

Brief Report

Epitope coverage of anti-SARS-CoV-2 nucleocapsid IgA and IgG antibodies correlates with protection against re-infection by new variants in subsequent waves of the COVID-19 pandemic

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Abstract: The COVID-19 pandemic continues to affect individuals across the globe, with some individuals experiencing more severe disease than others. The relatively high frequency of re-infections and breakthrough infections observed with SARS-CoV-2 highlights the importance of extending our understanding of immunity to COVID-19. Here we aim to shed light on the importance of antibody titers and epitope utilization in protection from re-infection. Health care workers are highly exposed to COVID-19 and are therefore also more likely to become re-infected. We utilized quantitative, multi-antigen, multi-epitope SARS-CoV-2 protein microarrays to measure IgG and IgA titers against various domains of the nucleocapsid and spike proteins. Potential re-infections in a large, diverse health care worker cohort (N=300) during the second wave of the pandemic were identified by assessing the IgG anti-N titers before and after the second wave. We assessed epitope coverage and antibody titers between the 'single infection' and 're-infection' groups. Clear differences were observed in the breadth of the anti-N response before the second wave, with the epitope coverage for both IgG ($p=0.019$) and IgA ($p=0.015$) being significantly increased in those who did not become re-infected compared to those who did. Additionally, the IgG anti-N ($p=0.004$) and anti-S titers ($p=0.018$) were significantly higher in those not re-infected. These results highlight the importance of the breadth of elicited antibody epitope coverage following natural infection in protection from re-infection and disease in the COVID-19 pandemic.

Keywords: immunoassay; epitope coverage; quantitative antibody binding; protein microarray; SARS-CoV-2 antibodies; humoral response; SARS-CoV-2 re-infection

1. Introduction

Despite extensive research, development and roll-out of vaccines to protect against COVID-19 disease, SARS-CoV-2 infections, re-infections, and breakthroughs have continued across the globe, albeit with the pandemic taking apparently different courses in different geographic regions. As part of efforts to curb the pandemic, it is important to gain a more complete understanding of the role of both the innate and adaptive immune responses in COVID-19 disease. COVID-19 presents as a spectrum of disease, with some

individuals experiencing more severe symptoms and a worse disease prognosis than others [1-3], suggesting that host factors play a key role in determining the outcome of infection. Amongst others, the humoral immune response between highly exposed individuals differs drastically in titers, epitope utilization, functionality, and longevity.

Furthermore, it is evident that previously infected individuals are becoming re-infected [4-10], suggesting: (1) a suboptimal immune response; (2) that some individuals are not mounting an immune response; (3) waning levels and decreased avidity [11-13] of antibodies post infection, as previously seen for SARS-CoV-1 [14] and MERS-CoV [15]; or (4) that the immune response is not protective across all the SARS-CoV-2 strains that have emerged. Identifying immunoglobulins that correlate with protection from infection or re-infection and disease may assist us in understanding the mechanisms behind an optimal immune response. This may also shed additional light on the reasons behind varying disease prognosis.

Many mono-epitope, qualitative antibody tests are available, however these cannot provide detailed information regarding the immune targets, how these vary between individuals and over time, and the functionality of the different immunoglobulins. Here, we used a high-throughput, quantitative SARS-CoV-2 multi-antigen, multi-epitope immunoassay to shed light on differential antibody responses in individuals who became re-infected in subsequent waves of the COVID-19 pandemic, compared to those who did not. This assay utilizes the KREX protein folding technology which uses biotin carboxyl carrier protein (BCCP) as a marker for correct folding and to immobilize individual domains of recombinant SARS-CoV-2 nucleocapsid and spike antigens [2, 16].

Healthcare workers (HCW) are assumed to be exposed to SARS-CoV-2 at a higher rate than the general population and are therefore considered to be at a higher risk of contracting COVID-19. Due to this increased exposure, it is also likely that these individuals represent an enriched cohort in which to study re-infection. Here we aimed to determine the rate of infection and re-infection amongst a South African HCW cohort and to use this information to identify IgG and IgA correlates of protection against re-infection.

2. Materials and Methods

2.1. Cohort information and sample collection

A prospective observational healthcare worker cohort was used for this study. The clinical characteristics of this cohort is summarized in Table 1. A total of 300 participants were enrolled from Groote Schuur, Victoria, and Somerset Hospitals. Blood samples were collected longitudinally from the enrolled participants. The Sisonke vaccine roll out of the Ad26.COV2.S vaccine to HCWs began in South Africa on 17 February 2021, which was post visit 6 (Figure 1).

