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Article

# Allogenic Synovia Derived Mesenchymal Stem Cells for Treatment of Equine Tendinopathies and Desmopathies – The Proof-of-Concept

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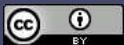
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**Simple Summary:** Horses are high level sport athletes prone to musculoskeletal injuries. Tendon/ligament injuries are one of the most frequent type of injuries and their treatment is very difficult to achieve. Usually, a fibrous scar tissue is achieved instead of tissue regeneration leading to decreased functionality of the organ, endangering their sportive life. The aim of regenerative medicine is finding a treatment that promotes tissue regeneration and patient return to the same level of sportive performance in the most reduced time gap. In this study, we developed a solution of equine synovial membrane stem cells and autologous serum, to be injected at the lesion site to promote tissue regeneration. The process of tissue collection, preparation, isolation of synovial stem cells, expansion, culture, cryopreservation and posterior preparation with autologous serum was described. This solution was tested in 16 tendons and ligaments of equines. After treatment all patients underwent a physical rehabilitation program and were monitored with physical and ultrasonographic exams. Results were very promising, encouraging the use of equine synovial stem cells and autologous serum in the treatment of tendonitis and desmitis.

**Abstract:** Tendons and ligaments injuries are frequent in sport horses and humans, representing a great therapeutic challenge. Tissue regeneration and function recovery are the paramount of tendon and ligament lesions management. Nowadays, several regenerative treatments are being developed, based on the use of stem cell and stem cell-based therapies. In the present study, the preparation of equine synovial membrane mesenchymal stem cells (eSM-MSCs) for clinical use - collection, transport, isolation, differentiation, characterization, and application is described. These cells are fibroblast-like and grow in clusters. They retain an osteogenic, chondrogenic and adipogenic differentiation potential. 16 clinical cases of tendonitis and desmitis, treated with allogenic eSM-MSCs and autologous serum are also presented, including their evaluation, treatment and follow-up. The use of autologous serum as a vehicle concern to a decreased



immunogenic response after the administration of this combination and to its healing properties, due to the presence of growth factors and immunoglobulins. Most of the cases (14/16), healed in 30 days and presented a good outcome. Treatment of tendon and ligament lesions with the mixture of eSM-MSCs and autologous serum appears to be a promising clinical option for this category of lesions in equine patients.

**Keywords:** allogenic; cell-based therapies; clinical trials; ligament; mesenchymal stem cells; sport horses; synovial mesenchymal stem cell; tendon.

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## 1. Introduction

Tendonitis and desmitis are defying clinical challenges in equine patients requiring long recovery periods, and ineffective tendon repair entails their sportive careers. Tendons operate near its functional limit during maximal exercise. Their ability to adapt to stress and self-repair is limited. A controlled exercise program alone or in combination with a variety of conservative treatments, such as corrective shoeing and nonsteroidal anti-inflammatory drugs (NSAIDs), is still the gold-standard therapy for equine tendon disease [1]. Current treatments often do not fully repair or regenerate the injured or affected tendon nor lead to its total functional recovery [1,2].

The aim on tendinopathy treatment is the achievement of tissue regeneration to provide return to complete organ function and performance. Tissue engineering has gained a special interest over the last years for tissue repair. Among this, the development of mesenchymal stem cell-based therapies has boosted, being a promising approach to tissue repair and regeneration including tendinopathy and desmitis [1,3–6].

Mesenchymal Stem Cells (MSCs) can be isolated from several tissue sources, such as bone marrow, peripheral blood, dental pulp, umbilical cord, and amniotic fluid [7]. MSCs characteristics have been defined by the Mesenchymal and Tissue Stem Cell Committee of the International Society for Cellular Therapy (ISCT), and those include being plastic-adherent when maintained in standard culture conditions, expressing cluster of differentiation (CD), such as CD44, CD90 and CD105 and no expression of major histocompatibility complex (MHC)-class II markers and of hematopoietic-related markers (CD45 and CD34) [8]. Finally, MSCs must be able to differentiate *in vitro* into, at least, osteoblasts, adipocytes and chondroblasts, in the presence of adequate differentiation culture media [8].

Synovial membrane mesenchymal stem cells (SM-MSCs) were initially isolated, in 2001 by *De Bari et al.* [9], from human knee joints showing a great proliferative ability in culture, even after passage 10 (P10), and multilineage differentiation potential *in vitro* [9]. These cells represent a good source of MSCs and a promising therapeutic tool mostly for musculoskeletal pathologies [10]. *Sakagushi et al.*, compared the properties of different sources of human stem cells - bone marrow, synovium, periosteum, skeletal muscle, and adipose tissue- and observed the superiority of synovium as a MSCs source for treatment of musculoskeletal pathologies as they had more ability to chondrogenesis. Pellets of synovium derived stem cells were larger and more stained for cartilage matrix [11].

SM-MSCs chondrogenic capacity is higher than other studied sources of MSCs, such as bone marrow (BM-MSCs) [12,13]. Cartilage pellets from SM-MSCs were significantly larger than those from BM-MSCs [12]. SM-MSCs have a higher production of Uridine Diphosphate Glucose Dehydrogenase (UDPGD) [13], an enzyme that converts UDP-glucose into UDP-glucuronate, one of the two substrates required by hyaluronan synthase for hyaluronan polymer assembly. Besides, Sox-9, collagen type -II (Col-II), aggrecan, specific markers for chondrogenesis, as well as cartilage-specific molecules such as cartilage oligomeric matrix protein (COMP), were also found in high amount on equine synovial fluid derived MSCs and extracellular matrix respectively by reverse transcription polymerase chain reaction (RT-PCR) [13].

In a recent study, using a rabbit model, *Bami et al.*, highlight the superiority of SM-MSC's in terms of chondrogenesis, osteogenesis, myogenesis and tenogenesis [14]. Also, a study of xenogenic

implantation of SM-MSCs in equine articular defects confirmed a better healing of the cartilage of affected knees as well as a higher expression of collagen type II, indicating the presence of hyaline cartilage in the healed defect [15].

SM-MSCs were defined as MSCs due to their phenotypic profile and differentiation potential. Even though there are no specific antibodies markers to identify these MSCs, there is a general agreement that MSCs should be negative to hematopoietic markers CD34 e CD45 and positive to CD44, CD73, CD90 and CD105 [16]. *Mochizuki et al.*, found that SM-MSCs maintain their proliferative ability, despite the part of the synovium they are collected from [17].

In 2003, *Fickert et al.*, reported that the markers CD9, CD44, CD54, CD90, and CD166 can be used to identify MSCs isolated from the synovium of human patients with osteoarthritis (OA), and also confirmed that CD9/CD90/CD166 triple-positive cell subgroups have obvious chondrogenic and osteogenic differentiation ability [18].

*Prado et al.*, confirmed the mesenchymal nature of equine synovial membrane and fluid-derived stem cells through the expression of significant markers of hematopoietic (CD45, CD34, CD117 and CD133) and mesenchymal (CD105, CD90), pluripotency (Oct3/4 and Nanog), embryonic (Tra-1-81), inflammatory and angiogenesis markers (vascular endothelial growth factor (VEGF-R1) and LY6a) [19]. Although the presence of hematopoietic and inflammatory markers was not expected, variations may occur and must be considered the influence of acute or chronic stages of osteochondrosis expression and/or inflammatory events [19,20].

Nevertheless, the immunophenotype characterization of equine MSCs (eMSCs), as well as in other veterinary species, is not yet completely established [19]. This is a major challenge as the expression of certain adult stem cell markers may differ between species. For that reason, it is mandatory to define a set of CD markers which can be uniformly applied for the identification of eMSCs [8] [20].

Horses are high performance athletes prone to musculoskeletal diseases – osteoarticular, tendon/ligament lesions and fractures - of various degrees due to sport and age-related injuries. These pathologies resemble human musculoskeletal conditions, turning horses into a valuable animal model for assessing stem cell and cell-based therapies prior to the translation of results into humans [21]. The use of a therapy able to regenerate these structures and restoring their complete functionality instead of an ordinary healing is the aim of our study and of the equine practitioners among the world.

Recent studies suggest that MSCs can self-renewal, to migrate to injury sites (homing), to perform multilineage differentiation and to secrete bioactive factors, increasing proliferation and migration of tendon stem/progenitor cells via paracrine signaling and increasing regeneration ability of tissues with poor aptitude [1,3–5,22,23].

In fact, the knowledge of the importance of this paracrine action has opened doors to cell-free therapeutic strategies in regenerative medicine. The soluble factors (cytokines, chemokines and growth factors) and non-soluble factors (extracellular vesicles and exosomes) released in the extracellular space by MSC's, commonly known as secretome, became the focus of the novel therapeutic approaches due to their key role in cell to cell communication, their active influence on immune-modulation and pro-regenerative capacity both *in vitro* and *in vivo* [23]. For this reason, in this study, secretome was also analysed with the prospect of being used therapeutically, in the future, in similar clinical cases.

In the present study, equines used as show jumping and dressage athletes, as well as leisure horses with acute and chronic lesions, were treated with intralesional administrations of the considered combination – autologous serum and eSM-MSC's. This treatment consisted of two injections, 15 days apart. Pre- and post-treatment evaluations consisted of clinical, orthopedic and tendon/ligament ultrasound exams. None of the selected patients have received any other regenerative treatment before.

## 2. Materials and Methods

### 2.1. Study Design and Horses Selection

A prospective longitudinal study was performed in Portugal between February 2016 and January 2019. Sixteen horses, from 5 to 22 years old with acute and chronic signs of lameness were enrolled in this study - 11 males and 5 mares, whose sportive activity was distributed over show jumping (14), dressage (1) and leisure (1). These animals were all outpatients from equine ambulatory clinic. This study includes the treatment of 16 tendons -14 superficial digital flexor tendons and 2 deep digital flexor tendons - and 4 suspensory ligaments.

Lameness was scored based on the American Association of Equine Practitioners (AAEP) scale (Table 1) and confirmed by a positive regional nerve block. Flexion test and pain to pressure were also evaluated [24].

