

<i>L. major</i>	1	MVFDTDFFVNVCSQMAANVLQSYIVAALHSAATQLPSSVQPYAMMLTRED	50
		. : : : :	
<i>L. donovani</i>	1	-----MAANALHSHIVAALHNAATKLPSSVQPYAMMLTRED	36
<i>L. major</i>	51	MVSTTLATAIATAVILYTVITVVLVPLRMDFYLSKLPTIKNSIPFLGHAL	100
		: : : : :	
<i>L. donovani</i>	37	MVSTTLATAIATAVILYTVITVVLVPLRMDFYLSKLPTIKNGIPFLGHAL	86
<i>L. major</i>	101	LLAGPSPWSKMSNNSLYPEKNLPQKKKSVDGPQTSRLVTYNVAGHRVIYI	150
		: : : : :	
<i>L. donovani</i>	87	LLAGPSPWSKMSNNSLYPEKNLPQKKKSADGPQTSRLVTYNVAGHRVIYI	136
<i>L. major</i>	151	NEPRLLRRVLLTHQRNYRKALAAAYKHFMCLLGTGLVTSEDEQWKKGRLL	200
		: : : : :	
<i>L. donovani</i>	137	NEPRLLRRVLLTHQRNYRKALAAAYKHFMCLLGTGLVTSEDEQWKKGRLL	186
<i>L. major</i>	201	LSHAMRIDILDSVPEMAMKAVDRILLKLDVDAKNPSVDLNEEYRHMTLQ	250
		: : : : :	
<i>L. donovani</i>	187	LSHAMRIDILDSVPEMAMKAVDRILLKLDVDAKNPSVDLNEEYRHMTLQ	236
<i>L. major</i>	251	VISESALSLSAEESDRIFPALYLPVHECNKRVWAPWRAYMPFLHGSRRMR	300
		: : : : :	
<i>L. donovani</i>	237	VISESALSLSAEESDRIFPALYLPVHECNKRVWAPWRAYMPFLQGSRRVR	286
<i>L. major</i>	301	NRCLSELNKLVRDIICRRWEQRNDSNYTAKPDILALCISQIDRIDEKMIV	350
		. : : : : :	
<i>L. donovani</i>	287	NHCLSELNKLVRNIICRRWEQRNDSNCTGKPDILALCISQIDRMDEKMIV	336
<i>L. major</i>	351	GLIDDVKTIILAGHETSAALLTFATYEVLRHPEIRQKILEEATRLFDPAR	400
		: : : : :	
<i>L. donovani</i>	337	GLIDDVKTIILAGHETSAALLTFATYEVLRHPEIRQKILEEATRLFDPAR	386
<i>L. major</i>	401	CTCTVQTRYGPRGVPALNDVRSVLVWTPAVLRETLRRHSVWPLVMRYAAKD	450
		. : : : : :	
<i>L. donovani</i>	387	CTRTVQTRYGPRGVPVNDVRDLVWTPAVLRETLRRHSVWPLVMRYAAKD	436
<i>L. major</i>	451	DVWPAADTGLDADVRIPAGCTIAVGIEGVHNNPDVWINKPEVFDPTRFIDA	500
		: : : : :	
<i>L. donovani</i>	437	DVWPAADTGLDADVRIPAGCTIAVGIEGVHNNPDVWINKPEVFDPTRFIDA	486
<i>L. major</i>	501	EIANDTNYLNQSTKDVKFAKKIDPYAFIPFINGPRNCLGQHLSMIETQVA	550
		: : : : :	
<i>L. donovani</i>	487	EIANDTNYLNQSTKDVKFAKKIDPYAFIPFINGPRNCLGQHLSMIETQVA	536
<i>L. major</i>	551	LAYHVLSDYDLTIYRDPSTYKGDVAAYEDAVGRHHDFIIPQVPHDGLKVNVT	600
		: : : : :	
<i>L. donovani</i>	537	LAYHVLNYDLTIYRDPSTYKGDAAAYEDAVGRHHDFIIPQVPHDGLKVNVT	586
<i>L. major</i>	601	PNKLFM	606
<i>L. donovani</i>	587	PNKLFM	592

Figure S1. Pair-wise alignment of *L. major* CYP5122A (LmjF.27.0090) and *L. donovani* CYP5122A1 (LdBPK_270090.1) using EMBOSS Needle. Identity: 94.4%. Similarity: 95.9%.

