

Essay

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Posted Date: 2 April 2024

doi: 10.20944/preprints202404.0116.v1

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Essay

Molecules, Up Your Spins!

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Abstract: The last 50 years witnessed the emergence of various so called “hyperpolarization technologies” that allow generating large magnetizations of spin ensembles. This *Editorial Essay* provides a brief overview of the use of these technologies in various fields, including medicine, chemistry, and quantum sensing. It is concluded that critical questions still need to be addressed to harness benefits of nuclear hyperpolarization in clinical medical imaging while interdisciplinary innovation can unlock potential of novel (yet unexplored) methods. My hope is that this *Editorial Essay* provides valuable insights into the current state and future directions of spin technologies.

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1. Introduction

Nuclear magnetic resonance (NMR) spectroscopy and magnetic resonance imaging (MRI) are indispensable tools in science and medicine, offering insights into functions of biological processes. Traditional analytical methods, such as mass spectroscopy and chromatography are significantly more sensitive, but they require destruction of samples during analysis.[1–3] In contrast, NMR and MRI rely on the detection of signals from nuclear spins without altering samples, making these modalities truly non-invasive and ideal for studying biological systems.

The non-invasiveness of NMR spectroscopy and imaging (NMR/MRI) stems from the low energy of spin-field interaction quanta ($h\nu$) compared to thermal energy of the environment (kT). Here, $\nu = |\gamma B|$ is a frequency of spin precession (γ is a gyromagnetic ratio of the spins, B is a static magnetic field used in NMR/MRI, typically several tesla), T is temperature, h and k are Planck's and Boltzmann's constants, respectively. In magnetic resonance, *polarization* (P) is defined as a dimensionless quantity directly proportional to the ratio of the above-mentioned energies[4]: $P \sim h\nu/kT$. At room temperature, kT is about $5 \cdot 10^{-21}$ joules while $h\nu$ for protons is on the order of 10^{-25} joules even at magnetic fields of modern high-field NMR spectrometers. It is this low interaction energy between spins and the external field that is a cornerstone to noninvasiveness of NMR/MRI: spins report on their environment without disturbing it.[5,6] At the same time, low spin-field interaction energy dictates overall poor sensitivity of magnetic resonance techniques. Given typical P values of 0.001-0.0001%, only relatively large concentrations of spins (>1 mM) can be measured with sufficient signal-to-noise ratio (SNR) in a conceivable amount of time.[7–9] This sets stringent limits on applications of NMR/MRI and constrains studies to large sample volumes as compared to other analytical methods.

Hyperpolarization refers to situations in which P can be much higher than a thermal equilibrium value, for example, >10%.[10,11] By enhancing polarization levels through hyperpolarization techniques (see below) NMR/MRI can achieve sensitivity to enable detection of

low-concentration samples (e.g., $<1 \mu\text{M}$) with high SNR. As an example of hyperpolarization, see **Figure 1**. At the top, an ^{15}N NMR spectrum of a fully labeled ^{15}N -pyridine as a neat liquid was measured at the magnetic field of 9.4 tesla. Since molecules are isotopically enriched with ^{15}N nuclei ($>99\%$) and present at high concentration ($\sim 12.4 \text{ M}$), a sufficient SNR is obtained in a single acquisition. For comparison, a sample of 50 mM metronidazole—a well-known antibiotic and hypoxia probe—at natural isotopic abundance of ^{15}N (0.35%) gives a strong ^{15}N NMR signal after being flushed with parahydrogen ($p\text{H}_2$) gas for ~ 30 seconds (Figure 1, bottom). For the same sample at thermal equilibrium to give an NMR signal with SNR comparable to the neat, ^{15}N -labeled pyridine, about 30 years of continuous signal averaging would be necessary. This example demonstrates the power of hyperpolarization, in this case, SABRE technique (SABRE = signal amplification by reversible exchange): molecules at low concentration and natural isotopic abundance can be detected with sufficient SNR on a time scale of seconds.[12]

This *Editorial Essay* briefly explores applications of hyperpolarization techniques in both medical diagnostics and emerging spin technologies. Current landscape of methods is examined with a particular focus on medical applications. Hyperpolarization-enhanced MRI is compared to positron-emission tomography (PET) and the promises in molecular imaging and disease monitoring are critically assessed. I also explore novel quantum sensing modalities empowered by spin hyperpolarization in biomedical research and beyond. Through this analysis, I hope to highlight promising research directions related to spin technologies which may refine our understanding of key (bio)molecular processes.

