

**Figure S1** Map of sampling sites in the northwestern area of Yunnan province, China (A). In the picture B, the green dot is sampling site for *Astragalus acaulis* (Aa) growing in high altitude grassland of 4,353 m; as shown in the yellow dot, *A. ernestii* (AE) grows in the steep slopes of flowstone beaches of 4,048 m; The blue dot shows the *A. forrestii* (Af) survival environment, surrounded by more pine trees and deciduous foliage of 3,225 m [1].



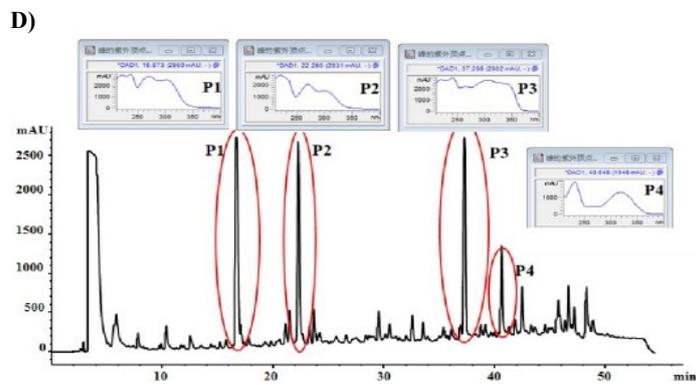
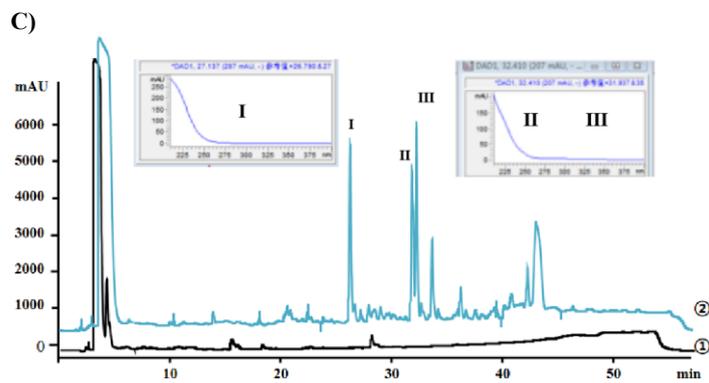
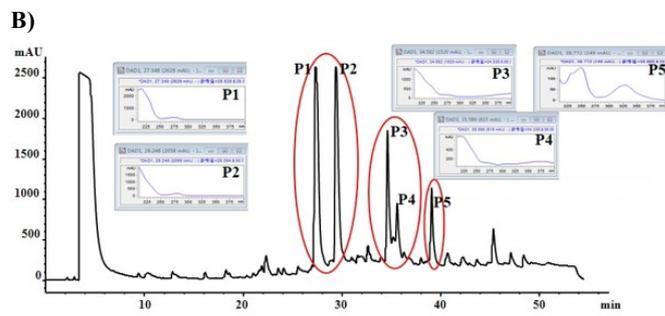
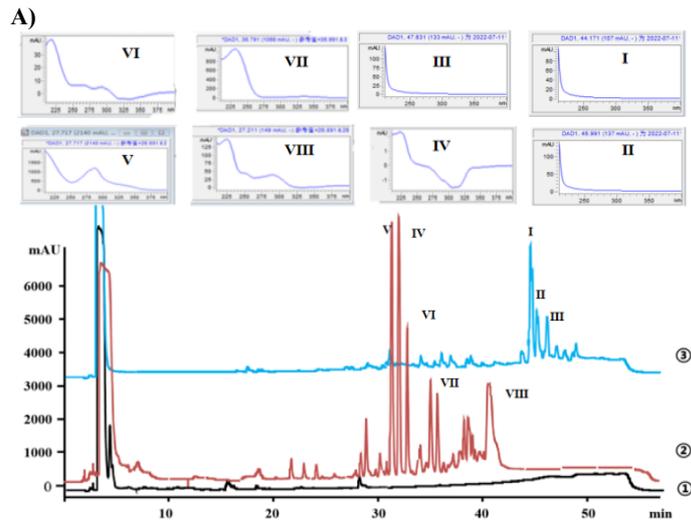
**Figure S2** The plants of *Astragalus acaulis* (Aa) grows in alpine grassland (A), *A. forrestii* (Af) is surrounded by more pine trees and deciduous foliage (B), *A. ernestii* (Ae) grows in the steep slopes of flowstone beaches (C) [1].