Lm	-----MV---LAKKQMKIPPIHKLRPGWSAVALHTVIYLLRHDF	37
At	-----	0
Sc	MSAVFNNATLSGLVQASTYSQTLQNVAHYQQLNFMKEYWAAWYSYM-----	47
Mm	MA---TNK-SVGVS--SASLAVEYVDSLLPENPLQEPFKNAWVYML-----	41
Lm	TAIVATSIVQPLYHRLILQNTYLQRLSDP--ALFTLVFAVLCHCVPWAFFNGIFLFFDSI	95
At	-----MLLLPFMVN---TYFS---	13
Sc	-----NNDV--LATGLMFLLHEFMYFFRCLPWFIIDQIPYFR-R-	84
Mm	-----DNYTKFQIATWGLIVHEAIFYLFLSLPGFLFQFIPYMR-K-	80
	: * . :	
Lm	HPQYGI EGLRNNALLAPLGRMAAYKLPRKPQQLPSAALIFSTMLHTAINHYLIIPVVLY	155
At	-----FVPM--QTKNNTPAAQGKCITRLLLYHFSVNLPML	47
Sc	-----WKLQ--PTKIPSAKEQLYCLKSVLLSHFLVEAIPW	118
Mm	-----YKIQ--KDKPETFEGQWKCLKKILFNHFFIQLPLIC	114
	: : : : : * : :	
Lm	AYLVHTNSCALRAPPPESAIVGFAPEDVWNMMSGSNLRQIPISLVTITSHFLIANVINEM	215
At	ASYPVFRA-----MG-----MRSSFP-LPSW-KEVSAQILFYFIIDF	83
Sc	TFHPMCEK-----LG-----ITVEVP-FPSL-KTMALEIGLFFVLEDT	154
Mm	GTYYFTEF-----FN-----IPYDWERMPrWYLTAR-CLGCAVIEDT	151
	. .. : * :: : :	
Lm	GFYIVHSMHSSPTLYRVFHKHHMYTGTISIAAEYATPLEGILANAIPPTAYFTFMFFH	275
At	VFYWGHRILHS-KWLYKNVHSVHHEYATPFGLTSEYAHPAEILFLGFATIVGPAL-----	137
Sc	WHYWAHRLFHY-GVFYKYIHKQHRYAAPFGLSAEYAHPAETLSLGFGTVGMPILY--VM	211
Mm	WHYFLHRLH-H-KRIYKYIHKVHHEFQAPFGIEAEYAHPLETLILGTGFFIGIVL-----	205
	. * * : * : * : * . * : : * * * * * : .	
Lm	YTREEASKSSFVTSARAWPLFITWMWARLWETYEVS HSGYCFSDTWLGKLGLLHGHRRARFH	335
At	-----TGPHLITLWLMVLRVLETVEAHCGYHFPWSLSNFLPLYG--GADFH	182
Sc	Y-----TGKLHLFTLCWITLRLFQAVDSHSGYDFPWSLNKIMPFWA--GAHH	258
Mm	-----LCDHVILLWAWVTIRLLETIDVHSGYDIPLNPLNLVPFYT--GARHH	250
	: * : * : : * . * : : * *	
Lm	DFHHTHN--VCNYGSGL-FMDALLNTMDPYLIYRYPDKHPHTTALKEEDLKEPRDLEVG	391
At	DYHHRLLYTKSGNYSSTFVYMDWIFGTDKGYRRL-----KTLKENG-----	223
Sc	DLHHHYF--IGNYASSFRWWDYCLDTESGPEAK-----ASREERMKKRAEN--	302
Mm	DFHHMNF--IGNYASTFTWWDKLFGTDAQYHAY-----IEKSKKLKGSKSD---	293
	* ** ** * : : * : *	
Lm	SEAMDQVRACS	402
At	--DMKQT----	228
Sc	-NAQKKTN---	309
Mm	-----	293

Figure S2. The alignment of ERG25 proteins (Lm: *L. major*; At: *Arabidopsis thaliana*; Sc: *Saccharomyces cerevisiae*; Mm: *Mus musculus*) using Clustal Omega. Asterisks (*): fully conserved residues; colons (:): highly similar residues; periods (.): weakly similar residues. *L. major* ERG25 shows 23-25% identity to other ERG25s.

