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Review

Viruses Identified in Shrews (Soricidae) and Their Biomedical Significance

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Abstract: Shrews (Soricidae) are common small wild mammals. Some species of shrews, such as Asian house shrews (*Suncus murinus*), have a significant overlap in their habitats with humans and domestic animals. Currently, over 190 species of viruses in 32 families, including *Adenoviridae*, *Arenaviridae*, *Arteriviridae*, *Astroviridae*, *Anelloviridae*, *Bornaviridae*, *Caliciviridae*, *Chuviridae*, *Coronaviridae*, *Filoviridae*, *Flaviviridae*, *Hantaviridae*, *Hepadnaviridae*, *Hepeviridae*, *Nairoviridae*, *Nodaviridae*, *Orthoherpesviridae*, *Orthomyxoviridae*, *Paramyxoviridae*, *Parvoviridae*, *Phenuiviridae*, *Picobirnaviridae*, *Picornaviridae*, *Polyomaviridae*, *Poxviridae*, *Rhabdoviridae*, *Sedoreoviridae*, *Spinareoviridae*, and three unclassified families, have been identified in shrews. Diverse shrew viruses, such as Borna disease virus 1, Langya virus, and severe fever with thrombocytopenia syndrome virus, cause diseases in humans and/or domestic animals, posing significant threats to public health and animal health. This review compiled fundamental information about shrews and provided a comprehensive summary of the viruses that have been detected in shrews, with the aim of facilitating a deep understanding of shrews and the diversity, epidemiology, and risks of their viruses.

Keywords: shrew; virus; diversity; public health; epidemiology; risk

1. Introduction of Shrews

Shrews, which are usually termed long-nosed mice, can be found in a variety of habitats, including forests, grasslands, deserts, and wetlands. Some species, such as American water shrews (*Sorex palustris*), are aquatic and can be found in or near water.

Asian house shrews (*Suncus murinus*) are the largest shrews, attaining a length of approximately 15 cm and a weight of around 100 g, while Etruscan shrews (*Suncus etruscus*) could represent the smallest extant terrestrial mammals, measuring a mere 3.5 cm in length and weighing approximately 1.8 g [1].

Shrews, hedgehogs, and moles share the same order, Eulipotyphla. They belong to the families Soricidae, Erinaceidae, and Talpidae, respectively. Eulipotyphla encompasses 593 species, second only to Rodentia (2,398 species) and Chiroptera (1,449 species) in mammals. These three orders comprise about 9.5%, 38.5%, and 23.2% of all mammalian species, respectively.

Of the known 163 mammalian families, Soricidae encompasses 488 species, only less than Muridae (775 species), Cricetidae (705 species), and Vespertilionidae (520). The 488 species of Soricidae belong to three subfamilies and 28 genera [2]. The three subfamilies are Crocidurinae (white-toothed shrews), Soricinae (red-toothed shrews), and Myosoricinae (African shrews). These three subfamilies comprise 10 genera with 263 species, 15 genera with 200 species, and 3 genera with 25 species, respectively. The genus *Crociodura* is the largest genus in Crocidurinae, covering 220 species. The genus *Sorex* is the largest genus in Soricinae, covering 89 species [2].

Notably, tree shrews in the order Scandentia, otter shrews in the order Afrosoricida, elephant shrews in the order Macroscelidea, and marsupial shrews in the order Dasyuromorphia are not shrews, because they do not fall within the order Eulipotyphla [3]. West Indies shrews were in the order Eulipotyphla, but they were in the family Nesophontidae and have been extinct.

Shrews are distributed across major tropical and temperate landmasses on all continents except Oceania and Antarctica (Table 1). *Suncus murinus* was introduced on Guam, a part of Oceania, following World War II. In South America, shrews are confined to the northern Andes region and were introduced during the Great American Interchange.

Table 1. Geographic distribution and species number of 26 shrew genera.

Subfamily	Genus	Species	Geographical distribution
Crocidurinae	<i>Crocidura</i>	220 species	Asia, Africa, and Europe
	<i>Suncus</i>	18 species	Asia, Africa, Europe, and Oceania
	<i>Sylvisorex</i>	15 species	Africa
	<i>Diplomesodon</i>	1 species	Europe
	<i>Feroculus</i>	1 species	Asia
	<i>Paracrocidura</i>	2 species	Africa
	<i>Ruwenzorisorex</i>	1 species	Africa
	<i>Scutisorex</i>	2 species	Africa
	<i>Solisorex</i>	1 species	Asia
	<i>Palawanosorex</i>	2 species	Asia
	<i>Sorex</i>	89 species	Asia, Europe, North America and South America
	<i>Anourosorex</i>	4 species	Asia
	<i>Blarinella</i>	2 species	Asia
	<i>Neomys</i>	4 species	Europe and Asia
Soricinae	<i>Blarina</i>	5 species	North America
	<i>Cryptotis</i>	54 species	North America and South America
	<i>Chimarrogale</i>	8 species	Asia
	<i>Chodsigoa</i>	10 species	Asia
	<i>Episoriculus</i>	7 species	Asia
	<i>Nectogale</i>	2 species	Asia
	<i>Soriculus</i>	5 species	Asia
	<i>Megasorex</i>	1 species	North America
	<i>Notiosorex</i>	5 species	North America
	<i>Paracrocidura</i>	3 species	Asia
	<i>Pseudosoriculus</i>	1 species	Asia
	<i>Myosorex</i>	19 species	Africa
Myosoricinae	<i>Surdisorex</i>	3 species	Africa
	<i>Congosorex</i>	3 species	Africa

Shrews have a lifespan of 12 to 30 months, and they exhibit a relatively high metabolic rate, higher than some mammals of similar body sizes. Consequently, shrews have a relatively large food intake. Shrews do not hibernate but can enter a torpid state. During the winter, many shrews undergo significant changes in morphology, with body weight decreasing by 30% to 50% and both skeletal and organ sizes noticeably shrinking [4]. Shrews are mostly found in cool and humid environments, with many being terrestrial, while some are semi-aquatic or burrowing. They may be active during both day and night or primarily nocturnal. Shrews are typically solitary creatures, coming together only for mating purposes.

