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Clinically Effective Molecules of Natural Origin for Obesity Prevention or Treatment

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Abstract: The prevalence and incidence of obesity and the comorbidities linked to it are increasing in the world population. Current therapies for obesity and associated pathologies have proven to cause a broad number of adverse effects and often, they are overpriced or not affordable for all patients. Among the alternatives currently available, natural bioactive compounds stand out. These are frequently contained in pharmaceutical presentations, nutraceutical products, supplements, or functional foods. The clinical evidence for these molecules is increasingly solid, among which epigallocatechin-3-gallate, ellagic acid, resveratrol, berberine, anthocyanins, probiotics, carotenoids, curcumin, silymarin, hydroxy citric acid, and α -lipoic acid stand out. The molecular mechanisms and signaling pathways of these molecules have been shown to interact with the endocrine, nervous, and gastroenteric systems, as well as regulate the expression of multiple genes and, therefore, proteins involved in starvation-satiety processes, activation of brown adipose tissue, increased lipolysis, decreased lipogenesis, and inflammation, beneficial changes in metabolism and improved insulin sensitivity. This review provides a comprehensive view of nature-based therapeutic options to address the increasing prevalence of obesity. It offers a valuable perspective for future research and subsequent clinical practice, addressing everything from the molecular, genetic, and physiological bases to the clinical study of the bioactive compound.

Keywords: natural compounds; functional foods; carotenoids; resveratrol; ellagic acid; berberine; probiotics; obesity; treatment; cardiovascular risk factors

1. Introduction

Obesity is defined as an abnormal or excessive accumulation of fat that represents a health risk, and it is characterized by reaching a body mass index (BMI) equal to or greater than 30 kg/m2 [1]. The magnitude of this problem extends globally and poses a public health crisis. Its prevalence continues to increase in all age groups in Europe and some American countries [2]. It is estimated that more than 60% of the population is obese or overweight [1,2]. Ischemic heart disease, the main cause of death worldwide, is closely linked to obesity [3]. It is also related to ischemic or hemorrhagic strokes [4], which represent the second global cause of death, as well as high blood pressure, type 2 diabetes mellitus (DM2), metabolic syndrome, and cancer [3,5]. Furthermore, obesity is correlated

with significant causes of work disability, such as low back pain and osteoarticular diseases, which significantly impact the financial resources of healthcare systems [5]. Despite this, the general population still fails to fully grasp the severity of this metabolic disorder [2].

Obesity has a complex origin, but diet is crucial to its development. In addition to the energy imbalance, some foods promote obesity, including simple carbohydrates such as sucrose (the main disaccharide of cane sugar), glucose and fructose (monosaccharides), highly hydrolyzed starches found in flour, saturated fatty acids, polyunsaturated fatty acids that promote the pro-inflammatory pathway of eicosanoids, such as arachidonic acid and linoleic acid, ultra-processed foods that contain a large number of additives, sweeteners, food colorings, preservatives, among others, related to obesity and other chronic non-communicable diseases. On the other hand, there are foods with molecules capable of regulating gene expression and metabolic pathways in a favorable way, known as phytochemicals, which are characterized by having a high antioxidant and anti-inflammatory power, among which the secondary metabolites of plants stand out, vegetables and fruits such as those derived from phenolic compounds, derivatives of terpene compounds, some alkaloids, as well as some methylated purines (pseudoalkaloids). Likewise, plant-based primary metabolites are linked to health benefits and combating obesity. Some of these molecules deserve special attention since the treatment of obesity requires multiple interventions, not only with physical activity and nutritional therapy, but also with pharmacological therapy and/or surgery in specific cases, but the currently available drugs have a high incidence of adverse effects and have variable effectiveness among patients. Many naturally occurring molecules have shown favorable effects in vitro or in vivo, but only some have been studied in humans. It is important to understand the mechanisms of action of the natural compounds or molecules that have shown clinical efficacy and analyze the results obtained in the clinical trials carried out to date. This review aims to analyze the clinical evidence and assess the efficacy and tolerability of key natural molecules in treating obesity.

2. Etiopathogenic mechanisms of obesity

Obesity is a chronic disease of multifactorial etiology that involves an energy imbalance, genetic and epigenetic factors, alterations in glucose and lipid metabolism, disorders of adipose tissue functioning, neuroendocrine dysregulation, and alterations in the intestinal microbiota, among others [6–8].

2.1. Genetic factors

It is estimated that the genetic load may be responsible for the development of obesity by 40 to 70% [6]. So far, researchers have identified 227 gene variants and over 300 SNPs linked to obesity [9]. In genome-wide association studies, more than 1,100 independent obesity-associated loci have been identified. Although monogenic obesity can happen, polygenic alterations are more frequently observed.

Some of the most studied alterations include the mutation of the leptin (LEP) and leptin receptor (LEPR) genes, which causes a deficiency of the LEP protein and, therefore, alterations in the satiety stimulus with increased appetite, decreasing the caloric expenditure. Some rearrangements in the sequence of the proprotein convertase subtilisin/kexin type 1 (PCSK1), proopiomelanocortin (POMC), and melanocortin-4 receptor (MC4R) genes have also been studied, which cause the expression of the Agouti-related protein that when binding to the melanocortin-2 receptor accessory protein (MRAP) 4 increases the appetite. Changes in the Ras2 suppressor protein kinase gene (KSR2) affect energy consumption and expenditure by interacting with adenosine monophosphate-activated protein kinase (AMPK) and the cyclic-adenosine monophosphate response-element-binding protein 3 -regulatory factor (CREBRF), which participates in the storage and use of cellular energy [6].

Other widely studied alterations include loss-of-function variants of the gene that encodes adenylate cyclase 3, which is widely distributed, predominantly in adipose and subcutaneous tissue, but also participates in the correct functioning of the MC4R gene [10] and participates in the synthesis of cyclic adenosine monophosphate (cAMP), which functions as a second messenger in the glucagon-like peptide-1 (GLP-1) signaling pathway, the hormone ghrelin and the melanocortin-stimulating

hormone [10], POMC, FTO (Fat Mass and Obesity) [11], interleukin 6 (IL-6) [12], perilipin (PLIN) [13], adiponectin (ADIPOQ) [14], LEP [15], LEPR [16], POMC [17] genes, the family of peroxisome proliferator-activated receptors (PPAR) [18] and the uncoupling proteins (UCPs) [19], which are related to the activation of thermogenesis. These genes and their metabolic and signaling pathways have a close relationship with systemic oxidative stress and inflammation, both critically related to the development of obesity and the appearance of many linked comorbidities [6,20].

2.2. Epigenetic factors

Regardless of pre-existing genetic modifications, epigenetics plays a fundamental role in the expression of the obesity phenotype. Epigenetic modifications are modulations in gene expression without DNA modifications [21]. This modulation can be done through DNA modifications, histone alterations, non-coding RNAs, or ATP-dependent chromatin remodeling complexes [22].

Histone methylation or DNA methylation are common modifications through which some environmental factors can prevent the expression of specific genes or DNA segments [20,21]. Histone methylation consists of adding a methyl group to lysine and arginine of histones, which are DNA packaging proteins. Methylated histones adhere strongly to the DNA, keeping the chain condensed, and, in this way, do not allow the integration of transcription factors, which means that the information from that section of DNA cannot be expressed. This process is also known as transcriptional gene silencing (TGS). DNA methylation occurs because DNA-methyltransferase adds a methyl group to cytokines, generally in regions rich in guanine and cytosine. These regions are called "CpG islands" and are part of the non-coding DNA, but their importance lies in the fact that they are usually located in the promoter region of most genes [22,23]. DNA methylation of CpG islands, in addition to maintaining the compact configuration of chromatin, prevents the integration of transcription factors and thus also produces gene silencing. Other modifications include acetylation, which favors gene expression; deacetylation, which inhibits gene expression; or modulation of the expression of some non-coding RNAs, such as microRNAs (miRNAs) [20,22,24].

The main importance of epigenetics is that unlike genotype and genetic changes, epigenetic changes are modifiable and, in many cases, reversible [24]. However, epigenetic modifications that persist can be replicated and conserved in mitosis or meiosis [22,24,25] so they can be transmitted to subsequent generations [26,27]. Epigenetic modifications can be induced by external factors such as diet, physical activity, inflammatory processes, stress, etc., and they can occur during pregnancy or throughout life [21,25], with critical moments of greater susceptibility in the fetal and neonatal stages. Although some prenatal epigenetic modifications are unstable, others remain in adulthood but can be corrected with interventions, such as exercise or favorable dietary changes [21,25,28].

Maternal malnutrition or obesity can cause intrauterine modifications that affect DNA methylation [21,26]. Similarly, obesity *prior* to pregnancy or overnutrition during pregnancy favors a pro-inflammatory state or insulin resistance that stimulates the release of pro-inflammatory molecules such as interleukins, tumor necrosis factor (TNF-α), and the expression of adipogenic factors in the mother and the fetus [26]. Exposure to these alterations can cause histone modification and DNA methylation, with a consequent increase in the expression of adipogenic and lipogenic genes that favor obesity in other stages of extrauterine life [26,27,29]. One of the genes studied is POMC, which, in cases of hypermethylation, can disrupt the regulation of food intake even in the presence of leptin and insulin [21]. Maternal obesity also causes methylation and increases leptin expression, leading to hypersecretion and functional resistance to leptin from infancy or later in life [26]. Likewise, a high-fat diet during pregnancy produces greater acetylation of histone H3K14 and lower sirtuin-1 (SIRT1) gene expression in the liver and heart. This decrease in SIRT1 expression favors the presence of fatty liver, obesity, DM2, and diabetic cardiomyopathy [30,31].

The diet we have throughout our lives is crucial in epigenetics. Consuming high levels of saturated fat leads to DNA methylation in adipose tissue and alters the expression of 28 messenger RNAs. A diet with polyunsaturated fats induces multiple methylations, but these do not generate changes in gene expression, although both diets have a similar effect on weight gain [21]. High concentrations of palmitate increase histone acetyltransferase activity [27]. On the contrary, calorie

restriction for more than eight weeks reduces hypermethylation in some genes affected by a high-fat diet. However, the adverse effects of insufficient overfeeding are quickly noticeable, while the positive effects of a calorie-restricted diet take longer to impact DNA methylation changes [21,27]. The above partly explains the benefits produced by metabolic surgery (restrictive, malabsorptive or combined technique) [32], which causes a sudden and sustained reduction in caloric intake, contributing to energy balance, after six to 12 months induces favorable effects on promoter methylation of the pyruvate dehydrogenase kinase 4 (PDK4) gene and proliferator-activated receptor coactivator g-1 alpha (PGC-1a), among other genes [21,33]. Dietary folate deficiency has been related to a more significant volume of fat mass, insulin resistance, increased risk of DM2, decreased DNA methylation of lymphocytes, and changes in DNA methylation of 236 CpG sites and genes associated with obesity [27].

A sedentary lifestyle, exercise, and moderate or intense physical activity have also been recognized as epigenetic modifiers. In individuals with a history of low physical activity, exercise for six months produced changes in the DNA methylation of 7663 genes, including some related to obesity, such as the cAMP response element-binding protein 4 (CREB4), elongation of very long-chain fatty acid proteins (ELOV), glucose transporter 4 (GLUT4), and hormone-sensitive lipase (HSL) [23,27]. Furthermore, a relationship has been reported between exercise and the expression of proinflammatory cytokines such as IL-6 and TNF- α [23,34]. Subjects who exercise periodically have lower expression of proinflammatory cytokines and are better adapted when they are released. Although exercise-induced methylation changes can be observed from the first session, the duration of this epigenetic modification is longer with chronic exercise exposure [23].

2.3. Energy imbalance

The primary cause of obesity is excessive caloric intake compared to the individual's requirements [7]. This explains the growing incidence of obesity since the consumption of foods high in fat and sugar has increased globally, coupled with decreased physical activity [7,35,36]. However, the fundamental cause of obesity is a state of positive energy balance characterized by caloric intake greater than energy expenditure. This originates from a complex interaction of behavioral, environmental, physiological, genetic, and social factors influenced by interactions between individuals, families, institutions, organizations, and communities. Although each of these levels is managed autonomously, they all have strong interconnections, from the molecular to the social level [8].

Many elements and variables are directly and indirectly associated with altering the regulation of energy balance. Still, the fundamental problem is found in insufficient physical activity, known as a sedentary lifestyle, which leads to a marked decrease in total energy expenditure. A sedentary lifestyle also changes body composition since it causes loss of muscle tissue, decreased bone density and water percentage, and a gain in subcutaneous and visceral adipose tissue [23,35]. When obesity occurs during childhood or adolescence, it causes a significant increase in the number of adipocytes, which predisposes the individual to become obese more easily in later life stages, as well as more difficulty in losing fat mass [26].

2.4. Neuroendocrine dysregulation

The regulation of energy expenditure and intake involves several body systems, such as the nervous system, digestive system (including the liver and pancreas), and adipose tissue cells, which play a crucial role [37]. Obesity is correlated with the dysregulation of many signaling pathways, including hormones, cytokines, neurotransmitters, neuropeptides, adipokines, chemokines, growth factors, proliferation molecules, differentiation factors, antioxidant enzymatic systems, non-enzymatic antioxidants, and the overexpression or underexpression of many genes [38]. These alterations can begin gradually and progressively, activating other neuroendocrine responses and promoting dysfunction. We summarize the principal occurrences of this pathophysiology below.

The foods consumed are digested into macronutrients (carbohydrates, lipids, and proteins) to be used in the body, and the excess nutrients are stored for later use. In the presence of insulin, excess

glucose is polymerized into glycogen in the liver and striated muscle. Excess macronutrients can only be stored in adipose tissue, which requires being transformed into triglycerides in the liver. In its catabolic process, glucose generates acetyl coenzyme A (Acetyl-CoA) that can pass into the anabolic pathway for the synthesis of fatty acids (lipogenesis), mainly in adipose tissue and the liver [39]. Amino acids are catabolized to generate intermediates of the Krebs cycle from where they can leave the mitochondria as citrate and be metabolized to form Acetyl CoA for lipogenesis [40,41]. These fatty acids are re-esterified and linked to glycerol-3-phosphate to form triglycerides, which are transported from the liver in very low-density lipoprotein (VLDL) molecules and stored in adipose tissue [42]. Mature adipocytes can store a large number of triglycerides, increasing their volume up to 20 times through hypertrophy [39]. Furthermore, this excessive growth favors adipocyte hypoxia, the release of reactive oxygen species (ROS), and inflammatory cytokines that activate another mechanism to increase storage capacity, adipogenesis (adipocyte hyperplasia) [43].

This process of adipogenesis develops from multipotent mesenchymal stem cells related to preadipocytes, which remain available in the vascular stroma of adipose tissue and undergo mitotic clonal expansion and differentiation of adipocytes to white adipose tissue that has triglyceride storage functions, or to brown adipose tissue with thermogenesis functions [44]. The CCAAT/enhancer-binding protein β (C/EBP) β and C/EBP δ , which are highly sensitive to adipogenic stimuli, increase from the initial stages [43,45,46] and stimulate the expression of the C/EBP α , sterol regulatory element-binding binding protein 1 (SREBP-1) and the expression of the peroxisome proliferator-activated receptor γ (PPAR γ) which has an essential role in the final phases of adipogenesis [46] and regulates the increase of adipogenic factors including C/EBP α expression itself in a positive feedback mechanism [43]. Because adipose tissue is not a passive storage space but rather a tissue with endocrine, paracrine, and autocrine functions, new mature adipocytes are capable of not only increasing storage capacity but also releasing adipocytokines, some of which function as neurotransmitters and participate in the neuroendocrine regulatory complex [47].

In healthy individuals, leptin, an adipocytokine, is typically present at 5 to 15 ng/ml [48]. However, its expression is influenced by various factors, including diet, insulin secretion, glucocorticoids, cytokines, and the amount of adipose tissue [48,49]. Under normal conditions, it can cross the blood-brain barrier by facilitated diffusion and induce satiety and energy consumption by stimulating POMC and inhibiting neuropeptide Y. However, high concentrations induce a state of leptin resistance that favors the presence of obesity and other metabolic alterations by limiting the stimulation of extracellular signal-regulated kinases (ERK) 1 and 2, mitogen-activated protein kinase (MAPK), and other downstream factors [50].

Adiponectin is secreted mainly by adipose tissue and has multiple functions, including (a) its participation as a promoter of the substrate of the insulin receptor IRS 1 and 2, which promotes insulin sensitivity; (b) activator of AMPK, which also promotes insulin sensitivity and

the oxidation of fatty acids, and (c) MAPK activator, which stimulates glucose uptake [51,52]. However, obesity decreases serum adiponectin concentrations with a consequent decrease in insulin sensitivity [14,51]. By reducing insulin sensitivity, insulin-dependent glucose transport decreases, which favors the presence of high plasma glucose concentrations and the consequent glucotoxicity that increases insulin resistance, creating positive feedback with glucose concentrations. Because insulin performs other functions, a series of metabolic pathways are triggered, including increased adipogenesis and lipogenesis [39].

2.5. Gut microbiota

Another important factor in the etiopathogenesis of obesity is the intestinal microbiota. This is the set of communities of living microorganisms colonizing the intestine and plays a critical role in the health-disease balance in the individual. The composition of the human microbiome results from millions of years of coevolution and selective pressure, selecting a specialized community to live in the intestinal environment and achieving a mutualistic relationship and a state of balance with the host that is beneficial, both for humans and for the microorganisms they harbor [53]. Among all the ecological niches found in the human body, the gastrointestinal tract (GIT) is the one with the greatest

diversity and abundance of microbial taxa. In the proximal regions of the GIT, there is a low concentration of microorganisms due to the acidic pH and the rapid transit of the bolus; meanwhile, in the distal region (colon), the most significant number of microbes is found [54].

The intestinal microbiota has a vital role in energy balance since these microorganisms can metabolize an enormous amount of compounds from the diet, produce metabolites related to the hunger-satiety process, as well as a large number of hormones and neurotransmitters such as serotonin, which induces satiety [57–59]. In humans, the gut microbiota mainly comprises five distinct phyla: Actinobacteria, Bacteroidetes, Verrucomicrobia, Firmicutes, and Proteobacteria. Bacteroidetes and Firmicutes represent up to 90% of all intestinal bacteria [55]. The microbiota's impact on host physiology is supported by a growing body of evidence, which highlights multiple pathways, including improved energy utilization, immune system changes, metabolic signaling, and inflammatory pathways [53].