Table 1: Characteristics of the COVID-19 HCW cohort.

Characteristic		
Gender	Female	220
	Male	77
	Not declared	3
Age distribution	18-30	89
	31-40	80
	41-60	105
	61-73	5
	Not declared	21
Ethnicity	African	107
	Caucasian	69
	Coloured	99
	Asian	16
	Other	3
	Not declared	6

Plasma was isolated from samples by centrifugation and aliquots stored at -80°C until needed. Ethics approval (HERC Ref 210/2020) for this study was obtained from the Human Research Ethics Committee of the Faculty of Health Sciences, University of Cape Town, South Africa. Full consent was obtained from all participants prior to sample collection.

Fifty pre-pandemic HIV positive serum samples were used as true negative controls. In this cohort, no additional clinical annotations were provided.

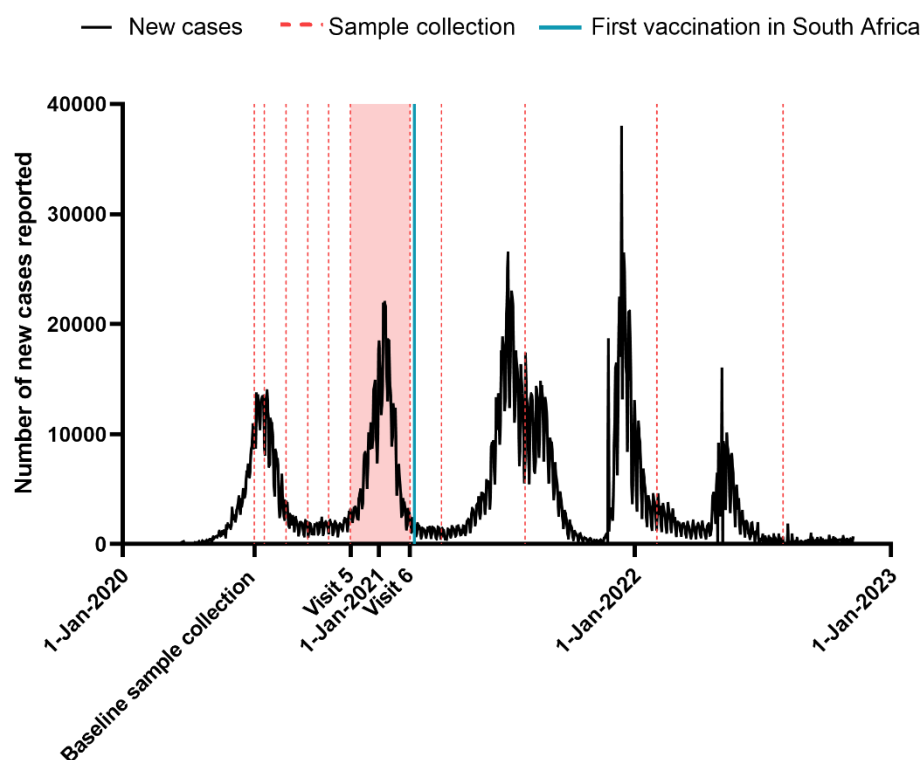


Figure 1: Number of COVID-19 cases in South Africa, between January 2020 and November 2023. Red dotted lines represent dates of sample collection from HCW cohort. The red shaded block represents the time period assessed in this study. The blue line represents start of the vaccine rollout in South Africa. (Data adapted from WHO [17]).

2.2. Protein Microarrays

SARS-CoV-2 multi-antigen, multi-epitope arrays containing component-resolved nucleocapsid (N) domains and intrinsically-disordered epitopes, as well as spike ectodomain trimer (S), S1 and RBD domains, were sourced commercially (ImmuSAFE array v4; Sengenics Corporation) and were used as per manufacturer's instructions.