Female shrews can give birth to as many as 10 litters each year. In tropical regions, they can reproduce year-round, while in temperate regions, they cease reproduction during the winter. The gestation period for shrews ranges from 17 to 32 days. Female shrews can become pregnant again

within one to two days after giving birth and can lactate and nurse their offspring during pregnancy [5].

Certain shrew species secrete venom [6], which contains various compounds. The venom of American short-tailed shrews can kill 200 mice when administered intravenously. Some shrews in China also produce venom [7]. Additionally, similar to bats and toothed whales, two genera, namely *Sorex* (long-tailed shrews) and *Blarina* (blarina shrews), possess echolocation abilities [8]. These two genera include Eurasian water shrews (*Neomys fodiens*), northern short-tailed shrews (*Blarina brevicauda*), *Sorex palustris*, and some others [9].

Diverse viruses in shrews have been identified, and many viruses in shrews have yet to be identified [10]. Currently, as detailed below and summarized in Figure 1, at least 190 species of viruses in 32 families have been identified in shrews, excluding plant viruses or invertebrate viruses that might be detected in shrews due to their existence in the food or surroundings of shrews. These viruses can be classified into negative-sense single-stranded RNA (–ssRNA), positive-sense single-stranded RNA (+ssRNA), double-stranded DNA (dsDNA), single-stranded DNA (ssDNA), and double-stranded RNA (dsRNA) viruses (Figure 1). The relevant sequences of these viruses were provided in the supplementary file (Table S1).

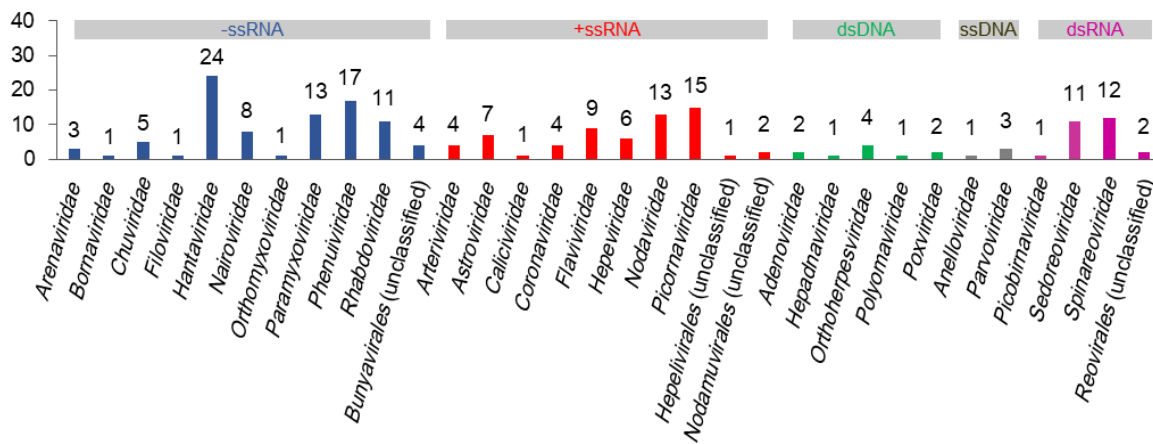


Figure 2. The numbers of virus species in 32 virus families having been identified in shrews.

2. ssRNA Viruses Detected in Shrews

Table 2 shows that 88 species of –ssRNA viruses in shrews have been detected.

Table 2. Detection of –ssRNA viruses in shrews using high-throughput sequencing (HTS), PCR, RT-PCR, serological methods, and/or virus isolation.

Virus family and genus	Virus species and name	Host species	Country	Detection and references
Arenaviridae, Mammarenavirus	Mammarenavirus wenzhouense* (Wēnzhōu virus)	Suncus murinus	China	diverse samples_RT-PCR+isolation [11,12]; multiple organs and feces_HTS [10]
	Mammarenavirus choriomeningitidis* (lymphocytic choriomeningitis virus)	Crociodura goliath	Gabon	sera_serology [13]
	unclassified species (shrew mammarenavirus)	Suncus murinus	China	diverse samples_RT-PCR [14]
Bornaviridae, Orthobornavirus	Orthobornavirus bornaense* (Borna disease virus 1)	Crociodura leucodon	Switzerland; Germany	heart_RT-PCR [15]; multiple organs_HTS [OR713886]#
Chuviridae, unclassified genera	unclassified species (Wufeng shrew chuvirus 1)	Crociodura attenuata; Chodsigoa smithii	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew chuvirus 2)	Chodsigoa smithii	China	multiple organs and feces_HTS [10]