Dysbiosis, an alteration in the composition of the microbiota, has been linked to three distinct mechanisms that can occur at the same time: (a) the loss of "beneficial" bacteria, (b) an overgrowth of potentially dangerous bacteria, and (c) a reduction in microbial diversity [56]. It can also lead to an increase in caloric intake to the detriment of basal and total energy expenditure, but a correlation has also been identified between the existing intestinal microbiota and metabolic and nutritional status. In this way, healthy individuals have a greater amount of the Bacteroidetes than Firmicutes, but industrialized diets low in dietary fiber cause a decrease in Bacteroidetes and an increase in Firmicutes [57,59]. In addition to bacteria, intestinal archaea, fungi, and viruses participate in the etiopathogenesis of obesity [57].

The microbiota can contribute significantly to promoting weight gain, fat storage, and generating insulin resistance [60]. DM2, a pathology associated with obesity, also involves dysbiosis, which can also favor infections due to changes in the regulation of receptors in immune cells. An example, is infection by fungal species of the genus Rhizopus, which is the causal agent of the severe infection called rhinocerebral mucormycosis [61]. The correlation between obesity and the bacterial composition of the human microbiome has been established through previous investigations, with particular attention given to members of the *Christensenellaceae* family, and the Methanobacteriales, *Lactobacillus*, Bifidobacteria, and *Akkermansia* genera [62].

More recently, it has been identified that the *Christensenellaceae* family is linked to weight loss, showing an inverse correlation between its relative presence and an individual's body mass index (BMI) [63]. In particular, it has been shown that *Akkermansia muciniphila* plays a crucial role in regulating body weight, and supplementation with this bacteria significantly improves the metabolic parameters of overweight and obese individuals [64].

The *Lactobacillus* and *Bifidobacterium* genera are recognized for promoting intestinal health. They have been used as traditional probiotics that significantly affect the balance of the intestinal microecology in humans, but the effects on body weight in overweight individuals vary depending on the species of *Lactobacillus*; specifically, a negative correlation was found between the abundance of *Lactobacillus paracasei* and obesity, while the abundance of *Lactobacillus reuteri* and *Lactobacillus gasseri* showed significant correlations for obesity development [65]. Therefore, we can summarize that the participation of the intestinal microbiota in weight control focuses on bacterial diversity, some specific species found, the metabolites that bacteria produce and are absorbed intestinally, the relationship between the microbiota, the immune system, inflammation, and oxidative stress, the participation of metabolites in gene expression, acting as transcription factors and the capacity of the microbiota to degrade, biotransform and metabolize compounds from the diet [66].

2.6. Other etiopathogenic factors

There are medications associated with the development of obesity, known as obesogenic medications, which include mineralocorticoids, glucocorticoids, some antidiabetic drugs, insulin, and some antihypertensive medications, among many others. On the other hand, ubiquitously, mainly in foods (due to agrochemicals) and plastic containers, there are obesogenic molecules known as endocrine disruptors, which interfere with the function and/or signaling of various hormones,

neurotransmitters and neuropeptides, among which contain androgenic and progestin sex hormones, peptides related to satiety and hunger such as LEP, ghrelin, peptide YY, neuropeptide Y, GLP-1, serotonin, etc. [67].

Circadian rhythms related to sleep-wake also play an important role in controlling energy balance, whose deregulation can cause greater hunger and lower basal energy expenditure. Several mediators regulate this system; however, the most important are cortisol, melatonin, growth hormone, and CLOCK genes [68]. There are viruses related to the obesity process, known as obesogenic viruses, among which is adenovirus 36, which increases insulin sensitivity in adipocytes, generating more significant lipogenesis while reducing the expression and secretion of leptin, thus increasing hunger [69]. Likewise, many psychological, cultural, social, economic, climatic, political, and geographical factors determine eating patterns, the type of food available, the amount of physical exercise performed, as well as the quantity and quality of the food consumed [70].

3. Molecules with demonstrated clinical efficacy in the prevention or treatment of obesity

Obesity requires comprehensive treatment that includes pharmacological interventions in many cases. The drugs currently available offer variable results and a high incidence of adverse effects, which motivates research into new molecules. Clinical trials carried out to date have shown that some molecules of natural origin can achieve favorable modifications for the management of obesity.

3.1. Epigallocatechin-3-gallate

Epigallocatechin-3-gallate (EGCG) is a polyphenol flavone-3-ol, catechin ester of epigallocatechin and gallic acid with three aromatic rings linked by a pyran ring, contributing to its functional benefits [71]. It is primarily found in green, red, white, black, and oolong teas derived from the tea plant (Camellia sinensis). Oral administration of EGCG has been shown to have high bioavailability. It is mainly absorbed in the small intestine, while the intestinal microbiota plays a fundamental role in its absorption in the colon [72]. It is suggested that EGCG is transported to the liver or metabolized by the gut microbiota to other potential metabolites because EGCG can be detected in plasma after ingestion, and its metabolites can be identified in bile, plasma, and urine [73].

EGCG exerts its effects mainly by regulating the suppression of hydrogen peroxide and ROS metabolism, the suppression of oxidative stress, the activation of fatty acid transport, and cholesterol oxidation and metabolism. Likewise, EGCG has been shown to significantly increase the expression of FOXO1, Sirt1, CAT, FABP1, GSTA2, ACSL1, and CPT2 proteins while decreasing the levels of NF-κB, ACC1, and FAS proteins in the liver [74].

Studies conclude that EGCG prolongs lifespan by improving free fatty acid metabolism and reducing inflammatory and oxidative stress levels in preclinical models [74]. EGCG also activates the PI3K/AKT molecular pathway, thus increasing the translocation of glucose into the cell and its utilization in cytoplasmic microtubules, improving glycolysis and energy metabolism [75]. EGCG has also been shown to significantly decrease levels of total free fatty acids (FFA), saturated fatty acids (SFA), and the n-6/n-3 ratio, as well as significantly increase n-3 FFA related to longevity and regulating inflammation (Table 1) [76].

EGCG has been shown to intervene in the regulation of various metabolic and molecular pathways involved in obesity, reducing glucose, serum lipids, inflammation, oxidative stress, and blood pressure, which are also involved in premature aging [77]. The results of clinical studies suggest that EGCG can prevent the increase in body weight, BMI, and visceral fat, mainly in individuals under 50 years of age [76]. However, it does not produce a decrease in these variables [76,77]. Table 1 shows the clinical effects of epigallocatechin-3-gallate on obesity and related comorbidities, according to clinical trials and systematic reviews with meta-analysis of clinical trials found in the scientific literature.

Table 1. Clinical effects of epigallocatechin-3-gallate (EGCG) on obesity and associated comorbidities, from clinical trials and systematic reviews with meta-analysis.

Compou nd	Type of study	Dose	Targeted populati on	Observed effect Δ	p	Adverse effects	Referen ce
Epigalloc atechin-3- gallate	Randomize d, double- blind, placebo- controlled clinical trial in 30 patients.	150 mg/day orally, for 8 weeks.	Men and women who are overweig ht or obese.	Significant decrease in plasma triacylglyceri des, systolic blood pressure, and diastolic blood pressure	<0.0 5 in all para meter s.	Unmenti oned	Chatree et al., 2021 [77]
Epigalloc atechin-3- gallate + α- glucosyl hesperidi n	Clinical trial, randomize d, placebocontrolled, doubleblind, and designed in parallel groups with 60 patients.	146 mg of EGCG + 178 mg of α- glucosy l hesperi din/day , orally, for 12 weeks.	Healthy Japanese men and women between 30 and 75 years old.	Reduction in BMI, triacylglyceri des, body fat, visceral fat, and LDL-c/HDL-c ratio.	<0.0 5 in all para met ers.	None.	Yoshito mi <i>et al.,</i> 2021 [76]

3.2. Ellagic acid

Ellagic acid is a polyphenol from the ellagitannin group; it is a derivative of chromene-dione and is a dimer of gallic acid. This compound exhibits a hydrophilic group consisting of four hydroxyl groups and two lactones and a lipophilic group incorporating two hydrocarbon rings [78]. These characteristics give ellagic acid the ability to act as an electron acceptor in various substrates and to participate in redox reactions [79]. Ellagic acid is found in a wide variety of fruits (pomegranates, persimmons, raspberries, strawberries, peaches, plums, kiwis), oilseeds (walnuts, almonds), and vegetables. It can be present in free form or as derivatives that can be hydrolyzed under physiological pH and the intestinal microbiota, thus increasing plasma levels of ellagic acid after ingestion of fruits and nuts [78].

Ellagic acid exerts antitumor, antioxidant, anti-inflammatory, antimutagenic, antibacterial, and antiallergic properties [80]. Its multiple pharmacological properties are because it acts on various signaling pathways, including the VEGFR-2, Notch, PKC, COX-2, PI3K/Akt, JNK (cJun) signaling pathway, and the mitochondrial pathways Bcl-2/Bax, TGF- β /Smad3, MMP SDF1 α /CXCR4, MAPK, PDK-1, mTOR, p-ERK, NRF2, p-JNK. Ellagic acid was also found to reduce the expression of histone

(

deacetylases that could inhibit neovascularization, a common process in cancer and obesity. Mechanism analysis revealed that ellagic acid inhibits hypoxia-induced angiogenesis by suppressing HDAC-6 in ECV304 cells. Furthermore, the knockdown of endogenous HDAC6 via small interfering RNA abolished hypoxia-induced expression of HIF-1 α and VEGF and blocked the Akt activation. This interaction explains the effect of ellagic acid in modulating inflammation and oxidative stress [81,82].

Ellagic acid inhibits adipogenesis in two phases: adipocyte differentiation and clonal expansion [83,84]. This effect is because it produces a downregulation of adipogenic factors such as C/EBPalpha, Krox20, KLF4, and 5 and inhibits the expression of PPAR gamma and aP2 through the decrease in the expression of mRNAs for these proteins, which varies with the concentration of ellagic acid [83]. Furthermore, in the presence of ellagic acid, the mRNA expression of the preadipocyte secretory factor called Pref-1 or Dlk1/FA1, increases, which prevents adipogenesis by activating the ERK/MAPK pathway [83,85]. These regulations produce inhibition of cyclin A expression and phosphorylation of retinoblastoma protein, key regulators of the cell cycle, which stops the progression between the G0/G1 and G1/S phases of preadipocytes found in these early phases, preventing their conversion to mature adipocytes [83,84]. Ellagic acid also interferes with triglyceride synthesis by decreasing the protein Lipin-1 and diacylglycerol acyltransferase-1 expression, which catalyzes the final acylation step in the triglyceride synthesis pathway [83]. This decrease may contribute to the prevention of hyperplasia of adipose tissue.

Table 3. Clinical effects of ellagic acid on obesity and associated comorbidities, from clinical trials and systematic reviews with meta-analysis.

Compou nd	Type of study	Dose	Targeted populati on	Observed effect Δ	p	Adverse effects	Referen ce
Ellagic acid	Double- blind randomize d clinical trial with 32 patients.	1000 mg/day 12 weeks	Men and women with metabolic syndrom e.	Decrease in waist circumferenc e H -2.7cm M -3.8cm	0.03	None	Hidalgo, et al., 2022 [86]
Ellagic acid	Double-blind randomize d clinical trial with 32 patients.	3 mg/day 12 weeks	Overweig ht men and women	Decrease in waist circumferenc e - 0.7cm	< 0.01	None	Shiojima , et al., 2020 [87]
Ellagic acid	Double-blind randomize d clinical trial with 150 patients.	50 mg/day 12 weeks	Overweig ht men	Decrease in waist circumferenc e - 1.5cm	< 0.01	None	Liu, et al., 2018 [88]

In clinical studies, ellagic acid has shown effectiveness in reducing waist circumference [86–88], which represents a decrease in central adiposity due to the correlation between them [89]. The reduction ranged between - 0.7 and - 3.8 cm. This effect is especially relevant due to the close relationship between abdominal fat and the risk of fatal and non-fatal cardiovascular disease [90,91]. Although the impact of ellagic acid on the percentage of body fat in humans has been reported [87], some authors suggest that ellagic acid causes a redistribution of body fat [92] and an increase in brown fat with a decrease in white adipose tissue [93], which can keep the amount of total body fat mass constant. To date, no adverse effects of ellagic acid have been reported, and it has a wide therapeutic margin [86,94]. However, the optimal dose still needs to be defined and subjected to evaluation in phase 3 clinical studies. Table 3 shows the clinical effects of ellagic acid on obesity and related comorbidities, according to clinical trials and systematic reviews with meta-analysis of clinical trials found in the scientific literature.

3.3. Resveratrol

Resveratrol [3,5,4'-trihydroxy-trans-stilbene) is a molecule belonging to phenolic compounds, which acts as phytoalexin in various plants and fruits. Specifically, it is classified within the group of stilbenes (existing in cis and trans configurations). It is found in a great diversity of plants and fruits; the best-known and most used to obtain it is the red grape (Vitis vinifera) [95].

Resveratrol can induce the expression of glutathione-S-transferase (GST) and glucuronyltransferase, both belonging to second-phase hepatic enzymes, and it inhibits several subfamilies of the cytochrome P450 (CYP450), including the CYP1A1 isoenzyme [96]. This comes with important changes in the metabolism of molecules related to cancer, inflammation, and oxidative stress due to the increase in detoxification mediated by conjugation, unlike first-phase hepatic metabolism, which is mediated by redox reactions. Resveratrol also has immunoregulatory properties mediated by decreasing the expression and activation of the TLR4 receptor in monocytes and macrophages, thereby reducing the activation of NF-kB and IL-1 beta, as well as inhibiting the proinflammatory JAK/STAT3 pathway and matrix metalloproteinases 3 and 9 [97]. Resveratrol also inhibits the expression of COX2, thereby reducing the synthesis of proinflammatory prostaglandins of the E2 series [98].

About 70% of dietary resveratrol is absorbed after ingestion. However, only 0.5% is ingested, and resveratrol becomes systemically bioavailable [99]. Resveratrol is perceived as a xenobiotic within the intestine. It passes to the enterocytes lining the small intestine, where it is metabolized (including conjugation with sulfate and glucuronate) with the generation of polar metabolites (to optimize excretion) [95]. Conjugated resveratrol is transported to the blood from the enterocyte by binding to an adenosine triphosphate (ATP)-binding cassette transporter called multidrug resistance protein 3 (MRP3). It is worth mentioning that the enzymes responsible for this Phase II metabolism (including sulfotransferases and glucuronosyltransferases) can manifest genetic polymorphisms that probably underlie some of the interindividual variability in the biological effects of resveratrol and its metabolites [100].

Current preclinical and clinical evidence on the positive effects of resveratrol in the treatment of obesity and related comorbidities is linked mainly to the regulation of various metabolic pathways and signal transduction, with resveratrol acting as a transcription factor, as well as an estrogenic hormone in some cases [101]. The main mechanisms of action of resveratrol are associated with its anti-inflammatory and antioxidant effects, helping to resolve inflammation and activate various endogenous antioxidant enzyme systems such as Nrf2 and ARE (antioxidant response element) for the synthesis of antioxidant enzymes such as superoxide dismutase (SOD), glutathione peroxidase (GSH-Px) and catalase (CAT) [102]. *In vitro* and *in vivo* studies have shown that different doses of resveratrol exert anti-obesity effects on 3T3-L1 adipocytes through various mechanisms, such as induction of apoptosis, decreased fat accumulation and adipogenesis, inhibition of cell differentiation through the activation of AMPK, and the induction of SIRT1-dependent cell apoptosis [103]. In randomized controlled clinical trials, resveratrol has shown significant reductions in weight and BMI compared to the placebo group (p < 0.05) [104-109]. Table 4 shows the clinical effects of resveratrol

on obesity and related comorbidities, according to clinical trials and systematic reviews with metaanalysis of clinical trials found in the scientific literature.

Table 4. Clinical effects of resveratrol on obesity and associated comorbidities, from clinical trials and systematic reviews with meta-analysis.

	terriatic reviews	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,					
Compou nd	Type of study	Dose	Targeted populatio n	Observed effect Δ	p	Adverse effects	Referen ce
Resveratr	Systemati c review and meta- analysis of 36 RCTs.	10 to 200 mg/day orally for 4-12 weeks.	Men and women with overweigh t or obesity, as well as comorbidi ties.	Decrease in body weight, BMI, fat mass, and waist circumference. Increase in lean mass. No significant changes in serum leptin and adiponectin levels.	0.03, 0.01, 0.03, 0.001 and <0.001, respect ively.	None	Tabrizi et al., 2020 [105]
Resveratr ol	Controlle d, randomiz ed, double- blind clinical trial with 71 participa nts.	1 g/day orally for 8 weeks.	Men and women with overweigh t and DM2.	It had no effect on hepatic steatosis or cardiovascula r indices.	Not signific ant.	None	Ali- Sangoun i <i>et al.</i> , 2022 [106]
Resveratr	Systemati c review and meta- analysis of 19 RCTs with a	>1 g/day orally for 4-12 weeks.	Men and women with DM2 and comorbidi ties such as	Reduction of fasting serum glucose and systolic and diastolic blood	<0.0000 1, <0.0000 1, 0.95, 0.66 and	None	Gu et al., 2022 [107]

	total of 1,151 patients.		overweigh t and obesity.	pressure. No significance in waist circumferenc e, serum triacylglyceri des, or HDL- c.	0.14, respect ively.		
Resveratr ol	Systemati c review and meta- analysis of 28 RCTs.	<500 mg/day orally for ≥ 3 months	Men and women with obesity.	Reduction in body weight, BMI, and waist circumferenc e. No effects on fat mass.	0.02, 0.02, 0.009 and 0.16, respect ively.	None	Mousavi et al., 2019 [109]

3.5. Berberine

Berberine is an alkaloid present in plants such as Berberis vulgaris, Berberis aristata, Mahonia aquifolium, Hydrastis canadensis, Xanthorhiza simplicissima, Phellodendron amurense and Rhizoma coptidis [110]. Its effect has been studied as a lipid-lowering, hypoglycemic, antibiotic, antioxidant, potassium channel blocker, and antineoplastic agent [111]. In obesity studies, berberine has decreased adipocyte differentiation through downregulation of C/EBP α and PPAR γ mRNA expression in visceral fat [112]. Furthermore, it upregulates the mRNA expression of the transcription factors GATA-2 and GATA-3 [113]. These factors control the transition from preadipocyte to adipocyte [114], participate in differentiation to brown adipose tissue, and suppress PPAR γ , the basal activity [115].