2.3. Serological assays

Microarray slides were blocked in blocking buffer (20% Glycerol, 25 mM HEPES buffer (pH 7.4), 50 mM KCl, 1% Triton X-100, 1 mM DTT and 50 μM Biotin) for 1 h and washed 2×5 min in PBST (PBS, 0.2% Tween-20, pH 7.4) and 2×5 min in PBS. The slides were dried by centrifugation at $1200 \times g$ at 23°C for 2 min. Individual arrays were isolated using ProPlate 24 plex multi-well chambers (GraceBio-Labs, Bend, OR, USA). Prior to assays, plasma samples were incubated with 0.1% Triton X-100 for 1 h on ice to deactivate potential live virions, then diluted 1:50 in assay buffer (PBST, 1% BSA, 1% milk powder). Each microarray was incubated with plasma for 1 h at room temperature at 100 RPM. The wells were briefly rinsed with PBST, and then the slides were removed from the gaskets and washed 2×5 min in PBST and 2×5 min in PBS. The slides were dried by centrifugation at $1200 \times g$ at 23°C for 2 mins. Slides were then incubated with detection antibodies (10 $\mu\text{g}/\text{ml}$ Cy3-labelled anti-human IgG and 10 $\mu\text{g}/\text{ml}$ AF647-labelled anti-human IgA in assay buffer) for 30 min at RT at 100 RPM. The wells were briefly rinsed with PBST, and

then the slides were removed from the gaskets and washed 2×5 min in PBST and 2×5 min in PBS. The slides were dried by centrifugation at $1200 \times g$ at 23°C for 2 min.

2.4. Bioinformatic Analysis

2.4.1. Image Analysis: Raw Data Extraction and Data Pre-Processing

Slides were scanned at a fixed gain setting using an InnoScan 710 (Innopsys, Carbone, France) fluorescence microarray scanner, generating a 16-bit TIFF file. A visual quality control check was conducted, and any arrays showing spot merging or other artefacts were re-assayed. A GAL (GenePix Array List) file containing information regarding the location and identity of all probed spots was used to aid with image analysis. Automatic extraction and quantification of each spot were performed using Mapix software (Innopsys) yielding the median foreground and local background pixel intensities for each spot.

The mean net fluorescence intensity of each spot was calculated as the difference between the raw mean intensity and its local background. Extrapolated data were filtered and normalized using an in-house developed software (CT100+ programme). Human IgG and IgA (detected by fluorescently labelled secondary antibody) and human anti-IgG (detected only when plasma or serum is added to the slide) were used as positive controls to assess image signal intensity.

Reciprocal titers per-antigen were determined from measured net fluorescence intensity, based on the projected further dilution of the sample required to reach the limit of detection in the assay, according to the following equation:

$$\text{Reciprocal Titer} = \frac{\text{Net intensity (RFU)} \times \text{initial serum dilution}}{\text{Limit of detection (RFU)}}$$

2.4.2. Statistical Tests

Statistical analyses and graphical representation were generated using GraphPad Prism (v 9.0; GraphPad Software, San Diego, CA, USA). A one-way ANOVA with Welch's correction was applied to determine the statistical significance of the differences observed between independent groups.

3. Results

3.1 Seroprevalence

The N protein is generally favoured over the S protein for seroprevalence studies due to the faster rates of clearance of the anti-S response post infection [18]. We therefore used the anti-N IgG titer to assess the seropositivity in our cohort across three different timepoints (Table 2), representing early in the first wave, prior to onset of the second wave and post-second wave of infections in South Africa. The Immusafe microarray platform has previously been reported to have 100% sensitivity and specificity in detecting anti-N antibody responses to SARS-CoV-2 [2].

Table 2: Seroprevalence based on anti-N IgG titers. Sample size: HCW: n=300. nd = not determined.

Sampling time	Baseline	Visit 5	Visit 6
Seroprevalence (%)	30,67	44,92	61,97
PCR+ (%)	14,00	nd	nd

3.2 Antibody profiles post potential re-infection

Due to new strains and sub-optimal immune responses, it is likely that individuals who are highly exposed to SARS-CoV-2 may become re-infected. Due to the relative stability of the anti-N IgG titers over time, observed here and elsewhere [18], it is possible that an increase in anti-N IgG titers > 1 month after previous infection could be indicative of re-infection. Therefore, we assessed antibody titers at Baseline (July/August 2020, early-mid- wave 1; driven by ancestral SARS-CoV-2), before the second wave (November 2020)

and late in the second wave (January 2021; driven by the β SARS-CoV-2 variant) to identify individuals who likely became re-infected during the second wave (Figure 1). Individuals who were seropositive prior to the second wave were selected and their anti-N titers were assessed at subsequent timepoints to identify individuals who showed an increase in antibody titers late in the second wave.

An epitope coverage score was calculated as the sum of the number of IgG or IgA positive N-protein epitopes (C- and N-terminal domains, and three intrinsically disordered regions) for each sample. This score represents the breadth of the antibody response (2). The epitope utilisation and antibody titers of the participants who appear to have become re-infected (n=28) were then compared to those who were seropositive but did not become re-infected (n=59) (Figure 2 and 3).