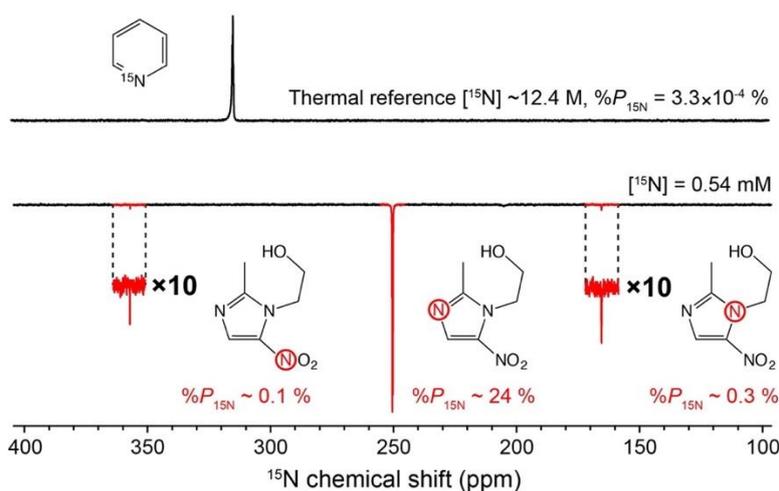


Figure 1. Example of NMR hyperpolarization. (Top) A typical ^{15}N NMR spectrum of ^{15}N -pyridine ($\sim 99\%$ ^{15}N labeling) recorded at 9.4 tesla. (Bottom) SABRE-enhanced ^{15}N NMR spectrum of metronidazole at 150 mM (^{15}N nuclei at natural isotopic abundance of 0.35%) after 30 seconds of parahydrogen bubbling (50% $p\text{H}_2$ enrichment fraction). Nitrogen-15 in the $-\text{NO}_2$ group is demonstrated to have T_1 of ~ 15 mins. Reproduced with permission from Ref. [13].

2. Hyperpolarization Techniques

Given the wealth of existing literature on the topic of nuclear and electron hyperpolarization, here I refrain from delving into the foundational principles of these techniques. However, it is important to note that hyperpolarization technologies broadly fall into two categories, although some methods may formally qualify to both groups:

(1) Techniques Utilizing Electromagnetic Fields

A significant fraction of hyperpolarization methods uses electromagnetic fields. Notably, in dynamic nuclear polarization (DNP) polarization transfer from unpaired electrons to spin-active nuclei is facilitated by the application of microwaves.[14–19] DNP allows generating strong NMR signals for molecules increased by orders of magnitude compared to thermal equilibrium, resulting

in dramatically decreased signal averaging times. DNP encompasses various methodologies such as Overhauser-DNP, dissolution-DNP, as well as emerging approaches like bullet-DNP.[19–21] Optical pumping techniques, involving visible or infrared radiation, play a major role in hyperpolarization of both electron and nuclear spins. Spin-exchange optical pumping (SEOP) and metastability-exchange optical pumping (MEOP) of noble gases[22] exemplify this group of methods along with optical pumping of defects in solids like NV-centers in diamond.[23–25] Visible light applied for generating hyperpolarization is a signature of chemically induced dynamic nuclear polarization (CIDNP) and related approaches[5,26,27].

In principle, pumping with electromagnetic radiation allows polarizing molecules in all phases of ordinary matter (gas, liquid, solid, and even plasma[28]). Direct pumping of nuclear magnetization with light seems to be possible in the gas or solid via optical pumping while hyperpolarization of molecules in solution necessitates more complex interactions.

(2) Techniques Utilizing Chemistry and Spin Statistics

Another subset of hyperpolarization techniques relies on intricate spin statistics facilitating the generation of hyperpolarized states. Chemical reactions and chemical exchange, notably in parahydrogen-induced polarization (PHIP), underpin these methodologies. PHIP variants like PASADENA, ALTADENA and SABRE demonstrate remarkable polarization levels (up to 50%) on various nuclei.[14,29] While challenges remain in clinical translation, specifically, the ability to control all stages of chemical transformations and fields at each moment of sample transfer, recent advancements have showcased reproducible polarization levels on biologically relevant nuclei, fostering optimism for future developments.[30–32]

For a comprehensive exploration of hyperpolarization, readers are encouraged to consult recent reviews that delve into the physicochemical principles of these techniques.[33] The semantic breadth of “hyperpolarization” terminology highlights its diverse manifestations which extend beyond simple magnetization to encompass complex spin orders with broad implications for both fundamental research and technological innovation.[10,34,35]

3. Medical Applications of Hyperpolarization

As of 2024, biomedical science remains a major driver for the hyperpolarization research if accounted by the number of peer-reviewed publication devoted to this subject in recent years[36–38]. The interest is not surprising due to immense applicability of MRI in medical diagnostics even without using hyperpolarization. While there are no yet clear avenues for generating hyperpolarization inside a living object without bringing hyperpolarized molecules from the outside (exogenous injections do not make hyperpolarization-enhanced MRI fully non-invasive), the existing alternative clinical approaches to monitor metabolism involve radioactive samples and, thus, MRI is freed from this complication. Coupled with the ability of selecting specific regions in the object under study and harnessing information from heteronuclei (e.g., ^{13}C and ^{15}N) hyperpolarization-enhanced MRI provides a novel toolkit for understanding chemical composition and functions of tissue, disease progression, and treatment[39–41].