Berberine also causes deacetylation and stimulation of AMPK expression in myotubes and adipocytes [115,116]. It also activates thermogenesis by activating the AMPK/SIRT1 signaling pathway [117]. On the other hand, it decreases insulin resistance by inhibiting the expression of $C/EBP\alpha$ and the phosphorylation of IKB kinase beta (IKKbeta) Ser (181) and IRS-1 Ser (307) [118–120]. It has been observed that berberine also reduces the expression of the genes LXR, PPAR, SREBP-1c, fatty acid synthase, acetyl-CoA carboxylase, acyl-CoA synthase, and lipoprotein lipase [115,121]. Additionally, other mechanisms associated with berberine have been identified, including its ability to inhibit hepatic gluconeogenesis through the phosphoenolpyruvate carboxykinase (PEPCK), glucose-6-phosphatase (G6Pase), and AMPK enzymes [122]. Many of the effects of berberine in obesity are related to its activity as an AMPK agonist, which also favors GLUT4 translocation and reduces glucotoxicity and insulin resistance, and therefore reduces the lipogenic stimulus.

Besides, berberine inhibits the activation of macrophages by oxidized LDL and the formation of foam cells by improving the expression and translocation of LXR α -ABCA1 and, thus, cholesterol efflux [121,122]. In addition, it modifies the intestinal microbiota, improving the Firmicutes/Bacteroidetes ratio, regulating the production of microbiota metabolites, and consequently causing weight loss and improvement in the lipid profile [123].

In human studies, berberine has been found to have a beneficial impact on gene regulation related to cholesterol absorption when administered at a daily dose of 300 mg and has improved glucose homeostasis with a daily dose of 1 gram [121]. Multiple studies in overweight or obese patients have reported the effects of berberine in reducing waist circumference and waist/hip ratio [124,125]. Some studies have also reported a decrease in body mass index, but the results on body

weight have not been significant in most studies [124,126]. Recent studies report a decrease in visceral fat [127] in overweight patients, in addition to other metabolic effects of berberine, such as an improvement in the glycemic profile and a decrease in insulin resistance, which make it interesting as an auxiliary in the treatment of obesity [119,127].

Table 5. Clinical effects of berberine on obesity and associated comorbidities, from clinical trials and systematic reviews with meta-analysis.

Compou nd	Type of study	Dose	Targeted populatio n	Observed effect Δ	p	Adverse effects	Referen ce
Berberine	Double- blind randomiz ed clinical trial with 49 patients.	1100 mg/day for 8 weeks.	Men and women with impaired fasting glucose and overweig ht.	Decrease in fat mass -1.0 kg, visceral fat - 93.2 g and waist circumference -1.42 cm.	<0.0 01 for the thre e vari able s	None	Rondan elli, et al., 2023 [127]
Berberine	Systemati c review with meta- analysis of 9 clinical trials with a total of 378 individua ls.	1000 to 1500 mg/day for 12-24 weeks.	Men and women who are overweig ht or obese in addition to T2DM or metabolic syndrome	Decrease in body weight. Decrease in BMI - 0.29 kg/m² and waist circumference - 1.78cm	0.00 4 0.00 1	Not mention ed.	Xiong, et al., 2020 [124]
Berberine	Systemati c review and meta- analysis of 12 clinical trials with a total of	1000 to 1500 mg/day for 12-24 weeks.	Men and women who are overweig ht or obese in addition to T2DM or	No change in body weight or BMI Decrease in waist/hip ratio - 0.03	<0.0 01	Not mention ed.	Amini, et al., 2020 [126]

14

849 metabolic individua syndrome ls. .

Although some clinical studies carried out with berberine have reported adverse effects such as constipation, diarrhea, nausea, or abdominal discomfort [128–130], to date, no serious adverse effects have been reported. There are toxicity studies [131] that report a wide range of adverse effects and have a wide therapeutic margin, so it is considered safe and well tolerated [127,132]. More studies are required to analyze whether its effectiveness may be greater in certain specific groups and determine its uses. Table 5 shows the clinical effects of berberine on obesity and related comorbidities, according to clinical trials and systematic reviews with meta-analysis of clinical trials found in the scientific literature.

3.6. Anthocyanins

Anthocyanins are flavonoid pigments of plant origin responsible for the colors of various fruits, flowers, and leaves. Anthocyanins range from orange-red to bright red, violet, and blue. Anthocyanins are water-soluble flavylium salts structurally composed by coupling a sugar unit to an anthocyanidin [133]. There are more than 600 types of anthocyanins, the main differences between them being the position and number of hydroxyl groups, the degree of methylation of the hydroxyl groups, the nature and number of sugar molecules present, and the acids attached to the sugars. Despite the large number of anthocyanin molecules already cataloged in the food matrix, only anthocyanins derived from six types of anthocyanidins: pelargonidin, cyanidin, peonidin, delphinidin, petunidin and malvidin [133,134].

The mechanism of action of anthocyanins is linked to their reducing capacity and their potential to act as a positive and negative transcription factor for many genes related to inflammation/anti-inflammation and oxidation-reduction. Cyanidin-3-glucoside, delphinidin-3-glucoside, and petunidin-3-glucoside can inhibit NF- κ B activities through mitogen-activated protein kinase (MAPK) pathways. In contrast, cyanidins inhibit NF- κ B activities of the enzyme cyclooxygenase type 2, involved in the production of proinflammatory series prostaglandins (E2 series) [135]. Furthermore, cyanidin-3-glucoside has reduced lung inflammation in rats [136]; in the same way, they have shown effects in reducing TNF- α , IL-6, MCP-1, iNOS, IL-10, serum glucose, total cholesterol, low-density lipoproteins (LDL), and C-reactive protein (CRP) [137]. Regarding their bioavailability, anthocyanins have very low absorption, bioavailability, and systemic distribution. However, the bioavailability of anthocyanin metabolites is 42 times greater than that of the original anthocyanins [138]. The identified anthocyanin metabolites have also been shown to contribute to improving human health [135,139].

Many preclinical and clinical studies have investigated the positive influence of anthocyanins on weight, obesity, and inflammation. In in vitro studies with cell cultures, strawberries have a notable anthocyanin richness and exhibit a high antioxidant capacity. One study found that strawberries, as opposed to bananas and oranges, experienced a significant reduction in oxidative stress-induced neurotoxicity in PC12 cells previously exposed to hydrogen peroxide (H2O2) [140,141].

In a 14-week preclinical study with six-week-old male C57BL/6J mice, a dose of *Murray* extract (*Lycium ruthenicum*) high in various anthocyanins at a 0.8% concentration dissolved in drinking water (orally), the extract increased the diversity of cecal bacterial communities, decreasing the Firmicutes/Bacteroidetes ratio, which has been related to lower weight gain and lower body mass index (BMI). Likewise, the species of the *Akkermansia* genus increased, and those of the Faecalibaculum genus decreased, resulting in a decrease in body mass [142]. Another 12-week study in nine-week-old male Wistar rats used raspberry extract and fructooligosaccharides (FOS) orally at a dose of 0.64% and 3% respectively of the diet, with raspberries being high in anthocyanins,

demonstrated an increase in bacterial species, genera and families in the intestinal microbiota of rats related to an increase in the production of short-chain fatty acids (SCFAs), which have been involved in improvements in energy metabolism and body weight [143].

In animal models, purple dye from potatoes demonstrated a protective capacity against the generation of ROS and restored glutathione levels in mice subjected to a high-fat diet [144]. Also, oral supplementation with anthocyanins from blackberry and blueberry mitigated oxidative stress and inflammation through an increase in the levels of first-line antioxidant enzymes, such as superoxide dismutase (SOD) and glutathione peroxidase (GPx). Likewise, its administration over 12 weeks prevented weight gain in C57BL/6 mice with obesity induced by a high-fat diet [145]. Meanwhile, anthocyanins derived from mulberry and cherry reduced body weight and improved SOD and GPx activities in mice fed with a high-fat diet [141,146].

In clinical research, healthy individuals who incorporated 500 grams of strawberries, rich in anthocyanins, daily for a month experienced a reduction in their risk of developing cardiovascular disease, evidenced by improvements in lipid profiles and antioxidant capacity [147]. In another trial carried out in participants with DM2 who received oral supplements of purified anthocyanins for 24 weeks, improved lipid profiles, antioxidant capacity, and insulin sensitivity were observed due to oral administration [141,148].

Table 6. Clinical effects of anthocyanins on obesity and associated comorbidities, from clinical trials and systematic reviews with meta-analysis.

Compou nd	Type of study	Dose	Targeted populatio n	Observed effect Δ	p	Adverse effects	Referenc e
Anthocya	Randomi zed, placebo- controlled clinical trial with 55 participa nts.	200 g/day of açaí- juçara juice [293.6 mg) orally for 12 weeks.	Men and women who are overweigh t or obese.	Decrease in arterial stiffness (pulse wave speed) and peripheral vascular resistance. No changes in flowmediated dilation (endothelial function).	0.002, 0.005 and >0.05, respec tively.	None.	Arisi et al., 2023 [149]
Anthocya nins	Systemati c review and meta- analysis of 11 RCTs with a total of 833	28.3- 500 mg/day orally for 4-24 weeks.	Women and men who are overweigh t or obese.	Reduction in BMI and body weight. No significant changes in waist circumference.	0.002, 0.04, and >0.05, respec tively.	Not mention ed.	Park <i>et</i> al., 2021 [150]

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patients.

Table 6 shows the clinical effects of anthocyanins on obesity and related comorbidities, according to clinical trials and systematic reviews with meta-analysis of clinical trials found in the scientific literature.

3.7. Probiotics

Probiotics are live microorganisms (such as bacteria and yeast) that provide health benefits when consumed. They are naturally present in some fermented foods, added to food products, and available as dietary supplements [151]. In the market for food, supplements, and medicines, there is a wide range of probiotics that must be categorized at the strain level due to the varying effects they have on human physiology [152]. Most probiotic bacteria belong to the genera *Lactobacillus* (recently, many of them were renamed into other genera according to new phylogenetic classifications) and *Bifidobacterium*, which are gram-positive. However, there are also gram-negative bacteria with probiotic benefits such as *Escherichia coli* Nissle 1917, *Akkermansia muciniphila*, *Christensenella minuta*, among others; many of which are under study since they are considered "second generation probiotics" [153]. Various yeasts are used as probiotics, among them *Saccharomyces boulardii* and *Saccharomyces cerevisiae*. Various probiotics belonging to the genera mentioned above have been shown to positively affect weight control, obesity, and associated comorbidities [154].

In clinical trials, the second-generation probiotic *Akkermansia muciniphila* improves insulin sensitivity and significantly reduces insulinemia, serum total cholesterol, total body weight, and fat mass [155]. Meanwhile, *Lactobacillus gasseri* has some interesting effects on obesity [156] since it is a habitual resident of the human intestine and can modulate the intestinal niche. Its presence confers beneficial effects related to biliary tolerance and the deconjugation of bile acids, improving hepatic and biliary metabolism. It has been shown, at an oral dose of 5×10¹⁰ CFU/day for 12 weeks, to reduce total serum cholesterol in people with hypercholesterolemia; however, its effect is more subtle than the effect of hypocholesterolemic drugs [157]. There is evidence that the administration of this probiotic reduces visceral and subcutaneous fat, decreasing the BMI [158].

Lactobacillus salivarius has antimicrobial effects and improves bile tolerance. Its administration in humans with an oral capsule dose of 10¹⁰ CFU/day for 12 weeks promotes an increase of *Bacteroides*, *Prevotella*, and Porphyromonas, which is decreased by the excessive presence of the bacterial phylum Firmicutes, which characterizes the microbiota in people with obesity [159]. *Bifidobacterium animalis* subspecies *lactis* could have beneficial effects in reducing body adipose tissue, improving lipid profile, increasing insulin sensitivity, as well as some effects in regulating redox balance and modulating satiety markers related to the metabolism of tryptophan, which includes the metabolism of serotonin and melatonin, an important regulator of the circadian sleep-wake rhythm [158]. There is clinical evidence that with the oral administration of this probiotic in a range of 10¹⁰ CFU/day, a reduction in abdominal, visceral, and subcutaneous fat is generated, and a reduction in anthropometric indicators of waist circumference is also evident in the BMI and taper index [160].

A clinical study was carried out with 45 patients with obesity, divided into three groups: single diet (low-carbohydrate and energy-reduced diet), prebiotics (30 g /day), and probiotics (a tablet containing *Bifidobacterium longum*, *Lactobacillus helveticus*, *Lactococcus lactis* and *Streptococcus thermophilus per* day). Age, sex, and BMI were used to categorize the three groups. All three groups significantly decreased weight, BMI, and waist circumference with p < 0.05. Only the prebiotic and probiotic groups showed a significant decrease in fat mass (p = 0.001) and a significant increase in muscle strength with p = 0.008 and 0.004. A significant reduction in fasting blood glucose was also demonstrated (p = 0.02).

Table 7. Clinical effects of probiotics on obesity and associated comorbidities, from clinical trials and systematic reviews with meta-analysis.

Compou nd	Type of study	Dose	Targeted populati on	Observed effect Δ	p	Adverse effects	Referenc e
Probiotics	Clinical trial, randomize d, controlled, and matched by age, sex, and BMI with 45 participant s.	One tablet/day orally with Bifidobacteri um longum, Lactobacillus helveticus, Lactococcus lactis and Streptococcus thermophilus, for 4 weeks.	Women and men with obesity.	Reduction in weight, BMI, and waist circumference. Decrease in fat mass. Increased muscle strength. Decrease in fasting blood glucose.	<0.0 5, 0.00 1, 0.00 4 and 0.02.	None	Ben Othman et al., 2023 [161]
Probiotics	Double-blind, randomize d, placebo-controlled clinical trial with 81 participant s.	One capsule/day orally with 13 million CFU/g of Bacillus subtilis (LMG P- 32899) and Bacillus coagulans (LMG P- 32921) for 12 weeks.	Overwei ght men and women between 18-45 years of age.	Reduction in body weight. No significant changes in BMI, waist circumference, blood pressure, or biomarkers.	0.02 7 and >0.0 5.	None	Danielss on <i>et al.</i> , 2023 [162]
Probiotics	Randomize d, double- blind, placebo- controlled clinical trial with 152 participant	Oral capsules with Lacticaseibaci llus rhamnosus HA-114 for 12 weeks.	Overwei ght adult men and women.	Significant decrease in plasma insulin, HOMA-IR, LDL cholesterol, and triacylglyceri	<0.0 5 and >0.0 5.	None	Choi et al., 2023 [163]

		18
S.	des. No	
	significant	
	changes in	
	body weight	
	and BMI.	

It was concluded that the prescription of prebiotics and probiotics, together with lifestyle measures, seems interesting for the management of obesity, especially if it is sarcopenic, in addition to the improvement of metabolic parameters and psychiatric disorders related to obesity [161]. Table 7 shows the clinical effects of probiotics on obesity and related comorbidities, according to clinical trials and systematic reviews with meta-analysis of clinical trials found in the scientific literature.

3.8. Carotenoids

Carotenoids are a class of more than 750 natural pigments synthesized by plants, algae, and photosynthetic bacteria [164]. They are terpenoid compounds with 40 carbon atoms derived biosynthetically from two geranyl-geranyl-pyrophosphate units, so they are considered tetraterpenes [165]. These richly colored molecules are the source of many plants' yellow, orange, and red colors. Fruits and vegetables provide most of the 40 to 50 carotenoids found in the human diet; the most common are α -Carotene, β -carotene, β -carotene, and β -cryptoxanthin, and lycopene. The human body can transform α -Carotene, β -carotene, and β -cryptoxanthin, which are provitamin A carotenoids, to retinol. Lutein, zeaxanthin, and lycopene are non-provitamin A carotenoids because they cannot be converted to retinol [164].

One of the most widely studied carotenoids is astaxanthin (ASTX), widely known for its potent antioxidant capacity. However, it has other biological properties, such as anti-inflammatory, antiaging, and anti-cancer. Recently, ASTX has been reported to inhibit the onset and development of fibrosis by regulating molecular signaling pathways, such as transforming growth factor β / and small mother against decapentaplegic (TGF-β1/Smad), SIRT1, and nuclear factor. Kappa-B (NF-κB), microRNA, nuclear factor E2-related factor 2/antioxidant response element (Nrf 2/ARE), and ROS pathways [166], which are molecular signaling pathways that are altered in obesity, contribute to the development of comorbidities. Another study demonstrated that ASTX improves the activation of the Nrf-2/HO-1 signaling pathway, which promotes the process of mitophagy, inhibits oxidative stress and ferroptosis of cartilage endplate chondrocytes, and, finally, improves the degradation of the extracellular matrix, the calcification of cartilage endplate and endplate chondrocytes by activating apoptosis. Furthermore, ASTX inhibits the NF-kB activity induced by oxidative stimulation and can improve the inflammatory response [167]. ASTX could also protect the endplate of vertebral cartilage against oxidative stress and degeneration by activating the Nrf-2/HO-1 pathway [167]. However, dysfunction of this molecular pathway is related to the development of many pathologies, such as obesity.

Lycopene, another carotenoid belonging to the xanthophylls, has anti-obesity and anti-diabetic activities in different organs and/or tissues, including adipose tissue, liver, kidneys, pancreas, brain, ovaries, intestine, and eyes. The underlying mechanism may be attributed to its antioxidant and anti-inflammatory properties and its ability to regulate AGE/RAGE, JNK/MAPK, PI3K/Akt, SIRT1/FoxO1/PPARγ signaling pathways, and AchE activity [168]. Epidemiological research supports that lycopene consumption may reduce the risk of obesity and T2DM. The cis isomers of lycopene are more bioavailable and better absorbed than trans-lycopene and are mainly distributed in the liver and adipose tissue. Lycopene has a good safety margin and can be obtained by plant extraction, chemical synthesis, and microbial fermentation [168].

Table 8. Clinical effects of carotenoids on obesity and associated comorbidities, from clinical trials and systematic reviews with meta-analysis.