**Table 1.** Score systems used by the veterinary surgeon to assess lameness, response to flexion test and pain to pressure [25] [26].

Parameter	Score	Clinical implication
AAEP Grading	0	No Lameness
	1	Lameness not consistent
	2	Lameness consistent under certain circumstances
	3	Lameness consistently observable on a straight line.
	4	Obvious lameness at walk: marked nodding or shortened stride
	5	Minimal weight bearing lameness in motion or at rest
Flexion Test	0	No flexion response
	1	Mild flexion response
	2	Moderate flexion response
	3	Severe flexion response
Pain to pressure	0	No pain to pressure
	1	Mild pain to pressure
	2	Moderate pain to pressure
	3	Severe pain to pressure

## 2.2. Inclusion and Exclusion Criteria

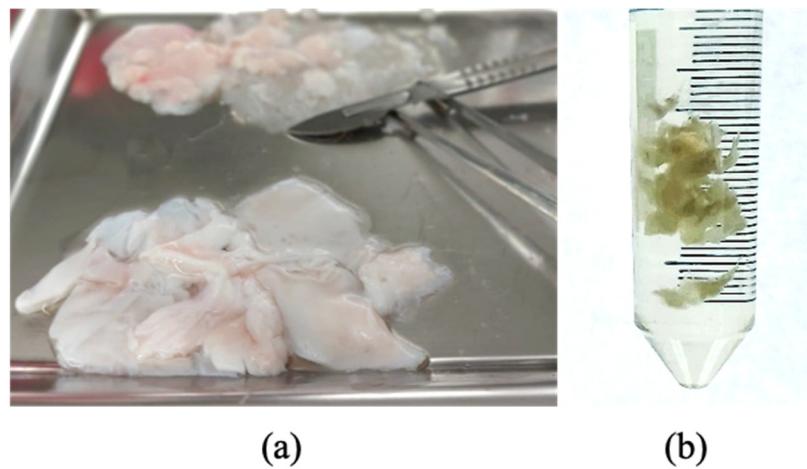
In this study, horses with acute or chronic lameness, with diagnosed tendonitis and/or desmitis and with no signs of systemic disease were accepted in the inclusion criteria. Injured horses were treated in acute stages of disease, except for one patient with chronic desmitis of suspensory ligament and tendonitis of deep and superficial digital tendon. Considering the established exclusion criteria, selected patients should not have been under any other medical treatment (including nonsteroidal anti-inflammatory drugs, intra-articular corticosteroids, hyaluronan, glycosaminoglycans, Platelet-rich plasma (PRP) and other MSC's preparations) at least 2 months before allogenic eSM-MSCs treatment didn't receive any additional medical treatment (except for the ones described in the treatment plan) for at least 2 months *post* the cell-based treatment.

## 2.3. Ethics and Regulation

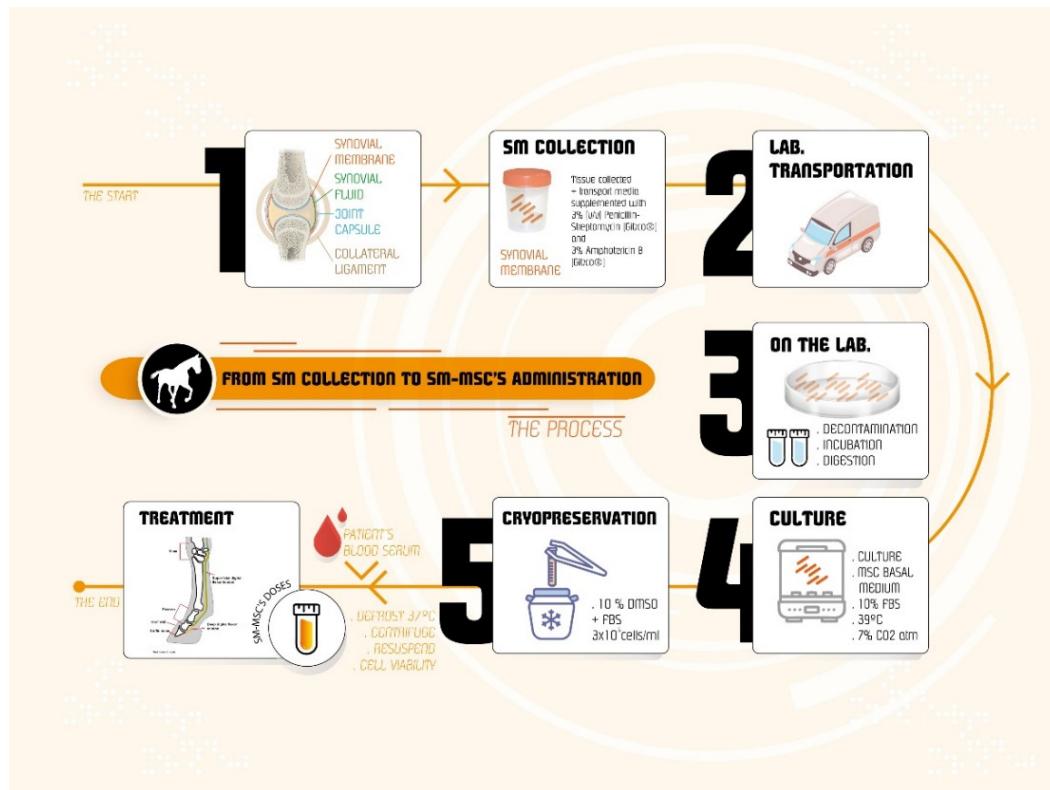
This study was carried out in accordance with *Organismo Responsável pelo Bem Estar Animal* (ORBEA) from ICBAS-UP, project number: P289/ORBEA/2018 recommendations and authorization. Treatments were performed with permission and signature of an informed consent from the patient's legal tutor, following a thorough explanation on the procedure itself and possible risks and associated effects, in accordance with national regulations and project approval from the competent authorities. In addition, no animals were euthanized for this study.

#### 2.4. Donor selection and SM collection

eSM-MSCs' donor was a young and healthy foal who died accidentally. The tutor authorized synovial membrane collection from hocks, knees and fetlocks. Skin covering the incisional field was surgically cleaned with chlorohexidine and alcohol. Skin and subcutaneous tissue were incised, and debrided, articular capsule was opened, and synovial membrane was isolated and extracted into a Dulbecco's Phosphate Buffered Saline (DPBS) container. Samples were transported to the laboratory with ice packs for refrigerated temperatures. Figure 1 (a) presents the fresh tissue arrival and Figure 1 (b) the preparation at the laboratory. Figure 2 synthetizes the process from eSM-MSCs collection to the administration of the combination: eSM-MSCs and autologous serum (1x10<sup>6</sup> cells/ml and 1ml of autologous serum in a total volume of 2ml).



**Figure 1.** Laboratory: arrival and preparation of the fresh tissue to start digestion, isolation and expansion. (a) Tissue collected in the field and (b) isolated synovial membrane.



**Figure 2.** Schematic representation of event sequence from the collection of synovial membrane to the administration of the therapeutic combination. After the collection, the synovial membrane is

transported to the laboratory where it is separated from the whole tissue, decontaminated, incubated and digested. Then cells are cultured and expanded and finally cryopreserved in a cell bank. When needed for treatment, cells are prepared with autologous serum and then applied in selected equine patient.

### 2.5. *eSM-MSCs isolation*

After collection, equine synovial membrane, was prepared at the Laboratory of Veterinary Cell-based Therapies from ICBAS-UP. The isolation protocol of eSM-MSCs had been developed by patented proprietary technology Regenera® (PCT/IB2019/052006, WO2019175773 – Compositions in use for the treatment of musculoskeletal conditions and methods for producing the same leveraging the synergistic activity of two different types of mesenchymal stromal/ stem cells - Regenera®). Fresh tissue was transported to the laboratory facilities in a hermetically sealed sterile container in transport media [supplemented with 3% (v/v) Penicillin-Streptomycin (Gibco®) and 3% Amphotericin B (Gibco®)] and processed within a period up to 48h. The synovial tissue was digested using collagenase and the isolated cells were incubated in a static monolayer culture using standard MSCs basal medium supplemented with 10% fetal bovine serum (FBS) and maintained in standard culture conditions (37°C, 5% CO<sub>2</sub> and humidified atmosphere) until they reached confluence. Cells from confluent cultures were cryopreserved in 10% dimethylsulphoxide (DMSO) and FBS, at a concentration of 3x10<sup>6</sup>cells/ml, using control rate temperature freezer (Sy-Lab Cryobiology, SY-LAB Geräte GmbH, Austria). For expansion optimization, cells were cryopreserved at passages (P) between P2 and P3 for constitution of suitable Master Cell Banks (MCB). Expansion thereafter was analyzed during a maximum of 20 cumulative population doublings (cCPDs). The range of cCPDs chosen allowed for enough expansion for maximizing the number of cells in working cell banks (WCB) but keeping cCPDs within the genomic stability range.

### 2.6. *SM-MSCs characterization*

#### 2.6.1. Tri-lineage differentiation protocols

For all differentiation protocols, cells in P4 after thawing were used

##### Adipogenic Differentiation and Oil Red O Staining

For the Adipogenic differentiation protocol, 1x10<sup>4</sup> cells/cm<sup>2</sup> were seeded in the wells of a 12-well plate (Cell culture plates, 12-well, VWR®), with addition of culture medium. The plate was incubated under standard conditions for 4 days. After this period, the culture medium of 10 wells was replaced by complete adipogenesis differentiation medium (StemPro® Adipogenesis Differentiation Kit, Gibco®), 2 wells were used as controls and maintained with the usual culture medium. Following the manufacturer's instructions, the medium was replaced every 3-4 days and the cells maintained in differentiation for 14 days. At the end of this period, Oil Red O staining protocol was performed using a handmade solution. The culture differentiation medium was removed, and the wells were gently washed with PBS. Cells were fixed with 4% formaldehyde (3.7 - 4% buffered to pH7, reference# 252931.1315, Panreac AppliChem®) for 10 minutes at room temperature, and the wells washed 3 additional times with Phosphate-buffered saline (PBS). Oil Red O solution was added to each well and the plate incubated for 10–20 minutes at room temperature. Oil Red O was discarded, and any excess dye removed by several washes with PBS. PBS was added to each well for visualization. The aim of this assay was the identification of rounded cells with intracytoplasmic lipid vacuoles and their red coloration due to the exposure to the Oil Red O solution.

