteria than the other patient groups. Like this, a study found a higher proportion of *Gemella* sp. in saliva from healthy patients than the saliva obtained from periodontitis patients. This study demonstrated that *Gemella* sp. in saliva is linked to periodontal health and the protein components in *Gemella*'s culture supernatant directly inhibited *P. gingivalis*'s growth in vitro [60].

To compare the findings with previous studies, *Cutibacterium* spp. and *Propionibacterium* spp. [61] were analyzed independently in the current study. Infections in neurosurgical shunts, bone, breast, and prostate infections, infective endocarditis, splenic and cutaneous abscesses, and chronic blepharitis and endophthalmitis are all linked to *Cutibacterium* spp. [62–64]. *C. acnes* is recognized for its ability to form biofilms on biomaterials in implanted medical device, as *C. albicans* [65]. These two microorganisms can establish polymicrobial biofilms that are synergistic, which enhances yeast resistance to micafungin [66]. The frequency of *C. albicans* and the percentage of biofilm were both higher in patients with periodontitis and OSA [9], in this scenario, *C. acnes* could participate as *C. albicans*' protector, encouraging the development of dysbiotic biofilms. This might lend to the notion that these opportunistic microorganisms behave in a way that makes it possible for periodontopathogen bacteria to colonize in periodontitis linked to OSA.

*Propionibacterium* spp. live on human skin as well as in the gastrointestinal and oral mucosa [67,68]. They have been linked to endocarditis and the infection of both natural and artificial valves [69], and is present in endodontic infections with a higher prevalence for secondary endodontic lesions [70]. Additionally, *P. acnes* plays a role in the onset and development of Alzheimer's disease (AD) [71] and Parkinson's disease (PD) [72]. *P. acnes* has been demonstrated to be able to cross the blood-brain barrier through transcellular invasion in an in vitro study [73]. In the current study, *Propionibacterium* spp. was identified in patients with OSA and patients with periodontitis and OSA, increasing 0.61% in G3, and 4.5% in G4. The presence of this microorganism in individuals with OSA and those who also had periodontitis and OSA is reported for the first time in this study. Other study determined the presence of *Propionibacterium* spp. in apical periodontitis, forming a network of interactions with *Lactobacillus* spp. and different species of *Streptococcus* [74]. It is crucial to emphasize the link between the presence of this microorganism in co-association with other periodontal pathogens in periodontitis and OSA. According to reports [75,76], OSA is associated with chronic diseases such as AD and PD, primarily due to its potential neurodegenerative effects. In a similar vein, studies have suggested that periodontitis and AD may have a potential bidirectional relationship, that may be caused by microorganisms [77]. The present study opens the possibility of establishing microbiological implications; for instance, the presence of *P. acnes* may play a role in the development and progression of this type of neurological disease in OSA patients.

*Bifidobacterium* spp. is usually related with dental carious, by acidogenic potential [78,79] and is well-adapted as commensal in the gastrointestinal tract. An earlier study determined that *Bifidobacterium dentium* is associated with an increased risk for Gastrointestinal Cancer [80], and along with *Lactobacillus*, *Bacteroides*, and *Prevotella*, was detected in OSA patients. It had also been implicated in obesity and diabetes [81]. Moreover, it had found a substantial correlation between *B. dentium* abundance and pregnant women's salivary progesterone concentrations [82] and also, pregnant women who had *B. dentium* had greater levels of IL-6 and IL-8 [83]. The G3 and G4 had the highest frequency of *Bifidobacterium* sp. compared to G1 and G2, and it was positively correlated with teeth with periodontitis (%) and PI in G3 and the presence of *Cutibacterium* spp. in G4, indicating its pathogenic effect in both diseases, and this microorganisms with *Atopobium* spp. [84,85], and *Parascardovia* spp. [86] showed association trend with severe OSA.

Despite these findings, more research will be required to fully understand the connections between cryptic oral microorganisms and known oral pathogens in periodontitis associated with OSA, as well as their participation as risk factors.

## 5. Conclusions

This study reveals the existence of cryptic microorganisms and confirms that some of them are connected to a microbial profile of health while others are more connected to

a microbial profile of sickness. While *Gemella* spp. were connected to healthy, *Cutibacteria* spp., *Propionibacterium* spp., and *Bifidobacterium* spp. were connected to disease profiles. *Staphylococcus* spp. prevalence in all patient categories may vary depending on the presence of other microorganisms. It was discovered that the illness profile contained microorganisms that might be categorized as possibly pathogenic and may have a role in the interaction between OSA, periodontitis, and other clinically significant chronic disorders or systemic infectious diseases. Understanding whether and how they will respond to health and disease is based on this reality.

**Supplementary Materials:** The following supporting information can be downloaded at: [www.mdpi.com/xxx/s1](http://www.mdpi.com/xxx/s1), Figure S1: Principal Coordinates Analysis (PCoA) calculated by the relative abundance of microorganisms in G1 (H). Figure S2: Principal Coordinates Analysis (PCoA) calculated by the relative abundance of microorganisms in G2 (P). Figure S3: Principal Coordinates Analysis (PCoA) calculated by the relative abundance of microorganisms in G3 (OSA). Figure S4: Principal Coordinates Analysis (PCoA) calculated by the relative abundance of microorganisms in G4 (P-OSA). Table S1: Species identified in each of group of patients. Table S2: Positive and negative correlations between the genera of microorganisms and periodontal parameters in each group of patients. Table S3: Identification of cryptic microorganism species and their relation to disease.