Compoun d	Type of study	Dose	Targeted populati on	Observed effect Δ	p	Adverse effects	Referenc e
Carotenoid s	Systemati c review and meta- analysis of 7 RCTs and 8 observati onal studies, with 28,944 participan ts.	1.2-60 mg/day orally for 20 days to 16 weeks.	Men and women who are overweig ht or obese.	Insufficiency of serum carotenoids is a risk factor for overweight and obesity (OR = 1.73). Reduction in body weight, BMI, and waist circumference.	<0.001 , <0.001 , <0.001 and <0.001 , respe ctivel y.	Not mentioned	Yao et al., 2021 [169]
Carotenoid s.	Systemati c review and meta- analysis of 12 RCTs with a total of 380 participan ts.	Oral supplem entation with astaxant hin in variable doses and periods.	Men and women with overweig ht, obesity, and/or DM2.	Significant reduction in blood malondialdehy de concentration and IL-6. Improvement of superoxide dismutase activity and reduction of serum isoprostane concentration.	<0.01, 0.02, <0.05 and >0.05.	Not mentioned	Ma et al., 2022 [170]
Carotenoid s.	Systemati c review and meta- analysis of 7 RCTs with a total of 321 participan	Oral supplem entation of 0.16- 20 mg/day for 8 weeks to 12	Men and women with overweig ht or obesity and metabolic syndrom	Significant reduction in LDL. No significant changes in BMI, fasting blood glucose, systolic and diastolic blood	<0.000 01 and ≥0.05.	No serious adverse effects were reported.	Leung et al., 2022 [171]

ts. months.

e. pressure, total cholesterol, HDL, or triacylglycerol

s.

In a systematic review with a meta-analysis of randomized, placebo-controlled clinical trials from 2021 [169], the association between carotenoid intake and the presence of overweight and obesity was analyzed. Seven randomized controlled trials and eight observational studies containing 28,944 subjects and data on multiple carotenoid subgroups, including lycopene, astaxanthin, cryptoxanthin, α -carotene, and β -carotene, were included. In placebo-controlled clinical trials, the intervention lasted for a minimum of 20 days and a maximum of 16 weeks, with a dosage range of 1.2 to 60 mg/day. The meta-analysis found that serum carotenoid insufficiency was a risk factor for overweight and obesity (OR=1.73, 95% CI [1.57, 1.91], p<0.001). Furthermore, carotenoid supplementation was significantly associated with reduced body weight (SMD = -2.34 kg, 95% CI [-3.8, -0.87] kg, p<0.001), decreased body mass index (BMI, SMD = -0.95 kg cm2, 95% CI [-1.88, -0.01] kg cm2, p<0.001) and losses in waist circumference (WC, SMD = -1.84 cm, 95% CI [-3.14, -0.54] cm, p<0.001). Therefore, it was concluded that carotenoids show promising effects in overweight or obese subjects. However, additional data from clinical trials are needed [169]. Table 8 shows the clinical effects of various carotenoids (carotenes and xanthophylls) on obesity and related comorbidities, according to clinical trials and systematic reviews with meta-analysis of clinical trials found in the scientific literature.

3.9. Curcumin

Curcumin, derived from the rhizome of Curcuma longa, is a natural pigment and the primary active component. It has been reported to have antioxidant, antiviral, antineoplastic, anti-inflammatory, and anti-obesity effects [172]. The mechanisms by which it can contribute to managing obesity are multiple since it can inhibit adipogenesis in both adipocyte differentiation and clonal expansion. It inhibits adipocyte differentiation by regulating the SREBP signaling pathway, increasing Rb phosphorylation, and cyclin D1 expression [173]. Curcumin activates the phosphorylation of AMPK and consequently inhibits the expression of PPAR γ , decreasing adipogenesis. Furthermore, curcumin inactivates fatty acid synthase by irreversible inhibition [174], which favors the prevention of hyperplasia of adipose tissue cells.

In humans, reports of the effect of curcumin on anthropometric parameters have been carried out in trials with patients with overweight or obesity combined with other pathologies. Although many studies have reported that curcumin reduces waist circumference [175,176] in patients with grade III obesity, no significant decrease in this variable or visceral fat was found [177]. In some clinical trials and meta-analyses, no significant reductions in BMI and weight have been found [175]; however, when performing the statistical analysis by groups, a decrease was found [176]. The results have been variable, and studies specifically designed to evaluate these parameters are required.

Although low doses of curcumin have not been related to adverse effects, some effects on anthropometric variables were obtained with high doses of this molecule [175,176]. High doses of curcumin, greater than 500 mg/day for 30 days, have been linked to nausea, dizziness, and liver damage [178]. Likewise, it has been reported that in populations with neoplastic risk, it can favor the appearance and development of malignant tumors through an oxidant effect more significant than the antioxidant [178,179]. In vitro studies report that it can inhibit the motility of healthy sperm, but clinical studies are required to determine its effect in healthy men after oral administration [178,180]. Caution is advised due to potential drug interactions, including with glibenclamide, as it inhibits the activity of CYP3A4 and may increase the risk of hypoglycemia [178]. Table 9 shows the clinical effects

of curcumin on obesity and related comorbidities, according to clinical trials and systematic reviews with meta-analysis of clinical trials found in the scientific literature.

Table 9. Clinical effects of curcumin on obesity and associated comorbidities, from clinical trials and systematic reviews with meta-analysis.

Compou nd	Type of study	Dose	Targeted populatio n	Observed effect Δ	p	Adverse effects	Referenc e
Curcumi	Systematic review and meta- analysis of 8 clinical trials with 520 individuals.	70 to 3000 mg/day for 8 to 12 weeks	Overweig ht men and women with nonalcoho lic fatty liver disease (NAFLD)	No change in body weight Decrease in BMI - 0.34 kg/m² and waist circumferenc e - 2.12 cm	<0.05 <0.01	Not mention ed.	Baziar, et al., 2019 [175]
Curcumi	Review and meta-analysis of 14 SRMAs with 39 RCTs with 8111 individuals.	70 to 3000 mg/day orally for 8 to 12 weeks	Men and women with overweigh t or obesity, NAFLD, T2DM, PCOS, or MetS	Decreased body weight -0.59kg BMI -0.24 kg/m²/ Waist circumferenc e -1.32cm	Data not show n	Not mention ed.	Unhapip atpong et al., 2023 [176]

3.10. Silymarin/silybin

Silymarin is the extract of Silybum marianum, commonly known as milk thistle, and comprises seven flavonolignans (including silibinin, isosilibinin, silicristin, isosylcristine and silidianin) and a flavonoid (taxifolin) [181]. Among these substances, silybin is the predominant one and has the most important biological effect. It constitutes approximately 70% of the total silymarin composition from two diastereoisomeric compounds: silybin A and silybin B [181]. Silymarin is a molecule with low bioavailability administered orally and poor solubility in water. This is due to its inefficient absorption in the intestine and a high metabolism of the first hepatic step after its absorption; both mechanisms reduce blood concentration and, consequently, arrive at the target organ [182]. However, this limitation has been effectively overcome with the introduction of complexes with phosphatidylcholine that have better absorption and new glycoconjugates of silibinin (gluco, manno, galacto, and lactoconjugates), which have high water solubility and strong antioxidant power [183]. No deaths or life-threatening adverse events have been reported in the medical and scientific literature [184].

Silymarin has been shown to inhibit the activity of the transcription factor NF-kB, reducing the expression of the genes encoding IL-1, IL-6, TNF-alpha, INF-gamma, and GMC-SF, which reduces

inflammation [185]. Likewise, it inhibits the activation of MAPK induced by TNF-alpha, as well as the c-Jun N-terminal kinase, generating changes in the ratio of Bax/Bcl-2 proteins and inducing the release of cytochrome c from the mitochondria, activating initiator caspases 3 and 9, to activate the effector caspase cascade and modulating IGF (insulin growth factor) signaling pathways, all of which regulate the correct apoptotic process [186]. Regarding its endocrine effects, silymarin partially activates estrogen receptors while presenting antiandrogenic activity in prostate cancer cells [187].

Additionally, silymarin can regulate various drug transporters by inhibiting membrane efflux proteins such as P-glycoprotein and the trypanosomal purine transporter TbAT1 [181]. Silymarin has also demonstrated antifibrotic effects, inhibiting the conversion of hepatic stellate cells into myofibroblasts, downregulating the expression of genes involved in fibrosis, such as procollagen III, alpha-SMA, and TGF-beta [181]. Silymarin also has a high antioxidant capacity, having the ability to neutralize free radicals, maintain the correct mitochondrial function, and activate the synthesis of protective molecules against oxidation and with reparative capacity, such as several HSPs (heat shock proteins), thioredoxin and sirtuins [188].

Silymarin has various beneficial metabolic effects, including serving as a PPAR-gamma agonist, improving the sensitivity of the insulin receptor type 1 substrate, increasing the activity of PI3K, Nrf2, and Akt, increasing the expression of GLUT4 on the cell surface and inhibiting HMG-CoA reductase [189,190]. Silymarin has demonstrated choleretic activity by generating upregulation of the bile salt export pump [191,192]. Table 10 shows the clinical effects of silymarin/silybin on obesity and related comorbidities, according to clinical trials and systematic reviews with meta-analysis of clinical trials found in the scientific literature.

Table 10. Clinical effects of silymarin/silybin on obesity and associated comorbidities, from clinical trials and systematic reviews with meta-analysis.

Compou nd	Type of study	Dose	Targeted populatio n	Observed effect Δ	p	Adverse effects	Referenc e
Silymarin (Silybum marianum).	Systemati c review and meta- analysis of 8 RCTs.	100-500 mg/day orally for 4-12 weeks.	Men and women with NAFLD (many of them overweigh t or obese).	Statistically significant reduction in BMI.	<0.05.	None.	Kalopitas et al., 2021 [193]
Silymarin (Silybum marianum).	Clinical, randomiz ed, and controlle d trial with 36 participa nts.	Two tablets/da y orally of silymarin + vitamin E for 12 weeks.	Men and women with NAFLD (many of them overweigh t or obese).	Decrease in body weight and anthropometri c parameters	<0.05 and >0.05.	None.	Aller <i>et</i> al., 2015 [194]

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Silymarin (Silybum marianum).	Clinical, randomiz ed, and controlle d trial with 78 participa nts.	Two tablets/da y of oral suppleme nt with Curcuma longa, silymarin , guggul, chlorogen ic acid, and inulin for 16 weeks.	Men and women with metabolic syndrome (many of them overweigh t or obese).	Significant body weight, BMI, waist circumference, fasting glucose and total cholesterol reduction.	<0.0001, 0.001, 0.0004, 0.014, 0.03 and >0.05, respecti vely.	None.	Patti <i>et al.</i> , 2015 [195]

3.11. Hydroxycitric acid

Hydroxycitric acid is an alpha-hydroxytribasic acid [1,2-dihydroxypropane-1,2,3-tricarboxylic acid) with two asymmetric centers, resulting in the formation of two pairs of diastereoisomers or four different isomers: acid (-) hydroxycitric (I), (+) hydroxycitric acid (II), (-) alo-hydroxycitric acid (III) and (+) alo-hydroxycitric acid (IV). The (-) hydroxycitric acid isomer (HCA) is found in the bark of the fruit of *Garcinia cambogia* (fam. *Clusiaceae*) [196,197].

Hydroxycitric acid is a competitive inhibitor of the enzyme ATP-citrate lyase, an enzyme that catalyzes the conversion of citrate and coenzyme A into oxaloacetate and acetyl coenzyme A in the cytosol. Acetyl CoA is necessary in the synthesis of fatty acids, cholesterol, and triglycerides, as well as in the synthesis of acetylcholine in the central nervous system. Inhibition of ATP-citrate lyase decreases acetyl CoA deposits, resulting in a reduction in malonyl CoA concentration, suppressing body fat accumulation through the activation of carnitine palmitoyl transferase I, an enzyme involved in the oxidation of fatty acids [197]. In some studies, it was observed that *G. cambogia* showed positive effects on the process of weight loss, lipogenesis, reduction of appetite, percentage of body fat, triglycerides, cholesterol, and glucose levels. In contrast, in others, it had no effect [198].

In a prospective, non-randomized controlled intervention trial, 214 overweight or obese subjects were treated with G. cambogia and glucomannan [500 mg twice daily, each) for six months, evaluating weight, fat mass, visceral fat, rate basal metabolic rate, and blood profiles of lipids and glucose, comparing them with basal values. Some patients were carriers of the PLIN4 polymorphisms -11482G > A-, fat mass and associated obesity (FTO) -rs9939609 A/T- and β -adrenergic receptor 3 (ADRB3) -Trp64Arg. The treatment produced weight loss, reducing fat mass, visceral fat, lipid profiles, and blood glucose while increasing basal metabolic rate. The results were independent of sex, age, or having hypertension, type 2 diabetes mellitus, or dyslipidemia and were attenuated in carriers of the PLIN4, FTO, and Trp64Arg polymorphisms [199]. However, multiple adverse effects have been reported [197,200]. A literature search study found 22 cases of liver injury caused by G. cambogia alone or in combination with green tea or Ashwagandha [201]. Patients taking G. cambogia were between 17 and 54 years old, and liver injury emerged between 13 and 223 days after onset. One patient died, one required a liver transplant, and 91% were hospitalized. The liver injury was hepatocellular with jaundice [201]. Table 11 shows the clinical effects of hydroxycitric acid (G. cambogia) on obesity and related comorbidities, according to clinical trials and systematic reviews with meta-analysis of clinical trials found in the scientific literature.

Table 10. Clinical effects of silymarin/silybin on obesity and associated comorbidities, from clinical trials and systematic reviews with meta-analysis.

Compound	Type of study	Dose	Targeted populatio	Observed effect Δ	p	Adverse effects	Referen ce
Hydroxycitr ic acid (G. cambogia).	Randomiz ed, placebo- controlled, double- blind, parallel- group clinical trial with 91 participant s.	Two tablets a day 30 minutes before meals for oral administr ation for 14 weeks.	Caucasian men and women who are overweigh t or obese.	Weight loss. Reduction of fat mass, waist circumference, and hip circumference.	0.00 2 and <0.0 5.	There were no serious events reported.	Chong et al., 2014 [202]
Hydroxycitr ic acid (G. cambogia).	Clinical, randomize d, and controlled trial with 86 participant s.	Oral tablets at 2 g/day for 10 weeks.	Overweig ht men and women.	No significant effect on adipocytokine s, non-HDL-c cholesterol, triacylglycerid es, antioxidants, body weight, HDL-c, or total cholesterol.	>0.0 5.	Not mentione d.	Kim et al., 2011 [203]
Hydroxycitr ic acid (G. cambogia).	Randomiz ed, double- blind, placebo- controlled clinical trial with 105 participant s.	Polyherb al oral suppleme nt in tablets with <i>G. cambogia</i> twice daily, for 12 weeks.	Men and women who are overweigh t or obese.	Significant change in the Body Composition Improvement Index. No significant changes in weight, BMI, and waist-hip ratio. Decrease in body fat.	0.01 2, >0.0 5 and 0.01 1.	Not mentione d.	Opala et al., 2006 [204]

Hydroxycitr Case series Oral Men and 22 cases of <0.0 Liver Vuppala ic acid in 1,418 suppleme women liver injury 18 injury. nchi et (G. patients ntation of between due to G. Hepatoce al., 2022 cambogia). enrolled in varying 17 and 54 cambogia alone the Drug-doses of Induced G. Liver cambogia Injury alone or Network in (DILIN) combinati from 2004 to 2018. green tea or Ashwaga ndha. Oral Men and 22 cases of <0.0 Liver injury 18 injury. nchi et (m = 5) or in combination with green tea (n = 16) or Ashwagandha (n = 1), arising between 13 and 223 days after onset. Significant increase in aminotransfer							
ases.	ic acid (<i>G</i> .	in 1,418 patients enrolled in the Drug- Induced Liver Injury Network (DILIN) from 2004	suppleme ntation of varying doses of G. cambogia alone or in combinati on with green tea or Ashwaga	women between 17 and 54	liver injury due to <i>G</i> . cambogia alone (n = 5) or in combination with green tea (n = 16) or Ashwagandha (n = 1), arising between 13 and 223 days after onset. Significant increase in	injury. Hepatoce Ilular injury with jaundice. Hospitali zation. Requirem ent of liver transplan	nchi <i>et</i> <i>al.,</i> 2022

3.12. α -lipoic acid

 α -Lipoic acid is a lipophilic thiol that, due to its chemical structure, is most easily identified by the name 6,8-dithio octanoic acid, although it is also often called 6,8-thioctic acid. The presence of two sulfhydryl radicals (-SH) in the structure of an eight-carbon fatty acid corresponds to its reduced form. Through an oxidation process, lipoic acid gives up two electrons and two protons [2 e- + 2 H+) to form an intramolecular disulfide bridge, which makes it an effective antioxidant [205]. It is an organosulfur compound produced in plants, animals, and humans. Naturally, α -lipoic acid is found in the mitochondria, which is used as a cofactor for pyruvate dehydrogenase (PDH) and α -ketoglutarate dehydrogenase complexes. Despite its diverse potential, the therapeutic efficacy of α -lipoic acid is relatively low due to its pharmacokinetic profile. Data suggest that α -lipoic acid has a short half-life and bioavailability (about 30%) caused by its hepatic degradation, reduced solubility, and instability in the stomach [206].

 α -Lipoic acid treatment stimulates PI3K activity and insulin receptor substrate 1 (IRS-1) phosphorylation in 3T3-L1 adipocytes. Insulin receptor substrate phosphorylation involves the activation of further intracellular mediators and, subsequently, GLUT4 translocation; therefore, α -lipoic acid can be considered an insulin mimetic agent since insulin receptor binding and IRS-1 phosphorylation could subsequently lead to GLUT4 translocation, increasing glucose uptake [207]. α -Lipoic acid also decreases hypothalamic AMPK activity and causes rodent weight loss by reducing food intake and increasing energy expenditure [208]. Side effects of α -lipoic acid may include headache, tingling sensation, rash, or muscle cramps. There have been some reports in Japan of a rare case known as autoimmune insulin syndrome in people using α -lipoic acid [209]. This condition causes hypoglycemia and causes antibodies to attack the insulin produced by the body without having previous insulin treatment. The safety of α -lipoic acid in pregnant or lactating women, children, or people with liver or kidney disease is unknown. α -Lipoic acid can also have other adverse effects, such as hives, abdominal pain, nausea, diarrhea, and vomiting, as well as foul-smelling urine. However, these effects are dependent on the dose of lipoic acid and the administration [210].