	unclassified species (Wufeng shrew chuvirus 5)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew chuvirus 6)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Crocidura lasiura chuvirus)	<i>Crocidura lasiura</i>	China	Lung_HTS [16]
<i>Filoviridae, Ebolavirus</i>	<i>Oblavirus percae*</i> (Ebola virus)	<i>Sylvisorex ollula</i>	Central African Republic	spleen_RT-PCR [17]
<i>Hantaviridae, Mobatvirus</i>	<i>Mobatvirus lenaense</i> (Lena virus)	<i>Sorex caecutiens; Sorex roboratus</i>	Russia	lung_RT-PCR [18]
	<i>Orthohantavirus artybashense</i> (Artybash virus)	<i>Sorex caecutiens</i>	Russia	lung_RT-PCR [19]
	<i>Orthohantavirus asikkalaense</i> (Asikkala virus)	<i>Sorex minutus</i>	Finland; Czech; Germany	diverse samples_RT-PCR [20]
	<i>Orthohantavirus boweense</i> (Bowé virus)	<i>Crocidura douceti</i>	Guinea	muscle_RT-PCR [21,30]
	<i>Orthohantavirus caobangense</i> (Cao Bằng virus; Wufeng shrew orthohantavirus 1; Wufeng shrew orthohantavirus 2)	<i>Anourosorex squamipes; Chodsigoa smithii</i>	Vietnam; China	diverse samples_RT-PCR [22]; multiple organs and feces_HTS [10]
	<i>Orthohantavirus jejuense</i> (Jeju virus)	<i>Crocidura shantungensis</i>	South Korea	lung+liver_RT-PCR [23]
	<i>Orthohantavirus kenkemeense</i> (Kenkeme virus)	<i>Sorex roboratus</i>	Russia	lung+liver_RT-PCR [24]
	<i>Orthohantavirus seoulense*</i> (Seoul virus)	<i>Crocidura lasiura</i>	China	lung_RT-PCR [25]
<i>Hantaviridae, Orthohantavirus</i>	<i>Orthohantavirus seewisense</i> (Seewis virus)	<i>Sorex araneus; Sorex tundrensis; Sorex daphaenodon; Sorex minutus; Neomys anomalus</i>	Switzerland	lung+liver_RT-PCR [26,27]
	unclassified species (Yákèshí virus)	<i>Sorex unguiculatus</i>	China	diverse samples_RT-PCR [22]
	unclassified species (Altai virus)	<i>Sorex araneus</i>	Russia	lung_RT-PCR+isolation [28]
	unclassified species (Xinyi virus)	<i>Anourosorex yamashinai</i>	China	multiple organs_RT-PCR [29]
	unclassified species (Ash River virus)	<i>Sorex cinereus</i>	United States	lung+liver_RT-PCR [30]
	unclassified species (Jemez Springs virus)	<i>Sorex monticolus</i>	United States	lung+liver_RT-PCR [30]
	unclassified species (Camp Ripley virus)	<i>Blarina brevicauda</i>	United States	lung+liver_RT-PCR [31]
	unclassified species (Boginia virus)	<i>Neomys fodiens</i>	Poland	lung_RT-PCR [32]
	unclassified species (Azagny virus)	<i>Crocidura obscurior</i>	Côte d'Ivoire	lung_RT-PCR [33]
	unclassified species (Qiān Hú Shān virus)	<i>Sorex cylindricauda</i>	China	lung_RT-PCR [34]
	<i>Thottimvirus imjinense</i> (Imjin virus)	<i>Crocidura lasiura</i>	South Korea	diverse samples_RT-PCR +isolation [35]
	<i>Thottopalayam thottimvirus</i> (Thottapalayam virus)	<i>Suncus murinus</i>	China	multiple organs and feces_HTS [10]
<i>Hantaviridae, Thottimvirus</i>	<i>Thottimvirus thottapalayamense</i> (Thottapalayam virus)	<i>Suncus murinus</i>	India	lung_RT-PCR+isolation [36]
	unclassified species (Kilimanjaro virus)	<i>Myosorex geata</i>	Tanzania	multiple organs_RT-PCR [37]
	unclassified species (Uluguru virus)	<i>Myosorex zinki</i>	Tanzania	multiple organs_RT-PCR [37]
<i>Nairoviridae, Orthonairovirus</i>	<i>Orthonairovirus erveense*</i> (Erve virus)	<i>Crocidura russula</i>	France, Germany	multiple organs_RT-PCR [38] [OR713866]†

Nairoviridae, Xinpirovirus	<i>Orthonairovirus lambarenense</i> (Lamusara virus)	<i>Crocidura</i> sp.	Gabon	kidney_RT-PCR [39]
	<i>Orthonairovirus thiaforaense</i> (Thiafora virus)	<i>Crocidura</i> sp.	Senegal	multiple organs_HTS [40]
	unclassified species (Rasenna virus)	<i>Crocidura russula</i>	Germany	multiple organs_HTS [OR713845]*
	unclassified species (Lamgora virus)	<i>Crocidura</i> sp.	Gabon	kidney_RT-PCR [39]
	unclassified species (Cencurut virus)	<i>Suncus murinus</i>	Singapore	lung+ spleen+kidney_HTS [41]
	unclassified species (<i>Crocidura tanakae</i> nairovirus 1)	<i>Crocidura tanakae</i>	China	Lung_HTS [16]
	unclassified species (<i>Crocidura tanakae</i> nairovirus 1)	<i>Crocidura tanakae</i>	China	Lung_HTS [16]
	<i>Alphainfluenzavirus</i> <i>influenzae</i> (influenza A virus)	<i>Anourosorex squamipes</i>	China	Lung_HTS [16]
	unclassified species (Daeryong virus)	<i>Crocidura lasiura</i> ; <i>Crocidura shantungensis</i>	South Korea	kidney_HTS [42]
	unclassified species (Langya virus)	<i>Crocidura lasiura</i> ; <i>Crocidura shantungensis</i>	China	diverse samples_HTS+isolation [43]
Paramyxoviridae, Henipavirus	unclassified species (Meliang virus)	<i>Crocidura grandiceps</i>	Guinea	kidney_HTS [44]
	unclassified species (Denwin virus)	<i>Crocidura russula</i>	Belgium; Germany	kidney_HTS [44] multiple organs_HTS; [OR713882]*
	unclassified species (Hasua virus)	<i>Crocidura russula</i>	Germany	multiple organs_HTS [OR713881]*
	unclassified species (Lechcodon virus)	<i>Crocidura russula</i>	Germany	multiple organs_HTS [OR713879]*
	unclassified species (Resua virus)	<i>Crocidura russula</i>	Germany	multiple organs_HTS [OR713876]*
	unclassified species (Ninorex virus)	<i>Sorex minutus</i>	Belgium	kidney_HTS [45]
	unclassified species (Jinmen shrew henipavirus 1)	<i>Crocidura shantungensis</i> ; <i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Jinmen shrew henipavirus 2)	<i>Crocidura shantungensis</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wenzhou shrew henipavirus 1)	<i>Suncus murinus</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wenzhou shrew henipavirus 2)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
Paramyxoviridae, Jeilongvirus	<i>Jeilongvirus beilongi</i> (Beilong virus)	<i>Crocidura shantungensis</i> ; <i>Suncus murinus</i>	China	diverse samples_RT-PCR [46]
Phenuiviridae, Bandavirus	<i>Bandavirus dabiense</i> * (severe fever with thrombocytopenia syndrome virus)	<i>Suncus murinus</i>	China	serum+spleen_RT-PCR+serology [47]
Phenuiviridae, unclassified genera	unclassified species (Wufeng shrew phenuivirus 1)	<i>Anourosorex squamipes</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew phenuivirus 2)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew phenuivirus 3)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]