**Author Contributions:** Conceptualization, M.A.T.C., E.H.D. and C.M.P.G.; methodology, M.A.T.C.; software, M.A.T.C. and E.H.D.; validation, M.A.T.C., N.S.R.M., L.O. M.E.C. and C.M.P.G.; formal analysis, M.A.T.C. and E.H.D.; resources, M.A.T.C., H.K.C.J., M.M.B., J.C.V.P and P.H.M.; data curation, M.A.T.C., H.K.C.J., M.M.B. and J.C.V.P.; writing—original draft preparation, M.A.T.C.; writing—review and editing, M.A.T.C., N.S.R.M., L.O. M.E.C. and C.M.P.G.; visualization, C.M.P.G.; supervision, N.S.R.M., L.O. M.E.C. and C.M.P.G.; project administration, M.A.T.C.; funding acquisition, M.A.T.C. and L.O. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by Pontificia Universidad Javeriana, grant number 008398.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Acknowledgments:** We greatly appreciate the support given by the Training Agreement for post-graduate studies of professors of the Pontificia Universidad Javeriana for their doctoral training.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

- (1) Khodadadi, N.; Khodadadi, M.; Zamani, M. Is Periodontitis Associated with Obstructive Sleep Apnea? A Systematic Review and Meta-Analysis. *J. Clin. Exp. Dent.* **2022**, *14* (4), 359–365. <https://doi.org/10.4317/jced.59478>.
- (2) Verhelst, A. R. E.; Kosho, M. X. F.; Aarab, G.; Loos, B. G. Screening for the Risk of OSA in Periodontitis Patients . A Pilot Study. *Oral Health Prev. Dent.* **2022**, *20*, 243–252. <https://doi.org/10.3290/j.ohpd.b3125665>.
- (3) Al-Jewair, T. S.; Al-Jasser, R.; Almas, K. Periodontitis and Obstructive Sleep Apnea's Bidirectional Relationship: A Systematic Review and Meta-Analysis. *Sleep Breath* **2015**, *19* (4), 1111–1120. <https://doi.org/10.1007/s11325-015-1160-8>.
- (4) Seo, W. H.; Cho, E. R.; Thomas, R. J.; An, S. Y.; Ryu, J. J.; Kim, H.; Shin, C. The Association between Periodontitis and Obstructive Sleep Apnea: A Preliminary Study. *J. Periodontal Res.* **2013**, *48* (4), 500–506. <https://doi.org/10.1111/jre.12032>.
- (5) Nizam, N.; Tasbakan, M. S.; Basoglu, O. K.; Lappin, D. F.; Buduneli, N. Is There an Association between Obstructive Sleep Apnea Syndrome and Periodontal Inflammation? *Clin. Oral Investig.* **2016**, *20* (4), 659–668. <https://doi.org/10.1007/s00784-015-1544-y>.
- (6) Wu, B. G.; Sulaiman, I.; Wang, J.; Shen, N.; Clemente, J. C.; Li, Y.; Laumbach, R. J.; Lu, S.; Udasin, I.; Le-hoang, O.; Perez, A.; Alimokhtari, S.; Black, K.; Plietz, M.; Twumasi, A.; Sanders, H.; Malecha, P.; Kapoor, B.; Scaglione, B. D.; Wang, A.; Blazoski, C.; Weiden, M. D.; Rapoport, D. M.; Harrison, D.; Chitkara, N. Severe Obstructive Sleep Apnea Is Associated with Alterations