Conceptually, in the context of *molecular imaging* (i.e., imaging of specific molecules and their transformations rather than imaging of bulk medium), hyperpolarization-enhanced MRI share similarities with positron-emission tomography (PET) and it is worth delving deeper into the comparative analysis of these two modalities. Both technologies in its current implementation require injection of exogenous contrast agents bearing a signal-generating nuclear isotope.

Comparison of PET and Hyperpolarization-Enhanced MRI

In PET, a radioactive agent is injected into the patient (ideally) immediately after its production. The radioactive decay typically happens on a timescale of minutes ($t_{1/2}$ ~110 mins for ^{18}F nuclei) generating positrons that annihilate with the nearby matter; measured signals are derived from the detection of γ -photons emitted upon this annihilation.[42] While PET offers high sensitivity for observing metabolic activity, its resolution is limited to 3-5 mm.[43] In hyperpolarization-enhanced

MRI, a hyperpolarized exogenous contrast agent (with polarization typically “stored” in the magnetization of heteronuclei such as ^{13}C) has a short in vivo lifetime providing a time-window of, at best, up to 5 min after injection. This can be used for angiography and perfusion (Figure 2) but is often not sufficient for monitoring metabolic processes of interest.[44] A time-window of at least a few hours would be more appropriate for studying unknown details of the Krebs cycle (such as its reversibility) and other metabolic transformations.[45] However, unlike PET, MRI faces no fundamental resolution limitations, with bottlenecks being practical, e.g., available SNR per voxel, ability to provide large field gradients in short time intervals etc. Typical resolution of conventional proton MRI is about 1 mm and sub-100- μm -resolution microimaging has been demonstrated with hyperpolarization.[46]

Despite these differences, both hyperpolarization-enhanced MRI and PET can visualize specific metabolic pathways by using tracer molecules and appropriate image reconstruction techniques. In MRI, ^{13}C -pyruvate is one of the most promising and well-developed hyperpolarized contrast agents for observing metabolism within the Krebs cycle[38] while PET-agents ^{18}F -FDG (fluorodeoxyglucose) and ^{13}N -ammonia are routinely used clinically (and numerous other agents have been tried in research[47]).

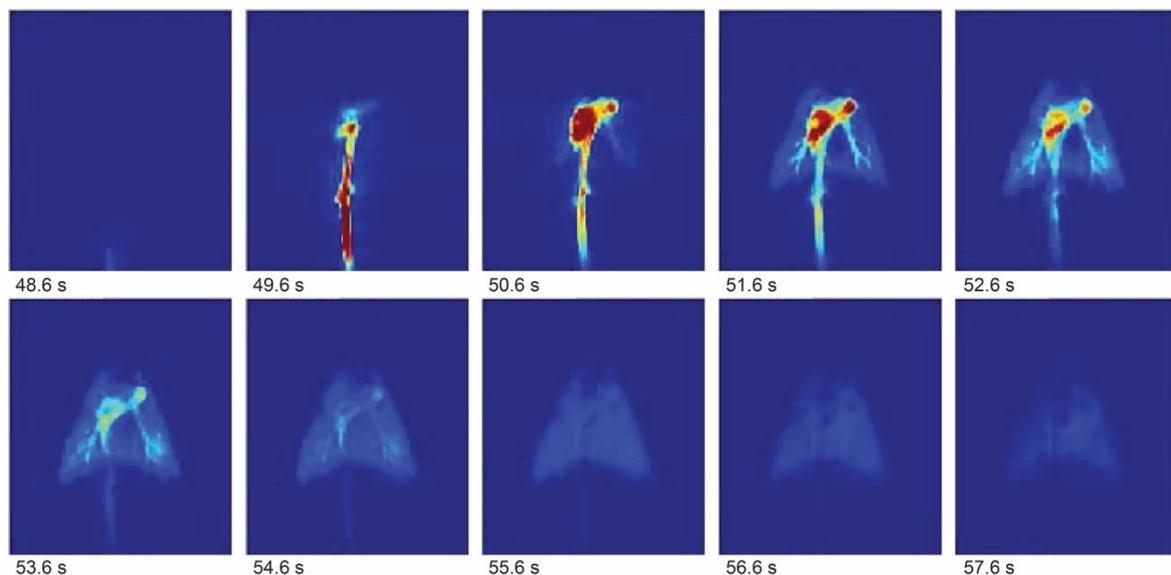


Figure 2. Hyperpolarization-enhanced ^{13}C magnetic resonance images showing lungs of a pig after injection of a hyperpolarized ^{13}C -2-hydroxyethylacrylate with 1 s time resolution. Adapted with permission from Ref. [48].

It is interesting to note that both PET and hyperpolarization-enhanced heteronuclear MRI exploit the lack of intrinsic background signal to observe molecular processes without interference. PET has no background signal owing to the absence radioactive positron-emitting nuclei in the body as well as quiet gamma-ray background in the environment. Similarly, signals from naturally abundant thermally polarized heteronuclei such as ^{13}C or ^{15}N are virtually absent in MRI. Monitoring metabolic changes by conventional ^1H MRI, on the other hand, is challenging due to the large background signal originating from thermally polarized protons in H_2O and lipids. Stargazing provides good analogy: observation of stars from inside a megapolis is challenging because of optical pollution; one would need to go to the mountains or far in the wilderness (where the background light is absent) to notice myriads of stars with a bear eye.