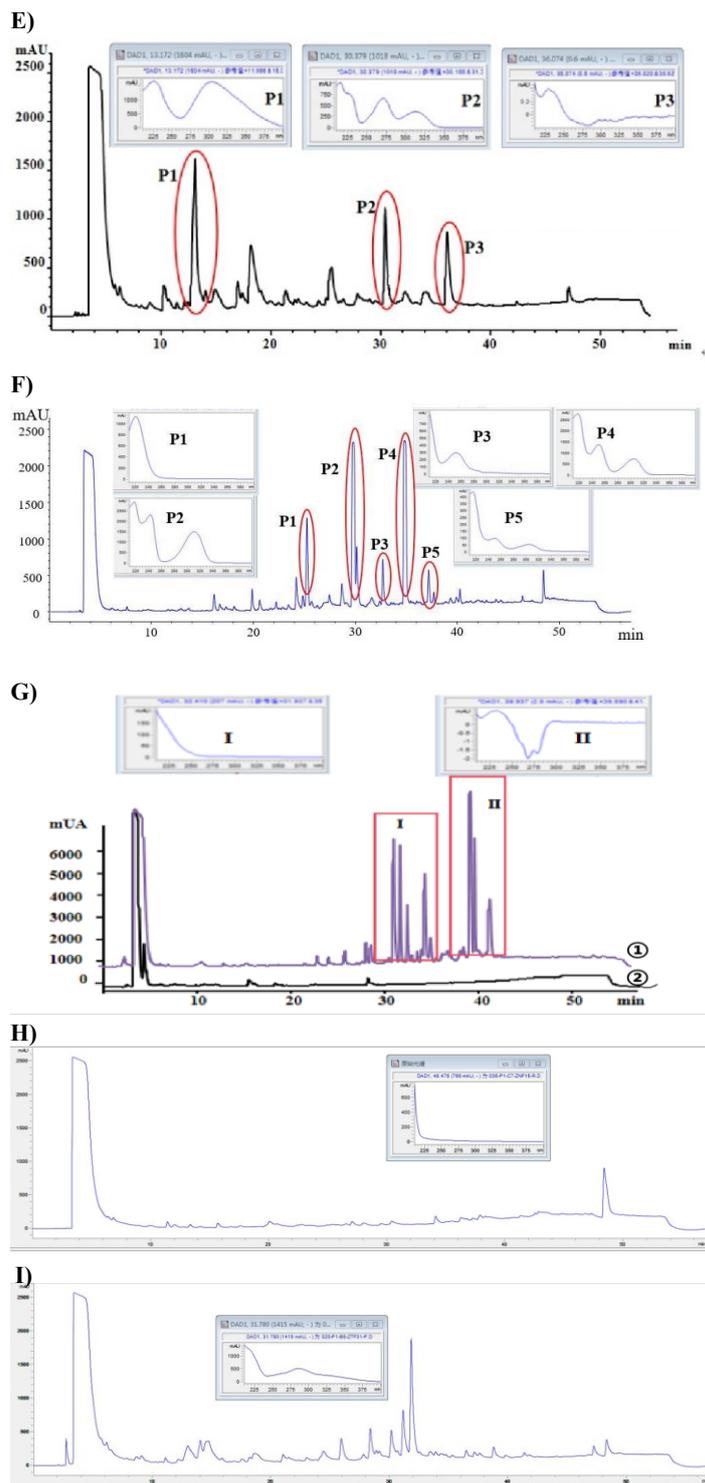
**Table S1** A total of 7, 2 and 4 unique genera identified in the rhizosphere fungal community of *Astragalus acaulis* (Aa), *A. forrestii* (Af) and *A. ernestii* (Ae), respectively.

<b>the name of fungal genera of Aa rhizosphere</b>	<b>the name of fungal genera of Af rhizosphere</b>	<b>the name of fungal genera of Ae rhizosphere</b>
<i>Truncatella</i>	<i>Lecanicillium</i>	<i>Leptosphaeria</i>
<i>Cordyceps</i>	<i>Umbelopsis</i>	<i>Fusarium</i>
<i>Exophiala</i>		<i>Nemania</i> ,
<i>Beauveria</i>		<i>Aporospora</i>
<i>Samsoniella</i>		
<i>Neonectria</i>		
<i>Pseudogymnoascus</i>		

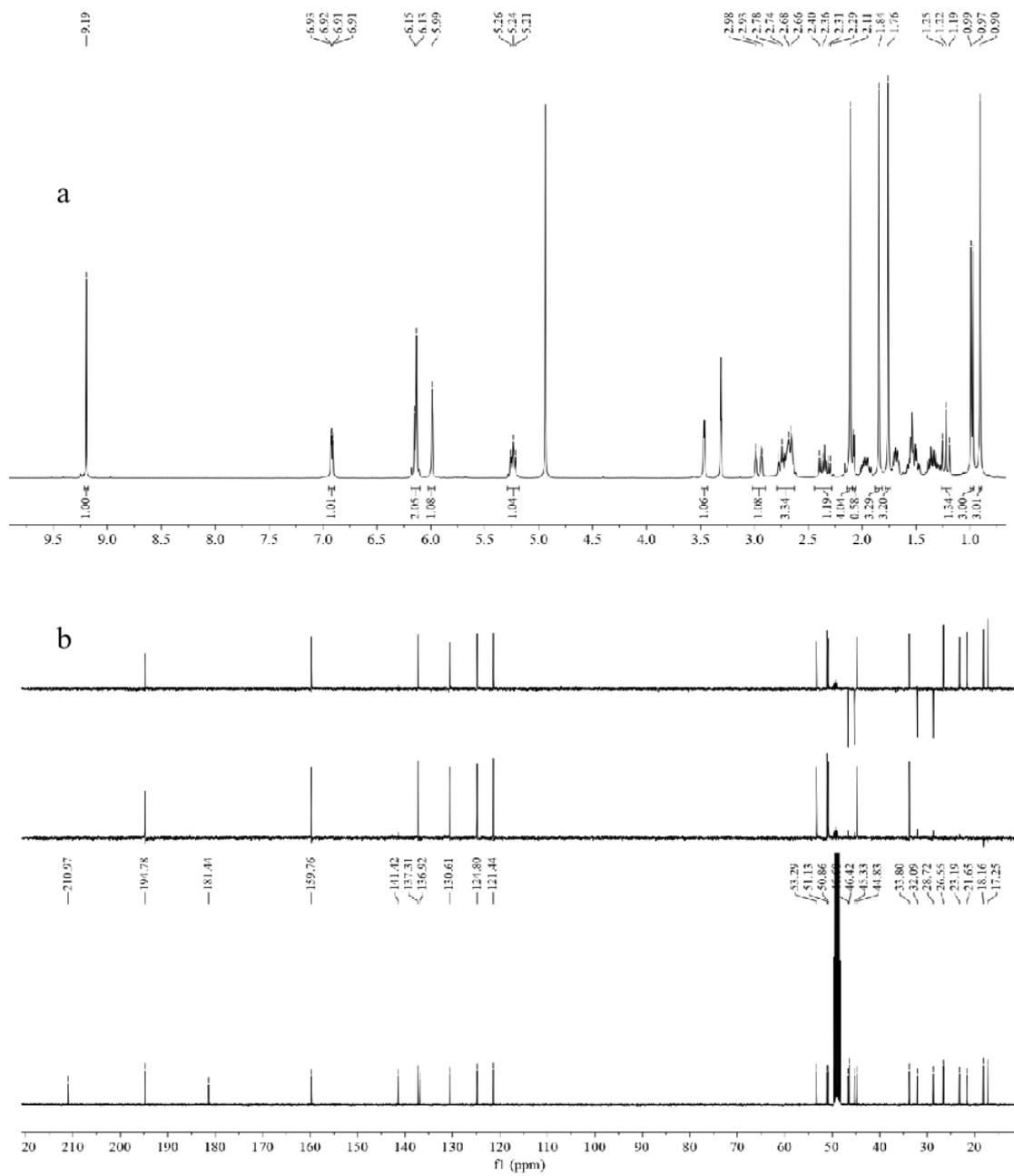
**Table S2** A total of 10, 15 and 7 unique species identified in the rhizosphere fungal community of *Astragalus acaulis* (Aa), *A. forrestii* (Af) and *A. ernestii* (Ae), respectively.