- in the Nasal Microbiome and an Increase in Inflammation. *Am. J. Respir. Crit. Care Med.* **2018**, *199*, 99–109. <https://doi.org/10.1164/rccm.201801-0119OC>.
- (7) Ko, C.; Hu, A.; Huang, D. C. L.; Su, H.; Yan, F.; Zhang, X.; Zhang, H.; Zeng, Y. Analysis of Oral Microbiota in Patients with Obstructive Sleep Apnea-Associated Hypertension. *Hypertens. Res.* **2019**, *42*, 1692–1700. <https://doi.org/10.1038/s41440-019-0260-4>.
- (8) Chen, Y.; Chen, X.; Huang, X.; Duan, Y.; Gao, H.; Gao, X. Analysis of Salivary Microbiome and Its Association With Periodontitis in Patients With Obstructive Sleep Apnea. *Front. Cell. Infect. Microbiol.* **2021**, *11* (December), 1–11. <https://doi.org/10.3389/fcimb.2021.752475>.
- (9) Tellez Corral, M. A.; Herrera Daza, E.; Cuervo Jimenez, H. K.; Arango Jimenez, N.; Morales Vera, D. Z.; Velosa Porras, J.; Latorre Uriza, C.; Escobar Arregoces, F. M.; Hidalgo Martínez, P.; Cortes, M. E.; Roa Molina, N. S.; Otero Mendoza, L. M.; Parra Giraldo, C. M. Patients with Obstructive Sleep Apnea Can Favor the Predisposing Factors of Periodontitis by the Presence P. Melaninogenica and C. Albicans, Increasing the Severity of the Periodontal Disease. *Front. Cell. Infect. Microbiol.* **2022**, *Sept.*, 1–15. <https://doi.org/10.3389/fcimb.2022.934298>.
- (10) Socransky, S. S.; Haffajee, A. D.; Cugini, M. A.; Smith, C.; Kent, R. L. Microbial Complexes in Subgingival Plaque. *J. Clin. Periodontol.* **1998**, *25* (2), 134–144. <https://doi.org/10.1111/j.1600-051X.1998.tb02419.x>.
- (11) Ogawa, Y.; Koizumi, A.; Kasahara, K.; Lee, S. T.; Yamada, Y.; Nakano, R.; Yano, H.; Mikasa, K. Bacteremia Secondary to *Alloscardovia Omnicolens* Urinary Tract Infection. *J. Infect. Chemother.* **2016**, *22* (6), 424–425. <https://doi.org/10.1016/j.jiac.2015.12.013>.
- (12) Stenfors, L. P.; Mayr, R.; Scherer, S.; Granum, P. E. Pathogenic Potential of Fifty *Bacillus Weihenstephanensis* Strains. *FEMS Microbiol. Lett.* **2002**, *215* (1), 47–51. [https://doi.org/10.1016/S0378-1097\(02\)00891-1](https://doi.org/10.1016/S0378-1097(02)00891-1).
- (13) Sárvári, K. P.; Schoblocher, D. The Antibiotic Susceptibility Pattern of Gas Gangrene-Forming *Clostridium* Spp. Clinical Isolates from South-Eastern Hungary. *Infect. Dis. (Auckl)*. **2020**, *52* (3), 196–201. <https://doi.org/10.1080/23744235.2019.1696472>.
- (14) Milano, V.; Biehle, L.; Patel, S.; Hammer, J. *Clostridium Tertium* Bacteremia and Hepatic Abscess in a Non-Neutropenic Patient. *IDCases* **2019**, *15*, e00510. <https://doi.org/10.1016/j.idcr.2019.e00510>.
- (15) Qudeimat, M. A.; Alyahya, A.; Karched, M.; Behbehani, J.; Salako, N. O. Dental Plaque Microbiota Profiles of Children with Caries-Free and Caries-Active Dentition. *J. Dent.* **2021**, *104* (December 2019), 103539. <https://doi.org/10.1016/j.jdent.2020.103539>.
- (16) Alibi, S.; Ferjani, A.; Boukadida, J.; Cano, M. E.; Fernández-Martínez, M.; Martínez-Martínez, L.; Navas, J. Occurrence of *Corynebacterium Striatum* as an Emerging Antibiotic-Resistant Nosocomial Pathogen in a Tunisian Hospital. *Sci. Rep.* **2017**, *7* (1), 1–8. <https://doi.org/10.1038/s41598-017-10081-y>.
- (17) Chong, K. K. L.; Tay, W. H.; Janela, B.; Yong, A. M. H.; Liew, T. H.; Madden, L.; Keogh, D.; Barkham, T. M. S.; Ginhoux, F.; Becker, D. L.; Kline, K. A. *Enterococcus Faecalis* Modulates Immune Activation and Slows Healing during Wound Infection. *J. Infect. Dis.* **2017**, *216* (12), 1644–1654. <https://doi.org/10.1093/infdis/jix541>.
- (18) Tyrrell, G. J.; Turnbull, L. A.; Teixeira, L. M.; Lefebvre, J.; Carvalho, M. da G. S.; Facklam, R. R.; Lovgren, M. *Enterococcus Gilvus* Sp. Nov. and *Enterococcus Pallens* Sp. Nov. Isolated from Human Clinical Specimens. *J. Clin. Microbiol.* **2002**, *40* (4), 1140–1145. <https://doi.org/10.1128/JCM.40.4.1140-1145.2002>.
- (19) Loke, W.; Girvan, T.; Ingmundson, P.; Verrett, R.; Schoolfield, J.; Mealey, B. L. Investigating the Association Between Obstructive Sleep Apnea and Periodontitis. *J. Periodontol.* **2015**, *86* (2), 232–243. <https://doi.org/10.1902/jop.2014.140229>.
- (20) Gamsiz-Isik, H.; Kiyan, E.; Bingol, Z.; Baser, U.; Ademoglu, E.; Yalcin, F. Does Obstructive Sleep Apnea Increase the Risk for Periodontal Disease? A Case-Control Study. *J. Periodontol.* **2017**, *88* (5), 443–449. <https://doi.org/10.1902/jop.2016.160365>.
- (21) Caton, J.; Armitage, G.; Berglundh, T.; Chapple, I. L. C.; Jepsen, S.; Kornman, K.; L. Mealey, B.; Papapanou, P. N.; Sanz, M.; S. Tonetti, M. A New Classification Scheme for Periodontal and Peri-Implant Diseases and Conditions – Introduction and