##### Chondrogenic Differentiation and Alcian Blue Staining

Thawed eSM-MSCs were automatically counted, and cell viability determined (%). The cells were then centrifuged, supernatant removed, and the pellet resuspended in culture medium to generate a cell suspension with 1.6x10<sup>7</sup> viable cells/ml. To generate micro-mass cultures, 5µl droplets

of the cell suspension were placed in the center of 10 wells of a 96-well plate (Cell culture plates, 96-well, VWR®), to induce chondrogenic differentiation. The plate was maintained under standard conditions for 2 hours. After this time, chondrogenic differentiation medium (StemPro® Chondrogenesis Differentiation Kit, Gibco®) was added to 8 wells, as 2 wells were considered as control and to these, usual culture medium was added. Following the manufacturer's instructions, medium was replaced every 3-4 days and cells maintained in differentiation for 14 days. At the end of this period, the Alcian Blue staining, pH2.5, protocol was performed (Alcian Blue 8GX, Sigma-Aldrich®). The culture differentiation medium was removed, and the wells were gently washed with PBS. Cells were fixed with 4% formaldehyde during 20 minutes at room temperature, and the wells washed 3 additional times with PBS. Alcian Blue solution was added to each well and the plate incubated for 30 minutes at room temperature. Alcian Blue was then discarded, and wells were rinsed 3 times with acetic acid 3% (v/v). For neutralization of acidity and for visualization by inverted phase contrast microscopy, distilled water was added to all wells. The aim of this assay was the identification of chondrogenic aggregates and their coloration in blue due to the exposure to Alcian Blue solution.

#### Osteogenic Differentiation and Alizarin Red S Staining

For osteogenic differentiation,  $8 \times 10^3$  cells/cm<sup>2</sup> were seeded into the wells of a 12-well plate. The plate was maintained under standard conditions for 4 days. After this period, the culture medium of 10 wells was replaced by complete osteogenic differentiation medium (StemPro® Osteogenic Differentiation Kit, Gibco®), and 2 wells were used as controls and maintained with the usual culture medium. Following the manufacturer's instructions, media was replaced every 3-4 days and the cells maintained in differentiation for 21 days. At the end of this period, Alizarin Red S Staining protocol was performed using a commercial solution (Alizarin-Red Staining Solution, Millipore®). The culture differentiation medium was removed, and the wells were gently washed with PBS. Cells were fixed with 4% formaldehyde for 30 minutes at room temperature, and the wells washed twice with distilled water. One ml of 40 mM of Alizarin Red solution was added to each well and the plate incubated for 30 minutes. Alizarin Red was then discarded, and wells were rinsed 3 times with distilled water until supernatant became clear. For visualization by inverted phase contrast microscopy, PBS was added to all the wells. The aim of this assay was to identify calcium containing osteocytes stained in red after exposure to Alizarin Red solution.

#### 2.6.2. Karyotype analysis

eSM-MSCs in two different passages (P4 and P7) were submitted to cytogenetic analysis to determine genetic stability in terms of chromosome number and occurrence of neoplastic changes. For both passages, 70 – 80% confluence was reached, culture medium was changed and supplemented with 10 µg/ml colcemid solution (KaryoMAX® Colcemid™ Solution, Gibco®). After 4 h, the eSM-MSCs were collected and resuspended in 8 mL of 0.075 M KCl solution, followed by incubation under standard conditions for 15 minutes. After centrifugation (1700 rpm), 8 ml of ice-cold fixative comprising methanol and glacial acetic at a proportion of 3:1, was added and mixed, afterwards, the cells were centrifuged again. Three fixation rounds were carried out. After the last centrifugation, the suspension of eSM-MSCs was spread over glass slides. Karyotype analysis was performed by one scorer on Giemsa-stained cells. For the different passages, a specific number of cells in metaphase were evaluated depending on the number of cells with a normal karyotype identified, guaranteeing a better representation of the population under study.

#### 2.6.3. Secretome- Cells Conditioned Medium (CM) Analysis

eSM-MSCs were harvested from equine synovial membrane and maintained in culture as previously described. Cells in P4 were subjected to an analysis of their Conditioned Medium (CM) to identify cytokines and chemokines secreted after conditioning. When in culture, after reaching a confluence of around 70–80%, the culture medium was removed, and the culture flasks were gently

washed with DPBS two to three times. Then, the culture flasks were further washed two to three times with the basal culture medium of each cell type, without any supplementation. To begin the conditioning, non-supplemented DMEM/F12 GlutaMAX™ (10565018, Gibco®, Thermo Fisher Scientific®, Waltham, MA, USA) culture medium was added to the culture flasks, which were then incubated under standard conditions. The culture medium rich in factors secreted by the cells (CM) was collected after 48 h. The collected CM was then concentrated five times. After collection, it was centrifuged for 10 min at 1600 rpm, its supernatant collected and filtered with a 0.2 µm Syringe filter (Filtropur S®, PES, Sarstedt, Nümbrecht, Germany). For the concentration procedure, Pierce™ Protein Concentrator, 3k MWCO, 5–20 mL tubes (88525, Thermo Scientific®, Waltham, MA, USA) were used. Initially, the concentrators were sterilized following the manufacturer's instructions. Briefly, the upper compartment of each concentrator tube was filled with 70% ethanol (v/v) and centrifuged at 300g for 10 min. At the end of the centrifugation, the ethanol was discarded, and the same procedure was carried out with DPBS. Each concentrator tube was subjected to two such centrifugation cycles, followed by a 10-min period in the laminar flow hood for complete drying. Finally, the upper compartment of the concentrator tubes was filled with plain CM (1× concentration) and subjected to new centrifugation cycle, under the conditions described above, for the number of cycles necessary to obtain the desired CM concentration (5×). The concentrated CM was stored at -20°C and subsequently subjected to a Multiplexing LASER Bead analysis (Eve Technologies, Calgary, AB, Canada) to identify a set of biomarkers present in the Equine Cytokine 8-Plex Assay (EQCYT-08-501). The list of searched biomarkers includes Basic Fibroblast Growth Factor (FGF-2), Granulocyte Colony Stimulating Factor (G-CSF), Granulocyte-macrophage Colony Stimulating Factor (GM-CSF), Monocyte Chemoattractant Protein-1 (MCP-1), Interleukins (IL) IL-6, IL-8, IL-17A and Human Growth-regulated oncogene/Keratinocyte Chemoattractant (GRO/KC). All samples were analyzed in duplicate.

#### 2.6.4. Immunohistochemistry

Early passages of eSM-MSCs - P0 and P3 – were maintained in culture until a confluence of 70 - 80% was reached and then enzymatic detachment was performed with 0.25% Trypsin-EDTA solution. A cytoblock was performed fixing the cells with Sure Thin® (Preserve Cell Solution®, Cytoglobe GbH®). Consecutive sections were cut at 2µm, deparaffinized, hydrated and submitted to immunohistochemical analysis using the Novolink™ Polymer Detection Systems (Leica Biosystems®) kit, according to the manufacturer's instructions. Information regarding the primary antibodies and antigen retrieval recovery methods used in the study are summarized in Table 2.

The antibodies were selected to confirm the pluripotent and mesenchymal origin of eSM-MSCs - Octamer-binding transcription factor 4 (Oct-4), Homeobox protein (NANOG), proto-oncogene receptor tyrosine kinase or stem cell factor receptor (c-kit), synovial origin (Lysozyme) and non-epithelial origin histogenesis (Vimentin). Additionally, pan-Cytokeratin (AE1 and AE3), Synaptophysin, CD31 and Glial fibrillary acidic protein (GFAP) were used to discard vascular, epithelial, neuronal and neuroendocrine origin, respectively. For each antibody, appropriate negative and positive controls were included, all primary antibodies were incubated overnight.

The final step consisted of microscopic cell observation, evaluation and photograph using the microscope Eclipse E600 (Nikon®) and the software Imaging Software NIS-Elements F Ver4.30.01 (Laboratory Imaging®). A semi-quantitative score was used for immunoexpression evaluation, consisting of the percentage of labeled cells (<5%; 5–80%; >80%) and labeling intensity (0, negative; +, weak; ++, moderate; +++, strong). Immunoreactivity was considered positive when distinct nuclear and cytoplasmic staining was recognized in at least 5% of the cells.

**Table 2.** List of antibodies investigated, dilutions, and antigen retrieval methods applied in the immunohistochemical analysis.