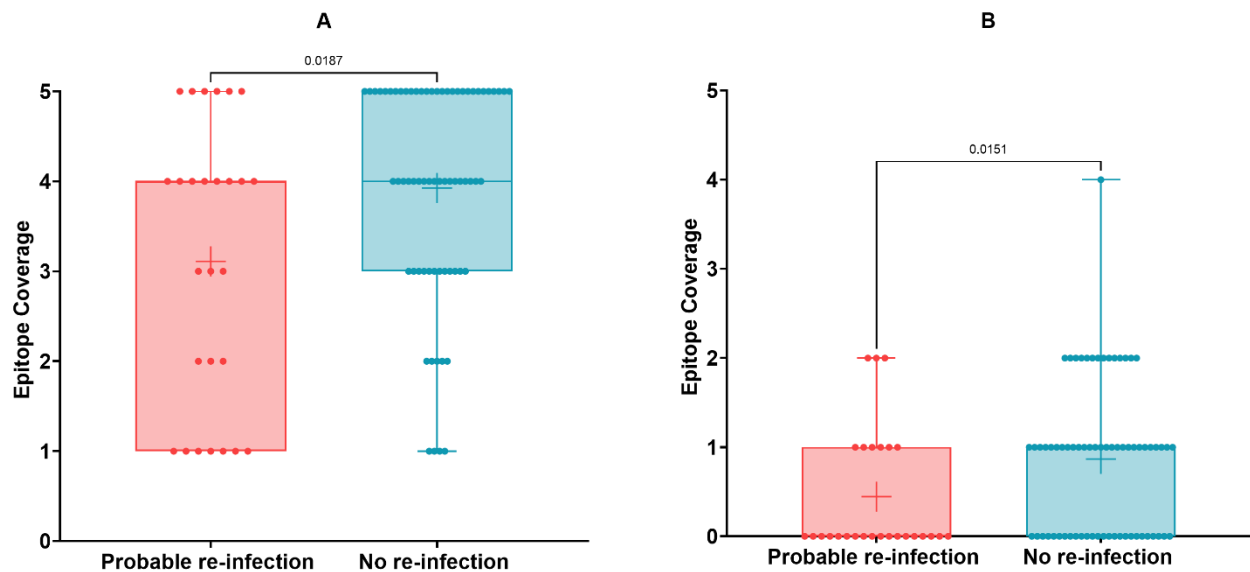


Figure 2: N protein epitope coverage by IgG and IgA specific antibodies in seropositive individuals before the second COVID-19 wave in South Africa. Participants were split into two groups based on potential re-infections. (A) IgG N protein epitope coverage. (B) IgA N protein epitope coverage. Pairwise comparisons were made using a one-way ANOVA, and p-values were calculated using Welch's correction to compare the mean of each category to the mean of each other category. Sample sizes: probable re-infection: n=28, no re-infection: n=59.

For both IgG and IgA responses, a significantly broader epitope coverage is observed in individuals who did not become re-infected during the second wave (Figure 2, A: p-value = 0.0187, B: p-value = 0.0151); thus, epitope utilisation appears to correlate with protection from re-infection. The mean IgG epitope coverage for re-infected and non-re-infected is 3.11 and 3.93 respectively, whereas the IgA epitope coverage for re-infected and non-re-infected is 0.44 and 0.87 respectively (Figure 2).

The relationship between antibody titers and re-infection was assessed. For both N- and S-protein, IgG titers are significantly higher in non-re-infected individuals (Figure 3A and B, p-value = 0.0042 and p-value = 0.0176 respectively). However, no statistically significant differences were observed for IgA titers (Figure 3C and D, p-value = 0.2433 and p-value = 0.4809 respectively).