- Key Changes from the 1999 Classification. *J. Clin. Periodontol.* **2018**, *45*, S1–S8. <https://doi.org/10.1111/jcpe.12935>.
- (22) Vieira Colombo, A. P.; Magalhães, C. B.; Hartenbach, F. A. R. R.; Martins do Souto, R.; Maciel da Silva-Boghossian, C. Periodontal-Disease-Associated Biofilm: A Reservoir for Pathogens of Medical Importance. *Microb. Pathog.* **2016**, *94*, 27–34. <https://doi.org/10.1016/j.micpath.2015.09.009>.
- (23) Sato, N.; Kakuta, M.; Hasegawa, T.; Yamaguchi, R.; Uchino, E.; Kobayashi, W.; Sawada, K.; Tamura, Y.; Tokuda, I.; Murashita, K.; Nakaji, S.; Imoto, S.; Yanagita, M.; Okuno, Y. Metagenomic Analysis of Bacterial Species in Tongue Microbiome of Current and Never Smokers. *npj Biofilms Microbiomes* **2020**, *6* (1), 1–9. <https://doi.org/10.1038/s41522-020-0121-6>.
- (24) Büyükcım, A.; Tuncer, Ö.; Gür, D.; Sancak, B.; Ceyhan, M.; Cengiz, A. B.; Kara, A. Clinical and Microbiological Characteristics of Pantoea Agglomerans Infection in Children. *J. Infect. Public Health* **2018**, *11* (3), 304–309. <https://doi.org/10.1016/j.jiph.2017.07.020>.
- (25) Morinaga, K.; Yoshida, K.; Takahashi, K.; Nomura, N.; Toyofuku, M. Peculiarities of Biofilm Formation by Paracoccus Denitrificans. *Appl. Microbiol. Biotechnol.* **2020**, *104* (6), 2427–2433. <https://doi.org/10.1007/s00253-020-10400-w>.
- (26) Freire, M. P.; De Oliveira Garcia, D.; Cury, A. P.; Spadão, F.; Di Gioia, T. S. R.; Francisco, G. R.; Bueno, M. F. C.; Tomaz, M.; De Paula, F. J.; De Faro, L. B.; Piovesan, A. C.; Rossi, F.; Levin, A. S.; David Neto, E.; Nahas, W. C.; Pierrotti, L. C. Outbreak of IMP-Producing Carbapenem-Resistant Enterobacter Gergoviae among Kidney Transplant Recipients. *J. Antimicrob. Chemother.* **2016**, *71* (9), 2577–2585. <https://doi.org/10.1093/jac/dkw165>.
- (27) Enigk, K.; Jentsch, H.; Rodloff, A. C.; Eschrich, K.; Stingu, C. S. Activity of Five Antimicrobial Peptides against Periodontal as Well as Non-Periodontal Pathogenic Strains. *J. Oral Microbiol.* **2020**, *12* (1). <https://doi.org/10.1080/20002297.2020.1829405>.
- (28) Armbruster, C. E.; Mobley, H. L. T.; Pearson, M. M.; Arbor, A.; States, U.; States, U. Pathogenesis of Proteus Mirabilis Infection. *EcoSal Plus.* **2018**, *8* (1), 1–123. <https://doi.org/10.1128/ecosalplus.ESP-0009-2017.Pathogenesis>.
- (29) Spielman, A. F.; Ghumman, A.; Panthaki, Z.; Lessard, A. S. Neisseria Elongata Osteomyelitis: Literature Review and Case Report in a 63-Year-Old Male Presenting with Progressive Right-Handed Redness, Swelling and Pain. *Int. J. Surg. Case Rep.* **2020**, *73*, 228–230. <https://doi.org/10.1016/j.ijscr.2020.07.022>.
- (30) Baniulyte, G.; Svirpliene, S.; Eccleston, A.; Arjunan, S.; Connor, M. Neisseria Oral Septicaemia in a Newborn: First Recorded Case. *Paediatr. Int. Child Health* **2021**, *41* (3), 226–227. <https://doi.org/10.1080/20469047.2020.1826780>.
- (31) Eribe, E. R. K.; Olsen, I. Leptotrichia Species in Human Infections II. *J. Oral Microbiol.* **2017**, *9* (1). <https://doi.org/10.1080/20002297.2017.1368848>.
- (32) Spohner, S. C.; Schaum, V.; Quitmann, H.; Czermak, P. Kluyveromyces Lactis: An Emerging Tool in Biotechnology. *J. Biotechnol.* **2016**, *222*, 104–116. <https://doi.org/10.1016/j.jbiotec.2016.02.023>.
- (33) Sękowska, A. Raoultella Spp. — Clinical Significance, Infections and Susceptibility to Antibiotics. *Folia Microbiol. (Praha)*. **2017**, *62* (3), 221–227. <https://doi.org/10.1007/s12223-016-0490-7>.
- (34) Finch, L. C.; Gerdzhikov, S.; Buttery, R. Haemophilus Parainfluenzae Endocarditis Presenting with Symptoms of COVID-19. *BMJ Case Rep.* **2021**, *14* (8), 4–9. <https://doi.org/10.1136/bcr-2021-245210>.
- (35) Modaweb, A.; Mansoor, Z.; Alsarhan, A.; Abuhammour, W. A Case of Successfully Treated Central Line-Associated Bloodstream Infection Due to Vancomycin-Resistant Leuconostoc Citreum in a Child With Biliary Atresia. *Cureus* **2022**, *14* (1), 12–15. <https://doi.org/10.7759/cureus.21227>.
- (36) Omori, R.; Fujiwara, S.; Ishiyama, H.; Kuroda, H.; Kohara, N. Leuconostoc Lactis- A Rare Cause of Bacterial Meningitis in an Immunocompromised Host. *Intern. Med.* **2020**, *59* (22), 2935–2936. <https://doi.org/10.2169/internalmedicine.5076-20>.
- (37) Meneguetti, M. G.; Gaspar, G. G.; Laus, A. M.; Basile-Filho, A.; Bellissimo-Rodrigues, F.; Auxiliadora-Martins, M. Bacteremia by Leuconostoc Mesenteroides in an Immunocompetent Patient with Chronic Chagas Disease: A Case Report. *BMC Infect. Dis.* **2018**, *18* (1), 13–15. <https://doi.org/10.1186/s12879-018-3452-7>.
- (38) Grenier, D. Porphyromonas Gingivalis Outer Membrane Vesicles Mediate Coaggregation and Piggybacking of Treponema