<b>the name of fungal species of Aa rhizosphere</b>	<b>the name of fungal species of Af rhizosphere</b>	<b>the name of fungal species of Ae rhizosphere</b>
<i>Penicillium wellingtonense</i>	<i>Penicillium vasconiae</i>	<i>Penicillium brevicompactum</i>
<i>Aspergillus calidoustus</i>	<i>P. suaveolens</i>	<i>Aspergillus niger</i>
<i>Mortierella clonocystis</i>	<i>P. thomii</i>	<i>Trichoderma longibrachiatum</i>
<i>Truncatella angustata</i>	<i>Aspergillus versicolor</i>	<i>Leptosphaeria sclerotioides</i>
<i>Cordyceps farinosa</i>	<i>A. tabacinus</i>	<i>Fusarium solani</i>
<i>Exophiala tremulae</i>	<i>Mortierella minutissima</i>	<i>Nemania diffusa</i>
<i>Beauveria pseudobassiana</i>	<i>M. verticillata</i>	<i>Aporospora terricola</i>
<i>Samsoniella hepiali</i>	<i>Trichoderma paraviridescens</i>	
<i>Neonectria radicola</i>	<i>T. koningiopsis</i>	
<i>Pseudogymnoascus roseus</i>	<i>T. viride</i>	
	<i>T. longipile</i>	
	<i>Lecanicillium aphanocladii</i>	
	<i>Umbelopsis vinacea</i>	
	<i>U. ramanniana</i>	
	<i>U. nana</i>	

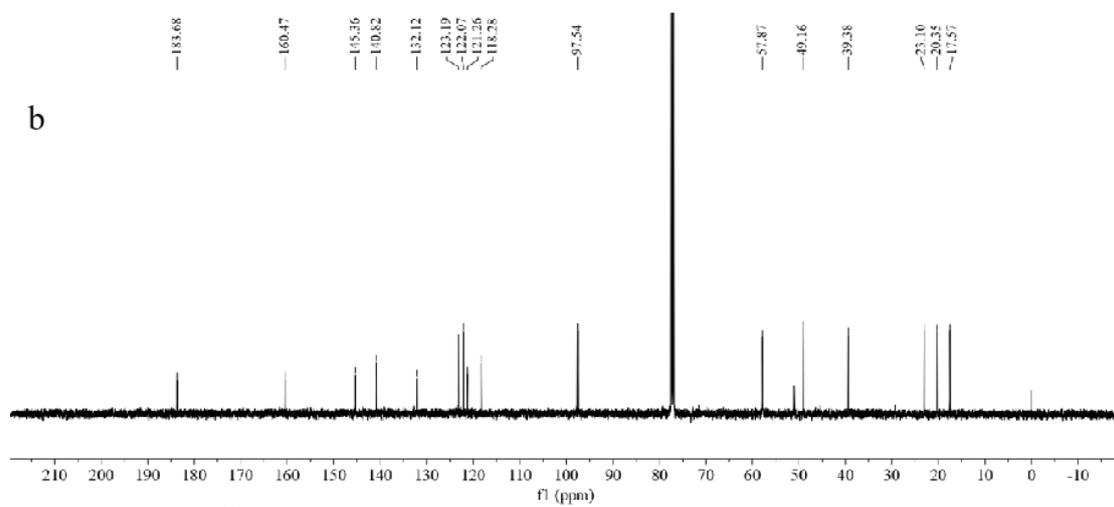
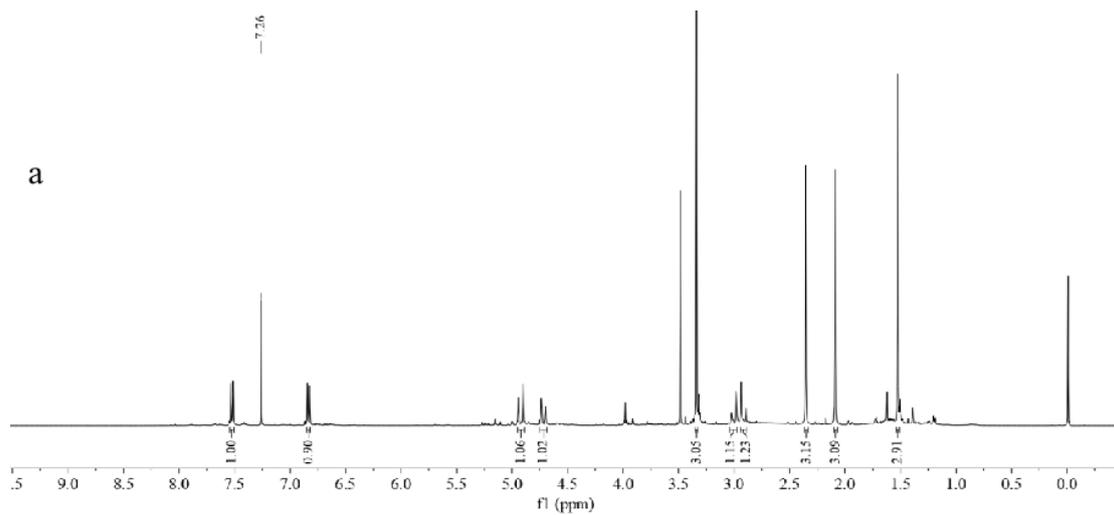