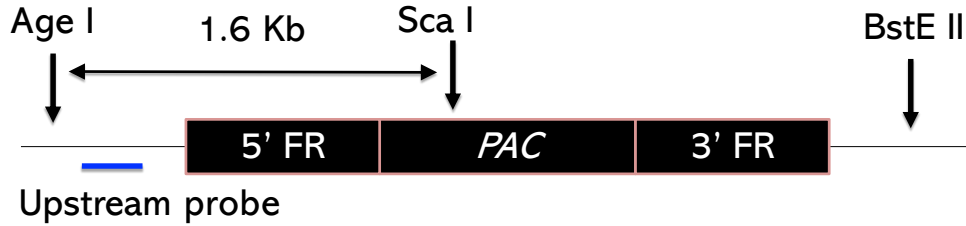
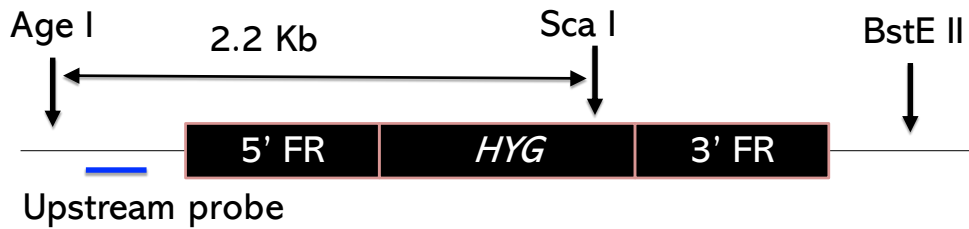
A**B****C**

Figure S3. Schematic depiction of the Southern blot for *L. major* *ERG 25*. (A) The genomic locus of *ERG25* in WT *L. major* along with its open reading frame (ORF), 5' and 3' flanking regions (FR), recognition sites of Age I, BseE II and Sca I, and the positions of ORF probe and upstream probe. (B-C) Replacement of *ERG25* ORF with hygromycin (HYG) and puromycin (PAC) resistance genes. The expected sizes of DNA fragments recognized by the ORF probe or upstream probe were indicated.