Developing endogenous hyperpolarized contrast agents generated on demand (or naturally produced) inside the object that is being investigated seems highly desirable. Green fluorescent protein (GFP) in combination with optical detection serves as an inspiration: generation of GFP is possible in various environments via genetic manipulations.[49] In the case of MRI, approaches of generating genetically encoded signal contrast in vivo have been proposed based on the use of

hyperpolarized ^{129}Xe gas.[50,51] Parahydrogen is another option since it offers a unique possibility of bringing latent nuclear spin order inside the object to be studied in such a way that magnetization is generated only in vivo and on demand (Figure 3).[52] While typically information encoding and signal detection are inseparable parts of the measurement, MRI fundamentally permits separating these two steps in time and/or space. Further interdisciplinary innovation is likely necessary to unlock opportunities provided by genetically-encoded hyperpolarized MRI sensors.[53]

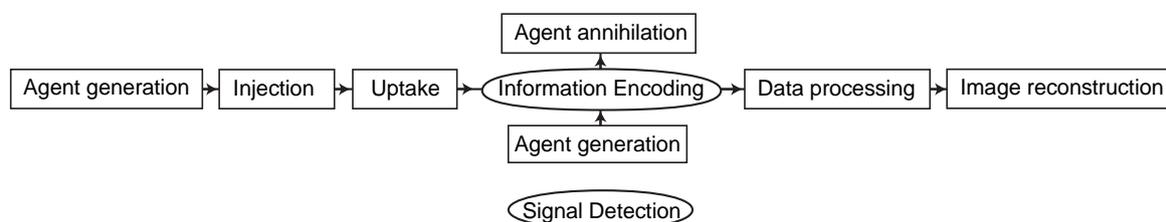


Figure 3. Stages of molecular imaging using exogenous and endogenous (dashed line) contrast agents.

Challenges of Hyperpolarized Molecular MRI.

Despite its immense potential, the widespread clinical adoption of hyperpolarization-enhanced MRI faces significant constraints. These limitations primarily stem from the prevalence of hardware optimized for detecting protons (^1H) and the absence of refined pulse sequences for effective polarization transfer, crucial for improving the signal-to-noise ratio (SNR) of heteronuclear signals. Additionally, hyperpolarization-enhanced MRI necessitate interdisciplinary working group and requires advanced infrastructure.[54]

The first proposals for using hyperpolarized contrast agents emerged in the early 1990s, but the steady stream of research publications has (up to date) not been sufficient to convince practicing physicians in their utility. As of 2024, the number of hospitals in the world equipped with the necessary devices and expertise to observe metabolic transformations using hyperpolarization-enhanced ^{13}C MRI remains fewer than 20.[54] While the principles of *d*DNP methodology are known since 2003,[19] the anticipated widespread clinical application of this method has not been materialized, despite advancements in other research areas driven by the Moore's Law.[55] Polarization levels are not universally high even for *d*DNP and can vary depending on the specific preparation method employed and are extremely technically challenging to maintain. But potentially the biggest drawback of the existing modality is short lifetime of hyperpolarized molecules in vivo—particularly concerning the most interesting molecules like pyruvate (T_1 of carbon-13 at 3 T is only ~30 s in vivo [56])—limiting applications to tissues with high cellularity and rapid transfer through cell membranes. It is essential for the research community to maintain a balanced perspective on this emerging technology since even niche applications without revolutionary clinical impact can still be valuable.

In summary, hyperpolarization-enhanced MRI is a molecular imaging modality offering sensitivity and resolution comparable to PET. However, resolving challenges related to polarization lifetime is critical for successful clinical adaptation. As hyperpolarization technology matures, the prospect of MRI with heteronuclear detection becoming commonplace holds promise for advancing our understanding of metabolic changes in both research and clinical contexts. Moreover, the development of joint modalities combining the sensitivity of PET with the resolution of MRI could further enhance diagnostic capabilities signaling exciting prospects toward future developments.

4. Emerging Spin Technologies

In today's world, appreciation of the technology may often outweigh the appreciation for the research that underpins it. Yet, it is crucial to recognize that without ongoing scientific exploration technological innovation would likely stagnate. This is particularly evident in the realm of spin technologies, where the fundamental quantum nature of spins opens doors to a plethora of

applications across diverse fields.[57] From enhancing chemical reactions dependent on nuclear spins to the development of quantum sensors utilizing single defects in crystal lattices, the breadth of potential applications is vast.[58]

In the context of MRI, quantum phenomena such as entanglement and long-lived spin states offer avenues for extending polarization lifetimes, thus enhancing imaging capabilities.[59] PHIP, SABRE, and, in general, magnetization transfer catalysis (MTC) demonstrate how transient molecular interactions can be leveraged to amplify spin signals, enabling novel detection schemes. In addition to hyperpolarization, principles of quantum metrology hold potential for enabling precise differentiation of chemicals and their transformations through high-resolution analysis of spectral frequencies and phases.[57] By leveraging key quantum concepts like squeezing and entanglement, spin techniques could improve MRI by achieving unprecedented resolution.