**Figure S3** HPLC spectra of the crude ethyl acetate (EA) extracts of fermented substrates of *Aspergillus calidoustus* AA12 (A③) and other nine target fungal strains of *A. fumigatus* AA15 (A②), *A. tabacinus* AF16 (B), *Penicillium polonicum* AA17 (C②), *Penicillium vasconiae* AF5 (D), *P. glabrum* AF3-1 (E), *P. brevicompactum* AE7 (F), *Cordyceps farinosa* AA8 (G), *Lecanicillium aphanocladii* AF15 (H), and *Umbelopsis nana* AF31 (I).



**Figure S4**  $^1\text{H}$  (a) and  $^{13}\text{C}$  (b) NMR spectrum of 6-epi-ophiobolin G (**1**).



**Figure S5**  $^1\text{H}$  (a) and  $^{13}\text{C}$  (b) NMR spectrum of penicisochroman A (**2**).

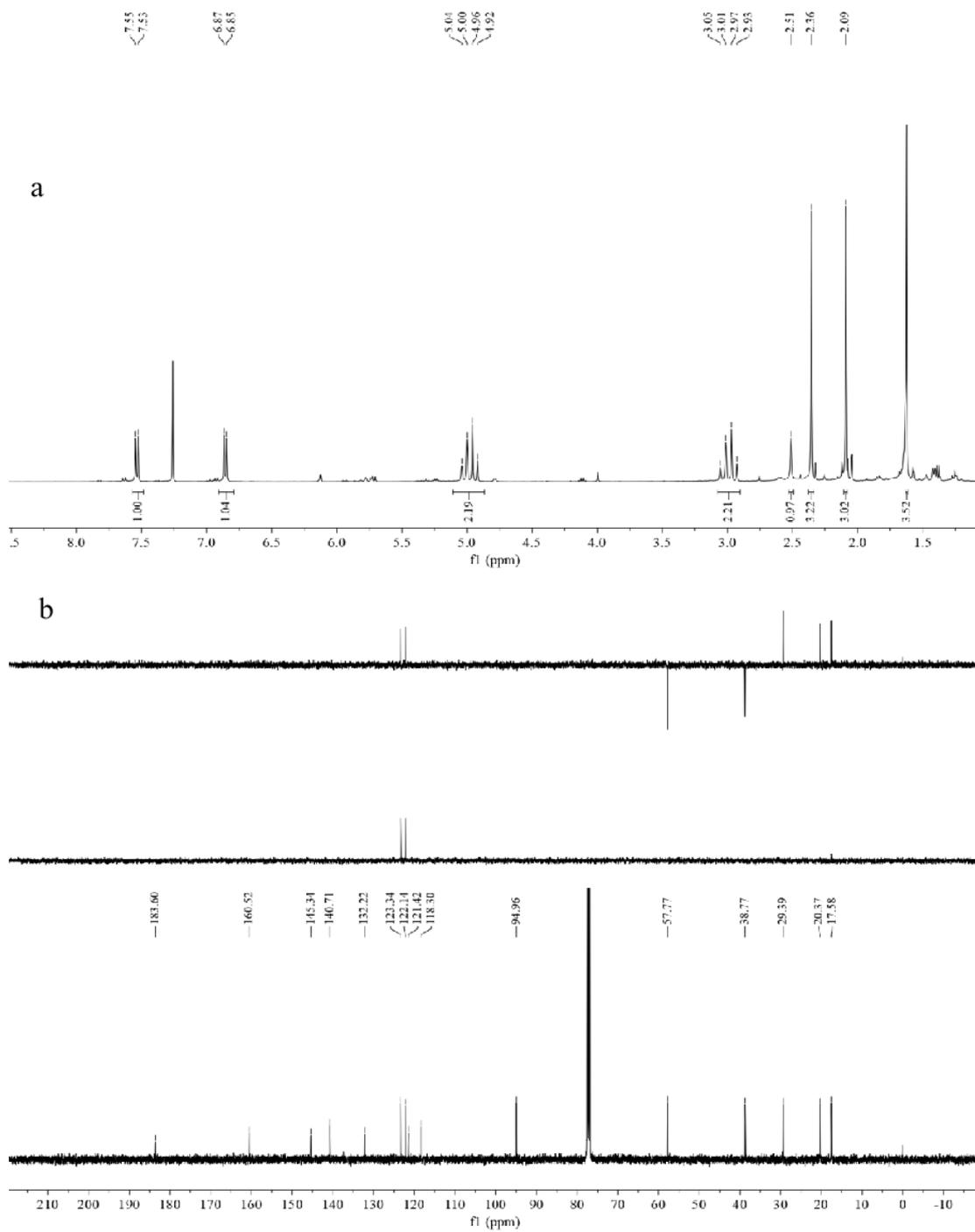
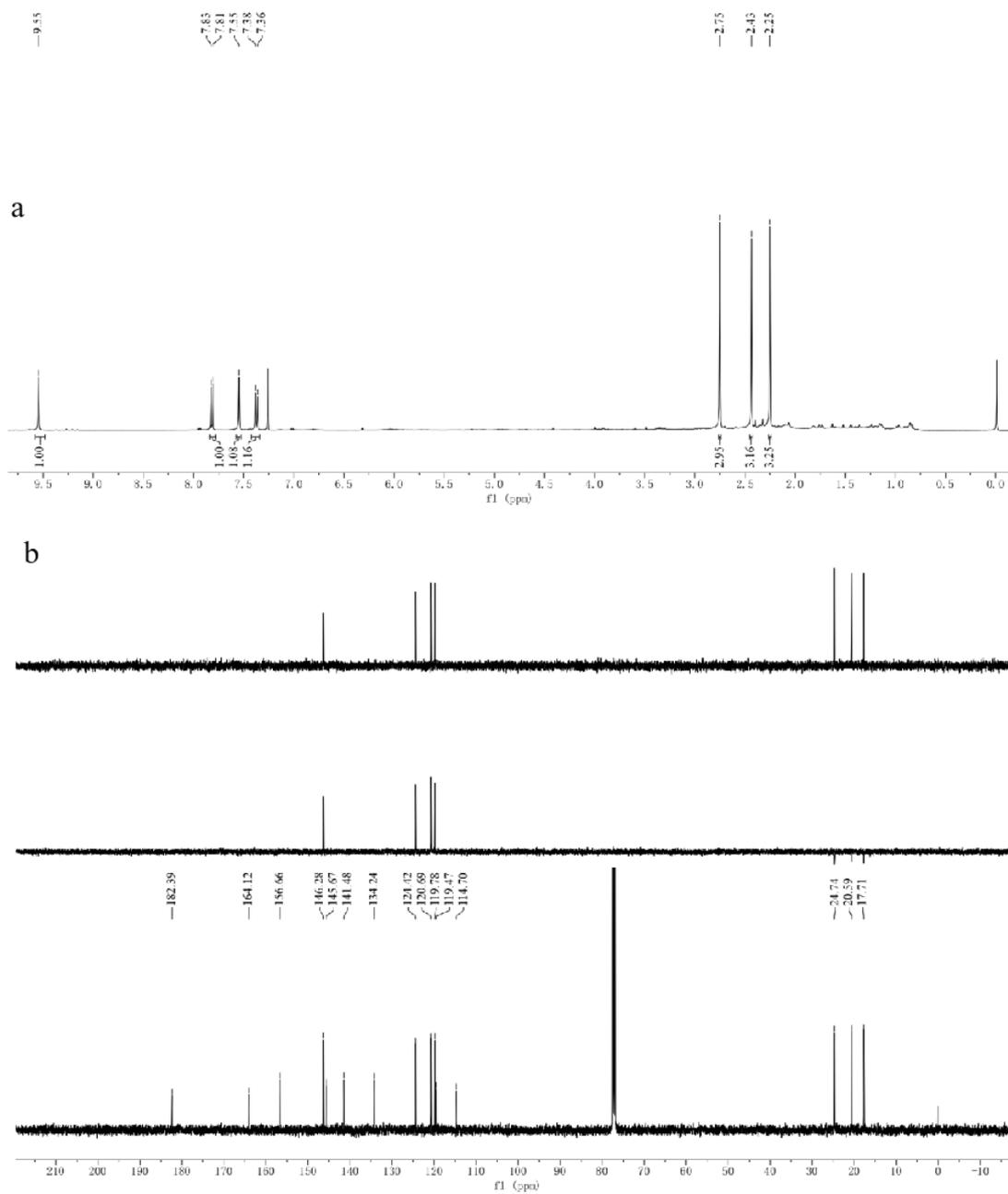
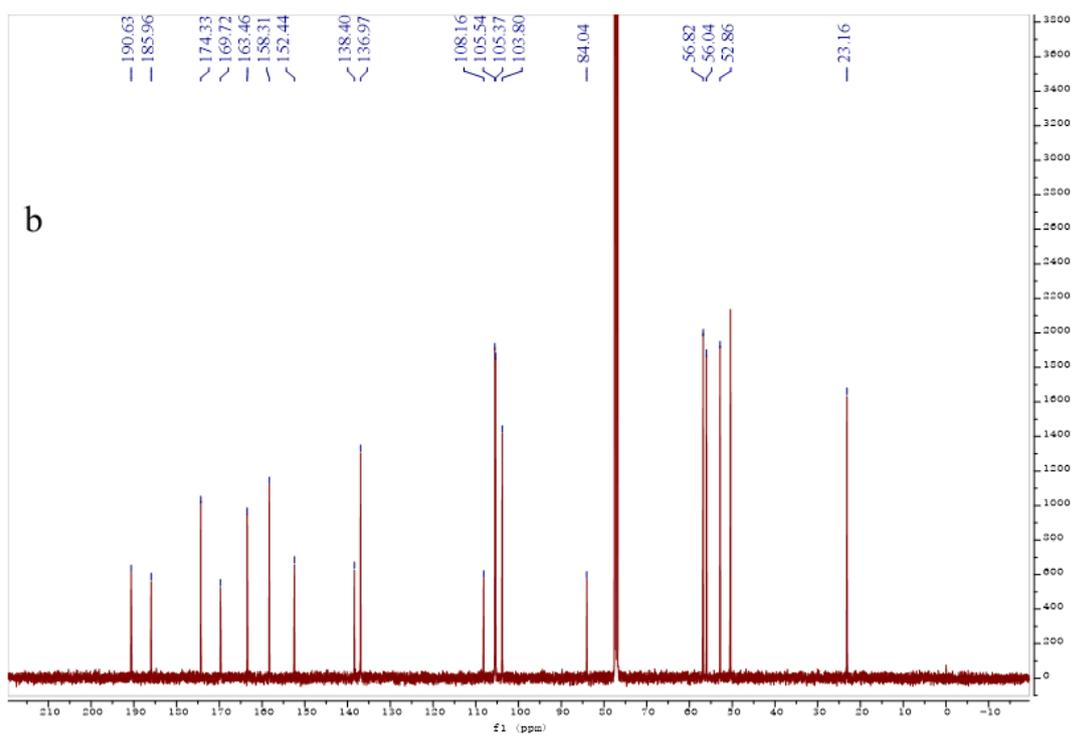
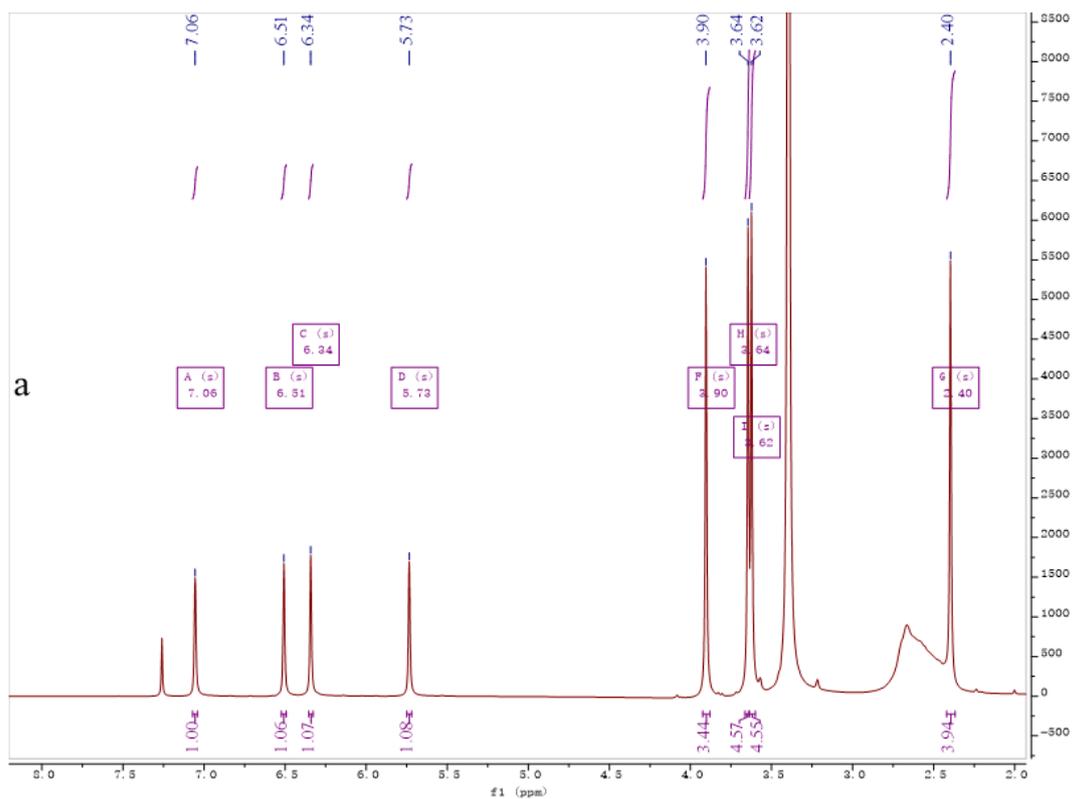