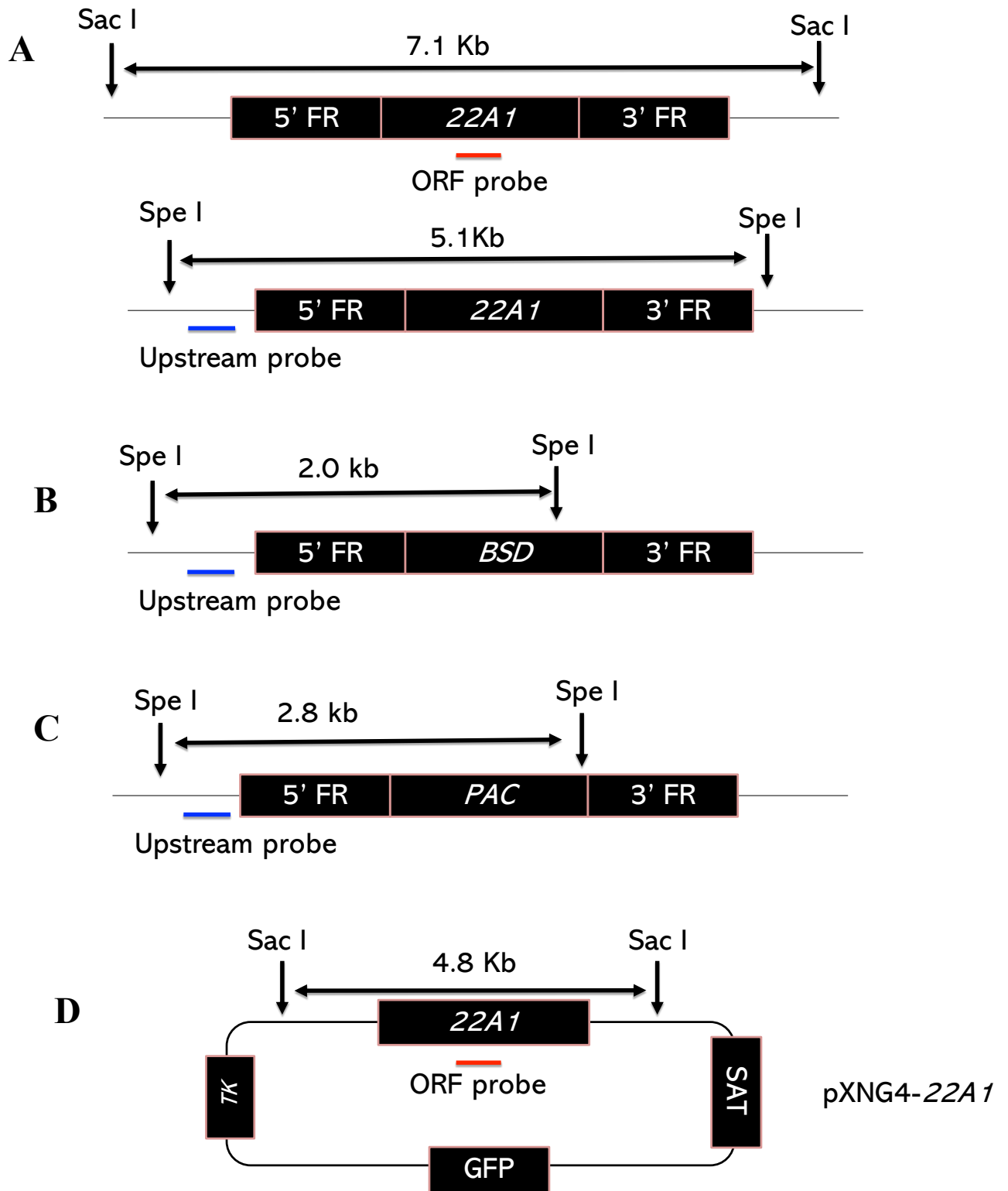


Figure S4. Schematic depiction of the Southern blot for *L. major* CYP5122A1 (22A1). (A) The genomic locus of CYP5122A1 in WT *L. major* along with its open reading frame (ORF), 5' and 3' flanking regions (FR), recognition sites of Sac I and Spe I, and the positions of ORF probe and upstream probe. (B-C) Replacement of 22A1 ORF with blasticidin (BSD) and puromycin (PAC) resistance genes. (D) Episomal expression of 22A1 from pXNG4-5122A1. TK: thymidine kinase, GFP: green fluorescent protein, SAT: nourseothricin resistance gene. The expected sizes of DNA fragments recognized by the ORF probe or upstream probe were indicated.

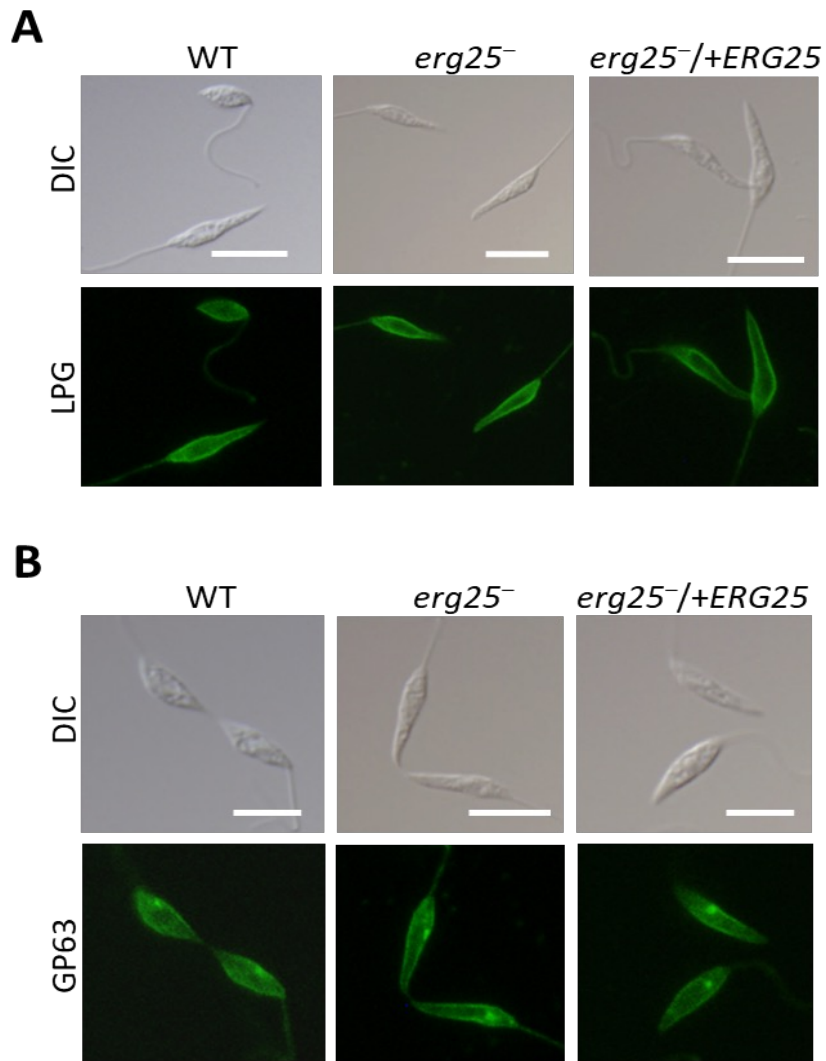


Figure S5. The expression of LPG and GP63 is not affected in *erg25*⁻ promastigotes. Log phase promastigotes were labeled with mouse anti-LPG (A) or anti-GP63 (B) monoclonal antibody followed by goat anti-mouse IgG-FITC. Scale bars: 10 μ m.