Marker	Type/Clone	Supplier	Dilution / Incubation period	Antigen unmasking	Positive control	Cells of interest	Reference
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Oct-4	Polyclonal	Abcam	1/100 ON	RS/WB	Canine mast cell tumour Canine testicular carcinoma	Stem cells	Ab18976
Nanog	Clone Mab	ABGENT	1/10 ON	RS/WB	Canine testicular carcinoma	Stem cells	AM1486b
c-Kit (CD117)	Polyclonal	Dako Denmark	1/450 ON	RS/WB	Canine mast cell tumour	Stem cells	A4502
Lysozyme	Polyclonal	Dako Denmark	1/400 ON	RS/WB	Canine synovial membrane	Synovial cells	A0099
Vimentin	Clone V9	Dako Denmark	1/500 ON	RS/WB	Canine mammary gland	Non-epithelial cells	M0725
Pan-cytokeratin	Cocktail AE1/AE3	Thermo Scientific	1/300 ON	RS/WB	Canine mammary gland	Epithelial cells	M3-343-P1
GFAP	Polyclonal	Merck Millipore	1/2000 ON	RS/WB	Mouse brain tissue	Neuronal cells	AB5804
Sinaptophysine	Clone SP11	Thermo Scientific	1/150 ON	RS/WB	Mouse brain tissue	Neuronal cells	RM-9111-S
CD31	Clone JC70A	Dako Denmark	1/50 ON	Pepsine	Canine spleen	Platelet endothelial cells	M0823

### 2.7. eSM-MSCs solution preparation

The eSM-MSCs solution for local clinical application in the 16 equine patients, was a combination of allogenic eSM-MSCs suspended in autologous serum. Prior to preparation of the final therapeutic combination, autologous serum was isolated from whole blood. 10 ml of whole blood were collected into dry blood collection tubes and after clotting, they were centrifuged at 2300 rpm for 10 minutes and their supernatant (serum) collected and transferred to a 15 ml falcon. Then, the serum was inactivated through a water bath at 56°C for 20 minutes followed by cooling on ice. Finally, the serum was centrifuged and filtered using a 0.22µm syringe filter and stored at -20°C until further use. Cryopreserved P3 eSM-MSC's batches were thawed in a 37°C water bath, its content transferred to a 10 ml tube with autologous serum and slowly diluted, followed by the addition of sterile DPBS until reaching 10mL. The mixture was then centrifuged at 1600rpm for 10 minutes. The supernatant was discarded, and the cell pellet was re-suspended in a mixture of autologous serum in a ratio of 0.8:1. Cell counting, and viability was determined by the Trypan Blue exclusion dye assay (Invitrogen™) using an automatic counter (Countess II FL Automated Cell Counter, Thermo Fisher Scientific®). Cell number was then adjusted to 5x10<sup>6</sup> cells/ ml. 2 ml of the solution of eSM-MSCs suspended in autologous serum was transferred to a perforable capped vial and preserved on ice until the time of administration.

### 2.8. Treatment Protocol

Twenty structures - tendons and ligaments - were treated with a mixture of allogenic eSM-MSCs and autologous serum. The same treatment protocol was used in every case. All patients were submitted to identification, anamnesis, physical examination (cardiac and respiratory frequency, body temperature, mucous membrane examination, inspection of the whole body and palpation), orthopedic examination (evaluation of the limbs, gait inspection and movements – walk, trot and gallop, and flexion test of the main joints for 60 seconds followed of trot). Lameness was evaluated at walk and trot on hard surface and scored in a scale of 0 to 5, according to AAEP parameters. Complementary diagnostic exams included regional nerve blocks (to identify the pain area), radiographs and ultrasound image as reported in other studies [21,24,25,27–32].

Following the assumptions of the exclusion criteria, the animals did not receive any treatment before or after the administration of the therapeutic.

Patients were monitored in the 48 hours after treatment and any occurrences were registered. Following the treatment, patients were assessed periodically to control patient's healing evolution and provide valid comparative data among patients within the same study group. Table 3 presents the lesion type casuistic.

**Table 3.** Lesion type casuistic.

Lesion type	Nº clinical cases	Total number (2019)
Tendonitis	16	
Desmitis	4	20

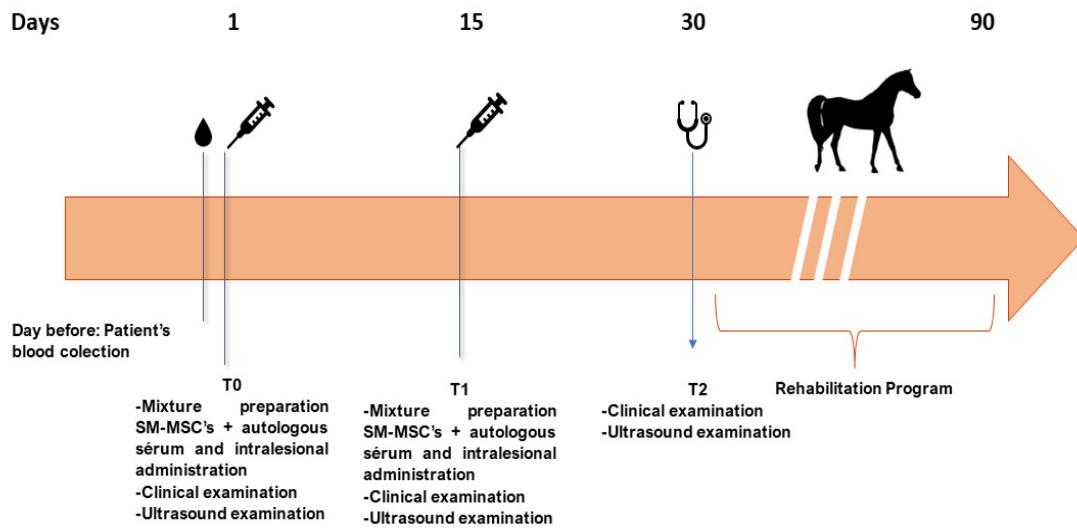
#### 2.8.1. Intralesional eSM-MSCs injection

Selected horses were sedated with detomidine (0.02 mg/kg), trichotomized, regional nerve block was performed with lidocaine 2% (20 mg/ml; 2 ml/point) and the surgical skin was disinfected with chlorohexidine and alcohol. The therapeutic combination was aspirated to a 2ml syringe and homogenized, ultrasound was used to identify the lesion site, and an ultrasound guided injection was performed at the lesion over three different points. Finally, a bandage was applied to the limb. All patients were injected with phenylbutazone (2.2 mg/kg, IV, SID) at the end of the treatment. The established protocol included a second eSM-MSCs administration 15 days after the first treatment using the same protocol.

#### 2.8.2. Clinical evaluation – Serial evaluations

Tissue regeneration was estimated through lameness evaluation, pain to pressure, limb inflammation, sensitivity and ultrasound image (reduction of hypoechoic area and fiber alignment). Lesion ultrasonographic evaluations were performed using a 7.5 MHz linear transductor probe (Ultrasound equipment - Sonoscape A5®). For each assessment, a complete examination of the structure was conducted by means of longitudinal and transverse scans. The obtained images were evaluated at each examination for two parameters: lesion echogenicity and lesion longitudinal fiber alignment (FA). The contralateral healthy limb was used as comparison. Evaluation was performed on treatment day (day 1) as well as on days 15, 30, and 45 post-treatments as presented in Figure 3. According to the classification proposed by *Guest et al*, this is a short term period study [33].

Rehabilitation program consisted of an exercise-controlled program with stall confinement and increasing time of exercise. Early mobilization included weight-bearing activities, strengthening, and flexibility, and stall rest alone was used as infrequently as possible as presented on Table 4 [34–38]. Regular ultrasound evaluations were also performed.



**Figure 3.** Timeline of eSM-MSCs treatment protocol and rehabilitation program. The day before the first treatment (T0), blood from the patient was collected to prepare autologous serum. At T0 the mixture of autologous serum and eSM-MSC's was injected intralesional after a clinical and ultrasound examination. After 15 days the same procedure was repeated. At day 30 (T2), a clinical and ultrasound examination was performed and if a favorable outcome was considered, the animal progressed to a physical rehabilitation program. During the physical rehabilitation program, the patient was also re-evaluated at days 60 and 90.

**Table 4.** Physical rehabilitation program. After eSM-MSCs treatment, all horses were submitted to a rehabilitation program consisting of two days of box rest followed by 13 days of 10 minutes hand-walk. Bandage applied on treatment day was removed 24h after treatment. At day 15 the second treatment was performed followed by another 15 days of rehabilitation, until day 30. Between day 30 and day 45 the work consisted of 20 min hand-walking, between day 45 and day 60 the work was 30 minutes of hand-walking, between day 60 and day 75, 30 minutes of hand walking plus 5 minutes trot and finally between day 75 and day 90, the patient was submitted to 30 minutes of hand-walking plus 10 minutes of trot. After this the patient could return to full work.

Week	Exercise
0-2	2 days: stall confinement Handwalk: 10 min Day 15: new treatment
3-4	2 days: stall confinement Handwalk: 10 min VET-CHECK – day 30
5	Handwalk: 15 min
6	Handwalk: 20 min VET-CHECK – day 45

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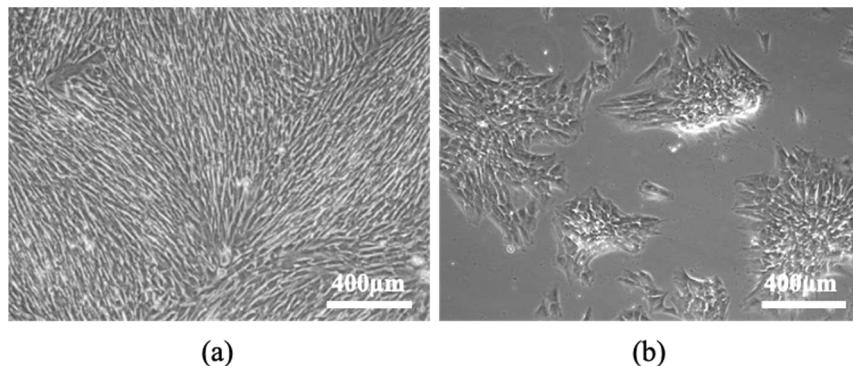
7	Handwalk: 25 min
8	Handwalk: 30 min
8	VET-CHECK – day 60
9-10	Handwalk: 30 min + 5min trot
11-12	Handwalk: 30 min + 10 min trot
11-12	VET-CHECK - day 90

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### 3. Results

#### 3.1. eSM-MSCs isolation

eSM-MSCs have been successfully isolated from equine synovial membrane samples and the averages total number of cells isolated from these samples is  $1.2 \times 10^5$  and  $5.6 \times 10^5$  at day 6 and 11, respectively, and expanded from the donor. Cells observed radiating from the explants and those identified in culture showed clear plastic adhesion and mostly fibroblast-like morphology, an essential feature to characterize cells as MSCs – Figure 4(a,b).