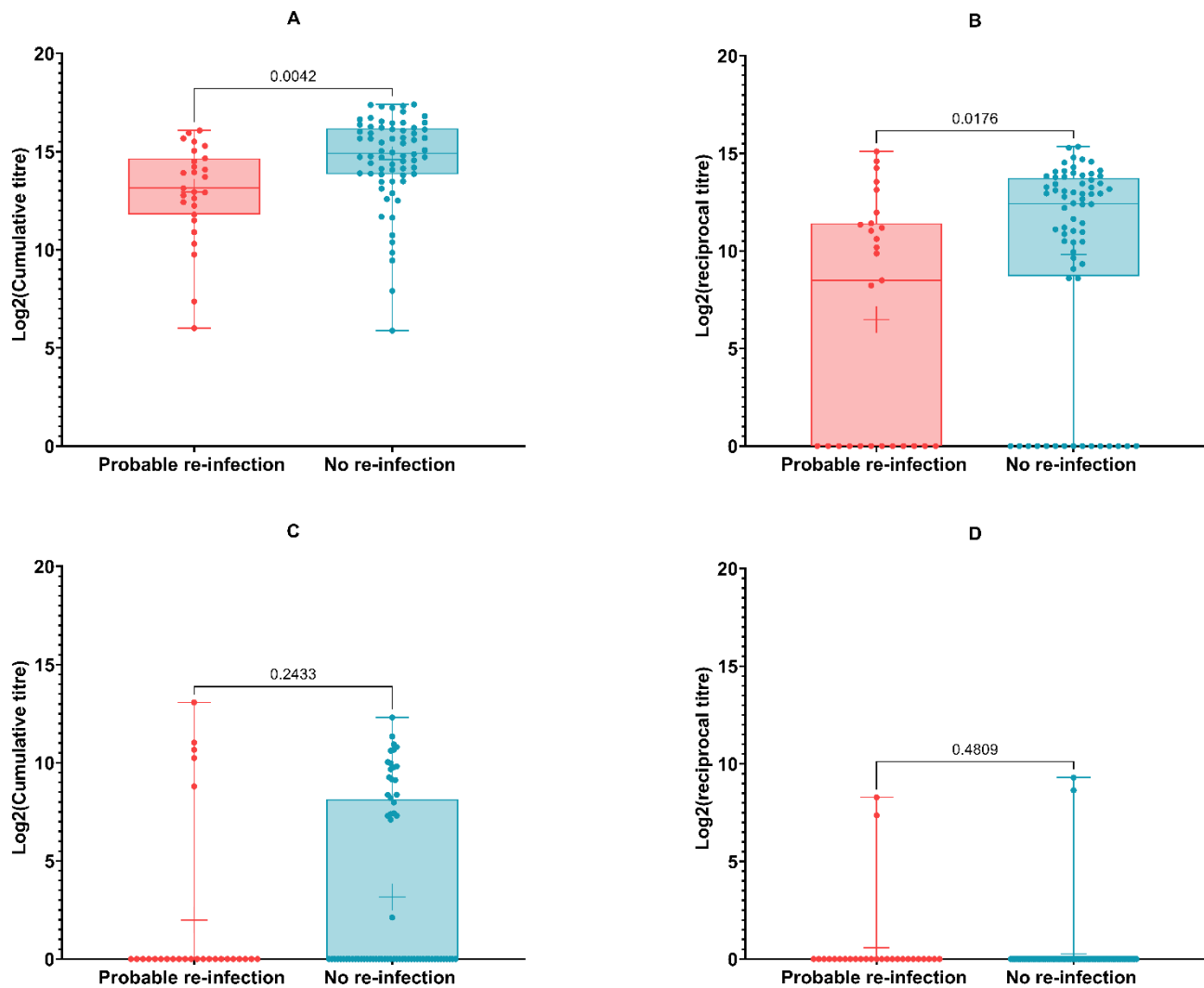


Figure 3: Anti-N and anti-S IgA and IgG titers in seropositive individuals prior to the second COVID-19 wave in South Africa. Participants were split into two groups based on potential re-infections. (A) Anti-N IgG cumulative reciprocal titer. (B) Anti-S IgG reciprocal titer. (C) Anti-N IgA cumulative reciprocal titer. (D) Anti-S IgA reciprocal titer. Pairwise comparisons were made using a one-way ANOVA, and p-values were calculated using Welch's correction to compare the mean of each category to the mean of each other category. Samples sizes: probable re-infection: n=28, no re-infection: n=59.

4. Discussion

As we face ongoing challenges of the COVID-19 pandemic, the global interest to understand the role of the T- and B-cell responses in protection from (re)-infection remains a priority. Despite high seroprevalence and vaccination rates, re-infections and breakthrough infections are continually reported [4-10]. Multiple factors could contribute to this incomplete protection from infection and disease. Serological assays can be used to determine the magnitude, breadth, and durability of the humoral immune response against SARS-CoV-2 and how these factors influence the rates of (re)-infection. Many studies have reported waning antibody levels post-infection and vaccination, with the duration of titers above the detectable level ranging from 5-8 months [19, 20]. Furthermore, the effects of age, ethnicity, sex, and disease severity on both infection rates, antibody titers and epitope coverage have been studied on cross-sectional cohorts [2, 21, 22], however the effects of these factors on the durability of protection remains largely unknown.