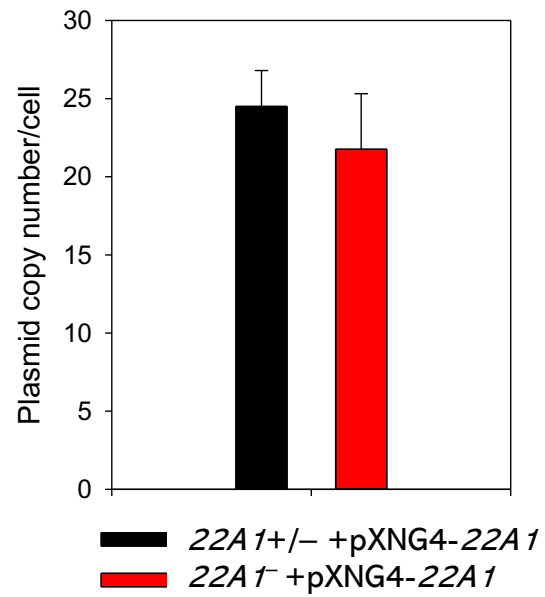


Figure S6. Plasmid levels in promastigotes. Stationary phase promastigotes of *22A1*^{+/-} +pXNG4-*22A1* and *22A1*⁻ +pXNG4-*22A1* were cultivated in the presence of SAT and their DNA was extracted to determine the average pXNG4-Lm22A1 plasmid copy numbers per cell by qPCR.

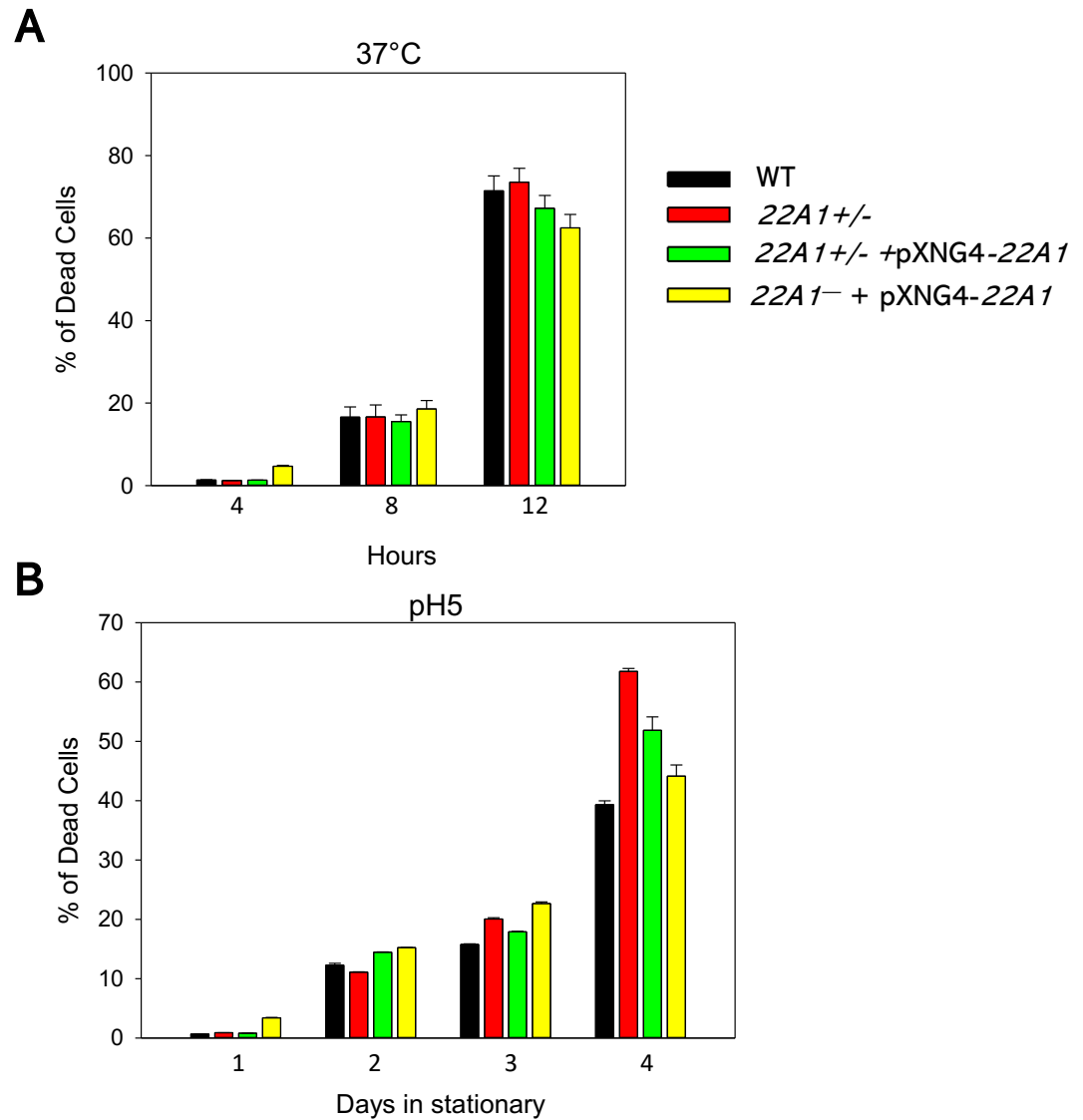
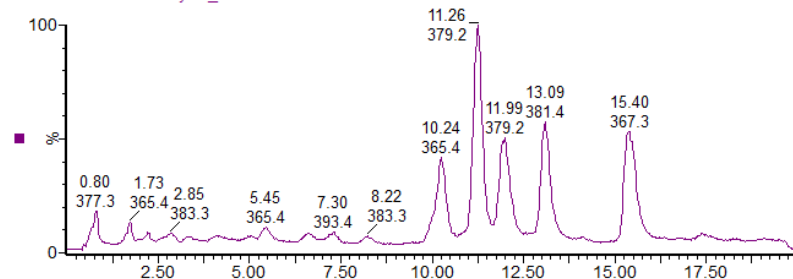


