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Article

Synergistic Effect of Postbiotic Yeast ABB C22® on Gut Inflammation, Barrier Function and Protection from Rotavirus Infection

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Abstract: Diarrhoea is a serious cause of mortality worldwide that can lead to dehydration, gut barrier function impairment, nutrient malabsorption, and alterations of the gut microbiota (dysbiosis). Current solutions for its management, such as oral rehydration salts (ORS), inhibitors of the gut motility, antibiotics, and living probiotics only partially counteract the mechanisms of the disease and do not provide a full coverage of the problem. Potential risks of the use of living probiotic strains, particularly in immunocompromised patients, can be eliminated with the use of tyndallized (heat-killed) postbiotic bacteria and yeast. ABB C22® is a postbiotic combination of three tyndallized yeasts, namely *Saccharomyces boulardii*, *Saccharomyces cerevisiae*, and *Kluyveromyces marxianus*. To assess the action of the postbiotic combination on diarrhoea, immune and gut epithelial cell signaling assays, gut barrier formation assay and rotavirus gene expression assay were performed. ABB C22® showed a strong anti-inflammatory effect, an induction of the build-up of the gut epithelium, and a degree of protection against rotavirus infection. These experimental studies support the use of the postbiotic ABB C22® as a solution for the management of diarrhoea and gastrointestinal conditions, alone or in combination with existing but incomplete treatments.

Keywords: diarrhoea; postbiotic; yeast; inflammation; gut protection; Rotavirus

1. Introduction

Diarrhoeal disease is a major social concern, especially in developing countries. It can affect adults and children, but the youngest are particularly vulnerable to its harsh effects and mortality. Diarrhoea accounts for 1 in 9 children's deaths worldwide, making it the second leading cause of mortality among children less than 5 years of age [1,2]. Data of the Centres for Disease Control (CDC) indicate that diarrhoea kills 2,195 children every day, more than AIDS, malaria and measles combined [2]. Diarrhoea is both preventable and treatable [1]. Acute diarrhoeal episodes are typically caused by infections, mainly rotavirus [3]. Chronic diarrhoea can have diverse causes, such as food allergies and intolerances, infections, inflammatory bowel disease (IBD) and functional gastrointestinal disorders [4,5]. The pathogenesis of diarrhoea is heterogenous, and may involve decreased water and electrolyte absorption, increased water secretion by the intestinal mucosa, increased luminal osmotic load and/or inflammation of the mucosa and increased permeability. Independently of the aetiology of diarrhoea, a strong alteration of the gut epithelial barrier and an acceleration of the intestinal transit lead to severe complications, such as dehydration [6], gut barrier function impairment with risk of septicaemia, nutrient malabsorption, and disturbances and loss of the gut microbial communities (dysbiosis) [7,8]. Diarrhoea is a recognized cause of dysbiosis, leading to changes in the diversity of gut bacterial and fungal (mycobiota) populations [9]. Healthy microbiota and mycobiota lead to gut homeostasis and drive a strong gut barrier function. Microbiota

and mycobiota dysbiosis compromise this equilibrium and drive a pro-inflammatory status in the lumen that leads to an altered immune response and to the disruption of the gut epithelium. This increases the susceptibility to pathogens favouring diarrhoea, and their toxins further affect gut microbiota disorders [8,10].

Current solutions for the management of diarrhoea, such as oral rehydration salts (ORS), inhibitors of the gut motility, antibiotics, and living probiotics only partially counteract the pathogenic mechanisms of the disease, and do not provide a full coverage of the problem [1,11–17]. Antibiotics eliminate bacterial-associated gastrointestinal infection causing diarrhoea but are not effective in case of resistant bacteria, infections caused by viruses or parasites, or in case of underlying diseases or food intolerance [18]. Moreover, antibiotics may cause antibiotic-associated diarrhoea due to dysbiosis [19]. Its use is a major disruptor of gut microbiota with negative effects reducing species diversity and allowing overgrowth of pathogens (i.e., *Clostridium difficile*) [19]. The indiscriminate use of antibiotics for the management of diarrhoea not only increases antibiotic resistance but impairs the homeostasis of the gut microbiome and may result in worsening diarrhoeal conditions or even to the establishment of chronic transit disturbances due to gut dysbiosis [20]. In this sense, there is an increasing focus on the development of nutritional supplements that show effects, help reduce the use of antibiotics and, at the same time, are safe and suitable for all populations.

Microbial interventions such as faecal microbiota transplantation (FMT) and the use of prebiotics, probiotics, and postbiotics are gaining focus as strategies to regulate the composition of the intestinal flora for the prevention and treatment of diarrhoeal episodes. The anti-diarrhoeal mechanism of probiotics mainly relies on regulation and improvement of the balance of gut microbiota, production of antibacterial compounds and improvement of intestinal defence barrier and immunity [10,21].