With the emergence of new SARS-CoV-2 variants, coupled with waning and incomplete immune responses, the rise in cases of re-infection is not surprising. There is a need

therefore for tools to predict whether natural- or vaccine-induced immunity offers protection against infection and/or disease.

The first wave in South Africa was driven by the ancestral SARS-CoV-2 virus, whereas the second wave was largely driven by the Beta variant (B.1.351). Here we have assessed anti-nucleocapsid and anti-spike protein IgG and IgA responses in the plasma of HCW before and after the second COVID-19 wave to identify probable re-infections. The antibody profiles of those re-infected and those not, were compared to identify a candidate correlate of protection. Our data on the breadth and magnitude of the antibody responses (Figure 2 and 3) clearly shows an importance for both the magnitude and breadth of the humoral immune response to elicit protection from re-infection with new SARS-CoV-2 strains.

It seems immunologically plausible that if multiple different epitopes on viral antigens are targeted by antibodies (including non-neutralising antibodies with Fc effector functions), then the subsequent variant strains may be less able to escape all antibodies present. Consistent with this, here we report a correlation between breadth of anti-N epitope coverage by IgG, as well as higher anti-N and anti-S IgG titers, and apparent protection from re-infection in this HCW cohort (Figure 2 and 3).

It is important to note that in this study we measured anti-N epitope coverage by serum antibodies, whereas protection against (re)-infection by SARS-CoV-2 seems likely to manifest through tissue resident B-cell-derived anti-S sIgA antibodies in mucosal tissues, rather than through bone marrow-derived monomeric IgG or IgA in blood. It remains to be determined therefore whether the breadth of anti-N epitope coverage by serum antibodies is reflected by anti-S epitope coverage by sIgA in mucosa, or whether there is some other immunological explanation (e.g. breadth of anti-SARS-CoV-2 T-cell responses) for the correlation between anti-N epitope coverage, titer and re-infection observed here. Further research on mucosal samples from this cohort is now underway to investigate this.

Limitations

While the study is statistically powered to identify longitudinal changes in antibody titers and epitope utilization, the ability to determine the influence of age, ethnicity, disease severity, and sex on the chances of re-infection is not possible due to cohort size and the number of participants we assume were re-infected. Additionally, whilst we have assumed re-infection in some individuals, this was not clinically proven, nor was the re-infecting virus sequenced to determine variant.

In individuals who did not become re-infected, we cannot prove re-exposure. However, given the increased rate of seropositivity observed at time point 5 (Table 2), it seems reasonable to infer on-going exposure in a significant proportion of the previously-infected-but-not-re-infected individuals in our cohort. Furthermore, if absence of re-infection was determined entirely by non-pharmaceutical interventions rather than by immunological factors, then it would seem unlikely that a statistically significant correlation between epitope coverage, antibody titer and re-infection would have been observed by chance. Importantly, since all time point 5 samples were collected prior to the Sisonke roll-out of the Ad26.COV2.S vaccine to HCWs in South Africa and, moreover, since all vaccines trialed in South Africa prior to time point 5 (principally the ChAdOx1, BNT162b2 and Ad26.COV2.S vaccines) would have elicited pure anti-S antibody responses, it seems reasonable to assume that vaccination status was not a confounding variable in this study.

Author Contributions: J.M.B. designed, conceived, and led the study. N.A.B.N. and W.A.B. provided the samples. M.M., H.M., M.S. and A.N. generated the data. M.M. performed the statistical analysis. M.M. and J.M.B. interpreted the data. M.M. and J.M.B. wrote the manuscript. All authors have read and agreed to the published version of the manuscript

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Institutional Review Board Statement: The study was conducted according to the guidelines of the Declaration of Helsinki and approved by the Ethics Committee of the University of Cape Town (UCT; HREC 210/2020; 15 April 2020). Ethical review and approval for the study on pre-pandemic HIV controls were waived, since it formed part of standard of care at Hospital Sungai Buloh, Selangor, Malaysia.

Informed Consent Statement: Informed consent was obtained from all subjects involved in the study.

Data Availability Statement: Data are contained within the article

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Conflicts of Interest: JMB is a Director of Sengenics Corporation, who marketed the Immusafe microarrays used in this study.

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