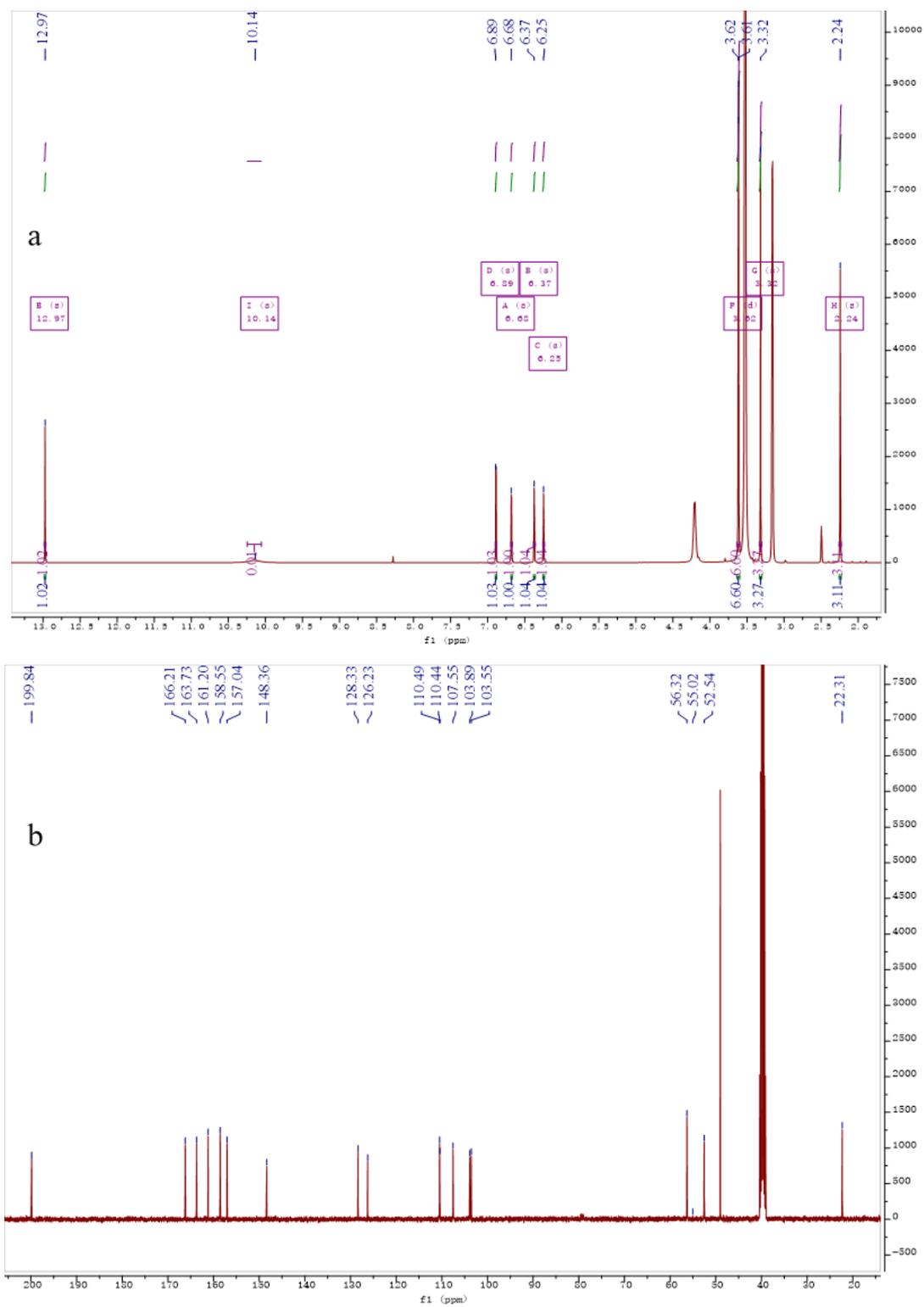
Figure S6  $^1\text{H}$  (a) and  $^{13}\text{C}$  (b) NMR spectrum of pergillin (3).