More than 50 clinical trials (more than 8,000 subjects involved in total) have been conducted to date regarding the efficacy of probiotic yeasts. Results point to a major advantage over bacterial probiotics in the prevention and treatment of many gastrointestinal conditions [22]. Cell wall components in yeasts, such as β -glucans, act as antigens recognized by receptors in the host's immune cells and can therefore exert immunomodulating effects. Although yeast represent only a minority of the organisms composing the intestinal flora, their cell size is 10 times larger than that of bacteria and they could represent a significant steric hindrance for bacteria. In addition, bacterial probiotics only modulate the bacterial microbiota but do not influence the gut mycobiota, which has a relevant role in maintaining gut homeostasis. Furthermore, yeasts are not affected by antibiotics, so yeast can be co-administered to exert a concomitant/immediate effect without compromising the efficacy or safety of the antibiotic treatment. Finally, yeast probiotics have better withstanding of the extreme environments of the stomach.

However, safety of the use of live probiotics has been a matter of concern. Risks associated with the use of live microorganisms have been reported, including systemic infection (septicaemia) caused by translocation, especially in paediatric populations and vulnerable patients, acquisition of antimicrobial resistant genes and interference with gut colonization in neonates [23]. Potential risks of the use of living probiotic strains, particularly in immunocompromised patients, has been eliminated with the use of tyndallized (heat-killed) probiotic bacteria and yeast, named postbiotics. After inactivation, dead cells can release bacterial or yeast components with key immunomodulating effects and with antagonizing properties against pathogens [23,24]. At the clinical level, products containing tyndallized probiotic strains have showed a role in gastrointestinal diseases, including diarrhoea.

ABB C22® is a postbiotic combination of three fractions of the yeast groups of *Saccharomyces boulardii*, *Saccharomyces cerevisiae* and *Kluyveromyces marxianus*. The product exhibits complementary mechanisms for the protection of the integrity and function of enterocytes. ABB C22® exerts microbiome modulating properties due to the intrinsic nature properties of its consortia of yeasts strains, which confer protection of the gut epithelium from the attachment of pathogens and can effectively compensate diarrhoea-related dysbiosis.

The aim of this study was to assess the synergistic mechanism of action by which ABB C22® exerts its benefits on different levels influencing diarrhoeal episodes: anti-inflammatory effect, protection of the gut barrier function and protection from rotavirus infection. Confirmation of the favourable effects of ABB C22® in these experimental studies may further support its use as a solution for improving diarrhoeal episodes and support gut health.

2. Materials and Methods

2.1. Investigational Product and Strains of Study

The following strains from AB Biotek Human Nutrition & Health's yeast strain collection were studied in the In vitro tests: *Saccharomyces boulardii* ABB S3, a tyndallized yeast strain with broad scientific evidence as a probiotic; zinc-enriched *Saccharomyces cerevisiae* ABB S6, a tyndallized yeast strain enriched with zinc salts that has proved to increase the bioavailability of this mineral; and *Kluyveromyces marxianus* ABB S8, a tyndallized yeast with anti-inflammatory and antioxidant properties. The investigation product (ABB C22®, AB Biotek Human Nutrition & Health, Peterborough, UK) is composed of a synergistic combination the aforementioned heat-treated postbiotic yeasts *Saccharomyces boulardii*, zinc-enriched *Saccharomyces cerevisiae* and *Kluyveromyces marxianus*. The yeast strains were tested at a concentration of 10^7 cells/mL in all experiments.

2.2. Immune Cell Signalling Assay

The In vitro immune-modulating activity of ABB C22® and individual yeasts was studied by analysis of the cytokine production by macrophages (human THP-1 cell line). The THP-1 cell line was cultured (1×10^5 cells/well) in 96-well plates in the presence of 100 nM phorbol 12-myristate 13-acetate (PMA, Sigma) and incubated for 48 hours to induce differentiation of the THP-1 monocytes into macrophages. Cells were washed and incubated for another 72 hours in culture medium. After this, cells were incubated for 1 hour with the test components (individual yeasts strains or ABB C22®). Cells were then incubated for another 16 hours with and without LPS (100 ng/ml, Sigma) in the presence of the test components. All conditions were tested in triplicates. Supernatants were collected after stimulation and stored at -20°C . ELISA assays (IL-10 Human Uncoated ELISA Kit, TNF-alpha Human Uncoated ELISA Kit, Life Technologies) were used to measure the TNF- α and IL-10 levels, according to the manufacturers protocol. The TNF- α /IL-10 ratio was used as a marker of pro-inflammatory status and was standardized against the effect of the negative control.

The metabolic activity of the cells for testing cytotoxicity of the test compounds was analysed by WST-1 assay (Roche), according to the manufacturers protocol, after collecting the culture supernatant of the immune cell signalling assay.