5. Conclusion

In the landscape of hyperpolarization-enhanced NMR/MRI, challenges and opportunities lie ahead. Key questions persist: will nuclear hyperpolarization unveil novel, previously unknown, dimensions of metabolism? How can spin order be efficiently preserved in molecules within biochemical processes over extended timeframes beyond a few minutes? Will innovative NMR detection methods, bolstered by hyperpolarization, transition to practical clinical applications? Could portable point-of-care NMR devices revolutionize healthcare diagnostics?

These questions not only underscore the ongoing evolution of hyperpolarization techniques but also point to potential avenues for future research and technological advancement. It is already evident that hyperpolarization represents a promising trajectory – one that complements established high-field NMR/MRI modalities – in our journey of improving magnetic resonance methods. The rallying cry remains 'Molecules, up your spins!' and with each discovery we shape the future of *truly quantum* molecular imaging.

Funding: The work is supported by Alexander von Humboldt Foundation in the framework of the Sofja Kovalevskaja Award.

Acknowledgments: The author thanks Prof. Dmitry Budker, Dr. Andrey Pravdivtsev, Dr. Sheng Chi Fan, Prof. Alexander Pines, Prof. Thomas Budinger, and Prof. Kev Salikhov for stimulating discussions.

Conflicts of Interest: The authors declare no conflict of interest.

Sample Availability: Samples of the compounds are available from the authors.

References

1. Nagornov, K. O.; Gorshkov, M. V.; Kozhinov, A. N.; Tsybin, Y. O., High-resolution Fourier transform ion cyclotron resonance mass spectrometry with increased throughput for biomolecular analysis. *Anal. Chem.* **2014**, *86* (18), 9020-9028.
2. Vasconcelos Soares Maciel, E.; de Toffoli, A. L.; Sobieski, E.; Domingues Nazário, C. E.; Lanças, F. M., Miniaturized liquid chromatography focusing on analytical columns and mass spectrometry: A review. *Anal. Chim. Acta* **2020**, *1103*, 11-31.
3. Huffman, R. G.; Leduc, A.; Wichmann, C.; Di Gioia, M.; Borriello, F.; Specht, H.; Derks, J.; Khan, S.; Khoury, L.; Emmott, E.; Petelski, A. A.; Perlman, D. H.; Cox, J.; Zanoni, I.; Slavov, N., Prioritized mass spectrometry increases the depth, sensitivity and data completeness of single-cell proteomics. *Nat. Methods* **2023**, *20* (5), 714-722.
4. Abragam, A., *The principles of nuclear magnetism*. Clarendon Press: Oxford, 1961.
5. Salikhov, K. M.; Molin, Y. N.; Sagdeev, R. Z.; Buchachenko, A. L., *Spin polarization and magnetic effects in chemical reactions*. Elsevier: Amsterdam, 1984.
6. Fisher, M. P. A.; Radzihovsky, L., Quantum indistinguishability in chemical reactions. *Proceedings of the National Academy of Sciences* **2018**, *115* (20), E4551-E4558.
7. Roose, B. W.; Zemerov, S. D.; Dmochowski, I. J., Nanomolar small-molecule detection using a genetically encoded (129)Xe NMR contrast agent. *Chem Sci* **2017**, *8* (11), 7631-7636.
8. Wen, L.; Meng, H.; Gu, S.; Wu, J.; Zhao, Y., Toward Nanomolar Multi-Component Analysis by 19F NMR. *Anal. Chem.* **2022**, *94* (22), 8024-8032.