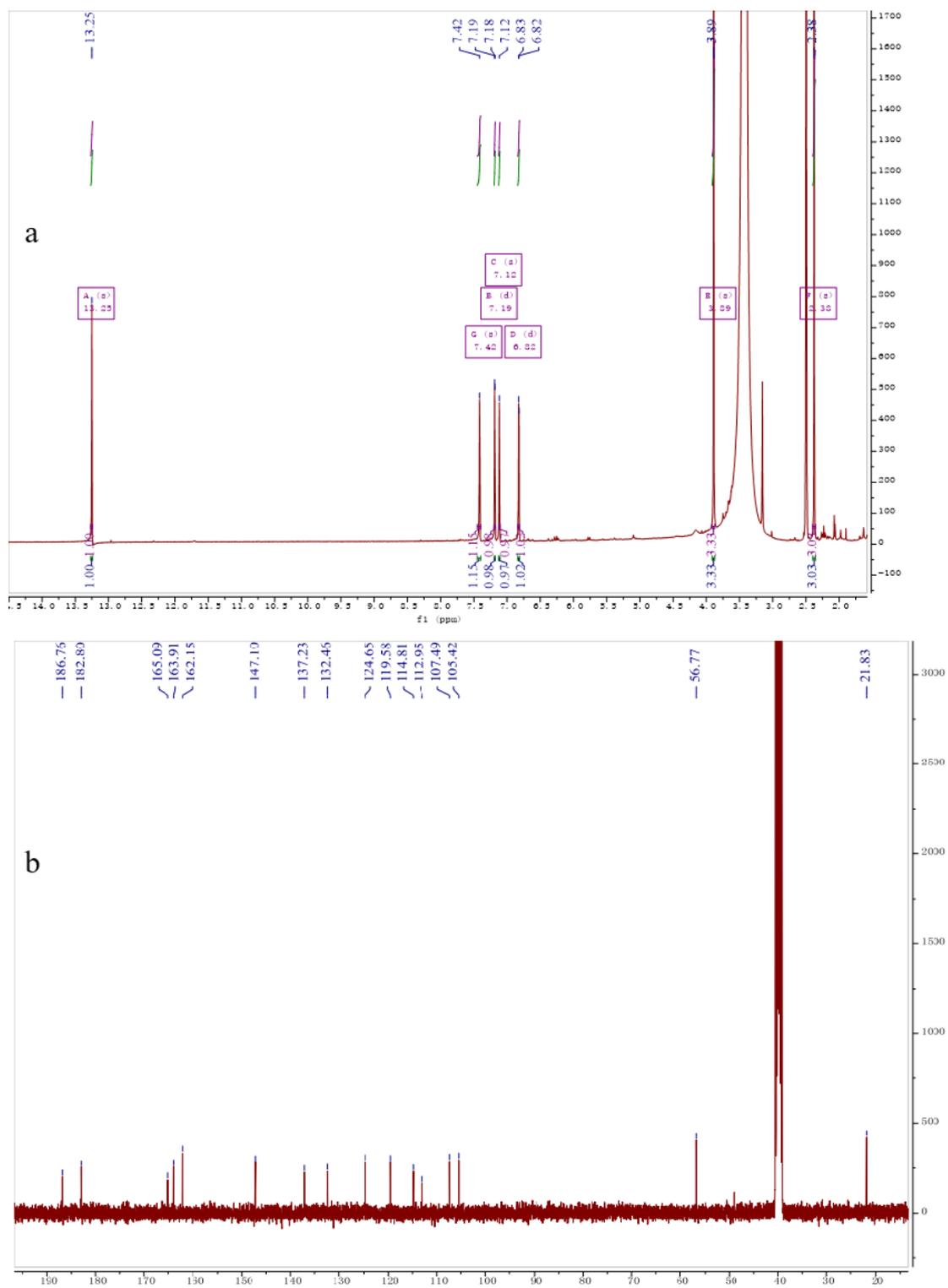
**Figure S7**  $^1\text{H}$  (a) and  $^{13}\text{C}$  (b) NMR spectrum of 7-methyl-2-(1-methylethylidene)-furo[3,2-H]isoquinoline-3-one (4).