2.3. Gut Epithelial Cell Signalling Assay

The anti-inflammatory effect of ABB C22® and individual yeasts on intestinal cells was studied by cytokine production by Caco-2 cells in the presence and absence of a pro-inflammatory stimulus. Caco-2 cells were cultured to confluence in 96-well plates in culture medium (modified Eagle's medium [MEM]), supplemented with 20% (v/v) foetal bovine serum (FBS), 1% non-essential amino acids (NEAA), 1% Glutamax™, 1% sodium pyruvate, with or without 1% penicillin-streptomycin and gentamicin (50 $\mu\text{g}/\text{mL}$) (all obtained from Invitrogen, Breda, The Netherlands). At the start of the experiment, cells were washed once with antibiotic-free culture medium. The monolayers were incubated with test components (individual yeasts strains or ABB C22®) in triplicate for 1 h at 37°C in antibiotic-free medium. Then, cells were further incubated for 24 h in the presence of the test components (individual yeasts strains or ABB C22®) and 50 $\mu\text{g}/\text{mL}$ gentamicin, with and without a mixture of recombinant tumour necrosis factor alpha (TNF- α) (10 ng/mL) and recombinant interferon (IFN- γ) (5 ng/mL) as a pro-inflammatory stimulus. Supernatants were collected 24 h after stimulation and stored at -20°C . A Bio-Plex Multiplex Immunoassay System (Bio-Rad, Hercules, CA, USA) was used to measure IFN- γ -induced protein-10 (IP-10), interleukin 8 (IL-8), and monocyte chemoattractant protein-1 (MCP-1) levels according to the manufacturer's instructions. IP-10, IL-8

and MCP-1 levels were exmeasured as pg/mL and then expressed as the percentual standardization against the effect of the negative control. All experiments were performed in triplicate.

The metabolic activity of the cells for testing cytotoxicity of the test compounds was analysed by WST-1 assay (Roche), according to the manufacturers protocol, after collecting the culture supernatant of the epithelial signalling assay.

2.4. Gut Barrier Integrity Assay

The protective effect of ABB C22® and the three individual yeasts on gut barrier integrity and functionality was studied by transepithelial electric resistance (TEER) over a gut cell monolayer. TEER was used to measure the spontaneous formation of the gut epithelium over time. Caco-2 cells were cultured in MEM medium in the same conditions as in the previous experiment. Then, the cells were seeded (2×10^4 cells/cm²) on Transwell polycarbonate cell culture inserts with a mean pore size of 0.4 µm and a diameter of 0.33 cm², until full differentiation (± 1000 ohms [Ω]) (Greiner Bio-one, Alphen aan de Rijn, The Netherlands). As an indicative measure of barrier integrity, TEER was measured with an EVOM2 Epithelial Volt/Ohm Meter (World Precision Instruments). On the day of the experiment, the cells were washed and incubated for 1 h at 37 °C with antibiotic- and serum-free medium containing the test components (individual yeasts strains or ABB C22®). TEER was measured before the start of the experiment ($t = -1$) every 2 days for 22 days. The TEER values of the individual conditions were compared to their own TEER value at $t = 0$ and expressed as Δ TEER (Ω/cm²). A negative control without the study product and a positive commercial control which had previously reported a strong effect on gut barrier formation were included. The experiment was performed in triplicate.

2.5. Rotavirus Gene Expression Assay

To assess the In vitro protective effects of ABB C22® and the individual yeasts on intestinal epithelial cells subjected to infection with rotavirus (RV) supernatants, gene expression assays were performed in MA104 Monkey African Green kidney cells subjected to rotavirus infection. For the generation of viral supernatants, MA104 cells were planted in a 75cm² culture flask at an 80 % confluence 24 h prior to infection. Once attached to the culture plastic, DMEM + 10 % FBS was removed and replaced with serum-free EMEM and cells were incubated overnight in serum starving to promote viral infection. The next day, a new vial of viral supernatant was thawed at room temperature and diluted up to 3 ml with serum-free EMEM and incubated in the presence of trypsin-EDTA 2 µg/ml for 30 min at 37 °C. After the incubation period, medium was removed from the culture flask and replaced by the viral supernatant. MA104 cells were incubated in the presence of viral supernatant for 4 h at 37 °C 5 % CO₂. After the incubation period, 7 ml of serum-free EMEM containing trypsin-EDTA 2 µg/ml were added to the culture flask and cells were incubated for 7 days to allow the accumulation of viral particles in the culture supernatant. From this culture, virus aliquots were prepared as follows: the flask containing the infected cells was subjected to three consecutive freeze-thaw cycles at -80°C to allow the lysis of cells and subsequent release of viral particles. Then, medium was harvested and centrifuged at 2000 rpm for 5 min to separate cell debris from the liquid fraction. This supernatant was recovered and stored in aliquots for further infection assays at -80°C. Confluent MA104 cells were detached after incubation with 0.5 % Trypsin for 1 min at 37 °C and further inactivation with DMEM containing 10 % FBS. Then, cells were seeded in 96-well plates at a density of 10^4 cells/well and incubated overnight at 37 °C. Culture medium was then replaced by the test components (individual yeast strains, their paired combination, or the combination of the three strains) suspended in serum-free EMEM. After incubation of MA104 cells in the presence of the test components for 24 h at 37°C, cell culture supernatants were removed and washed three times with 200 µl PBS per well. Parallely, viral aliquots were thawed and incubated for 30 min at 37 °C in the presence of trypsin/EDTA 2 µg/ml. Then, 25 µl of the trypsin-activated viral supernatants were added to each well and incubated with cells for 4 h at 37 °C and 5 % CO₂. After this period, 75 µl of serum-free EMEM containing trypsin/EDTA 2 µg/ml were added per well and cells were incubated for additional 72 h to allow viral expansion. Isolation of viral RNA and rotavirus