**Figure 4.** - eSM-MSCs in culture, isolated through enzymatic digestion. (a) - Passage 0 (P0) and (b) - Passage 1 (P1). Plastic adhesion, monolayer and fibroblast-like shape of eSM-MSC's may be observed. Magnification: 100x.

#### 3.2. eSM-MSCs characterization

##### 3.2.1. Tri-lineage differentiation

Tri-lineage differentiation was confirmed.

##### Adipogenic differentiation – Oil Red O Staining

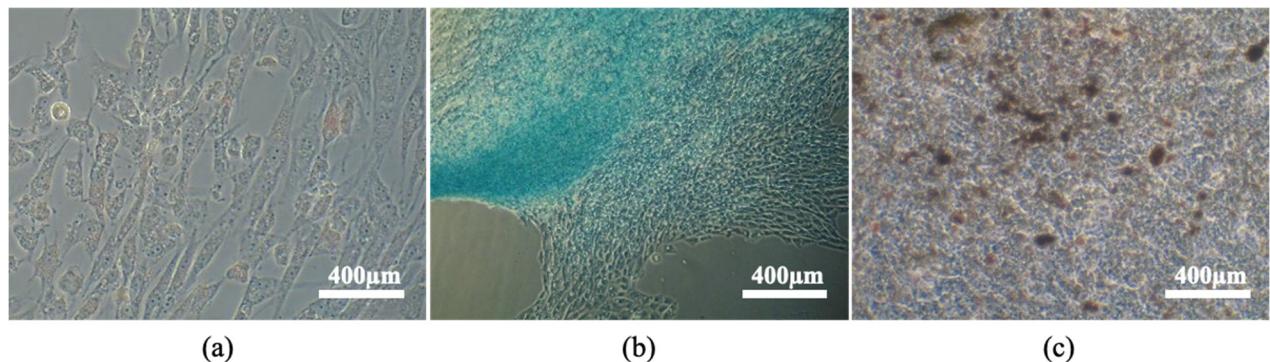
Adipogenic differentiation was confirmed by the presence of large red stained lipid vacuoles in the cytoplasm due to exposure of Oil Red O staining.

##### Chondrogenic differentiation – Alcian Blue Staining

Chondrogenic differentiation was confirmed by the presence of proteoglycans marked deposition in the extracellular matrix which stained blue, confirming the presence of chondrogenic aggregates.

### Osteogenic differentiation – Alizarin Red Staining

Osteogenic differentiation was demonstrated by the presence of extracellular calcium deposits red stained by Alizarin red solution, which dyes chelate complexes with calcium.



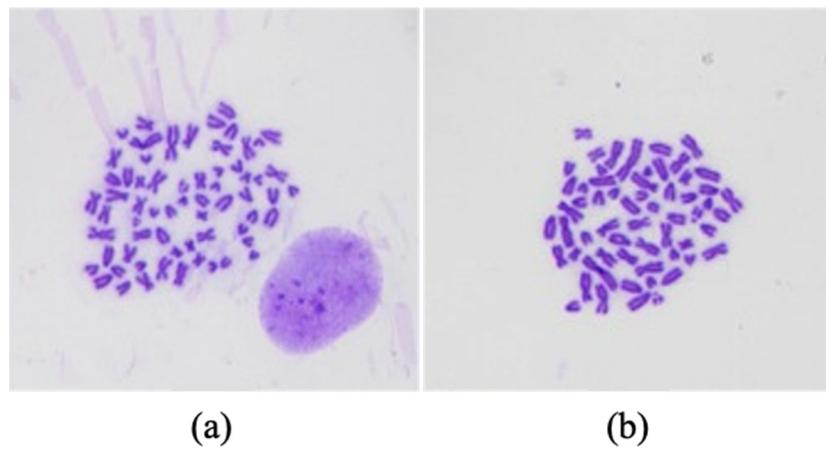
**Figure 5. Tri-lineage differentiation.** (a) eSM-MSC's Adipogenic differentiation: cytoplasmatic lipid vacuoles stained in red (Oil red staining). (b) eSM-MSC's Chondrogenic differentiation: proteoglycans in extracellular matrix stained in blue (Alcian Blue Staining). (c) eSM-MSC's Osteogenic differentiation: extracellular calcium deposits stained in red (Alizarin Red staining). Magnification 100X.

#### 3.2.2. Karyotype analysis

Cytogenetic analysis revealed the presence of 36% of normal cells at P4 and 32% at P7. Tetraploidy was present in 4% of P4 cells and 8% of P7 cells. Aneuploidy represented 60% of cells at both passages, being hypoploidy's the most representative (56%), as shown at Table 5 and Figure 7(a,b).

**Table 5.** Cytogenetic analysis in passages 4 and 7 (P4 and P7). Percentage of normal cells, tetraploid cells and aneuploid cells.

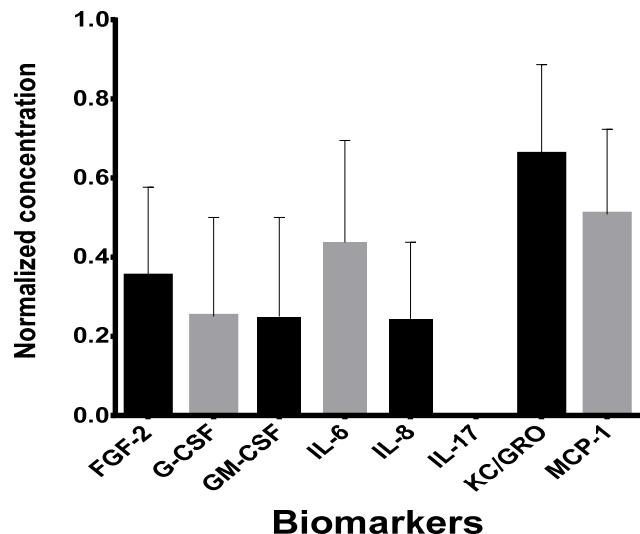
P4	Cytogenetic analysis	P7
36%	Normal cells 64, XY	32%
4%	Tetraploid cells 128 XXYY	8%
60%	Aneuploid cells:	60%
56%	Hipoploidy 54-63	56%
4%	Hiperploidy 71	4%



**Figure 6. Karyotype.** Images of eSM-MSC cytogenetic analysis. at Passage 7 (P7). (a) Normal karyotype: 64 chromosomes XY. (b) Hypoploid cell: 59 chromosomes: 3 acro and 2 submeta.

### 3.2.3. Secretome analysis

Analysis of CM revealed the production and secretion of several factors with immunomodulatory functions, capable of intervening beneficially in tissue regeneration. Results of eSM-MSCs CM analysis can be found in Figure 7. Seven biomarkers were identified: Keratinocyte chemoattractant/ Growth Regulated Oncogene (KC/GRO), Monocyte chemoattractant protein-1 (MCP-1), Interleukin-6 (IL-6), Fibroblast Growth Factor (FGF-2), Granulocyte colony stimulating factor (G-CSF), Granulocyte-macrophage colony stimulating factor (GM-CSF) and Interleukin 8 (IL-8). The most expressive were KC/GRO and MCP-1.

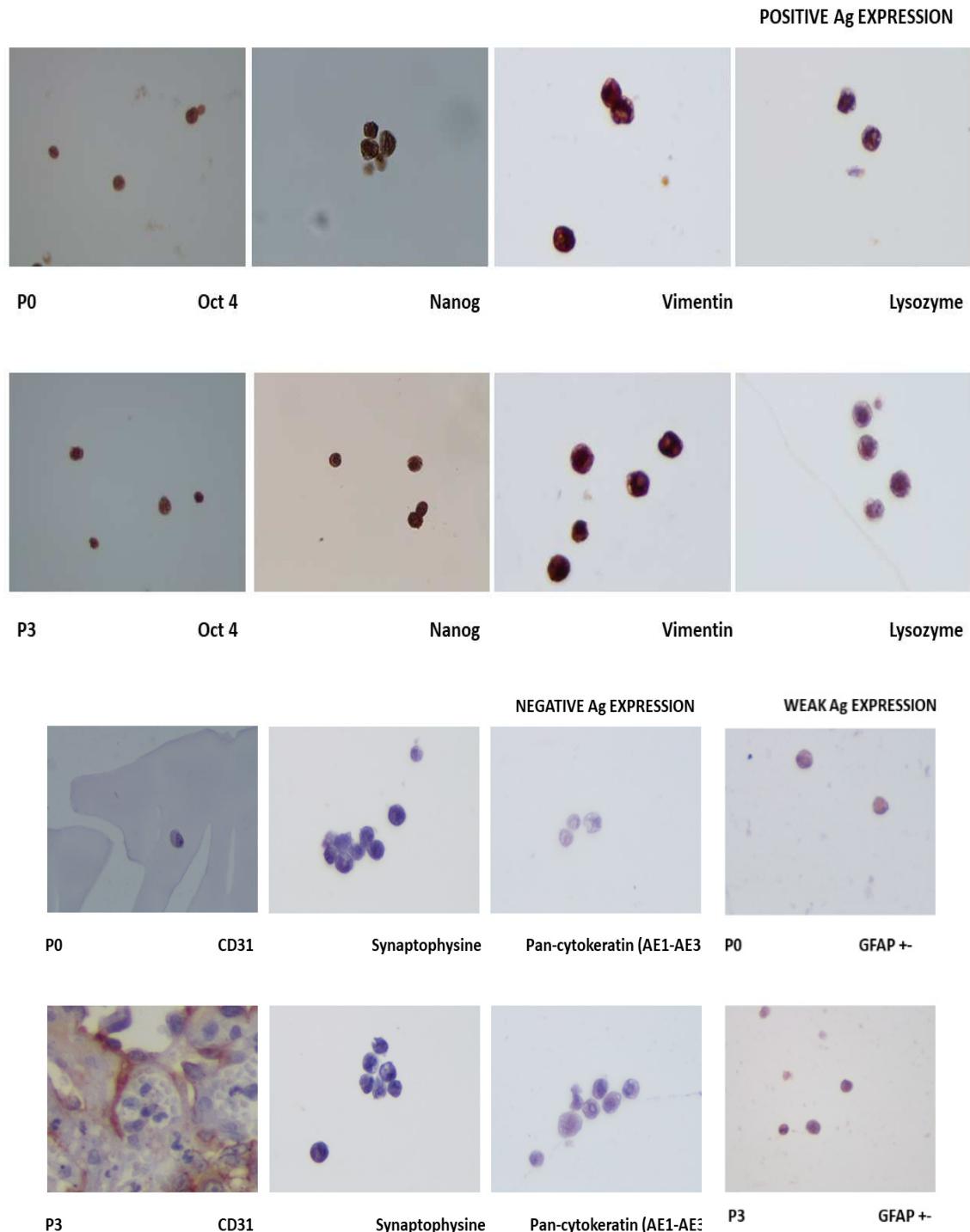


**Figure 7. Biomarkers.** Normalized concentration of each biomarker in the conditioned medium of eSM-MSCs in P4 (mean  $\pm$  SEM). \* Corresponds to  $0.01 \leq p < 0.05$ , \*\* to  $0.001 \leq p < 0.01$ , \*\*\* to  $0.0001 \leq p < 0.001$ , and \*\*\*\* to  $p < 0.0001$ .