	unclassified species (Wufeng shrew phenuivirus 4)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew phenuivirus 5)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew phenuivirus 6)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew phenuivirus 7)	<i>Anourosorex squamipes</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew phenuivirus 8)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew phenuivirus 9)	<i>Anourosorex squamipes</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew phenuivirus 10)	<i>Anourosorex squamipes</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew phenuivirus 11)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew phenuivirus 12)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew phenuivirus 13)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew phenuivirus 14)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew phenuivirus 15)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew phenuivirus 16)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	<i>Rhabdoviridae, Lyssavirus</i> <i>Lyssavirus mokola</i> (Mokola virus)	<i>Crocidura</i> sp.	Nigeria	multiple organs_isolation+serology+sequencing [48]
	<i>Rhabdoviridae, Lyssavirus</i> <i>Lyssavirus rabies</i> (rabies virus)	<i>Suncus murinus</i>	China	Lung_HTS [16]
<i>Rhabdoviridae, Tupavirus</i>	unclassified species (Wufeng shrew tupavirus 1)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew rhabdovirus 2)	<i>Anourosorex squamipes; Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	<u>unclassified species</u> (Wufeng shrew rhabdovirus 4)	<i>Anourosorex squamipes</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew rhabdovirus 6)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew rhabdovirus 7)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew rhabdovirus 10)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew rhabdovirus 15)	<i>Blarinella griselda</i>	China	multiple organs and feces_HTS [10]
<i>Rhabdoviridae, unclassified genera</i>				

Bunyavirales, unclassified families	unclassified species (Wufeng shrew rhabdovirus 18)	<i>Anourosorex squamipes</i>	China	multiple organs and feces_HTS [10]
	unclassified species (<i>Anourosorex squamipes</i> rhabdovirus)	<i>Anourosorex squamipes</i>	China	Lung_HTS [16]
	unclassified species (Wufeng shrew bunyavirus 1)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew bunyavirus 2)	<i>Anourosorex squamipes</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew bunyavirus 3)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	(Wufeng shrew bunyavirus 4)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]

Note: in the tables of this review, those viruses with known pathogenicity to humans or domestic animals are marked with asterisks; those shrew viruses supported by sequences rather than by reports were given the GenBank accession numbers of the relevant sequences and marked with #.

2.1. Arenaviridae Detected in Shrews

The family *Arenaviridae* currently comprises 5 genera and 69 species, which infect mammals, reptiles, or fish [49,50]. Their genomes consist of two or three single-stranded RNA molecules, some of which encode two proteins in non-overlapping ORFs of opposite polarities. Some viruses in this family, such as lymphocytic choriomeningitis virus and Lassa virus, can cause diseases in humans.

Three species in the genus *Mammarenavirus* (*M. wenzhouense*, *M. choriomeningitidis*, and an unclassified species) of this family were identified in shrews. *M. wenzhouense* and an unclassified species in *Suncus murinus* in China were reported in recent years [11,12,14], and *M. choriomeningitidis* in goliath shrews (*Crocidura goliath*) in Gabon was reported in 2021 [13]. Of them, *M. wenzhouense* (Wēnzhōu virus) can infect humans, causing influenza-like symptoms [11,12]. *M. choriomeningitidis* (lymphocytic choriomeningitis virus) can infect humans and primates, causing infection in the central nervous system. This virus can also cause other illnesses, such as conjunctivitis, hepatitis, pneumonia, meningitis, and sepsis [49,50]. Shrews could be accidental hosts or a minor reservoir of these two virus species, which mainly circulate in rodents [49,50].

2.2. Bornaviruses Detected in Shrews

The family *Bornaviridae* currently comprises the genera *Carbovirus*, *Cultervirus*, and *Orthobornavirus*, which infect mammals, birds, reptiles, or fish [51]. Several viruses in this family, such as *Orthobornavirus bornaense* (Borna disease virus 1, BDV-1) and variegated squirrel bornavirus 1, can cause diseases in humans or domestic animals.

BDV-1 in the genus *Orthobornavirus* of this family causes severe T-cell-mediated meningoencephalitis in horses, sheep, and other animals, especially in central Europe. Recent studies have identified white-toothed shrews (*Crocidura leucodon*) as the natural host and reservoir of BDV-1 [52,53]. This virus was detected in white-toothed shrews (*Crocidura russula*) in Germany in recent years (Table 2). BDV-1 can persist in the shrews for long periods without noticeable symptoms and can be shed through shrew saliva, urine, skin swabs, tears, and feces.