gene amplification assay: After infection with rotavirus supernatants, 100 μ l of RLT Plus buffer were added per well and mixed thoroughly with culture medium by repeated pipetting. Then, plates were frozen at -80°C to allow cell lysis and release of intracellular material. For RNA isolation, the whole volume of each well was incubated with 0.4 mg Dynabeads suspended in 80 μ l ethanol 100 % for 5 min at room temperature. After this period, Dynabeads were decanted with a DynaMagTM-96 Side Magnet. The liquid fraction was removed, and the Dynabeads-bound RNA was washed twice with 100 μ l ethanol 80 %. After the second wash step, ethanol was allowed to evaporate for 3 min at room temperature and RNA was eluted in 25 μ l of RNase-free H₂O. RNA was reverse transcribed to copy DNA (cDNA) with ABI High-Capacity kit from 206 ng of starting material. From the reverse transcription products, 45 ng were used as an input for qPCR reactions in the presence of fluorescent probe assays designed to amplify the rotavirus genes NSP3 [25] and VP7. Beta actin (ACTB) gene was used as housekeeping gene. Relative gene expression was calculated with the Pfaffl method [26].

2.6. Statistical Analysis

For all the experiments, quantitative data are expressed as mean and standard deviation (\pm SD). Student's t-test (two-sided) or one-way analysis of variance (ANOVA) with Dunnett's procedure was used for the comparison of data according to conditions of application.

For the viral gene expression assays, one biological replicate with five technical replicates per condition was set. Outliers were identified by Rout method ($Q = 5\%$) and excluded from the analysis. Cleaned data were statistically analysed through One-Way ANOVA and Dunnett's post hoc multiple comparisons test.

Statistical significance was declared at $p < 0.05$, 95% of confidence. Bars in the charts represent the mean value for each condition and error bars indicate the standard error of the mean (SEM) for each group of values.

3. Results

3.1. Immune-Modulating Activity of ABB C22[®] by Immune Cell Signalling Assay

In vitro immune-modulating activity of ABB C22[®] and individual yeast strains was studied by analysis of TNF- α and IL-10 cytokines production by macrophages (THP-1 cell line). Cytotoxicity of individual yeasts and ABB C22[®] was discarded by a WST-1 assay before assessing the anti-inflammatory effect on immune cells (data not shown). As shown in Figure 1A, ABB C22[®] caused a 71% reduction of the TNF- α /IL-10 ratio, a marker of pro-inflammatory status, compared to the negative control. Interestingly, none of the individual yeast strains had an anti-inflammatory effect when tested singularly. Moreover, the TNF- α /IL-10 ratio was unaffected in lipopolysaccharide (LPS)-challenged macrophages supplemented with ABB C22[®] (Figure 1B).

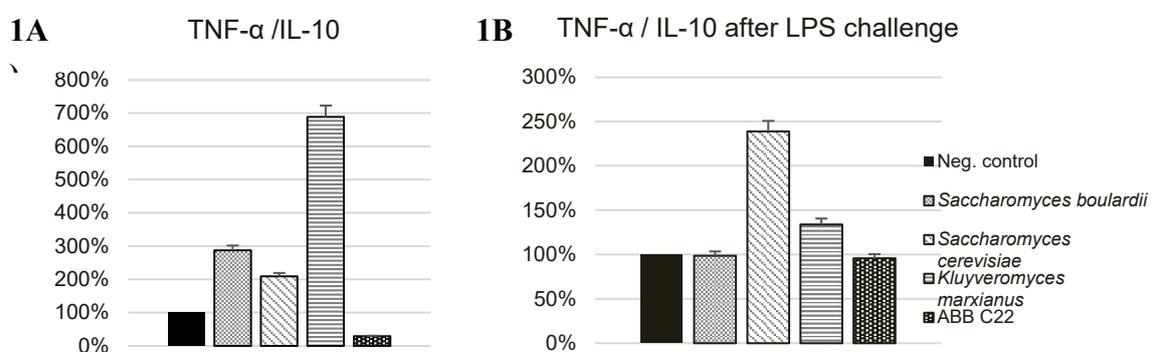


Figure 1. In vitro reduction of TNF- α /IL-10 ratio in human THP-1 cell line (macrophages). **A.** In the absence of a pro-inflammatory stimulus; **B.** In the presence of a pro-inflammatory stimulus (LPS challenge) simulating an infection. TNF- α /IL-10 ratio was calculated as a measure of the anti-inflammatory effect of the tested components, standardized against the effect of the negative control.

3.2. Anti-Inflammatory Effect of ABB C22® on Intestinal Cells by Gut Epithelial Cell Signalling Assay

In vitro immune-modulating activity of individual yeasts and ABB C22® in the gut was studied by cytokine production by Caco-2 cells in the presence and absence of a pro-inflammatory challenge with TNF- α /IFN- γ . Cytotoxicity of individual yeasts and ABB C22® was discarded by a WST-1 assay before assessing the anti-inflammatory effect on intestinal cells (data not shown). ABB C22® showed statistically significant reductions in IP-10, IL-8, and MCP-1 levels in the presence and absence of a pro-inflammatory challenge compared to the negative control and to the single yeast strains (Figure 2A–F). In the absence of a pro-inflammatory stimulus, IP-10 cytokine production by Caco-2 cells showed a 26% and 23% decrease after incubation with *K. marxianus* alone and ABB C22®, respectively (Figure 2A). A synergistic reduction of the pro-inflammatory cytokines IL-8 and MCP-1 in gut epithelial cells was particularly observed for ABB C22® compared to individual yeasts in the absence of a challenge (41% and 36%, respectively) (Figure 2C,E). On the other hand, after a pro-inflammatory challenge, ABB C22® synergistically reduced IP-10 and MCP-1 levels in gut epithelial cells (37% and 50%, respectively) (Figure 2B,F), and had an effect in reducing IL-8 levels by 22% (Figure 2D).