- Denticola and Lachnoanaerobaculum Saburreum. *Int. J. Dent.* **2013**, *2013*. <https://doi.org/10.1155/2013/305476>.
- (39) Meyburgh, C. M.; Bragg, R. R.; Boucher, C. E. Lactococcus Garvieae: An Emerging Bacterial Pathogen of Fish. *Dis. Aquat. Organ.* **2017**, *123* (1), 67–79. <https://doi.org/10.3354/dao03083>.
- (40) Baba, H.; Nada, T.; Ohkusu, K.; Ezaki, T.; Hasegawa, Y.; Paterson, D. L. First Case of Bloodstream Infection Caused by Rhodococcus Erythropolis. *J. Clin. Microbiol.* **2009**, *47* (8), 2667–2669. <https://doi.org/10.1128/JCM.00294-09>.
- (41) Greve, D.; Moter, A.; Kleinschmidt, M. C.; Pfäfflin, F.; Stegemann, M. S.; Kursawe, L.; Grubitzsch, H.; Falk, V.; Kikhney, J. Rothia Aeria and Rothia Dentocariosa as Biofilm Builders in Infective Endocarditis. *Int. J. Med. Microbiol.* **2021**, *311* (2). <https://doi.org/10.1016/j.ijmm.2021.151478>.
- (42) Franconieri, F.; Join-Lambert, O.; Creveuil, C.; Auzou, M.; Labombarda, F.; Aouba, A.; Verdon, R.; de La Blanchardière, A. Rothia Spp. Infective Endocarditis: A Systematic Literature Review. *Med. Mal. Infect.* **2020**, *51*, 228–235. <https://doi.org/10.1016/j.medmal.2020.10.021>.
- (43) Uranga, C. C.; Arroyo, P.; Gerwick, W.; Edlund, A. Commensal Oral Rothia Mucilaginosa Produces Enterobactin, a Metal-Chelating Siderophore. *Host-Microbe Biol.* **2020**, *5* (2), 1–14.
- (44) Begrem, S.; Jérôme, M.; Leroi, F.; Delbarre-Ladrat, C.; Grovel, O.; Passerini, D. Genomic Diversity of Serratia Proteamaculans and Serratia Liquefaciens Predominant in Seafood Products and Spoilage Potential Analyses. *Int. J. Food Microbiol.* **2021**, *354* (July). <https://doi.org/10.1016/j.ijfoodmicro.2021.109326>.
- (45) Jiang, B.; You, B.; Tan, L.; Yu, S.; Li, H.; Bai, G.; Li, S.; Rao, X.; Xie, Z.; Shi, X.; Peng, Y.; Hu, X. Clinical Staphylococcus Argenteus Develops to Small Colony Variants to Promote Persistent Infection. *Front. Microbiol.* **2018**, *9* (JUN), 1–10. <https://doi.org/10.3389/fmicb.2018.01347>.
- (46) Ahmad-Mansour, N.; Loubet, P.; Pouget, C.; Dunyach-Remy, C.; Sotto, A.; Lavigne, J. P.; Molle, V. Staphylococcus Aureus Toxins: An Update on Their Pathogenic Properties and Potential Treatments. *Toxins (Basel)*. **2021**, *13* (10), 1–22. <https://doi.org/10.3390/toxins13100677>.
- (47) Cui, B.; Smooker, P. M.; Rouch, D. A.; Daley, A. J.; Deighton, M. A. Differences between Two Clinical Staphylococcus Capitis Subspecies as Revealed by Biofilm, Antibiotic Resistance, and Pulsed-Field Gel Electrophoresis Profiling. *J. Clin. Microbiol.* **2013**, *51* (1), 9–14. <https://doi.org/10.1128/JCM.05124-11>.
- (48) Brescò, M. S.; Harris, L. G.; Thompson, K.; Stanic, B.; Morgenstern, M.; O'Mahony, L.; Richards, R. G.; Moriarty, T. F. Pathogenic Mechanisms and Host Interactions in Staphylococcus Epidermidis Device-Related Infection. *Front. Microbiol.* **2017**, *8*, 1401. <https://doi.org/10.3389/fmicb.2017.01401>.
- (49) Szczuka, E.; Krzysińska, S.; Bogucka, N.; Kaznowski, A. Multifactorial Mechanisms of the Pathogenesis of Methicillin-Resistant Staphylococcus Hominis Isolated from Bloodstream Infections. *Antonie van Leeuwenhoek, Int. J. Gen. Mol. Microbiol.* **2018**, *111* (7), 1259–1265. <https://doi.org/10.1007/s10482-017-1007-3>.
- (50) Parthasarathy, S.; Shah, S.; Raja Sager, A.; Rangan, A.; Durugu, S. Staphylococcus Lugdunensis: Review of Epidemiology, Complications, and Treatment. *Cureus* **2020**, *12* (6), 6–13. <https://doi.org/10.7759/cureus.8801>.
- (51) Wang, P.; Liu, Y.; Xu, Y.; Xu, Z. Staphylococcus Saccharolyticus Infection: Case Series with a PRISMA-Compliant Systemic Review. *Medicine (Baltimore)*. **2020**, *99* (26), e20686. <https://doi.org/10.1097/MD.00000000000020686>.
- (52) Kanuparth, A.; Challa, T.; Meegada, S.; Siddamreddy, S.; Muppidi, V. Staphylococcus Warneri: Skin Commensal and a Rare Cause of Urinary Tract Infection. *Cureus* **2020**, *12* (5). <https://doi.org/10.7759/cureus.8337>.
- (53) Lisowska-Lysiak, K.; Lauterbach, R.; Międzobrodzki, J.; Kosecka-Strojek, M. Epidemiology and Pathogenesis of Staphylococcus Bloodstream Infections in Humans: A Review. *Polish J. Microbiol.* **2021**, *70* (1), 13–23. <https://doi.org/10.33073/PJM-2021-005>.
- (54) Lima, B. P.; Hu, L. I.; Vreeman, G. W.; Weibel, D. B.; Lux, R. The Oral Bacterium Fusobacterium Nucleatum Binds Staphylococcus Aureus and Alters Expression of the Staphylococcal Accessory Regulator SarA. *Microb. Ecol.* **2019**, *78* (2), 336–