9. Eshuis, N.; Hermkens, N.; van Weerdenburg, B. J. A.; Feiters, M. C.; Rutjes, F. P. J. T.; Wijmenga, S. S.; Tessari, M., Toward Nanomolar Detection by NMR Through SABRE Hyperpolarization. *J. Am. Chem. Soc.* **2014**, *136* (7), 2695-2698.
10. Levitt, M. H.; Bengs, C., Hyperpolarization and the physical boundary of Liouville space. *Magn. Reson.* **2021**, *2* (1), 395-407.
11. Barskiy, D. A.; Kovtunov, K. V.; Koptuyug, I. V.; He, P.; Groome, K. A.; Best, Q. A.; Shi, F.; Goodson, B. M.; Shchepin, R. V.; Truong, M. L.; Coffey, A. M.; Waddell, K. W.; Chekmenev, E. Y., In Situ and Ex Situ Low-Field NMR Spectroscopy and MRI Endowed by SABRE Hyperpolarization. *ChemPhysChem* **2014**, *15* (18), 4100-4107.
12. Kircher, R.; Xu, J.; Barskiy, D. A., In Situ Hyperpolarization Enables ¹⁵N and ¹³C Benchtop NMR at Natural Isotopic Abundance. *J. Am. Chem. Soc.* **2024**, *146* (1), 514-520.
13. Barskiy, D. A.; Shchepin, R. V.; Coffey, A. M.; Theis, T.; Warren, W. S.; Goodson, B. M.; Chekmenev, E. Y., Over 20% N-15 Hyperpolarization in Under One Minute for Metronidazole, an Antibiotic and Hypoxia Probe. *J. Am. Chem. Soc.* **2016**, *138* (26), 8080-8083.
14. Kirill, K.; Ekaterina, P.; Oleg, S.; Samuel, C.; Dennis, K.; Basile, V.; Sami, J.; Eduard, C.; Boyd, G.; Danila, B.; Igor, K., Hyperpolarized NMR: d-DNP, PHIP, and SABRE. *Chemistry – An Asian Journal* **2018**.
15. Ardenkjaer-Larsen, J. H., On the present and future of dissolution-DNP. *J. Magn. Reson.* **2016**, *264*, 3-12.
16. Jähnig, F.; Kwiatkowski, G.; Ernst, M., Conceptual and instrumental progress in dissolution DNP. *J. Magn. Reson.* **2016**, *264*, 22-29.
17. Buratto, R.; Bornet, A.; Milani, J.; Mammoli, D.; Vuichoud, B.; Salvi, N.; Singh, M.; Laguerre, A.; Passemard, S.; Gerber-Lemaire, S.; Jannin, S.; Bodenhausen, G., Drug Screening Boosted by Hyperpolarized Long-Lived States in NMR. *ChemMedChem* **2014**, *9* (11), 2509-2515.
18. Comment, A.; Rentsch, J.; Kurdzesau, F.; Jannin, S.; Uffmann, K.; van Heeswijk, R. B.; Hautle, P.; Konter, J. A.; van den Brandt, B.; van der Klink, J. J., Producing over 100ml of highly concentrated hyperpolarized solution by means of dissolution DNP. *J. Magn. Reson.* **2008**, *194* (1), 152-155.
19. Ardenkjaer-Larsen, J. H.; Fridlund, B.; Gram, A.; Hansson, G.; Hansson, L.; Lerche, M. H.; Servin, R.; Thaning, M.; Golman, K., Increase in signal-to-noise ratio of > 10,000 times in liquid-state NMR. *Proc. Natl. Acad. Sci. U. S. A.* **2003**, *100* (18), 10158-10163.
20. Kouřil, K.; Kouřilová, H.; Bartram, S.; Levitt, M. H.; Meier, B., Scalable dissolution-dynamic nuclear polarization with rapid transfer of a polarized solid. *Nat. Commun.* **2019**, *10* (1), 1733.
21. Abragam, A., *The principles of nuclear magnetism*. Oxford university press: 1961.
22. Barskiy, D. A.; Coffey, A. M.; Nikolaou, P.; Mikhaylov, D. M.; Goodson, B. M.; Branca, R. T.; Lu, G. J.; Shapiro, M. G.; Telkki, V. V.; Zhivonitko, V. V.; Koptuyug, I. V.; Salnikov, O. G.; Kovtunov, K. V.; Bukhtiyarov, V. I.; Rosen, M. S.; Barlow, M. J.; Safavi, S.; Hall, I. P.; Schroder, L.; Chekmenev, E. Y., NMR Hyperpolarization Techniques of Gases. *Chem. Eur. J.* **2017**, *23* (4).
23. Kehayias, P.; Jarmola, A.; Mosavian, N.; Fescenko, I.; Benito, F. M.; Laraoui, A.; Smits, J.; Bougas, L.; Budker, D.; Neumann, A.; Brueck, S. R. J.; Acosta, V. M., Solution nuclear magnetic resonance spectroscopy on a nanostructured diamond chip. *Nat. Commun.* **2017**, *8* (1), 188.
24. Budker, D.; Barskiy, D.; Lenz, T., Kernspinresonanz ohne Magnetfeld. *Physik in unserer Zeit* **2023**, *54* (6), 294-301.
25. King, J. P.; Jeong, K.; Vassiliou, C. C.; Shin, C. S.; Page, R. H.; Avalos, C. E.; Wang, H.-J.; Pines, A., Room-temperature in situ nuclear spin hyperpolarization from optically pumped nitrogen vacancy centres in diamond. *Nat. Commun.* **2015**, *6* (1), 8965.
26. Goetz, M., Photo-CIDNP spectroscopy. *Annu. Rep. NMR Spectrosc.* **2009**, *66*, 77-147.
27. Morozova, O. B.; Ivanov, K. L., Time-Resolved Chemically Induced Dynamic Nuclear Polarization of Biologically Important Molecules. *ChemPhysChem* **2019**, *20* (2), 197-215.
28. Maul, A.; Blümler, P.; Nacher, P. J.; Otten, E.; Tastevin, G.; Heil, W., Nuclear hyperpolarization of ³He by magnetized plasmas. *Phys. Rev. A* **2018**, *98* (6), 063405.
29. Rayner, P. J.; Burns, M. J.; Oлару, A. M.; Norcott, P.; Fekete, M.; Green, G. G. R.; Highton, L. A. R.; Mewis, R. E.; Duckett, S. B., Delivering strong ¹H nuclear hyperpolarization levels and long magnetic lifetimes through signal amplification by reversible exchange. *Proc. Natl. Acad. Sci. U. S. A.* **2017**, *114* (16), E3188-E3194.
30. Knecht, S.; Blanchard, J. W.; Barskiy, D.; Cavallari, E.; Dagys, L.; Van Dyke, E.; Tsukanov, M.; Bliemel, B.; Münnemann, K.; Aime, S.; Reineri, F.; Levitt, M. H.; Buntkowsky, G.; Pines, A.; Blümler, P.; Budker, D.; Eills, J., Rapid hyperpolarization and purification of the metabolite fumarate in aqueous solution. *Proceedings of the National Academy of Sciences* **2021**, *118* (13), e2025383118.