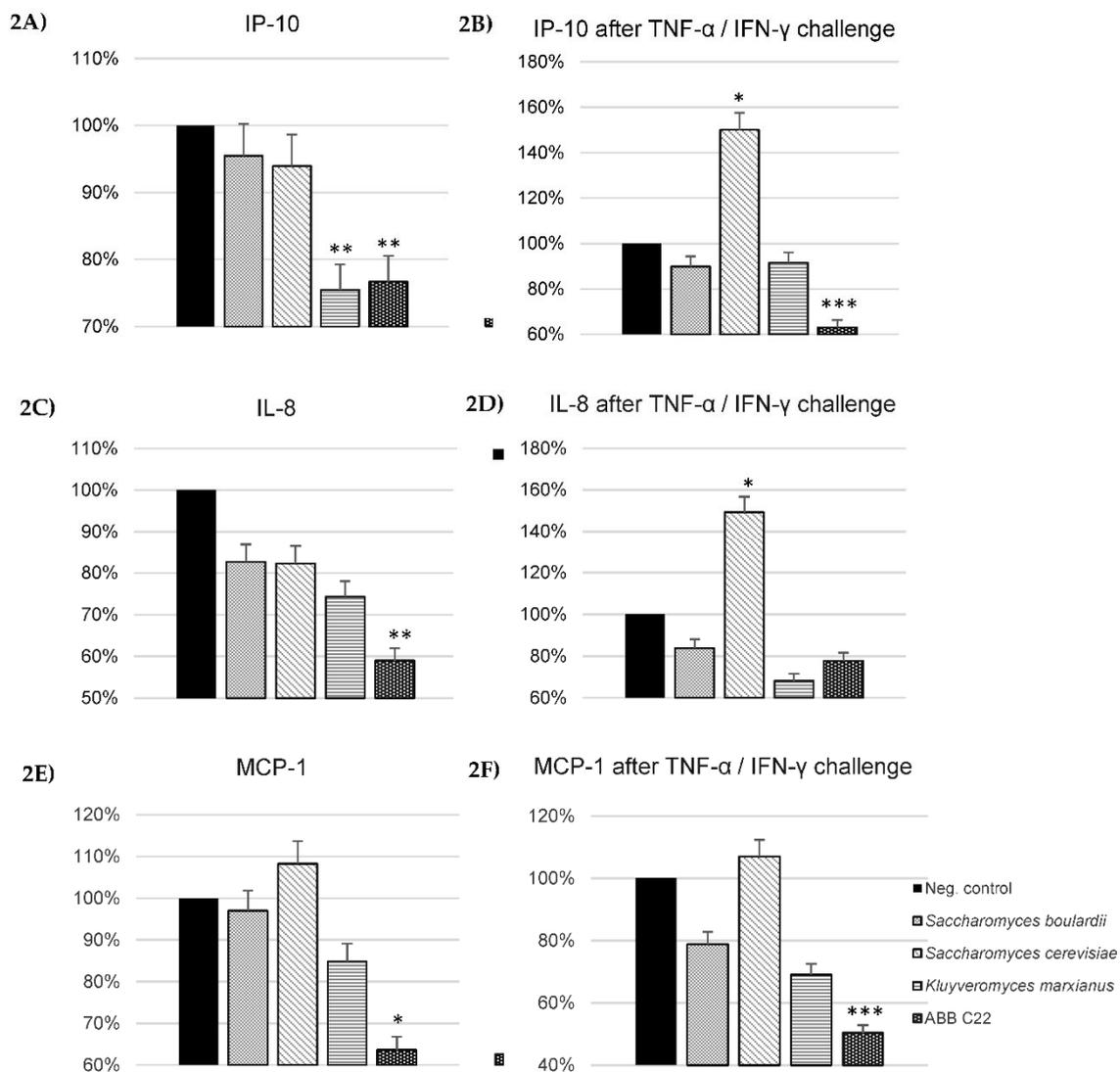


Figure 2. Production of pro-inflammatory cytokines by Caco-2 cells after incubation with ABB C22® and three yeasts. *, ** and *** represent statistical significance with p-value < 0.05, 0.01 and 0.001, respectively, for between-group comparisons (one-way analysis of variance, ANOVA and Dunnett's post hoc test). **A.** IP-10 cytokine production in the absence of pro-inflammatory stimulus; **B.** IP-10 cytokine production in the presence of a pro-inflammatory stimulus (TNF- α / IFN- γ) simulating an inflamed gut epithelium; **C.** IL-8 cytokine production in

the absence of pro-inflammatory stimulus; D. IL-8 cytokine production in the presence of a pro-inflammatory stimulus (TNF- α /IFN- γ) simulating an inflamed gut epithelium; E. MCP-1 cytokine production in the absence of pro-inflammatory stimulus; F. MCP-1 cytokine production in the presence of a pro-inflammatory stimulus (TNF- α /IFN- γ) simulating an inflamed gut epithelium. The results of each cytokine are expressed as the standardization against the effect of the negative control.