2.3. Chuviridae Detected in Shrews

The family *Chuviridae* currently comprises 16 genera and 43 species, which infect arachnids, barnacles, crustaceans, insects, fish, or reptiles in Africa, Asia, Australia, Europe, North America, and South America [54].

Five unclassified species of *Chuviridae* with unknown pathogenicity were identified in Asian grey shrews (*Crocidura attenuata*), Smith's shrews (*Chodsigoa smithii*), and Ussuri white-toothed shrews (*Crocidura lasiura*) in China in recent years [10,16].

2.4. Filoviruses Detected in Shrews

The family *Filoviridae* currently comprises 9 genera and 16 species, which infect mammals, reptiles, or fish [55]. Several viruses in this family, such as Ebola virus and Marburg virus, can cause severe diseases in humans and animals.

Ebola virus in the genus *Orthoebolavirus* of this family is highly pathogenic to humans and has been detected in greater forest shrews (*Sylvisorex ollula*) in the Central African Republic, but shrews could be intermediate or incidental hosts of this virus [56].

2.5. Hantaviruses Detected in Shrews

The family *Hantaviridae* currently comprises 4 subfamilies, 8 genera, and 53 species, which infect mammals, birds, reptiles, or fish [57]. Multiple viruses in this family, such as Hantaan virus and Seoul virus, can cause diseases in humans.

One species in the genus *Mobatvirus* of this family, *M. lenaense* (Lena virus), was discovered in Laxmann's shrews (*Sorex caecutiens*) and flat-skulled shrews (*Sorex roboratus*) captured between 2018 and 2019 in Russia [18].

Diverse species in the genus *Orthohantavirus* of this family, such as *O. artybashense*, *O. asikkalaense*, *O. boweense*, *O. caobangense*, *O. jejuense*, *O. kenkemeense*, *O. seewisense*, *O. seoulense*, and several unclassified species, were detected in multiple countries (Table 2). Of them, *O. seoulense* (Seoul virus) is highly pathogenic to humans with the reservoir of rodents. This virus was discovered in *Crocidura lasiura* captured in 2014 in China [25]. It remains unknown whether shrews are the natural hosts or a reservoir of the virus.

In the genus *Orthohantavirus*, *O. seewisense* (Seewis virus) was first detected in Eurasian common shrews (*Sorex araneus*) captured in 2006 in Switzerland [20]. This virus is widely distributed throughout the geographic ranges of its sorcid hosts, including *Sorex araneus*, tundra shrews (*Sorex tundrensis*), Siberian large-toothed shrews (*Sorex daphaenodon*), and Mediterranean water shrews (*Neomys anomalus*). In addition, some genetic variants of this virus, termed Artybash virus and Amga virus, were detected in *Sorex caecutiens* [26]. *O. caobangense* (Cao Bằng virus) was first detected in the lung tissue of Chinese mole shrews (*Anourosorex squamipes*) captured in 2006 in Vietnam [58]. *O. asikkalaense* (Asikkala virus) was first discovered in Eurasian pygmy shrews (*Sorex minutus*) captured in Finland [20]. Subsequently, this virus was identified in shrews in Czech and Germany. *O. boweense* (Bowé virus) was discovered in the muscle tissue of Doucet's musk shrews (*Crocidura douceti*) captured in 2012 in southwestern Guinea [21]. *O. jejuense* (Jeju virus) was first discovered in Asian lesser white-toothed shrews (*Crocidura shantungensis*) captured between 2007 and 2010 in South Korea [23]. *O. kenkemeense* (Kenkeme virus) was first discovered in *Sorex roboratus* captured in 2006 in Russia [24].

In the genus *Orthohantavirus*, the unclassified Altai virus was first detected in the tissue of *Sorex araneus* in the Altai Republic of Russia in 2007 [28]. The unclassified Ash River virus was discovered in masked shrews (*Sorex cinereus*) captured in 1983 and 1994 in Minnesota. The unclassified Jemez Springs virus was discovered in *Sorex monticolus* captured in 1994 in Colorado and between 1996 and 2000 in New Mexico [30]. The unclassified Artybash virus was first detected in the lung of *Sorex caecutiens* captured between 2006 and 2014 in eastern Siberia, Russia, and Hokkaido, Japan [19]. The unclassified Azagny virus was first discovered in West African pygmy shrews (*Crocidura obscurior*) captured in 2009 in Côte d'Ivoire [33]. The unclassified Boginia virus was first reported in 2013. It was discovered in *Neomys fodiens* captured between 2010 and 2012 in central Poland [32]. The unclassified Camp Ripley virus is the first hantavirus discovered in *Blarina brevicauda* captured in Minnesota in 1998 [31]. The unclassified Qiān Hú Shān virus was discovered in the lung tissue of stripe-backed shrew (*Sorex cylindricauda*) captured in 2005 in China [34]. The unclassified Xinyi virus was first discovered in Taiwanese mole shrews (*Anourosorex yamashinai*) captured in 1985 in Taiwan, China [29]. Phylogenetic analysis suggested that Xinyi virus shares a common ancestor with the Cao Bằng virus. The unclassified Yákèshí virus was discovered in long-clawed shrews (*Sorex unguiculatus*) captured in 2006 in Yákèshí, Inner Mongolia. The unclassified Liánghé virus was found in *Anourosorex squamipes* captured between 2010 and 2011 in China [22].