### 3.2.4. Immunohistochemistry

eSM-MSCs showed strong expression of Oct-4, Nanog, Vimentin and Lysozyme who mark stem cells, non-epithelial cells and synovial cells respectively. Weak expression of GFAP and no expression of CD31, Synaptophysin and Pan-cytokeratin, as seen in Figure 7, discarded vascular, neuronal and epithelial origin of the cells. Except for GFAP, in which a smaller number of cells exhibited weaker cytoplasmic immunolabelling at P3 when compared with P0, there was preservation of immunoexpression of all the antibodies between passages P0 and P3. These combination of the

positive and negative expression of these different markers, confirm the expected mesenchymal origin of the cells. Figure 8 presents immunolabeling of eSM-MSCs.



**Figure 8.** Immunolabelling of eSM-MSCs P0 and P3. Magnification 600x. Images present positive Ag expression to Oct-4 and Nanog confirming stem cell origin, to Vimentin confirming non-epithelial origin of the cells and to Lysozyme confirming synovial origin of the cells. Positive expression was revealed by cytoplasmatic staining of the cells. CD31, Synaptophysin and Pan-cytokeratin had negative expression, did not stain, and confirmed these cells had no vascular, neuronal or epithelial origin. GFAP represents a neuronal origin and had a weak expression in P0, which reduced in P3.

### 3.2. Treatment Results

No horse had any adverse event that required study cessation, unplanned procedures or additional treatments. All intra-tendinous injections and follow-up procedures had no adverse reactions (inflammation, infection, deterioration of the lesion, increased lameness), as shown by Godwin *et al*, 2012 [39]. No horse had abnormalities identified on the weeks following the injection.

Tendon/Ligament regeneration occurred in a time frame inferior to 30 days in 80% of the cases and between 30-90 days in 20% of the cases. In this study, 8 animals had lesion on right front limb, 6 on left front limb and 2 on right hind limb. There were 14 acute cases and 2 chronic cases. Chronic cases were diagnosed 6 months before our approach.

After day 90, meaning they had completed the proposed rehabilitation physical program, horses started cantering and started returning to their usual work plan. By day 120 *post* first treatment, 87.5% of the animals were back to full work, with exception of the 12.5% who needed more 30 days to return to full work.

All horses returned to the same level of sports activity they had before injury. Tables 6 and 7 summarize recovery progress. In the group that fully recovered at day 30, ultrasound cross-sectional area was fulfilled as well as a good fiber alignment. There was also no evidence of pain and lameness. Below it is presented a transversal and longitudinal ultrasound image of 4 cases on day 1 and day 30. After eSM-MSCs treatment, all horses were submitted to a rehabilitation program as explained in Table 4.

Radiograph exams were performed to rule out the presence of other associated pathologies and regional nerve blocks were performed to better localize the pain/injured region.

**Table 6.** Equine patient and Lesion characterization. Left column characterizes Equine patient: Sex - male (M) or female (F); age measured in years old (yo); sports modality- SM: Show jumping (SJ), Dressage (Dre) and Leisure (Lsr). Right column characterizes Lesion: Structure affected - Superficial Digital Flexor Tendon (SDFT), Deep Digital Flexor Tendon (DDFT) and Suspensory Ligament (SL) – Left branch (LB); affected Limb - Right Frontlimb (RF), Right Hindlimb (RH), Left Frontlimb (LF) and Left Hindlimb (LH).

Equine Patient				Lesion: Tendonitis / Desmitis			
ID	Sex	Age (yo)	SM	Structure	Type	Limb	Evolution
1	M	22	SJ	SDFT DDFT	Acute	LF	Favorable in 30 days
2	F	5	SJ	SDFT	Acute	RH	Favorable in 30 days
3	F	14	SJ	SDFT	Chronic	RF	Favorable in 30 days
4	M	8	SJ	SDFT	Acute	RF	Favorable in 30 days
5	M	7	SJ	LB SL	Acute	LF	Favorable in 30 days
6	M	13	Lsr	SDFT DDFT SL	Chronic	RH	Tendons: favorable evolution in 30 days; SL in 90 days.
7	M	15	SJ	SFDT	Acute	RF	Favorable in 90 days
8	M	11	SJ	SDFT	Acute	RF	Favorable in 30 days
9	F	10	SJ	SDFT SL	Acute	RF	Favorable in 30 days
10	M	9	SJ	SDFT	Acute	LF	Favorable in 30 days
11	F	10	SJ	SDFT	Acute	RF	Favorable in 30 days

12	M	12	Dre	SDFT	Acute	LF	Favorable in 30 days
13	M	14	SJ	SL	Acute	LF	Favorable in 30 days
14	M	7	SJ	SDFT	Acute	RF	Favorable in 30 days
15	M	12	SJ	SFDT	Acute	LF	Favorable in 30 days
16	F	6	SJ	SFDT	Acute	RF	Favorable in 30 days

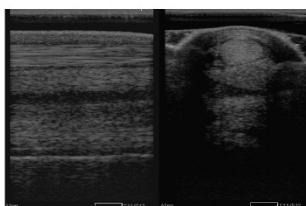
**Table 7.** Ultrasonographic lesion characterization at day 1, 15 and 30. Patient identification, structure affected, lesion ultrasonographic location, cross sectional area and longitudinal fiber pattern are characterized. Assessment outcome is also evaluated. Affected structure superficial digital flexor tendon (SDFT), deep digital flexor tendon (DDFT), Suspensory Ligament (SL) and Left Branch (LB). Lesion-ultrasonographic location (zones 1A-1B, 2A-2B, 3A-3B) [27], Cross Sectional Area % (0 = 0%, 1 = <25%, 2 = >25-50%, 3 = >50-75%, 4>75%) Longitudinal Fiber Pattern (0=0%, 1=<25%, 2=>25-50%, 3=>50-75%, 4>75%) [27] and assessment outcome (full function, acceptable function and unacceptable function) [33].

Patient ID	Day	Structure	Location	Cross Sectional Area	Longitudinal Fiber Pattern (%)	Assessment Outcome
1	1	SDFT	1A-1B	1	1	
		DDFT	1A-1B	1	1	
	15	SDFT	1A-1B	1	1	
		DDFT	1A-1B	1	1	
	30	SDFT	1A-1B	0	0	Full function
		DDFT	1A-1B	0	0	
2	1	SDFT	1A-1B	1	1	
	15	SDFT	1A-1B	1	1	
	30	SDFT	1A-1B	0	0	Full function
3	1	SDFT	1A-1B	1	1	
	15	SDFT	1A-1B	1	1	
	30	SDFT	1A-1B	0	0	Full function
4	1	SDFT	1A-1B	2	2	
	15	SDFT	1A-1B	2	2	
	30	SDFT	1A-1B	0	0	Full function
5	1	LB SL	3A-3B	2	2	
	15	LB SL	3A-3B	2	2	
	30	LB SL	3A-3B	0	0	Full function
6		SDFT	2A-2B	2	2	
	1	DDFT	2A-2B	2	2	
		SL	2A-2B	2	2	
		SDFT	2A-2B	1	1	
	15	DDFT	2A-2B	1	1	
		SL	2A-2B	2	2	
7		SDFT	2A-2B	1	1	Unacceptable function.
	30	DDFT	2A-2B	1	1	
		LS	2A-2B	1	1	Only at day 90.
7	1	SDFT	2A-2B	3	3	
	15	SDFT	2A-2B	3	3	
	30	SDFT	2A-2B	2	2	Unacceptable function.