3.3. Gut Barrier Integrity Assay

The capacity of ABB C22® to induce spontaneous formation of the gut epithelium over time was evaluated through TEER in a Caco-2 cell monolayer. ABB C22® shows the ability to increase TEER until day 22 when compared to a negative control and a positive commercial control of a live *S. boulardii* CNCM I-745® (Figure 3).

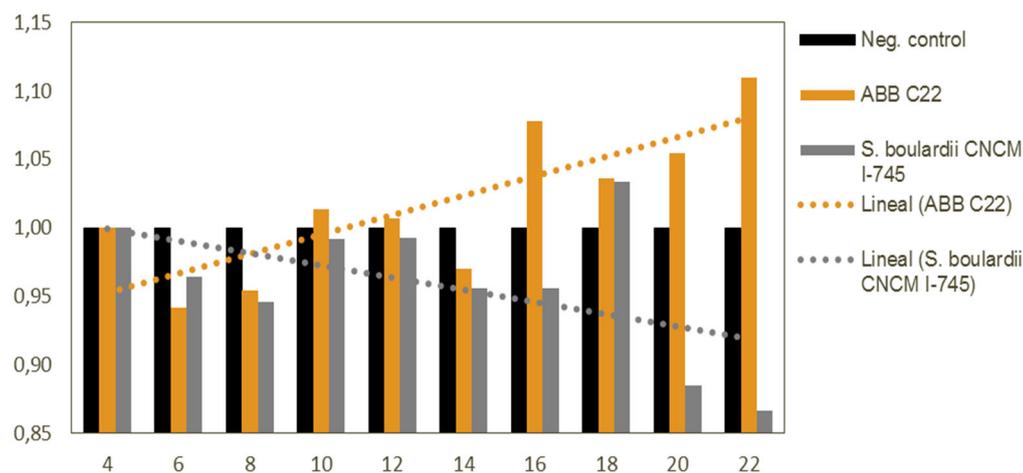


Figure 3. Bars and slopes of trendlines for TEER increases associated with incubation of Caco-2 monolayer cells with a negative control, ABB C22® and a positive commercial control of live *S. boulardii* CNCM I-745®. The comparison of Δ TEER values over the course of 22 days indicates a higher spontaneous build-up of the epithelium monolayer for the ABB C22® condition versus the controls.

3.4. Rotavirus Antagonism

Results from the gene expression assay revealed a statistically significant decrease in the relative expression of NSP3 after pre-treatment with all test components compared to the untreated cells ($p < 0.0001$ for all treatments). A significant decrease of NSP3 was also observed for single inactivated *Saccharomyces boulardii*, single inactivated *Kluyveromyces marxianus* and ABB C22®, but no significant effect was observed for gene VP7 for these strains. Moreover, pre-treatment with single live *Saccharomyces boulardii* (positive control), single inactivated *Saccharomyces cerevisiae*, inactivated *Saccharomyces boulardii* + inactivated *Saccharomyces cerevisiae* and inactivated *Saccharomyces cerevisiae* + inactivated *Kluyveromyces marxianus* triggered a significant relative decrease in VP7 gene expression as well. In other words, all the test components had the ability to reduce the relative expression of one or the two rotavirus genes, and test components single live *Saccharomyces boulardii*, single inactivated *Saccharomyces cerevisiae*, inactivated *Saccharomyces boulardii* + inactivated *Saccharomyces cerevisiae* and inactivated *Saccharomyces cerevisiae* + inactivated *Kluyveromyces marxianus* had a remarkable protective effect against rotavirus infection reflected in a significant relative reduction of both RV genes. (Figure 4).