Five species in the genus *Thottimvirus* of this family, such as *T. imjinense*, *T. thottapalayamense*, *T. thottimvirus*, and two unclassified species, were detected in shrews (Table 2). Of them, *T. thottapalayamense* (thottapalayam virus) was first isolated in 1964 in *Suncus murinus* in India. This virus represents the earliest isolation of a hantavirus in shrews [36]. *T. imjinense* (Imjin virus) was first discovered in *Crocidura lasiura* captured between 2004 and 2005 near the DMZ along the Imjin River in South Korea [35]. The unclassified Kilimanjaro virus and Uluguru virus in *Thottimvirus* were first discovered in Geata mouse shrews (*Myosorex geata*) and Kilimanjaro mouse shrews (*Myosorex zinki*) captured between 1995 and 2002 in Tanzania [37].

Except for Seoul virus, all the above hantaviruses were only found in shrews with unknown pathogenicity.

2.6. *Nairoviruses Detected in Shrews*

The family *Nairoviridae* currently comprises 7 genera and 51 species, which infect arthropods, birds, or mammals [59]. Several viruses in this family, such as Crimean-Congo hemorrhagic fever virus and Nairobi sheep disease virus can cause diseases in humans and/or domestic animals [59].

Six species in the genus *Orthonairovirus* of this family, namely *O. erveense*, *O. lambarenense*, *O. thiaforaense*, and three unclassified species, were detected in *Crocidura* spp. in multiple countries (Table 2). Of them, *O. erveense* (Erve virus) was first detected in 1982 from multiple organs of *Crocidura russula* trapped in Western France. Serological surveys indicated that Erve virus had apparently a large geographical distribution in France and infects rodents, insectivores, wild boars, red deer, sheep, herring gulls, and humans, leading to neurological symptoms in humans [38]. *O. lambarenense* (Lamusara virus) and the unclassified Lamgora virus in *Orthonairovirus* were discovered in *Crocidura* spp. captured between 2019 and 2020 in Gabon [39]. *O. thiaforaense* (Thiafora virus) was first discovered in *Crocidura* sp. captured in 1971 in Senegal [40].

Additionally, the unclassified Cencurut virus in *Orthonairovirus* was first discovered in *Suncus murinus* captured between 2012 and 2014 in Singapore [41]. An unclassified species of *Orthonairovirus* and an unclassified species of the genus *Xinspirovirus* of this family were detected in lungs of Taiwanese gray shrews (*Crocidura tanakae*) [16]. These orthonairoviruses have been found only in shrews with unknown pathogenicity.

2.7. *Othomyxoviruses Detected in Shrews*

The family *Orthomyxoviridae* currently comprises 9 genera and 21 species, which infect mammals and birds [60]. Several viruses in this family, such as influenza A virus and influenza B virus, can cause diseases in humans and/or domestic animals.

H5N6 subtype highly pathogenic avian influenza in the genus *Alphainfluenzavirus* of this family were detected in lungs of *Anourosorex squamipes* [16]. This virus is highly pathogenic to chickens, mainly circulating in wild and domestic birds. It has sporadically infected some humans [61]. The shrew was likely accidentally infected with the avian virus.

2.8. *Paramyxoviruses Detected in Shrews*

The family *Paramyxoviridae* currently comprises 4 subfamilies, 17 genera, and 78 species [62]. Numerous viruses in this family, such as measles virus and canine distemper virus, cause diseases in humans or domestic animals.

Gamak virus and Daeryong virus in the genus *Henipavirus* were first reported in South Korea in recent years. They were both discovered in *Crocidura lasiura* and *Crocidura shantungensis*. These two viruses were exclusively detected in shrews. Gamak virus can infect and replicate in human hypotriploid alveolar basal epithelial cell lines and elicit the production of type I/III interferon, interferon-stimulated genes, and proinflammatory cytokines [42].

Twelve unclassified species in the genus *Henipavirus* of this family have been identified. Among them, Langya virus was first reported in China in 2022 with the reservoir hosts of shrews. The virus was detected in *Crocidura lasiura* and *Crocidura shantungensis*. The virus is pathogenic to humans, and

35 acute Langya virus cases manifested influenza-like symptoms. Research encompassed 25 wild animal species and indicated a high virus infection rate among the shrews (71/262, 27%) [43].

In *Henipavirus*, Melian virus was first detected in large-headed shrews (*Crocidura grandiceps*) captured in 2018 in Guinea [44]. Denwin virus was first detected in *Crocidura russula* captured in 2019 in Belgium [44]. Ninorex virus was first detected in *Sorex minutus* captured in 2020 in Belgium [45]. Phylogenetic analysis of these viruses and some other relevant paramyxoviruses suggested that both genera could be divided into two clades, one covering bat-borne viruses and the other covering rodent- and shrew-borne viruses [63].

Beilong virus in the genus *Jeilongvirus* of this family has been detected in rodents and shrews. A study in China showed a positivity rate of Beilong virus 28.57% (2/7) in the Asian lesser white-toothed shrews (*Crocidura shantungensis*) and 17.57% (13/74) in the *Suncus murinus*. Currently, there is no evidence of human infection with Beilong virus, but its potential risk to humans and livestock should not be underestimated [46].

2.9. *Phenuiviruses Detected in Shrews*

The family *Phenuiviridae* currently comprises 23 genera and 159 species, which infect vertebrates, including mammals and birds, invertebrates, plants, and fungi [64]. Some viruses in this family, such as sand-fly fever Naples virus and Chikungunya virus, can cause diseases in humans and/or domestic animals. Phenuivirid genomes consist of two or three single-stranded RNA molecules, some of which encode two proteins in non-overlapping ORFs of opposite polarities.