In a preclinical study using a murine model, α -lipoic acid caused a notable reduction in the content of arachidonic acid, mainly in the phospholipid fraction, with a simultaneous decrease in the synthesis of proinflammatory mediators, that is, prostaglandin E2, leukotrienes B4 and C4 by

decreasing the expression of COX-2 and LOX-5. α -Lipoic acid also increased the level of antioxidants SOD2 and GSH and reduced the level of lipid peroxidation products, which improved the deterioration of the oxidative system in the tissue of the left ventricle of the heart. Therefore, α -lipoic acid has an important role in inflammation and the development of oxidative stress, decreasing the risk of obesity and cardiac dysfunction induced by a high-fat diet [211].

Clinical trials with α -Lipoic acid treatment have shown a statistically significant short-term reduction in weight, BMI, body fat, and waist circumference, compared to placebo [212-215]. Table 12 shows the clinical effects of α -lipoic acid on obesity and related comorbidities, according to clinical trials and systematic reviews with meta-analysis of clinical trials found in the scientific literature.

Table 12. Clinical effects of α -lipoic acid on obesity and associated comorbidities, from clinical trials and systematic reviews with meta-analysis.

Compou nd	Type of study	Dose	Targeted populati on	Observed effect Δ	p	Adverse effects	Referenc e
α-Lipoic acid	Clinical, controlled, and randomized trial with 92 participants	Oral admini stration for 8 weeks	Men and women with NAFLD and obesity	Reduction of alanine aminotransfera se	0.01	Not mentione d	Tutunchi et al., 2023 [213]
α-Lipoic acid	Clinical, controlled, and randomized trial with 100 participants.	1,200 mg/day orally for 8 weeks.	Men and women with obesity.	Weight reduction, BMI, body fat, and waist circumference.	<0.0 5.	Not mentione d.	Moham madshah i <i>et al.,</i> 2022 [214]
α-Lipoic acid	Clinical, controlled, and randomized trial with 88 participants	600 mg/day orally for 16 weeks	Overwei ght women and men.	Reduction in weight, waist circumference, and C-reactive protein. Maintenance of lost weight	<0.0 5.	Not mentione d.	Nasiri <i>et</i> al., 2021 [215]

4. Conclusions

Globally, obesity is one of the most widespread pathologies. In the same way, it demands a comprehensive and interdisciplinary strategy encompassing nutrition, physical education, anthropology, sociology, psychology, epidemiology, and medical treatments that can involve surgery and/or medication. The medications currently authorized for the treatment of obesity mainly generate a wide range of adverse effects, ranging from headaches to cardiac disorders that can cause the death of the individual. An alternative of interest is phytopharmaceuticals and molecules of natural origin. In this review, we analyze some natural molecules that have a promising future for

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obesity therapy and are primarily found in the usual human diet, which can contain hundreds to thousands of compounds with biological activity, depending on the typology and diversity of the diet. Due to their low concentrations in the conventional diet, these molecules often need to achieve therapeutic objectives or offer a minimal effect. Standardized presentations of these molecules may be an alternative for their administration. This review finds that some molecules have few clinical research publications available.

All the molecules included in this review have been evaluated in humans, demonstrating their clinical efficacy in vitro and in vivo. Some have results equal to or superior to those of currently authorized drugs, with the benefit of greater safety and lower incidence of adverse effects. However, some cause favorable modifications that reach statistical significance but lack clinical significance, making it essential to evaluate the synergies offered by different compounds to develop more complete and promising pharmaceutical formulations. The safety of some molecules, such as ellagic acid and resveratrol, places them in scientific interest since they have a wide therapeutic range and have not shown adverse effects. On the other hand, some molecules, such as hydroxycitric acid, must be carefully evaluated as they carry the risk of serious adverse reactions. Therefore, some natural molecules deserve to be considered for further analysis as pharmacological treatment options, and others may be effective as a complementary therapy for the treatment of obesity, but more phase II and III studies are required to expand the evaluation of their safety and efficacy.

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References

- 1. Tutor, A.W.; Lavie, C.J.; Kachur, S.; Milani, R.V.; Ventura, H.O. Updates on obesity and the obesity paradox in cardiovascular diseases. *Prog Cardiovasc Dis* **2023**, *78*, 2; DOI: 10.1016/j.pcad.2022.11.013
- 2. Boutari, C.; Mantzoros, C.S. A 2022 update on the epidemiology of obesity and a call to action: as its twin COVID-19 pandemic appears to be receding, the obesity and dysmetabolism pandemic continues to rage on. *Metabolism* 2022, 133, 155217; DOI: 10.1016/j.metabol.2022.155217
- 3. McPherson, R. Obesity and ischemic heart disease. *Circ Res* **2015**, *116*, 570; DOI: 10.1161/CIRCRESAHA.115.305826
- 4. Shiozawa, M.; Kaneko, H.; Itoh, H.; Morita, K.; Okada, A.; Matsuoka, S.; et al. Association of body mass index with ischemic and hemorrhagic stroke. *Nutrients* **2021**, *13*, 2343; DOI: 10.3390/nu13072343
- 5. Smith, K.B.; Smith, M.S. Obesity statistics. Prim Care 2016, 43, 121; DOI: 10.1016/j.pop.2015.10.
- 6. Loos, R.J.F.; Yeo, G.S.H. The genetics of obesity: from discovery to biology. *Nat Rev Genet* **2022**, 23, 120; DOI: 10.1038/s41576-021-00414-z
- 7. Blüher, M. Obesity: global epidemiology and pathogenesis. *Nat Rev Endocrinol* **2019**, *15*, 288; DOI: 10.1038/s41574-019-0176-8
- 8. Kaufer-Horwitz, M.; Pérez-Hernández, J.F.; Kaufer-Horwitz, M.; Pérez-Hernández, J.F. La obesidad: aspectos fisiopatológicos y clínicos. *Inter disciplina* **2022**, 10, 147; DOI:10.22201/ceiich.24485705e.2022.26.80973
- 9. Wen, X.; Zhang, B.; Wu, B.; Xiao, H.; Li, Z.; Li, R.; et al. Signaling pathways in obesity: mechanisms and therapeutic interventions. *Sig Transduct Target Ther***2022**, *7*, 1; DOI: 10.1038/s41392-022-01149-x
- Grarup, N.; Moltke, I.; Andersen, M.K.; Dalby, M.; Vitting-Seerup, K.; Kern, T.; et al. Loss-of-function variants in ADCY3 increase risk of obesity and type 2 diabetes. *Nat Genet* 2018, 50, 172; DOI: 10.1038/s41588-017-0022-7
- 11. Yin, D.; Li, Y.; Liao, X.; Tian, D.; Xu, Y.; Zhou, C.; et al. FTO: a critical role in obesity and obesity-related diseases. *Br J Nutr* **2023**, *1*; DOI: 10.1017/S0007114523000764
- 12. Wu, O.; Yuan, C.; Leng, J.; Zhang, X.; Liu, W.; Yang, F.; et al. Colorable role of interleukin (IL)-6 in obesity hypertension: A hint from a Chinese adult case-control study. *Cytokine* **2023**, *168*:156226; DOI: 10.1016/j.cyto.2023.156226

- 13. Bombarda-Rocha, V.; Silva, D.; Badr-Eddine, A.; Nogueira, P.; Gonçalves, J.; Fresco, P. Challenges in pharmacological intervention in perilipins (PLINs) to modulate lipid droplet dynamics in obesity and cancer. *Cancers (Basel)* **2023**, *15*, 4013; DOI: 10.3390/cancers15154013
- 14. Ramakrishnan, N.; Auger, K.; Rahimi, N.; Jialal, I. *Biochemistry, Adiponectin*. StatPearls Publishing; USA 2023 [cited on September 29, 2023]. Available in: http://www.ncbi.nlm.nih.gov/books/NBK537041/
- 15. Ayed, K.; Nabi, L.; Akrout, R.; Mrizak, H.; Gorrab, A.; Bacha, D.; et al. Obesity and cancer: focus on leptin. *Mol Biol Rep* **2023**, *50*, 6177; DOI: 10.1007/s11033-023-08525-y
- 16. Wang, Y.; Wan, R.; Hu, C. Leptin/obR signaling exacerbates obesity-related neutrophilic airway inflammation through inflammatory M1 macrophages. *Mol Med* **2023**, 29, 100; DOI: 10.1186/s10020-023-00702-w
- 17. Lechner, L.; Opitz, R.; Silver, M.J.; Krabusch, P.M.; Prentice, A.M.; Field, M.S. et al. Early-set POMC methylation variability is accompanied by increased risk for obesity and is addressable by MC4R agonist treatment. *Sci Transl Med* **2023**, *15*, 1659; DOI: 10.1126/scitranslmed.adg1659
- 18. Shaji, A.; Jayasri, M.A. A review of the role of liposome-encapsulated phytochemicals targeting PPAR Y and associated pathways to combat obesity. *3 Biotech* **2023**, *13*, 313; DOI: 10.1007/s13205-023-03740-7
- Kim, D.S.; Lee, H.Y.; Kim, H.J.; Lee, G.H.; Lim, Y.J.; Ko, B.M. et al. Combined treatment of mori folium and mori cortex radicis ameliorate obesity in mice via UCP-1 in brown adipocytes. *Nutrients* 2023, 15, 3713; DOI: 10.3390/nu15173713
- 20. Rohde, K.; Keller, M.; la Cour-Poulsen, L.; Blüher, M.; Kovacs, P.; Böttcher, Y. Genetics and epigenetics in obesity. *Metabolism* **2019**, 92, 37; DOI: 10.1016/j.metabol.2018.10.007
- 21. Ouni, M.; Schürmann, A. Epigenetic contribution to obesity. *Mamm Genome* **2020**, *31*, 134; DOI: 10.1007/s00335-020-09835-3
- 22. Pagiatakis, C.; Musolino, E.; Gornati, R.; Bernardini, G.; Papait, R. Epigenetics of aging and disease: a brief overview. *Aging Clin Exp Res.* **2021**, *33*, 737; DOI: 10.1007/s40520-019-01430-0
- 23. Plaza-Diaz, J.; Izquierdo, D.; Torres-Martos, Á.; Baig, A.T.; Aguilera, C.M.; Ruiz-Ojeda, F.J. Impact of physical activity and exercise on the epigenome in skeletal muscle and effects on systemic metabolism. *Biomedicines* **2022**, *10*, 126; DOI: 10.3390/biomedicines10010126
- 24. Zhang, L.; Lu, Q.; Chang, C. Epigenetics in health and disease. In: Epigenetics in Allergy and Autoimmunity, Chang, C., Lu, Q., Eds. Springer: Singapore, Singapore, 2020; pp. 3-55. DOI: 10.1007/978-981-15-3449-2_1
- 25. Wu, F.Y.; Yin, R.X. Recent progress in epigenetics of obesity. *Diabetol Metab Syndr* **2022**, *14*, 171; DOI: 10.1186/s13098-022-00947-1
- 26. Şanlı, E.; Kabaran, S. Maternal obesity, maternal overnutrition and fetal programming: effects of epigenetic mechanisms on the development of metabolic disorders. *Curr Genomics* **2019**, 20, 419; DOI: 10.2174/1389202920666191030092225
- 27. Ling, C.; Rönn, T. Epigenetics in human obesity and type 2 diabetes. *Cell Metab* **2019**, 29, 1028; DOI: 10.1016/j.cmet.2019.03.009
- 28. Grijalva-Avila, J.; Villanueva-Fierro, I.; Lares-Asseff, I.; Chairez-Hernández, I.; Rivera-Sanchez, G.; Martínez-Estrada, S. et al. Milk intake and IGF-1 rs6214 polymorphism as protective factors to obesity. *Int J Food Sci Nutr* **2020**, *71*, 388; DOI: 10.1080/09637486.2019.1666805
- 29. Chiu, Y.; Fadadu, R.P.; Gaskins, A.J.; Rifas-Shiman, S.L.; Laue, H.E.; Moley, K.H. et al. Dietary fat intake during early pregnancy is associated with cord blood DNA methylation at IGF2 and H19 genes in newborns. *Environ Mol Mutagen* **2021**, *62*, 388; DOI: 10.1002/em.22452
- 30. Kim, J.Y.; Mondaca-Ruff, D.; Singh, S.; Wang, Y. SIRT1 and autophagy: Implications in endocrine disorders. *Front Endocrinol (Lausanne)* **2022**, *13*, 930919; DOI: 10.3389/fendo.2022.930919
- 31. Hernández-Bedolla, M.; Barajas, A.; Cortés, F.; Alonso, D.; Hernández, A.; Ostoa, Z. et al. Papel pleiotrópico y homeostático de las sirtuinas en la función biológica humana. *Ciencia Huasteca Boletín Científico de la Escuela Superior de Huejutla* **2020**, *8*, 6; DOI: https://doi.org/10.29057/esh.v8i16.5721
- 32. Lahsen, M. R.; Kuzmanic, V. A. Cirugía metabólica 10 años después: una mirada desde la diabetología. *Rev Med Clin Condes* **2016**, 27, 188; DOI: 10.1016/j.rmclc.2016.04.008
- 33. Nicoletti, C.F.; Cortes-Oliveira, C.; Pinhel, M.A.S.; Nonino, C.B. Bariatric surgery and precision nutrition. *Nutrients* **2017**, *9*, 974; DOI: 10.3390/nu9090974
- 34. Hunter, D.J.; James, L.S.; Hussey, B.; Ferguson, R.A.; Lindley, M.R.; Mastana, S.S. Impacts of eccentric resistance exercise on DNA methylation of candidate genes for inflammatory cytokines in skeletal muscle and leukocytes of healthy males. *Genes (Basel)* **2023**, *14*, 478; DOI: 10.3390/genes14020478
- 35. Milton, K.; Gomersall, S.R.; Schipperijn, J. Let's get moving: The Global Status Report on Physical Activity 2022 calls for urgent action. *J Sport Health Sci* **2023**, *12*, 5; DOI: 10.1016/j.jshs.2022.12.006
- 36. Sung, H.; Siegel, R.L.; Torre, L.A.; Pearson-Stuttard, J.; Islami, F.; Fedewa, S.A. et al. Global patterns in excess body weight and the associated cancer burden. *CA Cancer J Clin* **2019**, *69*, 88; DOI: 10.3322/caac.21499
- 37. González-Jiménez, E. Obesidad: análisis etiopatogénico y fisiopatológico. *Endocrinol Nutr* **2013**, *60*, 17; DOI: 10.1016/j.endonu.2012.03.006

- 38. Santos, A.L.; Sinha, S. Obesity and aging: Molecular mechanisms and therapeutic approaches. *Ageing Res Rev* **2021**, *67*, 101268; DOI: 10.1016/j.arr.2021.101268
- 39. Hafidi, M.E.; Buelna-Chontal, M.; Sánchez-Muñoz, F.; Carbó, R. Adipogenesis: A necessary but harmful strategy. *Int J Mol Sci* **2019**, *20*, 3657; DOI: 10.3390/ijms20153657
- 40. Zolla, L. On the need to distinguish between insulin-normal and insulin-resistant patients in testosterone therapy. *Int J Mol Sci* **2022**, 23, 12730; DOI: 10.3390/ijms232112730. PMID: 36361519; PMCID: PMC9657366.
- 41. Malfacini, D.; Pfeifer, A. GPCR in adipose tissue function-focus on lipolysis. *Biomedicines* **2023**, *11*, 588; DOI: 10.3390/biomedicines11020588.
- 42. Lytle, K. A.; Bush, N. C.; Triay, J. M.; Kellogg, T. A.; Kendrick, M. L.; Swain, J. M.; Gathaiya, N. W.; Hames, K. C.; Jensen, M. D. Adipocyte proteins and storage of endogenous fatty acids in visceral and subcutaneous adipose tissue in severe obesity. *Obesity (Silver Spring)* **2021**, *29*, 1014; DOI: 10.1002/oby.23149
- 43. Horwitz, A.; Birk, R. Adipose tissue hyperplasia and hypertrophy in common and syndromic obesity— The case of BBS obesity. *Nutrients* **2023**, *15*, 3445; DOI: 10.3390/nu15153445
- 44. Gupta, A.; Efthymiou, V.; Kodani, S.D.; Shamsi, F.; Patti, M.E.; Tseng, Y.H. et al. Mapping the transcriptional landscape of human white and brown adipogenesis using single-nuclei RNA-seq. *Molecular Metabolism* **2023**, 74, 101746; DOI: 10.1016/j.molmet.2023.101746
- 45. Lee, E.O.; Joo, H.K.; Lee, Y.R.; Kim, S.; Lee, K.H.; Lee, S.D. et al. APE1/Ref-1 inhibits adipogenic transcription factors during adipocyte differentiation in 3T3-L1 cells. *Int. J. Mol. Sci* **2023**, 24, 3251; DOI: 10.3390/ijms24043251
- 46. Guo, L.; Li, X.; Tang, Q.Q. Transcriptional regulation of adipocyte differentiation: a central role for CCAAT/Enhancer-binding Protein (C/EBP) β. *J Biol Chem* **2015**, 290, 755; DOI: 10.1074/jbc.R114.619957
- 47. Kahn, C.R.; Wang, G.; Lee, K.Y. Altered adipose tissue and adipocyte function in the pathogenesis of metabolic syndrome. *J Clin Invest* **2019**, 129, 3990; DOI: 10.1172/JCI129187
- 48. Yang, R.; Barouch, L.A. Leptin signaling and obesity. *Circ Res* **2007**, *101*, 545; DOI: 10.1161/CIRCRESAHA.107.156596
- 49. Gjermeni, E.; Kirstein, A.S.; Kolbig, F.; Kirchhof, M.; Bundalian, L.; Katzmann, J.L. et al. Obesity—an update on the basic pathophysiology and review of recent therapeutic advances. *Biomolecules* **2021**, *11*, 1426; DOI: 10.3390/biom11101426
- 50. Gruzdeva, O.; Borodkina, D.; Uchasova, E.; Dyleva, Y.; Barbarash, O. Leptin resistance: underlying mechanisms and diagnosis. *Diabetes Metab Syndr Obes* **2019**, 12, 191; DOI: 10.2147/DMSO.S182406
- 51. Achari, A.E.; Jain, S.K. Adiponectin, a therapeutic target for obesity, diabetes, and endothelial dysfunction. *Int J Mol Sci* **2017**, *18*, 1321; DOI: 10.3390/ijms18061321
- 52. Ruan, H.; Dong, L.Q. Adiponectin signaling and function in insulin target tissues. *J Mol Cell Biol* **2016**, *8*, 101; DOI: 10.1093/jmcb/mjw014
- 53. Puljiz, Z.; Kumric, M.; Vrdoljak, J.; Martinovic, D.; Ticinovic, K.T.; Krnic. M.O. Obesity, gut microbiota, and metabolome: from pathophysiology to nutritional interventions. *Nutrients* **2023**, *15*, 2236; DOI: 10.3390/nu15102236
- 54. Lee, J.Y.; Tsolis, R.M.; Bäumler, A.J. The microbiome and gut homeostasis. *Science* **2022**, 377, eabp9960; DOI: 10.1126/science.abp9960.
- 55. Milani, C.; Duranti, S.; Bottacini, F.; Casey, E.; Turroni, F.; Mahony, J. The first microbial colonizers of the human gut: composition, activities, and health implications of the infant gut microbiota. *Microbiol Mol Biol Rev* **2017**, *81*, e00036-17; DOI: 10.1128/MMBR.00036-17
- 57. Cheng, Z.; Zhang, L.; Yang, L.; Chu, H. The critical role of gut microbiota in obesity. *Front Endocrinol (Lausanne)* **2022**, *13*,1025706; DOI: 10.3389/fendo.2022.1025706
- 58. Fontané, L.; Benaiges, D.; Goday, A.; Llauradó, G.; Pedro-Botet, J. Influencia de la microbiota y de los probióticos en la obesidad. *Clin Investig Arterioscler*. **2018**, *30*, 271; DOI: 10.1016/j.arteri.2018.03.004
- 59. Amabebe, E.; Robert, F.O.; Agbalalah, T.; Orubu, E.S.F. Microbial dysbiosis-induced obesity: role of gut microbiota in homoeostasis of energy metabolism. *Br J Nutr.* **2020**, *123*, 1127; DOI: 10.1017/S0007114520000380
- 60. Cardinelli, C.S.; Sala, P.C.; Alves, C.C.; Torrinhas, R.S.; Waitzberg, D.L. Influence of intestinal microbiota on body weight gain: a narrative review of the literature. *Obes Surg.* **2015**, *25*, 346; DOI: 10.1007/s11695-014-1525-2
- 61. Morales-Franco, B.; Nava-Villalba, M.; Medina-Guerrero, E.O.; Sánchez-Nuño, Y.A.; Davila-Villa, P.; Anaya-Ambriz, E.J. Host-pathogen molecular factors contribute to the pathogenesis of Rhizopus spp. in diabetes mellitus. *Curr Trop Med Rep.* **2021**, *8*, 6; DOI: 10.1007/s40475-020-00222-1
- 62. Liu, B.N.; Liu, X.T.; Liang, Z.H.; Wang, J.H. Gut microbiota in obesity. *World J Gastroenterol.* **2021**, *27*, 3837; DOI: 10.3748/wjg.v27.i25.3837
- 63. Waters, J.L.; Ley, R.E. The human gut bacteria Christensenellaceae are widespread, heritable, and associated with health. *BMC Biol.* **2019**, *17*, 83; DOI: 10.1186/s12915-019-0699-4