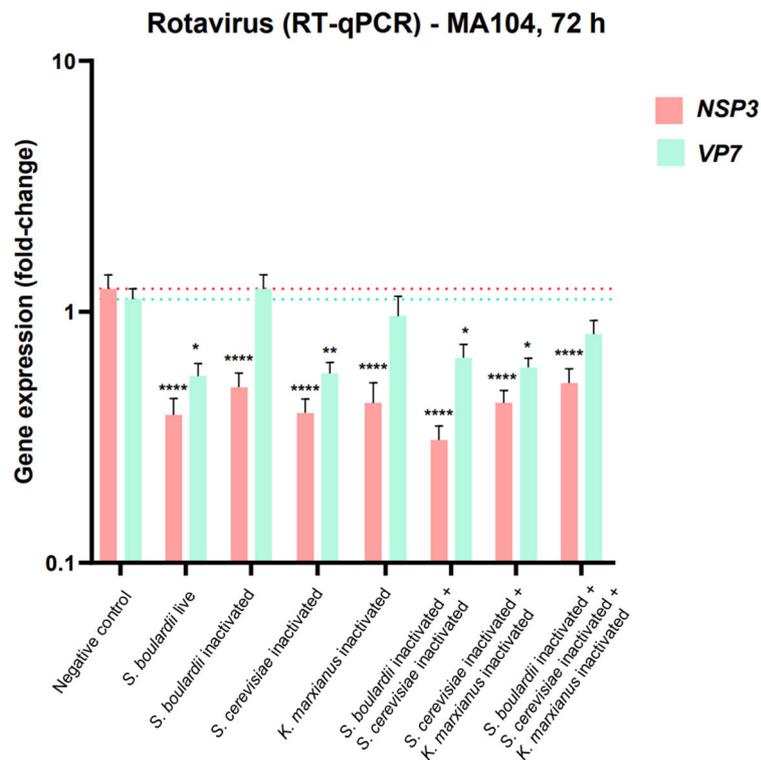


Figure 4. Bar diagram showing relative NSP3 and VP7 gene expression in RNA extracts from MA104 cells subjected to a 24 h pre-treatment with the selected combinations of yeast strains and to infection with rotavirus for 72 h. *, **, *** and **** represent statistical significance with p-value < 0.05, 0.01, 0.001 and 0.0001, respectively.

4. Discussion

Inflammatory diarrhoea is typically caused by cytotoxin-producing and invasive pathogens (e.g., enteroaggregative *E. coli*, enterohaemorrhagic *E. coli*, *C. difficile*) [27]. These organisms cause disease either by secreting noxious cytotoxins intraluminally or by invading the intestinal epithelium resulting in an acute inflammatory reaction in the mucosa with disruption of the epithelium [28]. By contrast, non-inflammatory diarrhoeas are usually caused by pathogens (e.g., rotavirus) that adhere to the small intestine mucosa disrupting the absorptive and/or secretory process of the enterocyte [29]. The aim of this study was to assess the synergistic mechanism of action by which ABB C22® exerts its benefits on different levels influencing both inflammatory and non-inflammatory diarrhoeal episodes: anti-inflammatory effect, protection of the gut barrier function and protection from rotavirus infection.

In this study, ABB C22® has demonstrated a markedly beneficial anti-inflammatory effect both on gut epithelial cells and on immune cells. These *In vitro* experiments have shown that ABB C22® is able to reduce the TNF α /IL-10 ratio, a parameter to assess immune homeostasis maintenance, in macrophages. Excessive production of TNF- α induces an exacerbated inflammatory response and tissue damage. In physiological conditions, an increase in TNF- α levels is counterbalanced with the synthesis of IL-10, an anti-inflammatory cytokine which suppresses production of many activating and regulatory inflammatory mediators. The balance between the two molecules prevents exacerbated reactions of the inflammatory response and manages immune homeostasis. Our results indicate that ABB C22® suppresses TNF α release while stimulating IL-10 production, contributing to immune homeostasis. Interestingly, the TNF α /IL-10 ratio remains unaffected after ABB C22® supplementation under lipopolysaccharide (LPS) challenge (simulating infection). The reason relies on the capacity of LPS-activated macrophages to promote a strong pro-inflammatory state to fight the LPS-secreting agent. In this case, overwhelming expression of IL-10 is undesirable since it may contribute to an impaired immune reaction to infection [30]. If the macrophages response under these

conditions was to increase the presence of IL-10, the effect of TNF α would be counteracted by IL-10 and the macrophage-induced inflammatory response would not be effective against the LPS-secreting agent. Therefore, ABB C22 $\text{\textcircled{R}}$ prevents an overwhelming expression of IL-10 after LPS challenge, enabling macrophages to effectively exert their pro-inflammatory properties against the infection. Very importantly, this effect was not seen when the strains were tested singularly, proving that the modulatory power relies on the synergy of the combined components in ABB C22 $\text{\textcircled{R}}$. These findings show the strong effects of ABB C22 $\text{\textcircled{R}}$ in modulating the inflammatory response to counteract infection while preventing exacerbated immune responses in patients suffering from inflammatory diarrhoeal episodes.

The evidence from various studies indicates the significant role of IP-10, IL-8, and MCP-1 cytokines in inflammatory processes that participate in diarrhoea. Research has shown increased expression of these cytokines in conditions like ulcerative colitis (UC), where they are involved in the recruitment and activation of inflammatory cells, contributing to tissue damage. Furthermore, IL-8 has been detected in samples from UC patients with inflammation, highlighting its role in inflammatory processes [31]. ABB C22 $\text{\textcircled{R}}$ has been shown to exhibit further anti-inflammatory effects on gut epithelium cells, by decreasing the release of the pro-inflammatory cytokines IP-10, IL-8, and MCP-1 in the gut epithelium cells in a healthy state (26%, 41% and 36% reduction of IP-10, IL-8, and MCP-1, respectively) and in a model of inflamed epithelium challenged with TNF- α /IFN- γ (37%, 22% and 50% reduction, respectively). Particularly, ABB C22 $\text{\textcircled{R}}$ had a synergistic anti-inflammatory effect on IL-8 and MCP-1 compared to the single strains. ABB C22 $\text{\textcircled{R}}$'s synergistic power was also observed on IP-10 and MCP-1 after a pro-inflammatory challenge. These findings sustain the crucial power of the combination of the three strains, as contained in ABB C22 $\text{\textcircled{R}}$, to counteract inflammation in the gut, which is a critical factor in inflammatory diarrhoeas.