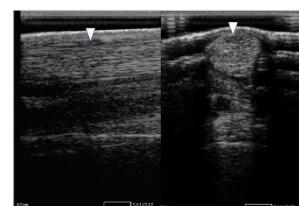
	1	SDFT	2A-2B	2	3	
8	15	SDFT	2A-2B	2	2	
	30	SDFT	2A-2B	0	0	Full function
	1	SDFT	2A-2B	2	1	
9	15	SDFT	2A-2B	1	1	
	30	SDFT	2A-2B	0	0	Full function
	1	SDFT	1A-1B	3	3	
10	15	SDFT	1A-1B	2	2	
	30	SDFT	1A-1B	0	0	Full Function
	1	SDFT	1A-1B	2	2	
11	15	SDFT	1A-1B	2	2	
	30	SDFT	1A-1B	0	0	Full function
	1	SDFT	2A-2B	1	1	
12	15	SDFT	2A-2B	1	1	
	30	SDFT	2A-2B	0	0	Full function
	1	SL	1A-1B	2	2	
13	15	SL	1A-1B	1	1	
	30	SL	1A-1B	0	0	Full function
	1	SDFT	2A-2B	1	1	
14	15	SDFT	2A-2B	1	1	
	30	SDFT	2A-2B	0	0	Full function
	1	SDFT	2A-2B	2	2	
15	15	SDFT	2A-2B	2	2	
	30	SDFT	2A-2B	0	0	Full function
	1	SDFT	2A-2B	2	2	
16	15	SDFT	2A-2B	2	2	
	30	SDFT	2A-2B	0	0	Full function

**Lesion type 0**

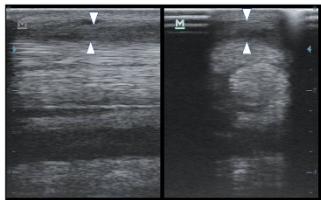
Fiber alignment and cross sectional area loss of echogenicity: 0%

**Lesion type 1**

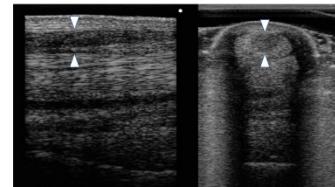
Fiber alignment and cross sectional área echogenicity loss &lt;25%

**Lesion type 2**

Fiber alignment and cross sectional área echogenicity loss: 25-50%

**Lesion type 3**

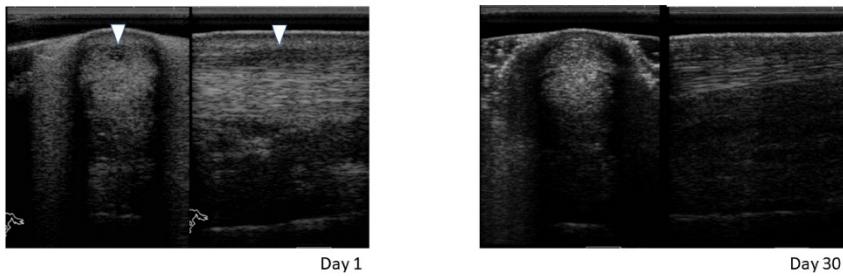
Cross Sectional Area and Fiber alignment hipoechogenicity: 50-75%



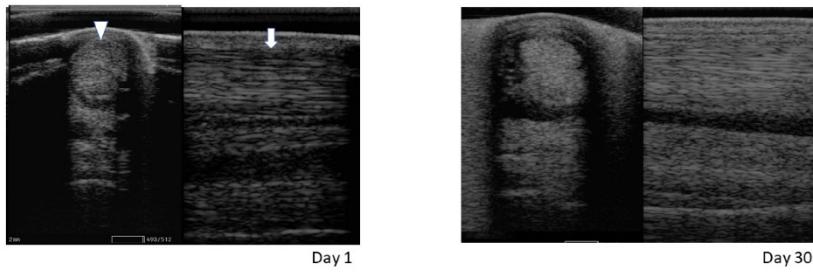
**Figure 9.** Illustration of ultrasonographic lesion characterization mentioned at table 7. Longitudinal fiber alignment and cross sectional area echogenicity loss is presented [27].

Ultrasound images at day 1 and day 30 are very clarifying of the evolution of tendon regeneration. Changes in echogenicity, fiber alignment and cross-sectional area are evident as may be seen in Figure 10.

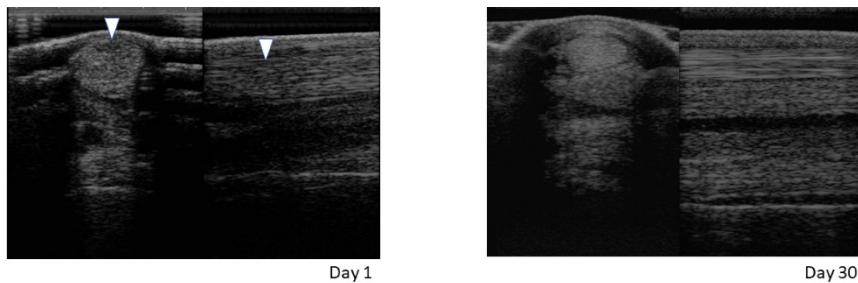
Case number 4: Transverse and Longitudinal ultrasound images at day 1 and day 30 after first treatment.



Case number 14: Transverse and Longitudinal ultrasound images at day 1 and day 30 after first treatment.



Case number 16: Transverse and Longitudinal ultrasound images at day 1 and day 30 after first treatment.



**Figure 10.** Ultrasound images of cases number: 4, 14 and 16 that represent the clinical cases concerning Superficial Digital Flexor Tendon (SDFT). Transversal and longitudinal images at day 1 and day 30 after first treatment with eSM-MSCs. These ultrasounds are representative of the cases and very illustrative of good fiber alignment and cross-sectional area reduction, evidencing tissue regeneration.

#### 4. Discussion

Over the last years, eSM-MSCs have become an interesting subject for those who study cellular and cell-based therapies due to their promising ability to promote tissue regeneration with high capacity of regeneration of articular structures, tendon and ligaments. Regarding the collection, isolation, expansion, freezing and thawing protocols used in this clinical trial, it was possible to use these cells in equine tendon regenerative treatments. The fully characterization of eSM-MSCs

presents itself a big challenge since eSM-MSCs are not as studied as MSCs from other species, namely human MSCs. However, in this study, their stemness and origin has been confirmed through different processes: trilineage differentiation, karyotype, secretome and immunohistochemistry. All SM-MSCs cultures presented monolayer culture, plastic-adherence capacity and fibroblast-like shape [40–43], accomplishing some of the minimal criteria defined by ISCT. Successful osteogenic, chondrogenic and adipogenic differentiation has also been demonstrated. *De Bari et al.*, [9] were to first group to isolate MSCs from synovial tissues.

Karyotype presented some genomic variations when the number of passages was increased. That is consistent with some studies regarding genomic variations along cell passages.[44–48] DNA replication is a critical event for timely genome duplication. Errors in replication lead to genomic instability across evolution [49]. Prieto Gonzalez et al., consider that genomic instability, incurred during the process of stem cell isolation, culture expansion, and reprogramming, might be the most critical point of a stem cell-based therapeutic approach as a viable option in the clinical perspective [50]. Peterson et al., highlighted there is very little evidence linking genomic abnormalities, for example, in human Pluripotent Stem Cell (hPSC's) with tumorigeneses.[44] The frequency and effects of variations is increasing with the development of even more sensitive methods for detecting genomic variation [45].

As reported by *Simona Neri*, interpretation of genetic instability and senescence of cultured MSCs is controversial, but the increasing incidence of genetic alterations at advanced culture times clearly indicates that few culture passages correspond to a reduced chance to harbour dangerous alterations. Therefore, a prudential behaviour is desirable with reduction of culture times as much as possible to avoid safety concerns[51]. More studies must be performed in this area.

During the last decade it has been shown that MSCs therapeutic effectiveness is due mainly to the release of paracrine factors, namely CM, composed of soluble (cytokines, chemokines and growth factors) and non-soluble factors (extracellular vesicles) that are primarily secreted in the extracellular space by the stem cells [52]. CM's paracrine signaling can be considered as the primary mechanism by which MSCs contribute to healing processes, becoming their study an interesting subject [53,54].

In our study, eSM-MSCs revealed a CM with a high level of KC/GRO, MCP-1, IL-6, FGF-2, G-CSF, GM-CSF and IL-8. This highlights the fibroblast intense activity, producing KC/GRO that is chemotactic for neutrophils during inflammation. MCP-1 is essential for reperfusion and the successful completion of musculoskeletal tissue after ischemic injury [55]. Macrophages are tissue resident cells involved in tissue regeneration beside their inflammatory and infection response [56]. IL-6 is a pro-inflammatory and angiogenic interleukin capable of increasing the expression of growth factors, reactivating for example, intrinsic growth programs of neurons, promoting axonal regrowth, creating a link between inflammation and tissue regeneration [57,58]. FGF-2 is a recognized GF responsible for proliferation of tenogenic stem cells. FGF-2 signaling has been reported to produce a tendon progenitor population that expressed scleraxis during somite development [59]. FGF-2 plays a crucial role in cell proliferation and collagen production, becoming a useful GF for tissue regeneration by promoting stem cell proliferation [60]. G-CSF is a cytokine that mobilizes bone marrow-derived cells (BM-DCs) to peripheral blood. A study suggests that injection of G-CSF promoting BM-DCs release in the target area - rotator cuff - effectively enhanced rotator cuff healing by promoting tenocyte and cartilage matrix production [61]. *Wright et al.*, presented a study confirming skeletal muscle damage, including that following strenuous exercise, induces an elevation in plasma G-CSF, implicating it as a potential mediator of skeletal muscle repair [62]. Recent human trials have shown the benefits of G-CSF administration as a treatment for neuromuscular diseases, considering that G-CSF affects skeletal muscle, leading to functional improvements [63–68]. GM-CSF is an hematopoietic growth factor with pro-inflammatory functions [69]. Major sources of GM-CSF are T and B cells, monocyte/macrophage endothelial cells, and fibroblasts. Neutrophils, eosinophils, epithelial cells, mesothelial cells, Paneth cells, chondrocytes, and tumor cells can also produce GM-CSF [70]. *Paredes et al.*, evidenced that elevated levels of pro-inflammatory factors such as those found at these cells CM (GM-CSF, G-CSF, IL-6, IL-8 and IL-17), were implicated in the activation of resident tendon cells for effective healing, stimulating tendon cell proliferation [71,72]. IL-8 is one of the major

mediators of inflammatory response and is a potent angiogenic factor. This is similar to IL-6 but has a longer half-life [73].

A recent study highlights hematopoietic factors promote tendon healing in aged mouse tendons. Histochemical results demonstrated vascularization of the injury site was significantly elevated. It was concluded that vascular endothelial growth factor (VEGF) not only plays an important role in decreasing adipocyte accumulation but also improves vascularization of the tendon during aged tendon healing. Active regulation of VEGF may improve the treatment of age-related tendon diseases and tendon injuries [74].