Figure S7. Stress response of *22A1* mutants. Day 1 stationary phase promastigotes were incubated under various conditions and percentages of dead cells were determined by flow cytometry at the indicated times. **A:** in complete M199, pH7.4, 37 °C. **B:** in complete M199, 27 °C, pH5.0. Error bars represent standard deviations from three experiments.

LV39 WT-119:05:5713-Aug-2019

20190813 sterol analysis_014

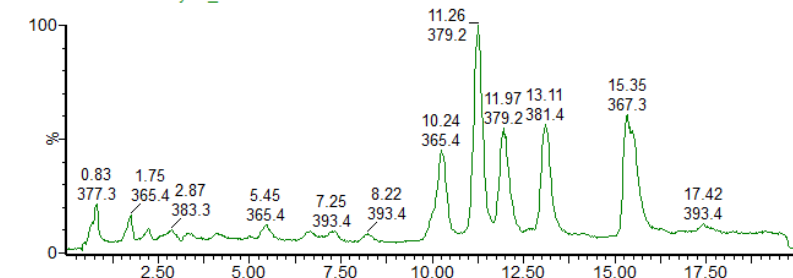
MRM of 27 Channels AP+
TIC
7.00e6



WT

20190813 sterol analysis_017

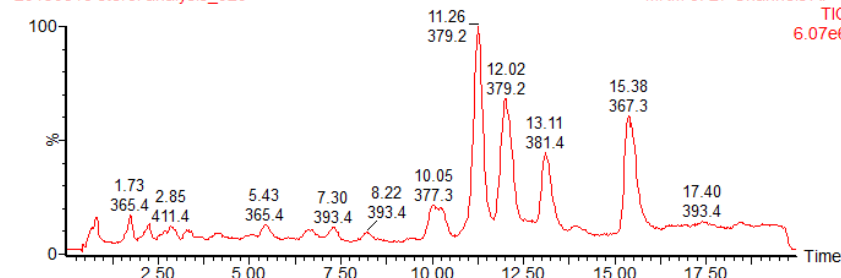
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TIC
6.21e6



22A1+/- #4

20190813 sterol analysis_020

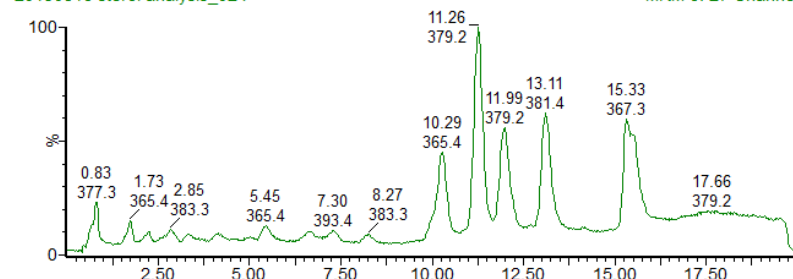
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TIC
6.07e6