Severe fever with thrombocytopenia syndrome (SFTS) is an emerging hemorrhagic fever caused by SFTS virus (SFTSV), a species (*Dabie Bandavirus*) in the genus *Bandavirus* in *Phenuiviridae* [47]. SFTS has been reported in humans in China, South Korea, and Japan since 2010 [65]. The main clinical manifestations of SFTS include acute fever, thrombocytopenia, leukopenia, and gastrointestinal and neurological symptoms [6–8]. Moreover, multiple organ failure may occur in severe cases, with a maximum mortality rate of 30%. A survey suggested that shrews are more likely than other wild animals to be the reservoir of SFTSV because the prevalence of SFTSV in *Suncus murinus* is significantly higher than in other animals [47].

Diverse unclassified species in *Phenuiviridae* have been identified in *Anourosorex squamipes*, *Crocidura attenuata*, and *Chodsigoa smithii* in China in recent years [10]. It remains unknown whether these viruses were shrew viruses or the viruses of plants or fungi in shrew food.

2.10. *Rhabdoviruses Detected in Shrews*

The family *Rhabdoviridae* currently comprises 56 genera that are assigned to four subfamilies and 434 species, which infect vertebrates, invertebrates, and plants [66]. A few rhabdoviruses, such as rabies virus and vesicular stomatitis virus, can cause diseases in humans and domestic animals.

Members of the genus *Lyssavirus* of this family infect a variety of mammals causing rabies-like diseases [67]. The species *L. mokola* was detected in Nigeria in 1968 from pooled lung, liver, spleen, kidney, and heart of shrews (*Crocidura* sp.) [67]. The species *L. rabies* (rabies virus) was identified in lungs of *Suncus murinus* in China using HTS [16]. This finding is of public health significance, as it suggests that shrews could be involved in the ecology and epidemiology of rabies virus.

An unclassified species in the genus *Tupavirus* and eight unclassified species in unclassified genera of *Rhabdoviridae* were identified in shrews, such as *Chodsigoa smithii*, *Anourosorex squamipes*, *Crocidura attenuata*, *Blarinella griselda* (Indochinese short-tailed shrews), and *Anourosorex squamipes*, in China [10,16]. The pathogenicity of these shrew viruses remains unknown.

3. +ssRNA Viruses Detected in Shrews

Table 3 shows that 62 species of +ssRNA viruses in shrews have been detected.

Table 3. Detection of +ssRNA viruses in shrews using high-throughput sequencing (HTS), PCR, RT-PCR, serological methods, and/or virus isolation.

Virus family and genus	Virus species and name	Host species	Country	Detection and references
Arteriviridae, Muarterivirus	Muarterivirus afrigant (Olivier's shrew virus 1)	Crocidura olivieri guineensis	Guinea	diverse sample_HTS [68]
	unclassified species (Wufeng shrew arterivirus 1)	Anourosorex squamipes	China	multiple organs_HTS [10]
	unclassified species (Jingmen shrew arterivirus 1)	Crocidura shantungensis	China	multiple organs_HTS [10]
	unclassified species (Jingmen shrew arterivirus 2)	Crocidura shantungensis	China	multiple organs_HTS [10]
Arteriviridae, unclassified genus	unclassified species (Crocidura shantungensis arterivirus 1)	Crocidura shantungensis	China	Lung_HTS [16]
	unclassified species (Crocidura shantungensis arterivirus 2)	Crocidura shantungensis	China	Lung_HTS [16]
	unclassified species (Crocidura shantungensis arterivirus 3)	Crocidura shantungensis	China	Lung_HTS [16]
Astroviridae, Bastrovirus	unclassified species (Wufeng shrew bastrovirus 1)	Chodsigoa smithii	China	multiple organs and feces_HTS [10]
Astroviridae, Bastrovirus	unclassified species (Wufeng shrew bastrovirus 2)	Chodsigoa smithii	China	multiple organs and feces_HTS [10]
Astroviridae, Bastrovirus	unclassified species (Wufeng shrew bastrovirus 3)	Chodsigoa smithii	China	multiple organs and feces_HTS [10]
Astroviridae, Mamastrovirus	unclassified species (shrew astrovirus)	Episoriculus leucops; Crocidura attenuata	China	feces_RT-PCR [69]
Astroviridae, Mamastrovirus	unclassified species (Jingmen_shrew_astrovirus 1)	Crocidura shantungensis	China	multiple organs and feces_HTS [10]
Astroviridae, Mamastrovirus	unclassified species (Wufeng shrew_astrovirus 1)	Anourosorex squamipes	China	multiple organs and feces_HTS [10]
Astroviridae, unclassified genera	unclassified species (Wenzhou rodent astrovirus 1)	Suncus murinus	China	multiple organs and feces_HTS [10]
Caliciviridae, Lagovirus	Lagovirus europaeus * (rabbit hemorrhagic disease virus)	Crocidura russula	Spain	liver_RT-PCR [70]
Coronaviridae, Alphacoronavirus	unclassified species (shrew alphacoronavirus)	Sorex araneus	United Kingdom	liver_RT-PCR [71]
	unclassified species (Wencheng shrew coronavirus)	Suncus murinus	China	rectum_RT-PCR [72]
	unclassified species (Common shrew coronavirus)	Sorex araneus	China	pharyngeal and anal swabs_HTS [73]
	unclassified species (Wufeng shrew alphacoronavirus 1)	Suncus murinus	China	multiple organs and feces_HTS [10]
Flaviviridae, Orthoflavivirus	Orthoflavivirus encephalitis * (tick-borne encephalitis virus)	Sorex araneus	Slovakia	sera_RT-PCR+isolation [74,75]
	Orthoflavivirus powassanense* (Powassan virus 2)	Blarina brevicauda	United States	spleen+brain_PCR [76]
	Orthoflavivirus usutuense* (Usutu virus)	Crocidura sp.	Senegal	brain_RT-PCR+isolation [77]
Flaviviridae, Pegivirus	Pegivirus neotomae* (rodent pegivirus)	Suncus murinus	China	sera_RT-PCR [78]
Flaviviridae, Pegivirus	unclassified species (Wufeng shrew pegivirus 2)	Chodsigoa smithii	China	multiple organs and feces_HTS [10]
Flaviviridae, Hepacivirus	unclassified species (Suncus murinus hepacivirus)	Suncus murinus	China	multiple organs and feces_HTS [10]
Flaviviridae, Hepacivirus	unclassified species (Wufeng rodent hepacivirus 3)	Chodsigoa smithii	China	multiple organs and feces_HTS [10]