Data of the In vitro assays on TEER have shown that ABB C22 $\text{\textcircled{R}}$ exhibits a superior effect in stimulating the build-up of the digestive epithelium compared to *Saccharomyces boulardii* CNCM I-745 $\text{\textcircled{R}}$ (a commonly studied yeast probiotic in gut health that had previously shown a strong protective effect on the gut barrier [32,33]). The effect of ABB C22 $\text{\textcircled{R}}$ was stronger and lasted for a longer time. Gut microbiota modifications, mucus layer alterations and epithelial damage can alter the intestinal permeability of the gut barrier function, worsening diarrhoeal episodes [34]. ABB C22 $\text{\textcircled{R}}$ has demonstrated to enhance and protect the gut barrier function and integrity, contributing to the recovery of the microbiota balance and gut homeostasis during diarrhoeal episodes.

The goal of the Rotavirus gene expression assay was to assess the In vitro protective effects of the proposed combinations of yeast strains and ABB C22 $\text{\textcircled{R}}$ on intestinal epithelial cells subjected to infection with RV supernatants. The results showed that all the test components had the ability to reduce the relative expression of one or the two rotavirus genes. Moreover, the test components single inactivated *Saccharomyces cerevisiae*, inactivated *Saccharomyces boulardii* + inactivated *Saccharomyces cerevisiae* and inactivated *Saccharomyces cerevisiae* + inactivated *Kluyveromyces marxianus* showed a remarkable protective effect against rotavirus infection reflected in a significant relative reduction of both RV genes expression. This highlights the synergistic effect of the paired combination of *S. boulardii* with *S. cerevisiae* and *K. marxianus* strains. These results are remarkable evidence for the combination of these yeasts strains to counteract rotavirus infection, and therefore prevent or manage non-inflammatory diarrhoea.

In current literature, individual yeasts components of ABB C22 $\text{\textcircled{R}}$ have shown different protective mechanisms of action against diarrhoea. *S. boulardii* has been long studied as a treatment for diarrhoea and the evidence on its effectivity is wide. It has shown potent antisecretory properties versus water and electrolyte secretion in a model of rat colon with castor-oil induced diarrhoea [35]. In a model of cholera toxin-induced secretion in rat jejunum cells, *S. boulardii* inhibited Cl-secretion through both cAMP- and Ca $^{2+}$ -mediated signalling pathways [36]. In a model of pig intact jejunal epithelia, *S. boulardii* showed a specific duration-response effects after stimulation with the secretagogue theophylline [37]. *S. boulardii* exerts an anti-inflammatory action and reduces chloride secretion, decreases IL-8 production via inhibition of the activation of MAP kinases Erki/2 and JNK/SAPK, prevents oxidative stress via inhibition of reactive oxygen species (ROS) formation and

restoration of the balance of the glutathione-dependent redox system [38,39]. In an adult mice model, *S. cerevisiae* extracts containing rotavirus-like particles were successfully used for producing an immunological response capable of reducing the replication of rotavirus after viral shedding infection [40]. The third component of ABB C22®, *K. marxianus*, has shown control of intestinal inflammatory pathways and reduction of ROS in different murine and colitis-induced models [41].

Overall, with the experiments exposed in this paper, we have proven evidence that the combination of these three yeast strains, each with their own independent evidence in the management of diarrhoea, provides added value for the management of diarrhoea compared to the use of the single strains alone.

5. Conclusions

ABB C22® is a postbiotic combination of the tyndallized yeasts *S. boulardii* ABB S3, *S. cerevisiae* ABB S6, and *K. marxianus* ABB S8. In this study, ABB C22® synergistic combination has demonstrated markedly beneficial anti-inflammatory effects both on gut epithelial cells and on immune cells, modulating the inflammatory response to counteract infection while preventing exacerbated immune responses in patients suffering from inflammatory diarrhoeal episodes. Very importantly, this effect was not seen when the strains were tested singularly, proving that the modulatory power relies on the synergy of the combined components in ABB C22®. ABB C22® has demonstrated to enhance and protect the gut barrier function and integrity, contributing to the recovery of the microbiota balance and gut homeostasis during diarrhoeal episodes. The yeast strains in ABB C22® have proven to have a protective effect against rotavirus infection when combined, particularly the paired combination of *S. boulardii* with *S. cerevisiae* and *K. marxianus*.

Showing efficacy in the different underlying mechanisms involved in gastrointestinal homeostasis, the postbiotic ABB C22® has proven to be an ideal solution for the management of diarrhoea and gastrointestinal conditions.

6. Patents

Three patent applications have resulted from the work reported in this manuscript: WO2023275325: Physiologically acceptable yeast compositions and uses thereof; WO2023275293: Physiologically acceptable yeast compositions and uses thereof; WO2023275323A9: Physiologically acceptable yeast compositions for use in the treatment of a gastro-intestinal disorder.

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Conflicts of Interest: Lydia Carrera Marcolin, Jordi Cuñé Castellana, Laia Martí Melero, Carlos de Lecea and Maria Tintoré Gazulla are full-time employees of AB Biotek Human Nutrition and Health.

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