22A1+/- #4 +pXNG4-
22A1

20190813 sterol analysis_024

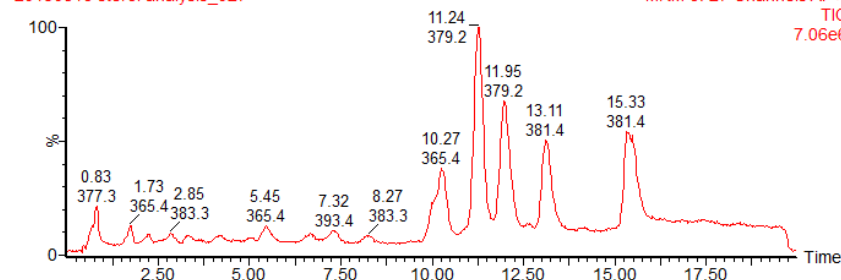
MRM of 27 Channels AP+
TIC
6.32e6



22A1+/- #5

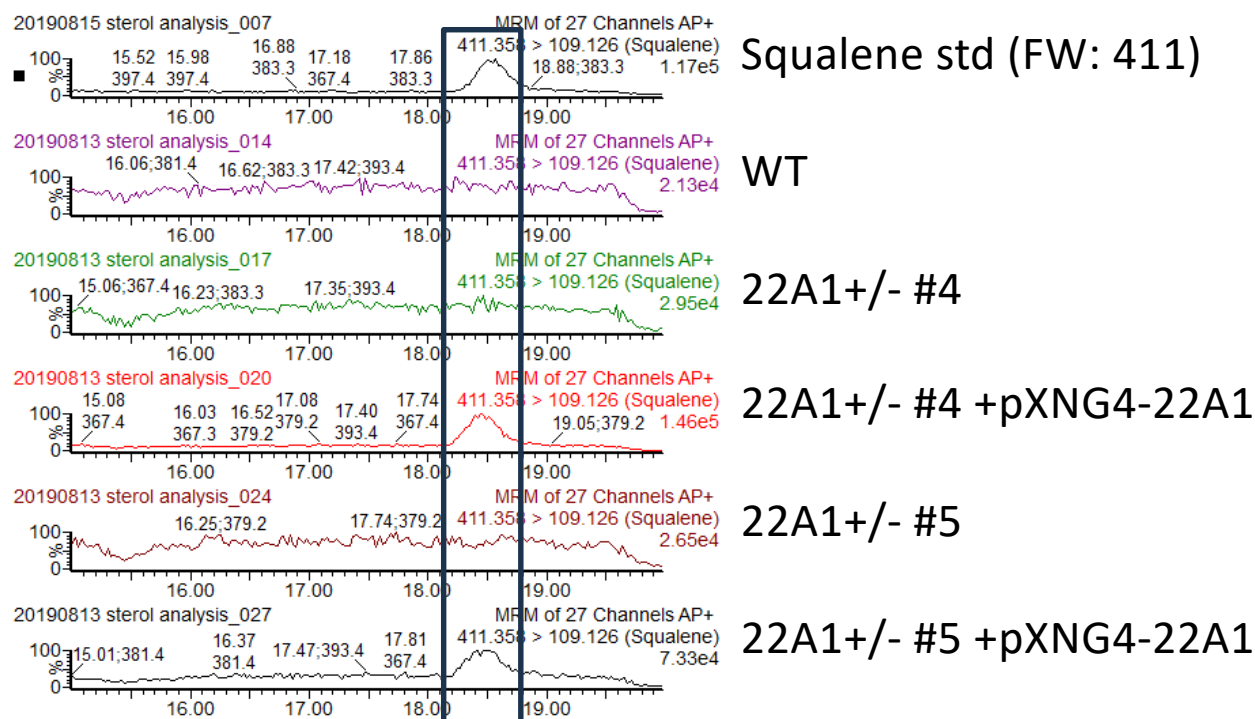
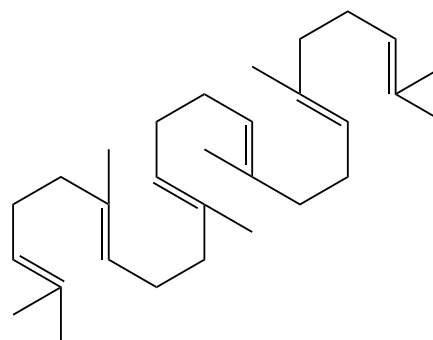
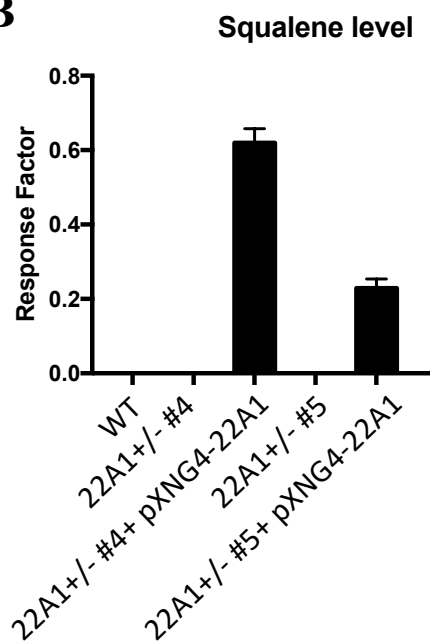
20190813 sterol analysis_027

MRM of 27 Channels AP+
TIC
7.06e6



22A1+/- #5 +pXNG4-
22A1

Figure S8. 22A1 half knockout and overexpression do not affect bulk sterol composition in *L. major*. Partial LC-MS chromatograms of free sterols from WT, 22A1+/- #4, 22A1+/- #4 +pXNG4-22A1, 22A1+/- #5, and 22A1+/- #5 +pXNG4-22A1 promastigotes.