31. Ellermann, F.; Sirbu, A.; Brahm, A.; Assaf, C.; Herges, R.; Hövener, J.-B.; Pravdivtsev, A. N., Spying on parahydrogen-induced polarization transfer using a half-tesla benchtop MRI and hyperpolarized imaging enabled by automation. *Nat. Commun.* **2023**, *14* (1), 4774.
32. de Maissin, H.; Groß, P. R.; Mohiuddin, O.; Weigt, M.; Nagel, L.; Herzog, M.; Wang, Z.; Willing, R.; Reichardt, W.; Pichotka, M.; Heß, L.; Reinheckel, T.; Jessen, H. J.; Zeiser, R.; Bock, M.; von Elverfeldt, D.; Zaitsev, M.; Korchak, S.; Glöggler, S.; Hövener, J.-B.; Chekmenev, E. Y.; Schilling, F.; Knecht, S.; Schmidt, A. B., In Vivo Metabolic Imaging of [1-13C]Pyruvate-d3 Hyperpolarized By Reversible Exchange With Parahydrogen**. *Angewandte Chemie International Edition* **2023**, *62* (36), e202306654.
33. Eills, J.; Budker, D.; Cavagnero, S.; Chekmenev, E. Y.; Elliott, S. J.; Jannin, S.; Lesage, A.; Matysik, J. r.; Meersmann, T.; Prisner, T., Spin hyperpolarization in modern magnetic resonance. *Chem. Rev.* **2023**, *123* (4), 1417-1551.
34. Xu, J.; Budker, D.; Barskiy, D. A., Visualization of dynamics in coupled multi-spin systems. *Magn. Reson.* **2022**, *3* (2), 145-160.
35. Barskiy, D. A.; Pravdivtsev, A., Magnetization and Polarization of Coupled Nuclear Spin Ensembles. *arXiv preprint arXiv:2308.15837* **2023**.
36. Nikolaou, P.; Goodson, B. M.; Chekmenev, E. Y., NMR Hyperpolarization Techniques for Biomedicine. *Chem. Eur. J.* **2015**, *21* (8), 3156-3166.
37. Meier, S.; Jensen, P. R.; Karlsson, M.; Lerche, M. H., Hyperpolarized NMR Probes for Biological Assays. *Sensors* **2014**, *14* (1), 1576-1597.
38. Kurhanewicz, J.; Vigneron, D. B.; Brindle, K.; Chekmenev, E. Y.; Comment, A.; Cunningham, C. H.; DeBerardinis, R. J.; Green, G. G.; Leach, M. O.; Rajan, S. S.; Rizi, R. R.; Ross, B. D.; Warren, W. S.; Malloy, C. R., Analysis of Cancer Metabolism by Imaging Hyperpolarized Nuclei: Prospects for Translation to Clinical Research. *Neoplasia* **2011**, *13* (2), 81-97.
39. Nagana Gowda, G. A.; Raftery, D., NMR Metabolomics Methods for Investigating Disease. *Anal. Chem.* **2023**, *95* (1), 83-99.
40. Shchepin, R. V.; Birchall, J. R.; Chukanov, N. V.; Kovtunov, K. V.; Koptuyug, I. V.; Theis, T.; Warren, W. S.; Gelovani, J. G.; Goodson, B. M.; Shokouhi, S.; Rosen, M. S.; Yen, Y.-F.; Pham, W.; Chekmenev, E. Y., Hyperpolarizing Concentrated Metronidazole 15NO2 Group over Six Chemical Bonds with More than 15 % Polarization and a 20 Minute Lifetime. *Chemistry – A European Journal* **2019**, *25* (37), 8829-8836.
41. Svyatova, A.; Skovpin, I. V.; Chukanov, N. V.; Kovtunov, K. V.; Chekmenev, E. Y.; Pravdivtsev, A. N.; Hövener, J. B.; Koptuyug, I. V., 15N MRI of SLIC-SABRE hyperpolarized 15N-labelled pyridine and nicotinamide. *Chemistry–A European Journal* **2019**, *25* (36), 8465-8470.
42. Budinger, T. F.; Van Brocklin, H. F., *Positron emission tomography*. CRC Press inc.–1995: 1985.
43. Moses, W. W., Fundamental Limits of Spatial Resolution in PET. *Nucl Instrum Methods Phys Res A* **2011**, *648* Supplement 1, S236-s240.
44. Angelovski, G., What We Can Really Do with Bioresponsive MRI Contrast Agents. *Angewandte Chemie International Edition* **2016**, *55* (25), 7038-7046.
45. Lane, N., *Transformer: The Deep Chemistry of Life and Death*. Profile: 2022.
46. Coffey, A. M.; Kovtunov, K. V.; Barskiy, D. A.; Koptuyug, I. V.; Shchepin, R. V.; Waddell, K. W.; He, P.; Groome, K. A.; Best, Q. A.; Shi, F.; Goodson, B. M.; Chekmenev, E. Y., High-Resolution Low-Field Molecular Magnetic Resonance Imaging of Hyperpolarized Liquids. *Anal. Chem.* **2014**, *86* (18), 9042-9049.
47. Zhu, L.; Ploessl, K.; Kung, H. F., PET/SPECT imaging agents for neurodegenerative diseases. *Chem. Soc. Rev.* **2014**, *43* (19), 6683-6691.
48. Golman, K.; Olsson, L. E.; Axelsson, O.; Mansson, S.; Karlsson, M.; Petersson, J. S., Molecular imaging using hyperpolarized 13C. *British Journal of Radiology* **2003**, *76* (Spec. Iss. 2), S118-S127.
49. Zimmer, M., Green fluorescent protein (GFP): applications, structure, and related photophysical behavior. *Chem. Rev.* **2002**, *102* (3), 759-782.
50. Shapiro, M. G.; Ramirez, R. M.; Sperling, L. J.; Sun, G.; Sun, J.; Pines, A.; Schaffer, D. V.; Bajaj, V. S., Genetically encoded reporters for hyperpolarized xenon magnetic resonance imaging. *Nature Chem.* **2014**, *6* (7), 629-634.
51. Jayapaul, J.; Schröder, L., Molecular Sensing with Host Systems for Hyperpolarized 129Xe. *Molecules* **2020**, *25* (20), 4627.
52. Hövener, J. B.; Schwaderlapp, N.; Lickert, T.; Duckett, S. B.; Mewis, R. E.; Highton, L. A. R.; Kenny, S. M.; Green, G. G. R.; Leibfritz, D.; Korvink, J. G.; Hennig, J.; von Elverfeldt, D., A hyperpolarized equilibrium for magnetic resonance. *Nat. Commun.* **2013**, *4*.
53. Bricco, A. R.; Miralavy, I.; Bo, S.; Perlman, O.; Korenchan, D. E.; Farrar, C. T.; McMahon, M. T.; Banzhaf, W.; Gilad, A. A., A Genetic Programming Approach to Engineering MRI Reporter Genes. *ACS Synthetic Biology* **2023**, *12* (4), 1154-1163.