347. <https://doi.org/10.1007/s00248-018-1291-0>.
- (55) Murdoch, F. E.; Sammons, R. L.; Chapple, I. L. C. Isolation and Characterization of Subgingival Staphylococci from Periodontitis Patients and Controls. *Oral Dis.* **2004**, *10* (3), 155–162. <https://doi.org/10.1046/j.1601-0825.2003.01000.x>.
- (56) Ko, C. Y.; Hu, A. K.; Zhang, L.; Lu, X. L.; Zeng, Y. M. Alterations of Oral Microbiota in Patients with Obstructive Sleep Apnea–Hypopnea Syndrome Treated with Continuous Positive Airway Pressure: A Pilot Study. *Sleep Breath.* **2022**, *26* (2), 811–814. <https://doi.org/10.1007/s11325-021-02428-7>.
- (57) Taimur, S.; Madiha, R.; Samar, F.; Bushra, J. Gemella Morbillorum Endocarditis in a Patient with a Bicuspid Aortic Valve. *Hell. J. Cardiol.* **2010**, *51* (2), 183–186. <https://doi.org/10.14744/nci.2020.39206>.
- (58) Maraki, S.; Plevritaki, A.; Kofteridis, D.; Scoulica, E.; Eskitzis, A.; Gikas, A.; Panagiotakis, S. H. Bicuspid Aortic Valve Endocarditis Caused by Gemella Sanguinis: Case Report and Literature Review. *J. Infect. Public Health* **2019**, *12* (3), 304–308. <https://doi.org/10.1016/j.jiph.2019.01.001>.
- (59) Collins, M. D. The Genus Gemella. In *Prokaryotes*; Stackebrandt, E. D. M. F. S. R. E. S. K., Ed.; Springer, New York, NY, 2006; Vol. 6, pp 197–214.
- (60) Miyoshi, T.; Oge, S.; Nakata, S.; Ueno, Y.; Ukita, H.; Kousaka, R.; Miura, Y.; Yoshinari, N.; Yoshida, A. Gemella Haemolysans Inhibits the Growth of the Periodontal Pathogen Porphyromonas Gingivalis. *Sci. Rep.* **2021**, *11* (1), 1–12. <https://doi.org/10.1038/s41598-021-91267-3>.
- (61) Broly, M.; Ruffier d’Epenoux, L.; Guillouze, A.; Le Gargasson, G.; Juvin, M. E.; Leroy, A. G.; Bémer, P.; Corvec, S. Propionibacterium/Cutibacterium Species–Related Positive Samples, Identification, Clinical and Resistance Features: A 10-Year Survey in a French Hospital. *Eur. J. Clin. Microbiol. Infect. Dis.* **2020**, *39* (7), 1357–1364. <https://doi.org/10.1007/s10096-020-03852-5>.
- (62) Corvec, S. Clinical and Biological Features of Cutibacterium (Formerly Propionibacterium) Avidum, an Underrecognized Microorganism Stéphane. *Clin. microbiology Rev.* **2018**, *31* (3), 1–42.
- (63) Mayslich, C.; Grange, P. A. Cutibacterium Acnes as an Opportunistic Pathogen: An Update of Its Virulence-Associated Factors. *Microorganisms* **2021**, *9* (303), 1–21.
- (64) Singh, M.; Teles, F.; Uzel, N. G.; Papas, A. Characterizing Microbiota from Sjögren’s Syndrome Patients. *JDR Clin. Transl. Res.* **2021**, *6* (3), 324–332. <https://doi.org/10.1177/2380084420940623>.
- (65) Bernard, C.; Lemoine, V.; Hoogenkamp, M. A.; Girardot, M.; Krom, B. P.; Imbert, C. Candida Albicans Enhances Initial Biofilm Growth of Cutibacterium Acnes under Aerobic Conditions. *Biofouling* **2019**, *35* (3), 350–360. <https://doi.org/10.1080/08927014.2019.1608966>.
- (66) Bernard, C.; Renaudeau, N.; Mollichella, M. L.; Quellard, N.; Girardot, M.; Imbert, C. Cutibacterium Acnes Protects Candida Albicans from the Effect of Micafungin in Biofilms. *Int. J. Antimicrob. Agents* **2018**, *52* (6), 942–946. <https://doi.org/10.1016/j.ijantimicag.2018.08.009>.
- (67) McDowell, A.; McLaughlin, J.; Layton, A. M. Is Cutibacterium (Previously Propionibacterium) Acnes a Potential Pathogenic Factor in the Aetiology of the Skin Disease Progressive Macular Hypomelanosis? *J. Eur. Acad. Dermatology Venereol.* **2021**, *35* (2), 338–344. <https://doi.org/10.1111/jdv.16789>.
- (68) Suzuki, H.; Arshava, E. V.; Ford, B.; Nauseef, W. M. Don’t Let Its Name Fool You: Relapsing Thoracic Actinomycosis Caused by Pseudopropionibacterium Propionicum (Formerly Propionibacterium Propionicum). *Am. J. Case Rep.* **2019**, *20*, 1961–1965. <https://doi.org/10.12659/AJCR.919775>.
- (69) Clayton, J. J.; Baig, W.; Reynolds, G. W.; Sandoe, J. A. T. Endocarditis Caused by Propionibacterium Species: A Report of Three Cases and a Review of Clinical Features and Diagnostic Difficulties. *J. Med. Microbiol.* **2006**, *55* (8), 981–987. <https://doi.org/10.1099/jmm.0.46613-0>.
- (70) Dioguardi, M.; Alovisi, M.; Crincoli, V.; Aiuto, R.; Malagnino, G.; Quarta, C.; Laneve, E.; Sovereto, D.; Russo, L. Lo; Troiano,