Studies with human BM-MSCs using a human-specific proteome profiler array with different angiogenic factors such as VEGF-A, IL-6, IL-8, platelet-derived growth factor A (PDGF-A), endothelin-1 (ET1), and urokinase plasminogen activator (uPA), which has not been previously reported in the CM of human MSCs, has also been identified in the equine one, confirming what we found in this study [75]. This factor has been proposed as a modulator of the different neovascularization stages, through the enhancement of VEGF gene promotor activity [75,76]. Schokry *et al.* [77] reported that BM-MSCs therapies have recovery times of 3-6 months and conservative therapeutic methods allow recovery in 12-18 months without regeneration but formation of fibrous scar tissue. Retrospectively, no re-injuries of tendons have occurred in horses treated with this new approach, during the study frame time. In the literature [78], Smith *et al.*, referred as a low percentage re-injury rate 27% for SFD tendonitis treated with bone marrow stem cells. Horses returned to "full function" as defined by Cook *et al* and modified by Guest *et al.*, [33,79].

A study using a murine osteoarthritis (OA) model demonstrated that an injection of MSCs CM, similarly to injection of MSCs, resulted in early pain reduction and had a protective effect on the development of cartilage damage in a murine OA model, by using the regenerative capacities of the MSCs-secreted factors [80].

Interestingly, the results accumulated so far have provided evidence that veterinary patients affected by naturally occurring diseases should provide more reliable outcomes of cell therapy than laboratory animals, thus allowing translating potential therapies to the human field. More recently, a cell-free therapy based on MSCs-CM has been proposed. Even though there are very few clinical reports to refer to in veterinary medicine, recent acquisitions suggest that MSCs-derived products may have major advantages compared to the related cells, e.g., they are considered safer and less immunogenic [52]. As evidenced before, eSM-MSCs CM factors are able to promote tendon healing by reducing inflammation and fatty infiltration, stimulating cell proliferation and tenogenic differentiation [81].

In this study we used a cell-based therapy instead of CM itself, but we are aware of its effect and potential on cell-based therapies, its advantages and therapeutic effects, reason why this study was performed.

To better characterize the cells under study, we carried out immunohistochemistry assays. The markers choice was based on a previous work [8] and include several of the criteria used for humans as determined by the ISCT. Results of our study demonstrated the presence of embryonic stem cell markers – Oct 4 and Nanog. Detection of these markers was previously described by Beltrami *et al.*, in multipotent adult stem cells (HMASC) from bone marrow human [82], as well as, by Riekstina *et al.*, who also demonstrated the presence of these markers in HMASC derived from bone marrow, adipose tissue, heart and dermis[83]. Greco *et al.*, also evidenced elevated expression of Oct 4 in P3 MSCs and hypothesized OCT 4 expression could be an indicator of MSC differentiation potential in clinical diagnostics [84]. In equine characterization of synovial fluid and membrane-derived MSCs, Prado *et al*, also evidenced the presence of Nanog and Oct 4 markers [19]. In contrast, Fulber *et al.*, had no positive results for these two markers, in equine mesenchymal stem cell of synovial tissues [43]. Vimentin, a mesenchymal stem cell marker was also detected, suggesting mesenchymal origin. The presence of Lysozyme confirms the synovial origin of the cells as stated by Fulber *et al* [43].

Immunohistochemistry analysis showed absence of expression of CD31, sinaptophysine and Pan-cytokeratin, discarding the vascular, neuronal and epithelial origin of our cells. GFAP was weakly expressed, being less expressed in P3 than in P0 cells. CD31 was performed to investigate the

presence of hematopoietic cells in eSM-MSCs. The expression of VEGF was not found, being this results similar with those from *Fulber et al.*, and other authors that evidenced the absence of hematopoietic markers [43,85]. The absence of neuronal and dermal markers was also consistent with other studies [19,43].

In our clinical trial, we manage to treat mainly early acute lesions. 87.5% of the cases were acute lesions of tendons or ligaments. This concerns with having a master bank cell of allogenic eSM-MSCs that allows treatments in early acute phases versus treatments with autologous cells where time of tissue collection, preparation and cell culture need to be considered. Furthermore, cells harvesting for autologous treatment is an invasive procedure unnecessary with this new product. The possibility of having a master cell bank enables faster healing of the organ and a quicker return to sportive life. Horses spend less time in recovery time and have a regenerated tissue instead of a fibrotic tissue. These are some advantages of this mixture. Another one concerns with the fact that in early stages of lesion there is inflammatory phase, the paracrine factors released by eSM-MSC's also have anti-inflammatory action, reducing inflammation.

Chronic cases represented 12.5% of the cases, involving 4 structures. Three of them recovered in 30 days and one of them had a delayed recovery time.

The delayed recovery time in 20% of the structures, meaning 12.5% of the animals, was due to, in case 6, increased number of involved structures (more than one tendon or ligament) and foot conformation abnormality, the horse had a fetlock hyperextension that was disabling the correct tendon healing. This was corrected with special shoeing. Inappropriate rehabilitation program (case number 7) was another cause of delayed recovery time. As soon as the corrective shoeing was performed, ligament regeneration started.

Kamm et al, 2021, conclude that based on the evidence to date, tendons appear to have improved healing when treated with allogeneic MSCs, and the use of these treatments in equine tendon and ligament lesions is warranted [86]. Colbath et al., 2020, claimed that some of the advantages of using allogenic stem cells, include the ability to bank cells and reduce the time to treatment, to collect MSCs from younger donor animals and the ability to manipulate banked cells prior to administration [87]. Some of disadvantages focused on the risk of immunological reactions. However, nowadays there are several studies in horses accumulating evidence that allogeneic MSCs maybe a safe alternative to autologous MSCs [87]. Nevertheless, donor's health must always be taken in attention as well as his age [88].

## 5. Conclusions

To sum up, this study accomplishes the criteria for reporting veterinary and animal medicine research for MSCs in orthopedic applications [33] and the ISCT perspective on immune assays for MSC's criteria for advanced phase clinical trials [89], confirmed by plastic adherence, tri-lineage differentiation, synovial membrane origin, spindle shaped cells, proliferative and immune modulatory capacity proven by immunohistochemistry and CM.

From a clinical point of view, the idea of having an allogenic eSM-MSCs cell bank is very interesting. Therefore, the possibility of having a universal donor who can provide a large amount of eSM-MSCs, to culture and preserve non-immunogenic cells whose availability is immediate, allowing a quick and effective therapeutic answer in acute stages of musculoskeletal lesion is the paramount of orthopedic medicine.

In a "One-health" perspective, equines play an important role as a model for human musculoskeletal disorders, the high-level analogy between human and equine structures may have a great translational value for both species for future clinical aspects [28,90]. There are great resemblances between equine SDFT and human Achilles tendon in the size of anatomical structure and load, function (energy store), pathophysiology of tendon injury, and the healing response under activity or traumatic rupture compared to other species [90]. Moreover, considering the result of tendinopathy in equine species which reflects the conditions encountered in human, horse is accepted as an appropriate model in this area by research community and other authorities such as the US Food and Drug Administration (FDA) and the European Medicines Agency (EMA).

Based on the clinical, ultrasonographic and performance outcomes identified in the present study, the use of eSM-MSCs together with autologous serum solution has proven its efficiency for tendon and ligament repair and contribute to reduce the recovery period and subsequent rapid return to athletic activity. The therapy demonstrated to be safe and had no adverse findings. The clinical results and sportive outcome of the horses were very positive. Comparing our study with others, using for example BM-MSCs, it seems our new approach has smaller recovery times and less re-injuries [39,77]. These results encourage the use of eSM-MSCs and autologous serum for the treatment of tendonitis and desmitis as they can regenerate tendon and ligament tissue and regain organ function, enhancing the return to competition in excellent time frames.

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## Abbreviations

AAEP	American Association of Equine Practitioners
AE1/AE3	Pan-Cytokeratin
BM-MSC	Bone Marrow mesenchymal stem cell
CD	Cluster differentiation
c-Kit	Proto-oncogene receptor tyrosine kinase or stem cell factor receptor
CM	Conditioned Medium
cm <sup>2</sup>	Square centimetre
Coll-II	Collagen type II
COMP	Cartilage oligomeric matrix protein

CPD	Cumulative population doublings
DMSO	Dimethylsulphoxide
DPBS	Dulbecco's Phosphate Buffered Saline
eSM-MSC	Equine synovial membrane mesenchymal stem cell
FBS	Fetal bovine serum
FGF-2	Basic Fibroblast Growth Factor
G-CSF	Granulocyte Colony Stimulating Factor
GFAP	<i>Glial fibrillary acidic protein</i>
GM-CSF	Granulocyte-macrophage Colony Stimulating Factor
GRO/KC	Human Growth-regulated oncogene/Keratinocyte Chemoattractant
ICBAS-UP	Instituto de Ciências Biomédicas Abel Salazar – Universidade do Porto
IL	Interleukins
ISCT	International Society for Cellular Therapy
IV	Endovenous
kg	Kilogram
MCB	Master Cell Banks
MCP-1	Monocyte Chemoattractant Protein-1
mg	milligram
MHC-II	Major Histocompatibility Complex
MHz	Megahertz
min	minutes
mL	millilitre
MSCs	Mesenchymal Stem Cell
NSAIDs	Non-steroidal anti-inflammatory drugs
OA	Osteoarthritis
OCT-4	Octamer-binding transcription factor 4
ORBEA	Organismo Responsável pelo Bem-estar Animal
P	Passage
PBS	Phosphate-buffered saline
PRP	Platelet-rich plasma
RT-PCR	Reverse transcription polymerase chain reaction
SID	Once a day
SM-MSC	Synovial Membrane Mesenchymal Stem Cell
UDP	Uridine Diphosphate
UDPGD	Uridine Diphosphate Glucose Dehydrogenase UDPGD
VEGF-R1	Vascular endothelial growth factor
WCB	Working cell banks

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