- 64. Depommier, C.; Everard, A.; Druart, C.; Plovier, H.; Van-Hul, M.; Vieira-Silva, S. Supplementation with *Akkermansia Muciniphila* in overweight and obese human volunteers: a proof-of-concept exploratory study. *Nat Med.* **2019**, 25, 1096; DOI: 10.1186/s12915-019-0699-4
- 65. Crovesy, L.; Ostrowski, M.; Ferreira, D.M.T.P.; Rosado, E.L.; Soares-Mota M. Effect of Lactobacillus on body weight and body fat in overweight subjects: a systematic review of randomized controlled clinical trials. *Int J Obes (Lond).* **2017**, *41*, 1607; DOI: 10.1038/ijo.2017.161
- 66. Moore, S.C.; Matthews, C.E.; Sampson, J.N.; Stolzenberg-Solomon, R.Z.; Zheng, W.; Cai, Q. Human metabolic correlates of body mass index. *Metabolomics* **2014**, *10*, 259; DOI: 10.1007/s11306-013-0574-1
- 67. Miller, G.D. Appetite regulation: hormones, peptides, and neurotransmitters and their role in obesity. *Am J Lifestyle Med.* **2019**, *13*, 586; DOI: 10.1177/1559827617716376
- 68. Albrecht, U. The circadian clock, metabolism and obesity. *Obesity Reviews* **2017**, *18*, 25; DOI: 10.1111/obr.12502
- 69. Marjani, A.; Khatami, A.; Saadati, H.; Asghari, M.; Razizadeh, M.H.; Abbasi, A. Association of adenovirus 36 infection and obesity; An updated meta-analysis of community-based studies. *Rev Med Virol.* **2022**, 32, e2255; DOI: 10.1002/rmv.2255
- Calloway EE, Parks CA, Bowen DJ, Yaroch AL. Environmental, social, and economic factors related to the intersection of food security, dietary quality, and obesity: an introduction to a special issue of the Translational Behavioral Medicine journal. *Transl Behav Med.* 2019, 9, 823; DOI: 10.1093/tbm/ibz097. PMID: 31682731.
- 71. James, A.; Wang, K.; Wang, Y. Therapeutic activity of green tea epigallocatechin-3-gallate on metabolic diseases and non-alcoholic fatty liver diseases: The current updates. *Nutrients.* **2023**, *15*, 3022; DOI: 10.3390/nu15133022
- 72. Andreu-Fernández, V.; Almeida-Toledano, L.; Pizarro-Lozano, N.; Navarro-Tapia, E.; Gómez-Roig, M.D.; De la Torre-Fornell, R. Bioavailability of epigallocatechin gallate administered with different nutritional strategies in healthy volunteers. *Antioxidants (Basel)* **2020**, *9*, 440; DOI: 10.3390/antiox9050440
- 73. Van-Amelsvoort, J.M.; Van-Hof, K.H.; Mathot, J.N.; Mulder, T.P.; Wiersma, A.; Tijburg, L.B. Plasma concentrations of individual tea catechins after a single oral dose in humans. *Xenobiotica*. **2001**, *31*, 891; DOI: 10.1080/00498250110079149
- 74. Yuan, H.; Li, Y.; Ling, F.; Guan, Y.; Zhang, D.; Zhu, Q. The phytochemical epigallocatechin gallate prolongs the lifespan by improving lipid metabolism, reducing inflammation and oxidative stress in high-fat dietfed obese rats. *Aging Cell.* **2020**, *19*, e13199; DOI: 10.1111/acel.13199
- 75. Savova, M.S.; Mihaylova, L.V.; Tews, D.; Wabitsch, M.; Georgiev, M.I. Targeting PI3K/AKT signaling pathway in obesity. *Biomed Pharmacother.* **2023**, *159*, 114244; DOI: 10.1016/j.biopha.2023.114244
- 76. Yoshitomi, R.; Yamamoto, M.; Kumazoe, M.; Fujimura, Y.; Yonekura, M.; Shimamoto, Y. The combined effect of green tea and α -glucosyl hesperidin in preventing obesity: a randomized placebo-controlled clinical trial. *Sci Rep.* **2021**, *11*, 19067; DOI: s41598-021-98612-6
- 77. Chatree, S.; Sitticharoon, C.; Maikaew, P.; Pongwattanapakin, K.; Keadkraichaiwat, I.; Churintaraphan, M. Epigallocatechin gallate decreases plasma triglyceride, blood pressure, and serum kisspeptin in obese human subjects. *Exp Biol Med (Maywood)*. **2021**, 246, 163; DOI: 10.1177/1535370220962708
- 78. Ríos, J.L.; Giner, R.M.; Marín, M.; Recio, M.C. A pharmacological update of ellagic acid. *Planta Med.* **2018**, 84, 1068; DOI: 10.1055/a-0633-9492
- 79. Clifford, M.N.; Scalbert, A. Ellagitannins nature, occurrence and dietary burden. *Journal of the Science of Food and Agriculture* **2000**, *80*, 1118; DOI: https://doi.org/10.1002/(SICI)1097-0010(20000515)80:7<1118::AID-JSFA570>3.0.CO;2-9
- 80. Lu, G.; Wang, X.; Cheng, M.; Wang, S.; Ma, K. The multifaceted mechanisms of ellagic acid in the treatment of tumors: State-of-the-art. *Biomed Pharmacother*. **2023**, *165*, 115132; DOI: https://doi.org/10.1016/j.biopha.2023.115132
- 81. Kowshik, J.; Giri, H.; Kishore, T.K.K.; Kesavan, R.; Vankudavath, R.N.; Reddy, G.B. Ellagic acid inhibits VEGF/VEGFR2, PI3K/Akt and MAPK signaling cascades in the hamster cheek pouch carcinogenesis model. *Anti-Cancer Agents in Medicinal Chemistry* **2014**, *14*, 1249; DOI: 10.2174/1871520614666140723114217
- 82. Polce, S.A.; Burke, C.; França, L.M.; Kramer, B.; de Andrade-Paes, A.M.; Carrillo-Sepulveda, M.A. Ellagic acid alleviates hepatic oxidative stress and insulin resistance in diabetic female rats. *Nutrients*. **2018**, *10*, F531; DOI: 10.3390/nu10050531
- 83. Woo, M.S.; Choi, H.S.; Seo, M.J.; Jeon, H.J.; Lee, B.Y. Ellagic acid suppresses lipid accumulation by suppressing early adipogenic events and cell cycle arrest. *Phytother Res.* **2015**, 29, 398; DOI: 10.1002/ptr.5264
- 84. Wang, L.; Li, L.; Ran, X.; Long, M.; Zhang, M.; Tao, Y. Ellagic acid reduces adipogenesis through inhibition of differentiation-prevention of the induction of rb phosphorylation in 3T3-L1 adipocytes. *Evid Based Complement Alternat Med.* **2013**, 2013, 287534; DOI: 10.1155/2013/287534
- 85. Hudak, C.S.; Sul, H.S. Pref-1, a gatekeeper of adipogenesis. *Front Endocrinol (Lausanne)*. **2013**, *4*, 79; DOI: 10.3389/fendo.2013.00079

- 86. Hidalgo-Lozada, G.M.; Villarruel-López, A.; Martínez-Abundis, E.; Vázquez-Paulino, O.; González-Ortiz, M.; Pérez-Rubio, K.G. Ellagic acid effect on the components of metabolic syndrome, insulin sensitivity and insulin secretion: a randomized, double-blind, placebo-controlled clinical trial. *J Clin Med.* **2022**, *11*, 5741; DOI: 10.3390/jcm11195741
- 87. Shiojima, Y.; Takahashi, M.; Kikuchi, M.; Akanuma, M. Effect of ellagic acid on body fat and triglyceride reduction in healthy overweight volunteers: a randomized, double-blind, placebo-controlled parallel group study. *Functional Foods in Health and Disease* **2020**, *10*, 180; DOI: 10.31989/ffhd.v10i4.702
- 88. Liu, Y.; Yu, S.; Wang, F.; Yu, H.; Li, X.; Dong, W. Chronic administration of ellagic acid improved the cognition in middle-aged overweight men. *Appl Physiol Nutr Metab.* **2018**, *43*, 266; DOI: 10.1139/apnm-2017-0583
- 89. Pasanta, D.; Htun, K.T.; Pan, J.; Tungjai, M.; Kaewjaeng, S.; Chancharunee, S. Waist Circumference and BMI are strongly correlated with mri-derived fat compartments in young adults. *Life (Basel)*, **2021**, *11*, 643; DOI: 10.3390/life11070643
- 90. Powell-Wiley, T.M.; Poirier, P.; Burke, L.E.; Després, J.P.; Gordon-Larsen, P.; Lavie, C.J. Obesity and cardiovascular disease: a scientific statement from the american heart association. *Circulation* **2021**, *143*, e984; DOI: 10.1161/CIR.0000000000000973
- 91. Mohammadi, H.; Ohm, J.; Discacciati, A.; Sundstrom, J.; Hambraeus, K.; Jernberg, T. Abdominal obesity and the risk of recurrent atherosclerotic cardiovascular disease after myocardial infarction. *Eur J Prev Cardiol.* **2020**, *27*, 1944 DOI: 10.1177/2047487319898019
- 92. Panchal, S.K.; Ward, L.; Brown, L. Ellagic acid attenuates high-carbohydrate, high-fat diet-induced metabolic syndrome in rats. *Eur J Nutr.* **2013**, *52*, 559; DOI: 10.1007/s00394-012-0358-9
- 93. Wang, L.; Wei, Y.;, Ning, C.; Zhang, M.; Fan, P.; Lei, D. Ellagic acid promotes browning of white adipose tissues in high-fat diet-induced obesity in rats through suppressing white adipocyte maintaining genes. *Endocrine Journal* **2019**, *66*, 923; DOI: 10.1507/endocrj.EJ18-0467
- 94. Tasaki, M.; Umemura, T.; Maeda, M.; Ishii. Y.; Okamura, T.; Inoue, T. Safety assessment of ellagic acid, a food additive, in a subchronic toxicity study using F344 rats. *Food and Chemical Toxicology* **2008**, *46*, 1119; DOI: 10.1016/j.fct.2007.10.043
- 95. Barber, T.M.; Kabisch, S.; Randeva, H.S.; Pfeiffer, A.F.H.; Weickert, M.O. Implications of Resveratrol in Obesity and Insulin Resistance: A State-of-the-Art Review. *Nutrients* **2022**, *14*, 2870; DOI: 10.3390/nu14142870
- 96. Diaz-Gerevini, G.T.; Repossi, G.; Dain, A.; Tarres, M.C. Das, U.N.; Eynard, A.R. Beneficial action of resveratrol: How and why? *Nutrition* **2016**, 32, 174; DOI: 10.1016/j.nut.2015.08.017
- 97. Zhang, M.; Xue, Y.; Chen, H.; Meng, L.; Chen, B.; Gong, H. Resveratrol inhibits MMP3 and MMP9 expression and secretion by suppressing TLR4/NF-κB/STAT3 activation in Ox-LDL-treated HUVECs. *Oxid Med Cell Longev.* **2019**, 2019, 9013169; DOI:10.1155/2019/9013169
- 98. Yang, C.M.; Chen, Y.W.; Chi, P.L.; Lin, C.C.; Hsiao, L.D. Resveratrol inhibits BK-induced COX-2 transcription by suppressing acetylation of AP-1 and NF-κB in human rheumatoid arthritis synovial fibroblasts. *Biochem Pharmacol.* **2017**, 132, 77; doi: 10.1016/j.bcp.2017.03.003
- 99. Walle, T.; Hsieh, F.; DeLegge, M.H.; Oatis, J.E.; Walle, U.K. High absorption but very low bioavailability of oral resveratrol in humans. *Drug Metab Dispos.* **2004**, *32*, 1377; DOI: 10.1124/dmd.104.000885
- 100. Springer, M.; Moco, S. Resveratrol and Its Human Metabolites Effects on Metabolic Health and Obesity. *Nutrients* **2019**, *11*, 143; DOI: 10.3390/nu11010143
- 101. Hoca, M.; Becer, E.; Vatansever, H.S. The role of resveratrol in diabetes and obesity associated with insulin resistance. *Arch Physiol Biochem.* **2023**, 129, 555; DOI: 10.1080/13813455.2021.1893338
- 102. Shahcheraghi, S.H.; Salemi, F.; Small, S.; Syed, S.; Salari, F.; Alam, W. Resveratrol regulates inflammation and improves oxidative stress via Nrf2 signaling pathway: Therapeutic and biotechnological prospects. *Phytother Res.* **2023**, *37*, 1590; DOI: 10.1002/ptr.7754
- 103. Chen, S.; Zhou, N.; Zhang, Z.; Li, W.; Zhu, W. Resveratrol induces cell apoptosis in adipocytes via AMPK activation. *Biochem Biophys Res Commun.* **2015**, 457, 608; DOI: 10.1016/j.bbrc.2015.01.034
- 104. Asghari, S.; Asghari-Jafarabadi, M.; Somi, M.H.; Ghavami, S.M.; Rafraf, M. Comparison of Calorie-Restricted Diet and Resveratrol Supplementation on Anthropometric Indices, Metabolic Parameters, and Serum Sirtuin-1 Levels in Patients With Nonalcoholic Fatty Liver Disease: A Randomized Controlled Clinical Trial. *J Am Coll Nutr.* **2018**, 37, 223; DOI: 10.1080/07315724.2017.1392264
- 105. Tabrizi, R.; Tamtaji, O.R.; Lankarani, K.B.; Akbari, M.; Dadgostar, E.; Dabbaghmanesh, M.H. The effects of resveratrol intake on weight loss: a systematic review and meta-analysis of randomized controlled trials. *Crit Rev Food Sci Nutr.* **2020**, *60*, 375; DOI: 10.1080/10408398.2018.1529654
- 106. Ali-Sangouni, A.; Abdollahi, S.; Mozaffari-Khosravi, H. Effect of resveratrol supplementation on hepatic steatosis and cardiovascular indices in overweight subjects with type 2 diabetes: a double-blind, randomized controlled trial. *BMC Cardiovasc Disord*. **2022**, 22, 212; DOI: 10.1186/s12872-022-02637-2