- G.; Muzio, L. Lo. Prevalence of the Genus Propionibacterium in Primary and Persistent Endodontic Lesions: A Systematic Review. *J. Clin. Med.* **2020**, *9* (3), 1–30. <https://doi.org/10.3390/jcm9030739>.
- (71) Emery, D. C.; Shoemark, D. K.; Batstone, T. E.; Waterfall, C. M.; Coghill, J. A.; Cerajewska, T. L.; Davies, M.; West, N. X.; Allen, S. J. 16S rRNA next Generation Sequencing Analysis Shows Bacteria in Alzheimer's Post-Mortem Brain. *Front. Aging Neurosci.* **2017**, *9*, 1–13. <https://doi.org/10.3389/fnagi.2017.00195>.
- (72) Leheste, J. R.; Ruvolo, K. E.; Chrostowski, J. E.; Rivera, K.; Husko, C.; Miceli, A.; Selig, M. K.; Brüggemann, H.; Torres, G. P. Acnes-Driven Disease Pathology: Current Knowledge and Future Directions. *Front. Cell. Infect. Microbiol.* **2017**, *7*, 1–9. <https://doi.org/10.3389/fcimb.2017.00081>.
- (73) Lu, X.; Qi, X.; Yi, X.; Jian, Z.; Gao, T. Transcellular Traversal of the Blood-Brain Barrier by the Pathogenic Propionibacterium Acnes. *J. Cell. Biochem.* **2019**, *120* (5), 8457–8465. <https://doi.org/10.1002/jcb.28132>.
- (74) Korona-Glowniak, I.; Piatek, D.; Fornal, E.; Lukowiak, A.; Gerasymchuk, Y.; Kedziora, A.; Bugla-Płoskonska, G.; Grywalska, E.; Bachanek, T.; Malm, A. Patterns of Oral Microbiota in Patients with Apical Periodontitis. *J. Clin. Med.* **2021**, *10* (12), 1–14. <https://doi.org/10.3390/jcm10122707>.
- (75) Elfil, M.; Bahbah, E. I.; Attia, M. M.; Eldokmak, M.; Koo, B. B. Impact of Obstructive Sleep Apnea on Cognitive and Motor Functions in Parkinson's Disease. *Mov. Disord.* **2021**, *36* (3), 570–580. <https://doi.org/10.1002/mds.28412>.
- (76) Ferini-Strambi, L.; Hensley, M.; Salsone, M. Decoding Causal Links Between Sleep Apnea and Alzheimer's Disease. *J. Alzheimers. Dis.* **2021**, *80* (1), 29–40. <https://doi.org/10.3233/JAD-201066>.
- (77) Liccardo, D.; Marzano, F.; Carraturo, F.; Guida, M.; Femminella, G. D.; Bencivenga, L.; Agrimi, J.; Addonizio, A.; Melino, I.; Valletta, A.; Rengo, C.; Ferrara, N.; Rengo, G.; Cannavo, A. Potential Bidirectional Relationship Between Periodontitis and Alzheimer's Disease. *Front. Physiol.* **2020**, *11*, 1–13. <https://doi.org/10.3389/fphys.2020.00683>.
- (78) Manome, A.; Abiko, Y.; Kawashima, J.; Washio, J.; Fukumoto, S.; Takahashi, N. Acidogenic Potential of Oral Bifidobacterium and Its High Fluoride Tolerance. *Front. Microbiol.* **2019**, *10* (MAY), 1–10. <https://doi.org/10.3389/fmicb.2019.01099>.