**Figure S8**  $^1\text{H}$  (a) and  $^{13}\text{C}$  (b) NMR spectrum of trypacidin (**5**).



**Figure S9**  $^1\text{H}$  (a) and  $^{13}\text{C}$  (b) NMR spectrum of 1,2-seco-trypacidin (**6**).



**Figure S10**  $^1\text{H}$  (a) and  $^{13}\text{C}$  (b) NMR spectrum of questin (7).

**Table S3** <sup>1</sup>H and <sup>13</sup>C NMR data of compounds **1** and **7** (δ in ppm).

No.	<b>1</b> (In CD <sub>3</sub> OD)		<b>7</b> (In DMSO-d <sub>6</sub> )	
	δ <sub>H</sub> <sup>a</sup>	δ <sub>C</sub> <sup>b</sup>	δ <sub>H</sub> <sup>a</sup>	δ <sub>C</sub> <sup>b</sup>
1	1.19 t	45.33 CH <sub>2</sub>		162.15 C
2	2.66 m	50.86 CH <sub>2</sub>	7.12 s	124.65 CH
3		181.44 C		147.10 C
3-Me			2.38 s	21.83 CH <sub>3</sub>
4	5.99 s	130.61 CH	7.42 s	119.58 CH
5		210.97 C	7.19 s	107.49 CH
6	3.46 d	51.13 CH		165.09 C
7		141.42 C	6.82 s	105.42 CH
8	6.91 d	159.76 CH		163.91 C
8-OMe			3.89 s	56.77 CH <sub>3</sub>
9	2.93 d	32.09 CH <sub>2</sub>		186.76 C
10	2.61 m	44.83 CH		182.80 C
11		46.68 C		137.23 C
12	1.49 m; 1.52 m	46.42 CH <sub>2</sub>		112.95 C
13	1.28 m	28.72 CH <sub>2</sub>		114.81 C
14	1.87 m	50.29 CH		132.46 C
15	2.68 m	33.80 CH		
16	5.21 t	137.31 CH		
17	6.13 t	124.80 CH		
18	5.99 t	121.44 CH		
19		136.92 C		
20	2.11 s	17.20 CH <sub>3</sub>		
21	9.19 s	194.78 CH		
22	0.90 s	23.19 CH <sub>3</sub>		
23	0.97 d	21.65 CH <sub>3</sub>		
24	1.76 s	18.16 CH <sub>3</sub>		
25	1.84 s	26.55 CH <sub>3</sub>		

a Recorded at 400 MHz; b Recorded at 100 MHz.

**Table S4** <sup>1</sup>H and <sup>13</sup>C NMR data of compounds **2–4** (δ in ppm).

No.	<b>2</b> (In CDCl <sub>3</sub> )		<b>3</b> (In CDCl <sub>3</sub> )		<b>4</b> (In CDCl <sub>3</sub> )	
	δ <sub>H</sub> <sup>a</sup>	δ <sub>C</sub> <sup>b</sup>	δ <sub>H</sub> <sup>a</sup>	δ <sub>C</sub> <sup>b</sup>	δ <sub>H</sub> <sup>a</sup>	δ <sub>C</sub> <sup>b</sup>
2		145.34 C		145.34 C		145.67 C
3		183.59 C		183.40 C		182.39 C
3a		121.28 C		121.42 C		119.47 C
4	7.53 d	122.02 CH	7.53 d	122.14 CH	7.78 d	124.42 CH
5	6.85 d	123.16 CH	6.87 d	123.34 CH	7.36 d	119.78 CH
5a		140.77 C		140.71 C		141.48 C
6	3.00 dd	39.38 CH <sub>2</sub>	2.93 dd	38.77 CH <sub>2</sub>	7.55 s	120.69 CH
7		97.50 C		94.96 C		156.66 C
9	4.93 dd	57.86 CH <sub>2</sub>	4.92 dd	57.77 CH <sub>2</sub>	9.51 s	146.28 CH
9a		118.29 C		118.30 C		114.70 C
9b		160.45 C		160.52 C		164.12 C
10	1.53 s	23.10 CH <sub>3</sub>	1.63 s	29.39 CH <sub>3</sub>	2.75 s	24.74 CH <sub>3</sub>
11		131.85 C		132.22 C		134.22 C
12	2.09 s	20.31 CH <sub>3</sub>	2.09 s	17.58 CH <sub>3</sub>	2.43 s	17.71 CH <sub>3</sub>
13	2.36 s	17.53 CH <sub>3</sub>	2.36 s	20.37 CH <sub>3</sub>	2.55 s	20.59 CH <sub>3</sub>
7-OMe	3.55 s	49.13 CH <sub>3</sub>				

a Recorded at 400 MHz; b Recorded at 100 MHz.

**Table S5** <sup>1</sup>H and <sup>13</sup>C NMR data NMR data of compounds **5** and **6** (δ in ppm).

No.	<b>5</b> (In CDCl <sub>3</sub> )		<b>6</b> (In DMSO-d <sub>6</sub> )	
	δ <sub>H</sub> <sup>a</sup>	δ <sub>C</sub> <sup>b</sup>	δ <sub>H</sub> <sup>a</sup>	δ <sub>C</sub> <sup>b</sup>
2		84.04 C		126.23 C
3		190.63 C		199.84 C
3a		108.16 C		110.44 C
4		158.31 C		161.20 C
5	6.34 s	105.37 CH	6.37 s	110.49 CH
6		152.44 C		148.36 C
6a	2.40 s	23.16 CH <sub>3</sub>	2.24 s	22.31 CH <sub>3</sub>
7	6.51 s	105.54 CH	6.68 s	103.89 CH
7a		174.33 C		163.73 C
1'		138.40 C		128.33 C
2'	7.06 s	136.97 CH	6.89 s	107.55 CH
3'		185.96 C		158.55 C
4'	5.73 s	103.80 CH	6.25 s	103.55 CH
5'		169.72 C		157.04 C
1'-COOMe		163.46 C		166.21 C
4-OMe	3.90 s	56.04 CH <sub>3</sub>	3.32 s	52.54 CH <sub>3</sub>
5'-OMe	3.62 s	56.82 CH <sub>3</sub>	3.62 s	56.32 CH <sub>3</sub>
1'-COOMe	3.64 s	52.86 CH <sub>3</sub>	3.61 s	55.02 CH <sub>3</sub>

a Recorded at 400 MHz; b Recorded at 100 MHz.

## References

1. Ding, J.J.; Zhou, G.J.; Chen, X.J.; Xu, W.; Gao, X.M.; Zhang, Y.Z.; Jiang, B.; Li, H.F.; Wang, K.L. Analysis of microbial diversity and community structure of rhizosphere soil of three *Astragalus* species grown in special high-cold environment of northwestern Yunnan, China. *Microorganisms* **2024**, *12*, 539.