- 107. Gu, W.; Geng, J.; Zhao, H.; Li, X.; Song, G. Effects of resveratrol on metabolic indicators in patients with type 2 diabetes: a systematic review and meta-analysis. *Int J Clin Pract.* **2022**, 2022, 9734738; DOI: 10.1155/2022/9734738
- 108. Delpino, F.M.; Figueiredo, L.M.; Caputo, E.L.; Mintem, G.C.; Gigante, D.P. What is the effect of resveratrol on obesity? A systematic review and meta-analysis. *Clin Nutr ESPEN* **2021**, 41, 59; DOI: 10.1016/j.clnesp.2020.11.025
- 109. Mousavi, S.M.; Milajerdi, A.; Sheikhi, A.; Kord-Varkaneh, H.; Feinle-Bisset, C.; Larijani, B. et al. Resveratrol supplementation significantly influences obesity measures: a systematic review and dose-response meta-analysis of randomized controlled trials. *Obes Rev.* **2019**, *20*, 487; DOI: 10.1111/obr.12775
- 110. Noh, J.W.; Jun, M.S.; Yang, H.K.; Lee, B.C. cellular and molecular mechanisms and effects of berberine on obesity-induced inflammation. *Biomedicines* **2022**, 10, 1739; DOI: 10.3390/biomedicines10071739
- 111. Han, Y.; Xiang, Y.; Shi, Y.; Tang, X.; Pan, L.; Gao, J.; Bi, R.; Lai, X. Pharmacokinetics and pharmacological activities of berberine in diabetes mellitus treatment. *Evid Based Complement Alternat Med.* **2021**, 2021, 9987097; DOI: 10.1155/2021/9987097
- 112. Hu, Y.; Davies, G.E. Berberine inhibits adipogenesis in high-fat diet-induced obesity mice. *Fitoterapia* **2010**, *81*, 358; DOI: 10.1016/j.fitote.2009.10.010
- 113. Tong, Q.; Tsai, J.; Tan, G.; Dalgin, G.; Hotamisligil, G.S. Interaction between GATA and the C/EBP family of transcription factors is critical in GATA-mediated suppression of adipocyte differentiation. *Mol Cell Biol.* **2005**, 25, 706; DOI: 10.1128/MCB.25.2.706-715.2005
- 114. Tong, Q.; Dalgin, G.; Xu, H.; Ting, C.N.; Leiden, J.M.; Hotamisligil, G.S. Function of GATA transcription factors in preadipocyte-adipocyte transition. *Science* **2000**, 290, 134; DOI: 10.1126/science.290.5489.134
- 115. Choi, B.H.; Ahn, I.S.; Kim, Y.H.; Park, J.W.; Lee, S.Y.; Hyun, C.K. et al. Berberine reduces the expression of adipogenic enzymes and inflammatory molecules of 3T3-L1 adipocyte. *Exp Mol Med.* **2006**, *38*, 599; DOI: 10.1038/emm.2006.71
- 116. Lee, Y.S.; Kim, W.S.; Kim, K.H.; Yoon, M.J.; Cho, H.J.; Shen, Y. et al. Berberine, a Natural Plant Product, Activates AMP-Activated Protein Kinase With Beneficial Metabolic Effects in Diabetic and Insulin-Resistant States. *Diabetes* **2006**, *55*, 2256; DOI: 10.2337/db06-0006
- 117. Xu, Y.; Yu, T.; Ma, G.; Zheng, L.; Jiang, X.; Yang, F. et al. Berberine modulates deacetylation of PPARγ to promote adipose tissue remodeling and thermogenesis via AMPK/SIRT1 pathway. *Int J Biol Sci.* **2021**, *17*, 3173; DOI: 10.7150/ijbs.62556
- 118. Wang, L.; Ye, X.; Hua, Y.; Song, Y. Berberine alleviates adipose tissue fibrosis by inducing AMP-activated kinase signaling in high-fat diet-induced obese mice. *Biomedicine & Pharmacotherapy* **2018**, , 105121; DOI: 10.1016/j.biopha.2018.05.110
- 119. Yang, J.; Yin, J.; Gao, H.; Xu, L.; Wang, Y.; Xu, L. et al. Berberine Improves Insulin Sensitivity by Inhibiting Fat Store and Adjusting Adipokines Profile in Human Preadipocytes and Metabolic Syndrome Patients. *Evid Based Complement Alternat Med.* **2012**, 2012, 363845; DOI: 10.1155/2012/363845
- 120. 120. Firouzi, S.; Malekahmadi, M.; Ghayour-Mobarhan, M.; Ferns, G.; Rahimi, H.R. Barberry in the treatment of obesity and metabolic syndrome: possible mechanisms of action. *Diabetes Metab Syndr Obes.* **2018**, *11*, 699; DOI: 10.2147/DMSO.S181572
- 121. Ilyas, Z.; Perna, S.; Al-Thawadi, S.; Alalwan, T.A.; Riva, A.; Petrangolini, G. et al. The effect of Berberine on weight loss in order to prevent obesity: A systematic review. *Biomed Pharmacother.* **2020**, *127*, 110137; DOI: 10.1016/j.biopha.2020.110137
- 122. Pei, C.; Zhang, Y.; Wang, P.; Zhang, B.; Fang, L.; Liu, B. et al. Berberine alleviates oxidized low-density lipoprotein-induced macrophage activation by downregulating galectin-3 via the NF-κB and AMPK signaling pathways. *Phytotherapy Research* **2019**, 33, 294; DOI: 10.1002/ptr.6217
- 123. Cheng, H.; Liu, J.; Tan, Y.; Feng, W.; Peng, C. Interactions between gut microbiota and berberine, a necessary procedure to understand the mechanisms of berberine. *J Pharm Anal.* **2022**, *12*, 541; DOI: 10.1016/j.jpha.2021.10.003
- 124. Xiong, P.; Niu, L.; Talaei, S.; Kord-Varkaneh H.; Clark, C.C.T.; Găman, M.A. et al. The effect of berberine supplementation on obesity indices: A dose–response meta-analysis and systematic review of randomized controlled trials. *Complement Ther Clin Pract.* **2020**, *39*, 101113; DOI: 10.1016/j.ctcp.2020.101113
- 125. Zhao, J.V.; Yeung, W.F.; Chan, Y.H.; Vackova, D.; Leung, J.Y.Y.; Ip, D.K.M. et al. Effect of Berberine on Cardiovascular Disease Risk Factors: A Mechanistic Randomized Controlled Trial. *Nutrients* **2021**, *13*, 2250; DOI: 10.3390/nu13082550
- 126. Amini, M.R.; Sheikhhossein, F.; Naghshi, S.; Djafari, F.; Askari, M.; Shahinfar, H. et al. Effects of berberine and barberry on anthropometric measures: A systematic review and meta-analysis of randomized controlled trials. *Complement Ther Med.* 2020, 49, 102337; DOI: 10.1016/j.ctim.2020.102337
- 127. Rondanelli, M.; Gasparri, C.; Petrangolini, G.; Allegrini, P.; Avenoso, D.; Fazia, T. et al. Berberine phospholipid exerts a positive effect on the glycemic profile of overweight subjects with impaired fasting blood glucose (IFG): a randomized double-blind placebo-controlled clinical trial. *Eur Rev Med Pharmacol Sci.* 2023, 27, 6718

- 128. Zhang, J.; Han, C.; Lu, W.Q.; Wang, N.; Wu, S.R.; Wang, Y.X. et al. A randomized, multicenter and noninferiority study of amoxicillin plus berberine vs tetracycline plus furazolidone in quadruple therapy for Helicobacter pylori rescue treatment. *J Dig Dis.* **2020**, *21*, 256; DOI: 10.1111/1751-2980.12870
- 129. Chen, Y.X.; Gao, Q.Y.; Zou, T.H.; Wang, B.M.; Liu, S.D.; Sheng, J.Q. et al. Berberine versus placebo for the prevention of recurrence of colorectal adenoma: a multicentre, double-blinded, randomised controlled study. *Lancet Gastroenterol Hepatol.* **2020**, *5*, 267; DOI: 10.1016/S2468-1253(19)30409-1
- 130. Zhang, Y.; Li, X.; Zou, D.; Liu, W.; Yang, J.; Zhu, N. et al. Treatment of type 2 diabetes and dyslipidemia with the natural plant alkaloid berberine. *J Clin Endocrinol Metab.* **2008**, 93, 2559; DOI: 10.1210/jc.2007-2404
- 131. Jahnke, G.D.; Price, C.J.; Marr, M.C.; Myers, C.B.; George, J.D. Developmental toxicity evaluation of berberine in rats and mice. *Birth Defects Research Part B: Developmental and Reproductive Toxicology* **2006**, 77, 191; DOI: https://doi.org/10.1002/bdrb.20075
- 132. Ye, Y.; Liu, X.; Wu, N.; Han, Y.; Wang, J.; Yu, Y. et al. Efficacy and safety of berberine alone for several metabolic disorders: a systematic review and meta-analysis of randomized clinical trials. *Front Pharmacol.* **2021**, *12*, 653887; DOI: 10.3389/fphar.2021.653887
- 133. Santamarina, A.B.; Calder, P.C.; Estadella, D.; Pisani, L.P. Anthocyanins ameliorate obesity-associated metainflammation: Preclinical and clinical evidence. *Nutr Res.* **2023**, *114*, 50; DOI: 10.1016/j.nutres.2023.04.004
- 134. Eker, M.E.; Aaby, K.; Budic-Leto, I.; Rimac-Brnčić, S.; El, S.N.; Karakaya, S. et al. A review of factors affecting anthocyanin bioavailability: possible implications for the inter-individual variability. *Foods* **2020**, 9, 2; DOI: 10.3390/foods9010002
- 135. Yoon, Y.; Yoon, H.; Park, H.M.; Song, S.; Yeum, K.J. Dietary anthocyanins against obesity and inflammation. *Nutrients* **2017**, *9*, 1089; DOI: 10.3390/nu9101089
- 136. Rossi, A.; Serraino, I.; Dugo, P.; Di-Paola, R.; Mondello, L.; Genovese, T. et al. Protective effects of anthocyanins from blackberry in a rat model of acute lung inflammation. *Free Radic Res.* **2003**, *37*, 891; DOI: 10.1080/1071576031000112690
- 137. DeFuria, J.; Bennett, G.; Strissel, K.J.; Perfield, J.W.; Milbury, P.E.; Greenberg, A.S. et al. Dietary blueberry attenuates whole-body insulin resistance in high fat-fed mice by reducing adipocyte death and its inflammatory sequelae. *J Nutr.* **2009**, *139*, 1510; DOI: 10.3945/jn.109.105155
- 138. Czank, C.; Cassidy, A.; Zhang, Q.; Morrison, D.J.; Preston, T.; Kroon, P.A. et al. Human metabolism and elimination of the anthocyanin, cyanidin-3-glucoside: a (13)C-tracer study. *Am J Clin Nutr.* **2013**, 97, 995; DOI: 10.3945/ajcn.112.049247
- 139. Manach, C.; Williamson, G.; Morand, C.; Scalbert, A.; Rémésy, C. Bioavailability and bioefficacy of polyphenols in humans. I. Review of 97 bioavailability studies. *Am J Clin Nutr.* **2005**, *81*, 230S; DOI: 10.1093/ajcn/81.1.230S
- 140. Heo, H.J.; Lee, C.Y. Strawberry and its anthocyanins reduce oxidative stress-induced apoptosis in PC12 cells. *J Agric Food Chem.* **2005**, *53*, 1984; DOI: 10.1021/jf048616l
- 141. Ngamsamer, C.; Sirivarasai, J.; Sutjarit, N. The Benefits of Anthocyanins against Obesity-Induced Inflammation. *Biomolecules* **2022**, *12*, 852; DOI: 10.3390/biom12060852
- 142. Li, N.; Liu, X.; Zhang, J.; Lang, Y.Z.; Lu, L.; Mi, J. et al. Preventive Effects of Anthocyanins from Lyciumruthenicum Murray in High-Fat Diet-Induced Obese Mice Are Related to the Regulation of Intestinal Microbiota and Inhibition of Pancreatic Lipase Activity. *Molecules* 2022, 27, 2141; DOI: 10.3390/molecules27072141
- 143. Fotschki, B.; Juśkiewicz, J.; Jurgoński, A.; Sójka, M. Fructo-Oligosaccharides and Pectins Enhance Beneficial Effects of Raspberry Polyphenols in Rats with Nonalcoholic Fatty Liver. *Nutrients* **2021**, *13*, 833; DOI: https://doi.org/10.3390/nu13030833
- 144. Esatbeyoglu, T.; Rodríguez-Werner, M.; Schlösser, A.; Winterhalter, P.; Rimbach, G. Fractionation, enzyme inhibitory and cellular antioxidant activity of bioactives from purple sweet potato (Ipomoea batatas). *Food Chem.* **2017**, 221, 447; DOI: 10.1016/j.foodchem.2016.10.077
- 145. Wu, T.; Gao, Y.; Guo, X.; Zhang, M.; Gong, L. Blackberry and Blueberry Anthocyanin Supplementation Counteract High-Fat-Diet-Induced Obesity by Alleviating Oxidative Stress and Inflammation and Accelerating Energy Expenditure. *Oxid Med Cell Longev.* 2018, 2018, 4051232; DOI: 10.1155/2018/4051232
- 146. Wu, T.; Yin, J.; Zhang, G.; Long, H.; Zheng, X. Mulberry and cherry anthocyanin consumption prevents oxidative stress and inflammation in diet-induced obese mice. *Mol Nutr Food Res.* **2016**, *60*, 687; DOI: 10.1002/mnfr.201500734
- 147. Alvarez-Suarez, J.M.; Giampieri, F.; Tulipani, S.; Casoli, T.; Di-Stefano, G.; González-Paramás, A.M. et al. One-month strawberry-rich anthocyanin supplementation ameliorates cardiovascular risk, oxidative stress markers and platelet activation in humans. *J Nutr Biochem.* **2014**, *25*, 289; DOI: 10.1016/j.jnutbio.2013.11.002
- 148. Li, D.; Zhang, Y.; Liu, Y.; Sun, R.; Xia, M. Purified anthocyanin supplementation reduces dyslipidemia, enhances antioxidant capacity, and prevents insulin resistance in diabetic patients. *J Nutr.* **2015**, *145*, 742; DOI: 10.3945/jn.114.205674

- 149. Arisi, T.O.P.; Gorski, F.; Eibel, B.; Barbosa, E.; Boll, L.; Waclawovsky, G. et al. Dietary intake of anthocyanins improves arterial stiffness, but not endothelial function, in volunteers with excess weight: A randomized clinical trial. *Phytother Res.* **2023**, 37, 798; DOI: 10.1002/ptr.7659
- 150. Park, S.; Choi, M.; Lee, M. Effects of Anthocyanin Supplementation on Reduction of Obesity Criteria: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Nutrients* **2021**, *13*, 2121; DOI: 10.3390/nu13062121
- 151. Stojanov, S.; Berlec, A.; Štrukelj, B. The Influence of Probiotics on the Firmicutes/Bacteroidetes Ratio in the Treatment of Obesity and Inflammatory Bowel disease. *Microorganisms*. **2020**, *8*, 1715; DOI: 10.3390/microorganisms8111715. PMID: 33139627; PMCID: PMC7692443.
- 152. Vallianou, N.G.; Kounatidis, D.; Tsilingiris, D.; Panagopoulos, F.; Christodoulatos, G.S.; Evangelopoulos, A. et al. The Role of Next-Generation Probiotics in Obesity and Obesity-Associated Disorders: Current Knowledge and Future Perspectives. *Int J Mol Sci.* **2023**, *24*, 6755; DOI: https://doi.org/10.3390/ijms24076755
- 153. Kobyliak, N.; Conte, C.; Cammarota, G.; Haley, A.P.; Styriak, I.; Gaspar, L. et al. Probiotics in prevention and treatment of obesity: a critical view. *Nutr Metab (Lond)* **2016**, *13*, 14; DOI: 10.1186/s12986-016-0067-0
- 154. Cai, Y.; Liu, P.; Zhou, X.; Yuan, J.; Chen, Q. Probiotics therapy show significant improvement in obesity and neurobehavioral disorders symptoms. *Front Cell Infect Microbiol.* **2023**, *13*, 1178399; DOI: 10.3389/fcimb.2023.1178399
- 155. Depommier, C.; Everard, A.; Druart, C.; Plovier, H.; Van Hul, M.; Vieira-Silva, S.; Falony, G.; Raes, J.; Maiter, D.; Delzenne, N. M.; de Barsy, M.; Loumaye, A.; Hermans, M. P.; Thissen, J. P.; de Vos, W. M.; Cani, P. D. Supplementation with Akkermansia muciniphila in overweight and obese human volunteers: a proof-of-concept exploratory study. *Nat Med.* **2019**, *25*, 1096; DOI: 10.1038/s41591-019-0495-2
- 156. Markowiak, P.; Śliżewska, K. Effects of Probiotics, Prebiotics, and Synbiotics on Human Health. *Nutrients* **2017**, *9*, 1021; DOI: 10.3390/nu9091021
- 157. Kadooka, Y.; Sato, M.; Imaizumi, K.; Ogawa, A.; Ikuyama, K.; Akai, Y. et al. Regulation of abdominal adiposity by probiotics (Lactobacillus gasseri SBT2055) in adults with obese tendencies in a randomized controlled trial. *Eur J Clin Nutr.* **2010**, *64*, *636*; DOI: 10.1038/ejcn.2010.19
- 158. Uusitupa, H. M.; Rasinkangas, P.; Lehtinen, M. J.; Mäkelä, S. M.; Airaksinen, K.; Anglenius, H.,; Ouwehand, A. C.; Maukonen, J. Bifidobacterium animalis subsp. lactis 420 for Metabolic Health: Review of the Research. *Nutrients* **2020**, *12*, 892; DOI: 10.3390/nu12040892
- 159. Larsen, N.; Vogensen, F.K.; Gøbel, R.J.;, Michaelsen, K.F.; Forssten, S.D.; Lahtinen, S.J. et al. Effect of Lactobacillus salivarius Ls-33 on fecal microbiota in obese adolescents. *Clin Nutr.* **2013**, *32*, 935; DOI: 10.1016/j.clnu.2013.02.007
- 160. Pedret, A.; Valls, R.M.; Calderón-Pérez, L.; Llauradó, E.; Companys, J.; Pla-Pagà, L. et al. Effects of daily consumption of the probiotic Bifidobacterium animalis subsp. lactis CECT 8145 on anthropometric adiposity biomarkers in abdominally obese subjects: a randomized controlled trial. *Int J Obes.* **2019**, 43, 1863; DOI: 10.1038/s41366-018-0220-0
- 161. Ben-Othman, R.; Ben-Amor, N.; Mahjoub, F.; Berriche, O.; El-Ghali, C.; Gamoudi, A. et al. A clinical trial about effects of prebiotic and probiotic supplementation on weight loss, psychological profile and metabolic parameters in obese subjects. *Endocrinol Diabetes Metab.* **2023**, *6*, e402; DOI: 10.1002/edm2.402
- 162. Danielsson, P.; Putri, R.R.; Marcus, C.; Hagman, E. Evaluating probiotic efficacy on weight loss in adults with overweight through a double-blind, placebo-controlled randomized trial. *Sci Rep.* **2023**, *13*, 18200; DOI: https://doi.org/10.1038/s41598-023-45395-7
- 163. Choi, B.S.Y.; Brunelle, L.; Pilon, G.; Cautela, B.G.; Tompkins, T.A.; Drapeau, V. et al. Lacticaseibacillus rhamnosus HA-114 improves eating behaviors and mood-related factors in adults with overweight during weight loss: a randomized controlled trial. *Nutr Neurosci.* **2023**, 26, 667; DOI: 10.1080/1028415X.2022.2081288
- 164. Mounien, L.; Tourniaire, F.; Landrier, J.F. Anti-obesity effect of carotenoids: direct impact on adipose tissue and adipose tissue-driven indirect effects. *Nutrients* **2019**, *11*, 1562; DOI: 10.3390/nu11071562
- 165. Bohn, T.; Bonet, M.L.; Borel, P.; Keijer, J.; Landrier, J.F.; Milisav, I. et al. Mechanistic aspects of carotenoid health benefits where are we now? *Nutr Res Rev.* **2021**, *34*, 276; DOI: 10.1017/S0954422421000147
- 166. Li, K.; Wang, W.; Xiao, W. Astaxanthin: A promising therapeutic agent for organ fibrosis. *Pharmacol Res.* **2023**, *188*, 106657; DOI: 10.1016/j.phrs.2023.106657
- 167. Yang, G.; Liu, X.; Jing, X.; Wang, J.; Wang, H.; Chen, F. et al. Astaxanthin suppresses oxidative stress and calcification in vertebral cartilage endplate via activating Nrf-2/HO-1 signaling pathway. *Int Immunopharmacol.* **2023**, *119*, 110159; DOI: https://doi.org/10.1016/j.intimp.2023.110159
- 168. Zhu, R.; Chen, B.; Bai, Y.; Miao, T.; Rui, L.; Zhang, H. et al. Lycopene in protection against obesity and diabetes: A mechanistic review. *Pharmacol Res.* **2020**, *159*, 104966; DOI: 10.1016/j.phrs.2020.104966
- 169. Yao, N.; Yan, S.; Guo, Y.; Wang, H.; Li, X.; Wang, L. et al. The association between carotenoids and subjects with overweight or obesity: a systematic review and meta-analysis. *Food Funct.* **2021**, *12*, 4768; DOI: 10.1039/d1fo00004g