- (79) Ventura, M.; Turrone, F.; Zomer, A.; Foroni, E.; Giubellini, V.; Bottacini, F.; Canchaya, C.; Claesson, M. J.; He, F.; Mantzourani, M.; Mulas, L.; Ferrarini, A.; Gao, B.; Delledonne, M.; Henrissat, B.; Coutinho, P.; Oggioni, M.; Gupta, R. S.; Zhang, Z.; Beighton, D.; Fitzgerald, G. F.; O'Toole, P. W.; Van Sinderen, D. The Bifidobacterium Dentium Bd1 Genome Sequence Reflects Its Genetic Adaptation to the Human Oral Cavity. *PLoS Genet.* **2009**, *5* (12). <https://doi.org/10.1371/journal.pgen.1000785>.
- (80) Wu, J.; Zhang, C.; Xu, S.; Xiang, C.; Wang, R.; Yang, D.; Lu, B.; Shi, L.; Tong, R.; Teng, Y.; Dong, W.; Zhang, J. Fecal Microbiome Alteration May Be a Potential Marker for Gastric Cancer. *Dis. Markers* **2020**, *2020*. <https://doi.org/10.1155/2020/3461315>.
- (81) Zhang, X.; Wang, S.; Xu, H.; Yi, H.; Guan, J.; Yin, S. Metabolomics and Microbiome Profiling as Biomarkers in Obstructive Sleep Apnoea: A Comprehensive Review. *Eur. Respir. Rev.* **2021**, *30* (160), 200–220. <https://doi.org/10.1183/16000617.0220-2020>.
- (82) Kato, S.; Nagasawa, T.; Uehara, O.; Shimizu, S.; Sugiyama, N.; Hasegawa-Nakamura, K.; Noguchi, K.; Hatae, M.; Kakinoki, H.; Furuichi, Y. Increase in Bifidobacterium Is a Characteristic of the Difference in the Salivary Microbiota of Pregnant and Non-Pregnant Women. *BMC Oral Health* **2022**, *22* (1), 1–13. <https://doi.org/10.1186/s12903-022-02293-0>.
- (83) Galley, J. D.; Mashburn-Warren, L.; Blalock, L. C.; Lauber, C. L.; Carroll, J. E.; Ross, K. M.; Hobel, C.; Coussons-Read, M.; Dunkel Schetter, C.; Gur, T. L. Maternal Anxiety, Depression and Stress Affects Offspring Gut Microbiome Diversity and Bifidobacterial Abundances. *Brain. Behav. Immun.* **2022**. <https://doi.org/10.1016/j.bbi.2022.10.005>.
- (84) Yachida, S.; Mizutani, S.; Shiroma, H.; Shiba, S.; Nakajima, T.; Sakamoto, T.; Watanabe, H.; Masuda, K.; Nishimoto, Y.; Kubo, M.; Hosoda, F.; Rokutan, H.; Matsumoto, M.; Takamaru, H.; Yamada, M.; Matsuda, T.; Iwasaki, M.; Yamaji, T.; Yachida, T.; Soga, T.; Kurokawa, K.; Toyoda, A.; Ogura, Y.; Hayashi, T.; Hatakeyama, M.; Nakagama, H.; Saito, Y.; Fukuda, S.; Shibata, T.; Yamada, T. Metagenomic and Metabolomic Analyses Reveal Distinct Stage-Specific Phenotypes of the Gut Microbiota in Colorectal Cancer. *Nat. Med.* **2019**, *25* (6), 968–976. <https://doi.org/10.1038/s41591-019-0458-7>.
- (85) Chen, J.; Wu, X.; Zhu, D.; Xu, M.; Yu, Y.; Yu, L.; Zhang, W. Microbiota in Human Periodontal Abscess Revealed by 16S rDNA

---

Sequencing. *Front. Microbiol.* **2019**, *10* (July), 1–12. <https://doi.org/10.3389/fmicb.2019.01723>.

- (86) Oshima, K.; Hayashi, J. I.; Toh, H.; Nakano, A.; Omori, E.; Hattori, Y.; Morita, H.; Honda, K.; Hattori, M. Complete Genome Sequence of *Scardovia Inopinata* JCM 12537T, Isolated from Human Dental Caries. *Genome Announc.* **2015**, *3* (3), 5–6. <https://doi.org/10.1128/genomeA.00481-15>.