A**B**

Squalene

Figure S9. Squalene were detected in 22A1 overexpressing cells. (A) Partial GC chromatograms of total lipids from WT, 22A1+/- #4, 22A1+/- #4 + pXNG4-22A1, 22A1+/- #5, and 22A1+/- #5 + pXNG4-22A1 promastigotes showed the presence of squalene in the boxed region. (B) The relative abundance of squalene was determined.

Table S1. List of oligonucleotides used in this study

Oligo #	Name	Purpose	Sequence
709	28s rRNA_forward	For qRT-PCR analysis of 28S rRNA	AAGATGGACCGGCTCTAGT
710	28s rRNA_reverse	For qRT-PCR analysis of 28S rRNA	ATCCTTCCCCGCTCCAGTAT
359	5' ORF C4 methyl oxidase	To amplify the ERG25 ORF	GATCATagatct TCGATGGTGCTCGCGAAAAAGC
360	3' ORF C4 methyl oxidase	To amplify the ERG25 ORF	GATCATagatct CCTTCACGAACACGCGCGC
361	for 5' UTR C4 methyl oxidase	To amplify the 5'- flanking sequence of ERG25	GATCATgaattc TCAATGTGACTTGCGTTTAACTCCCTCC
362	rev 5' UTR C4 methyl oxidase	To amplify the 5'- flanking sequence of ERG25	GTCAGCggatccGATCTAactagt CGAGTGCCGAGGGATAAGGTGGAG
363	for 3' UTR C4 methyl oxidase	To amplify the 3'- flanking sequence of ERG25	GATCATggatcc AGGTGCTCAGGCACAAAACGACTTC
364	rev 3' UTR C4 methyl oxidase	To amplify the 3'- flanking sequence of ERG25	GATCATaagett TCAACAGTTATGCGCATACATATTCACAG
602	Erg25_probe_F.P	To generate the ERG25 probe for Southern blot	GCGCACACCTTCAATGTG
603	Erg25_probe_R.P	To generate the ERG25 probe for Southern blot	CGAACACGACGTGTA
631	Erg25 probe out of 5'UTR F	To generate the ERG25 probe for Southern blot	CAGAGCTAAGTCGTGCTG
632	Erg25 probe out of 5'UTR R	To generate the ERG25 probe for Southern blot	GCTGTTCTGCGGATCATC
740	LmCYP-5'UTR-R	To amplify the 5'- flanking sequence of Lm22A1	GTC GCT actagt GTG AAA TGG GCG ACA AGA G
757	LmCYP-5'UTR-F	To amplify the 5'- flanking sequence of Lm22A1	CAT GCT gaattc CCT TTC TCT GTG CAC CCT TC
741	LmCYP-3'UTR-F	To amplify the 3'- flanking sequence of Lm22A1	CGG ACG actagt GGC TAG agatct AGC GAG CTG AGA ATT GCC
742	LmCYP-3'UTR-R	To amplify the 3'- flanking sequence of Lm22A1	GTC AAG aagett GCA TTC ACG CAC GCT CTC
754	LmCYPORF-F	To amplify the Lm22A1 ORF	TCA GTA ggatcc ACC ATG GTG TTT GAC ACC GAC
755	LmCYPORF-R	To amplify the Lm22A1 ORF	GCG CT ggatcc TTACATGAACAACCTTGTTTCG
756	LmCYPORF-nostop-R	To amplify the Lm22A1-GFP fusion	GCG GTC gatatc CAT GAA CAA CTT GTT CGG
768	Lm22A1 ORF probe FP	To generate the Lm22A1 and Ld22A1 probe for Southern blot	GTC GCA TGC GAT GCG CAT C
769	Lm22A1 ORF probe RP	To generate the Lm22A1 and Ld22A1 probe for Southern blot	GTA GCG AGT CTG CAC GGT GC
770	Lm22A1 5'UTR probe FP	To generate the Lm22A1 probe for Southern blot	CAG TGA GCG AGA GAA CGA G
771	Lm22A1 5'UTR probe RP	To generate the Lm22A1 probe for Southern blot	CCT ACA TGA GCG TAG GAC AC