54. Chaumeil, M. M.; Bankson, J. A.; Brindle, K. M.; Epstein, S.; Gallagher, F. A.; Grashei, M.; Guglielmetti, C.; Kaggie, J. D.; Keshari, K. R.; Knecht, S.; Laustsen, C.; Schmidt, A. B.; Vigneron, D.; Yen, Y.-F.; Schilling, F., New Horizons in Hyperpolarized ¹³C MRI. *Molecular Imaging and Biology* **2023**.
55. Pearl, R., Breaking The Rules Of Healthcare: Selecting The Best Technology. *Forbes* 2022.
56. Nelson, S. J.; Kurhanewicz, J.; Vigneron, D. B.; Larson, P. E.; Harzstark, A. L.; Ferrone, M.; van Criekinge, M.; Chang, J. W.; Bok, R.; Park, I., Metabolic imaging of patients with prostate cancer using hyperpolarized [1-¹³C] pyruvate. *Science translational medicine* **2013**, *5* (198), 198ra108-198ra108.
57. Pezze, L.; Smerzi, A.; Oberthaler, M. K.; Schmied, R.; Treutlein, P., Quantum metrology with nonclassical states of atomic ensembles. *Rev. Mod. Phys.* **2018**, *90* (3), 035005.
58. Budker, D., Extreme nuclear magnetic resonance: Zero field, single spins, dark matter.... *J. Magn. Reson.* **2019**.
59. Sonnefeld, A.; Razanahoera, A.; Pelupessy, P.; Bodenhausen, G.; Sheberstov, K., Long-lived states of methylene protons in achiral molecules. *Science Advances* **2022**, *8* (48), eade2113.

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