- 170. Ma, B.; Lu, J.; Kang, T.; Zhu, M.; Xiong, K.; Wang, J. Astaxanthin supplementation mildly reduced oxidative stress and inflammation biomarkers: a systematic review and meta-analysis of randomized controlled trials. *Nutr Res.* **2022**, *99*, 40; DOI: 10.1016/j.nutres.2021.09.005
- 171. Leung, L.Y.L.; Chan, S.M.N.; Tam, H.L.; Wong, E.S.W. Astaxanthin Influence on Health Outcomes of Adults at Risk of Metabolic Syndrome: A Systematic Review and Meta-Analysis. *Nutrients* **2022**, *14*, 2050; DOI: 10.3390/nu14102050
- 172. Hao, M.; Chu, Y.; Lei, J.; Yao, Z.; Wang, P.; Chen, Z. et al. Pharmacological Mechanisms and Clinical Applications of Curcumin: Update. *Aging Dis.* **2023**, *14*, 716; DOI: 10.14336/AD.2022.1101
- 173. Wu, L.Y.; Chen, C.W.; Chen, L.K.; Chou, H.Y.; Chang, C.L.; Juan, C.C. Curcumin Attenuates Adipogenesis by Inducing Preadipocyte Apoptosis and Inhibiting Adipocyte Differentiation. *Nutrients* **2019**, *11*, 2307; DOI: 10.3390/nu11102307
- 174. Zhao, J.; Sun, X.B.; Ye, F.; Tian, W.X. Suppression of fatty acid synthase, differentiation and lipid accumulation in adipocytes by curcumin. *Mol Cell Biochem.* **2011**, *351*, 19; DOI: 10.1007/s11010-010-0707-z
- 175. Baziar, N.; Parohan, M. The effects of curcumin supplementation on body mass index, body weight, and waist circumference in patients with nonalcoholic fatty liver disease: A systematic review and doseresponse meta-analysis of randomized controlled trials. *Phytother Res.* **2020**, *34*, 464; DOI: 10.1002/ptr.6542
- 176. Unhapipatpong, C.; Polruang, N.; Shantavasinkul, P.C.; Julanon, N.; Numthavaj, P.; Thakkinstian, A. The effect of curcumin supplementation on weight loss and anthropometric indices: an umbrella review and updated meta-analyses of randomized controlled trials. *The American Journal of Clinical Nutrition* **2023**, 117, 1005; DOI: 10.1016/j.ajcnut.2023.03.006
- 177. Hellmann, P.H.; Bagger, J.I.; Carlander, K.R.; Forman, J.; Chabanova, E.; Svenningsen, J.S. et al. The effect of curcumin on hepatic fat content in individuals with obesity. *Diabetes Obes Metab.* **2022**, 24, 2192; DOI: 10.1111/dom.14804
- 178. Liu, S.; Liu, J.; He, L.; Liu, L.; Cheng, B.; Zhou, F. et al. A Comprehensive Review on the Benefits and Problems of Curcumin with Respect to Human Health. *Molecules* **2022**, 27,4400; DOI: 10.3390/molecules27144400
- 179. Dance-Barnes, S.T.; Kock, N.D.; Moore, J.E.; Lin, E.Y.; Mosley, L.J.; D'Agostino, R.B. Jr. et al. Lung tumor promotion by curcumin. *Carcinogenesis* **2009**, *30*, 1016; DOI: 10.1093/carcin/bgp082
- 180. Rithaporn, T.; Monga, M.; Rajasekaran, M. Curcumin: a potential vaginal contraceptive. *Contraception* **2003**, *68*, 219; DOI: 10.1016/s0010-7824(03)00163-x
- 181. Federico, A.; Dallio, M.; Loguercio, C. Silymarin/Silybin and Chronic Liver Disease: A Marriage of Many Years. *Molecules* **2017**, 22, 191; DOI: 10.3390/molecules22020191
- 182. Hawke, R.L.; Schrieber, S.J.; Soule, T.A.; Wen, Z.; Smith, P.C.; Reddy, K.R. et al. Silymarin Ascending Multiple Oral Dosing Phase I Study in Noncirrhotic Patients With Chronic Hepatitis C. *J Clin Pharmacol.* **2010**, *50*, 434; DOI: 10.1177/0091270009347475
- 183. Zarrelli, A.; Romanucci, V.; Tuccillo, C.; Federico, A.; Loguercio, C.; Gravante, R. et al. New silibinin glycoconjugates: synthesis and evaluation of antioxidant properties. *Bioorg Med Chem Lett.* **2014**, 24, 5147; DOI: 10.1016/j.bmcl.2014.10.023
- 184. Saller, R.; Brignoli, R.; Melzer, J.; Meier, R. An updated systematic review with meta-analysis for the clinical evidence of silymarin. *Forsch Komplementmed*. **2008**, *15*, 9; DOI: 10.1159/000113648
- 185. Aghemo, A.; Alekseeva, O.P.; Angelico, F.; Bakulin, I.G.; Bakulina, N.V.; Bordin, D. et al. Role of silymarin as antioxidant in clinical management of chronic liver diseases: a narrative review. *Ann Med.* **2022**, *54*, 1548; DOI: 10.1080/07853890.2022.2069854
- 186. Kim, S.H.; Choo, G.S.; Yoo, E.S.; Woo, J.S.; Han, S.H.; Lee, J.H. et al. Silymarin induces inhibition of growth and apoptosis through modulation of the MAPK signaling pathway in AGS human gastric cancer cells. *Oncol Rep.* **2019**, *42*, 1904; DOI: 10.3892/or.2019.7295
- 187. Gao, X.; Xiao, Z.H.; Liu, M.; Zhang, N.Y.; Khalil, M.M.; Gu, C.Q. et al. dietary silymarin supplementation alleviates zearalenone-induced hepatotoxicity and reproductive toxicity in rats. *J Nutr.* **2018**, *148*, 1209; DOI: 10.1093/jn/nxy114
- 188. Lee, J.A.; Shin, M.R.; Choi, J.; Kim, M.; Park, H.J.; Roh, S.S. Co-treatments of gardeniae fructus and silymarin ameliorates excessive oxidative stress-driven liver fibrosis by regulation of hepatic Sirtuin1 activities using thioacetamide-induced mice model. *Antioxidants* (*Basel*) **2022**, *12*, 97; DOI: https://doi.org/10.3390/antiox12010097
- 189. Yassin, N.Y.S.; AbouZid, S.F.; El-Kalaawy, A.M.; Ali, T.M.; Elesawy, B.H.; Ahmed, O.M. Tackling of renal carcinogenesis in wistar rats by silybum marianum total extract, silymarin, and silibinin via modulation of oxidative stress, apoptosis, Nrf2, PPARγ, NF-κB, and PI3K/Akt signaling pathways. *Oxid Med Cell Longev.* **2021**, 2021, 7665169; DOI: 10.1155/2021/7665169
- 190. Du, Q.; Wu, X.; Ma, K.; Liu, W.; Liu, P.; Hayashi, T. et al. Silibinin alleviates ferroptosis of rat islet β cell INS-1 induced by the treatment with palmitic acid and high glucose through enhancing PINK1/parkin-mediated mitophagy. *Arch Biochem Biophys.* **2023**, 743, 109644; DOI: 10.1016/j.abb.2023.109644

- 191. Sohail, I.; Malkani, N.; Tahir, N.; Khalil, A.; Attar, R.; Mumtaz, S. Silymarin protects the liver from α-naphthylisothiocyanate-induced cholestasis by modulating the expression of genes involved in bile acid homeostasis. *Cell Mol Biol (Noisy-le-grand)*. **2022**, *68*, 208; PMID: 36495494
- 192. Bellavite, P.; Fazio, S.; Affuso, F. A descriptive review of the action mechanisms of berberine, quercetin and silymarin on insulin resistance/hyperinsulinemia and cardiovascular prevention. *Molecules* **2023**, *28*, 4491; DOI: 10.3390/molecules28114491
- 193. Kalopitas, G.; Antza, C.; Doundoulakis, I.; Siargkas, A.; Kouroumalis, E.; Germanidis, G. et al. Impact of silymarin in individuals with nonalcoholic fatty liver disease: a systematic review and meta-analysis. *Nutrition* **2021**, *83*, 111092; DOI: 10.1016/j.nut.2020.111092
- 194. Aller, R.; Izaola, O.; Gómez, S.; Tafur, C.; González, G.; Berroa, E. et al. Effect of silymarin plus vitamin E in patients with non-alcoholic fatty liver disease. A randomized clinical pilot study. *Eur Rev Med Pharmacol Sci.* **2015**, *19*, 3118; PMID: 26367736.
- 195. Patti, A.M.; Al-Rasadi, K.; Katsiki, N.; Banerjee, Y.; Nikolic, D.; Vanella, L. et al. Effect of a natural supplement containing curcuma longa, guggul, and chlorogenic acid in patients with metabolic syndrome. *Angiology* **2015**, *66*, 856; DOI: 10.1177/0003319714568792
- 196. Amini, M. R.; Rasaei, N.; Jalalzadeh, M.; Akhgarjand, C.; Hashemian, M.; Jalali, P.; Hekmatdoost, A. The effects of Garcinia cambogia (hydroxycitric acid) on lipid profile: A systematic review and meta-analysis of randomized controlled trials. *Phytother Res.* **2023**; DOI: 10.1002/ptr.8102
- 197. Saito, M.; Ueno, M.; Ogino, S.; Kubo, K.; Nagata, J.; Takeuchi, M. High dose of Garcinia cambogia is effective in suppressing fat accumulation in developing male Zucker obese rats, but highly toxic to the testis. *Food Chem Toxicol* **2005**, *43*, 411; DOI: 10.1016/j.fct.2004.11.008
- 198. Fassina, P.; Scherer-Adami, F.; Zani, V.T.; Kasper-Machado, I.C.; Garavaglia, J.; Quevedo-Grave, M.T. et al. El efecto de la Garcinia Cambogia como coadyuvante en el proceso de perdida de peso. *Nutr Hosp.* **2015**, 32, 2400; DOI: 10.3305/nh.2015.32.6.9587
- 199. Maia-Landim, A.; Ramírez, J.M.; Lancho, C.; Poblador, M.S.; Lancho, J.L. Long-term effects of Garcinia cambogia/Glucomannan on weight loss in people with obesity, PLIN4, FTO and Trp64Arg polymorphisms. *BMC Complement Altern Med.* **2018**, *18*, 26; DOI: 10.1186/s12906-018-2099-7
- 200. Zovi, A.; Langella, R.; Nisic, A.; Vitiello, A.; Musazzi, U.M. Liver injury and dietary supplements: Does hydroxycitric acid trigger hepatotoxicity? *J Integr Med.* **2022**, *20*, 473; DOI: 10.1016/j.joim.2022.05.003
- 201. Vuppalanchi, R.; Bonkovsky, H.L.; Ahmad, J.; Barnhart, H.; Durazo, F.; Fontana, R.J. et al. Garcinia cambogia, Either Alone or in Combination With Green Tea, Causes Moderate to Severe Liver Injury. *Clin Gastroenterol Hepatol.* **2022**, 20, e1416; DOI: 10.1016/j.cgh.2021.08.015
- 202. Chong, P.W.; Beah, Z.M.; Grube, B.; Riede, L. IQP-GC-101 reduces body weight and body fat mass: a randomized, double-blind, placebo-controlled study. *Phytother Res.* **2014**, *28*, 1520; DOI: 10.1002/ptr.5158
- 203. Kim, J.E.; Jeon, S.M.; Park, K.H.; Lee, W.S.; Jeong, T.S.; McGregor, R.A. et al. Does Glycine max leaves or Garcinia Cambogia promote weight-loss or lower plasma cholesterol in overweight individuals: a randomized control trial. *Nutr J.* 2011, *10*, 94; DOI: 10.1186/1475-2891-10-94
- 204. Opala, T.; Rzymski, P.; Pischel, I.; Wilczak, M.; Wozniak, J. Efficacy of 12 weeks supplementation of a botanical extract-based weight loss formula on body weight, body composition and blood chemistry in healthy, overweight subjects--a randomised double-blind placebo-controlled clinical trial. *Eur J Med Res.* **2006**, *11*, 343; PMID: 17052970
- 205. Pons, L. Ácido lipoico: un debate cosmético. Offarm. 2003, 22, 157; ISSN: 0212-047X
- 206. Salehi, B.; Berkay-Yılmaz, Y.; Antika, G.; Boyunegmez-Tumer, T.; Fawzi-Mahomoodally, M.; Lobine, D. et al. Insights on the Use of α -Lipoic Acid for Therapeutic Purposes. *Biomoleculess* **2019**, *9*, 356; DOI: 10.3390/biom9080356
- 207. Capece, U.; Moffa, S.; Improta, I.; Di Giuseppe, G.; Nista, E.C.; Cefalo, C.M.A. et al. Alpha-Lipoic Acid and Glucose Metabolism: A Comprehensive Update on Biochemical and Therapeutic Features. *Nutrient.s* **2022**, *15*, 18; DOI: 10.3390/nu15010018
- 208. Kim, M.S.; Park, J.Y.; Namkoong, C.; Jang, P.G.; Ryu, J.W.; Song, H.S. et al. Anti-obesity effects of alphalipoic acid mediated by suppression of hypothalamic AMP-activated protein kinase. *Nat Med.* **2004**, *10*, 727; DOI: 10.1038/nm1061
- 209. Ishida, Y.; Ohara, T.; Okuno, Y.; Ito, T.; Hirota, Y.; Furukawa, K. et al. α-Lipoic Acid and Insulin Autoimmune Syndrome. *Diabetes Care*. **2007**, 30, 2240; DOI: https://doi.org/10.2337/dc07-0689
- 210. Vigil, M.; Berkson, B.M.; Garcia, A.P. Adverse effects of high doses of intravenous alpha lipoic Acid on liver mitochondria. Glob *Adv Health Med.* **2014** , 3, 25; doi: 10.7453/gahmj.2013.011
- 211. Kucukgoncu, S.; Zhou, E.; Lucas, K.B.; Tek, C. Alpha-lipoic acid (ALA) as a supplementation for weight loss: results from a meta-analysis of randomized controlled trials. *Obes Rev.* **2017**, 18, 594; DOI: 10.1111/obr.12528
- 212. Sztolsztener, K.; Hodun, K.; Chabowski, A. α-lipoic acid ameliorates inflammation state and oxidative stress by reducing the content of bioactive lipid derivatives in the left ventricle of rats fed a high-fat diet. *Biochim Biophys Acta Mol Basis Dis.* **2022**, 1868, 166440; DOI: 10.1016/j.bbadis.2022.166440

- 213. Tutunchi, H.; Arefhosseini, S.; Ebrahimi-Mameghani, M. Clinical effectiveness of α-lipoic acid, myoinositol and propolis supplementation on metabolic profiles and liver function in obese patients with NAFLD: A randomized controlled clinical trial. *Clin Nutr ESPEN* **2023**, 54, 412; DOI: 10.1016/j.clnesp.2023.02.016
- 214. Mohammadshahi, M.; Zakizadeh, E.; Ahmadi-Angal,i K.; Ravanbakhsh, M.; Helli, B. The synergic effects of alpha-lipoic acid supplementation and electrical isotonic contraction on anthropometric measurements and the serum levels of VEGF, NO, sirtuin-1, and PGC1-α in obese people undergoing a weight loss diet. *Arch Physiol Biochem.* **2022**, 128(5). DOI: 10.1080/13813455.2020.1762660
- 215. Nasiri, G.; Bastani, A.; Haji-Aghamohammadi, A.A.; Nooshabadi, M.R.; Shahmirzalou, P.; Haghighian, H.K. Effects of probiotic and alpha-lipoic acid supplements, separately or in combination on the anthropometric indicators and maintenance of weight in overweight individuals. *Clin Nutr ESPEN* **2021**, 41, 242; DOI: 10.1016/j.clnesp.2020.12.007

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