Flaviviridae, Hepacivirus	unclassified species (Wufeng rodent hepacivirus 3)	Chodsigoa smithii	China	multiple organs and feces_HTS [10]
Flaviviridae, Hepacivirus	unclassified species (Wufeng shrew hepacivirus 1)	Crocidura attenuata	China	multiple organs and feces_HTS [10]
Flaviviridae, Pestivirus	unclassified species (Wufeng shrew pestivirus 1)	Chodsigoa smithii	China	multiple organs and feces_HTS [10]
Hepeviridae, Paslahepevirus	unclassified species (shrew hepatitis E virus)	Suncus murinus	China	serum_RT- PCR+serology [79]
Hepeviridae, unclassified genera	unclassified species (Wufeng shrew hepevirus 2)	Crocidura attenuata; Chodsigoa smithii	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew hepe-like virus 1)	Chodsigoa smithii; Crocidura attenuata	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew hepevirus_2)	Chodsigoa smithii; Crocidura attenuata	China	multiple organs and feces_HTS [10]
	unclassified species (shrew hepatitis E virus)	Crocidura russula	Germany	multiple organs_HTS [OR713884]#
	unclassified species (Crocidura lasiura hepevirus)	Crocidura lasiura	China	Lung_HTS [16]
	unclassified species (Jingmen shrew nodavirus 1)	Crocidura shantungensis	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew nodavirus 1)	Chodsigoa smithii	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew nodavirus 2)	Chodsigoa smithii; Crocidura attenuata	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew nodavirus 4)	Chodsigoa smithii	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew nodavirus 6)	Chodsigoa smithii	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew nodavirus 7)	Crocidura attenuata; Chodsigoa smithii	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew nodavirus 8)	Crocidura attenuata; Anourosorex squamipes	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew nodavirus 10)	Crocidura attenuata	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew nodavirus 12)	Anourosorex squamipes	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew nodavirus 13)	Crocidura attenuata; Blarinella griselda	China	multiple organs and feces_HTS [10]
Nodaviridae, unclassified genus	unclassified species (Wufeng shrew nodavirus 15)	Chodsigoa smithii	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew nodavirus 16)	Chodsigoa smithii	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew nodavirus 17)	Crocidura attenuata	China	multiple organs and feces_HTS [10]
	unclassified species (Crocidura lasiura picornavirus 2)	Crocidura lasiura	China	Lung_HTS [16]
	Hepatovirus ishrewi (shrew hepatovirus)	Sorex araneus	Germany	diverse samples_RT- PCR+isolation [80]
Picornaviridae, Dicapivirus				
Picornaviridae, Hepatovirus				

	unclassified species (Wufeng shrew_picornavirus 1)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
<i>Picornaviridae</i> , <i>Mischivirus</i>	<i>Mischivirus ehoushre</i> (Mischivirus E)	<i>Suncus murinus</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew picornavirus 3)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
<i>Picornaviridae</i> , <i>Parechovirus</i>	<i>Parechovirus beljungani</i> * (Ljungan virus)	<i>Crocidura leucodon</i> ; <i>Sorex antinorii</i>	Italy	multiple organs_RT-PCR [81]
<i>Picornaviridae</i> , <i>Parabovirus</i>	unclassified species (<i>Sorex caecutiens</i> picornavirus)	<i>Sorex caecutiens</i>	China	Lung_HTS [16]
	unclassified species (Wufeng shrew picornavirus 4)	<i>Anourosorex squamipes</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wenzhou shrew picornavirus 1)	<i>Suncus murinus</i> ; <i>Chodsigoa smithii</i> ; <i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
<i>Picornaviridae</i> , unclassified genera	unclassified species (Wufeng shrew picornavirus 2)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew_picornavirus 5)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew picornavirus 6)	<i>Blarinella griselda</i>	China	multiple organs and feces_HTS [10]
	unclassified species (<i>Crocidura lasiura</i> picornavirus 1)	<i>Crocidura lasiura</i>	China	Lung_HTS [16]
	unclassified species (<i>Suncus murinus</i> picornavirus 12)	<i>Suncus murinus</i>	China	Lung_HTS [16]
	unclassified species (<i>Suncus murinus</i> picornavirus 294)	<i>Suncus murinus</i>	China	Lung_HTS [16]
<i>Hepelivirales</i> unclassified family	unclassified species (Wufeng shrew hepe-like virus 1)	<i>Chodsigoa smithii</i> ; <i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
<i>Nodamuvirales</i> unclassified family	unclassified species (Wufeng shrew nodamuvirus 1)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
<i>Nodamuvirales</i> unclassified family	unclassified species (Wufeng shrew nodamuvirus 2)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]

3.1. Arteriviruses Detected in Shrews

The family *Arteriviridae* currently comprises 6 subfamilies, 13 genera, and 23 species, which infect vertebrates, such as pigs, horses, and non-human primates [82]. Several significant veterinary pathogens, such as equine arteritis virus and porcine respiratory and reproduction syndrome virus 2, can cause severe diseases in domestic animals and/or non-human primates.

The genus *Muarterivirus* of *Arteriviridae* contains a shrew arterivirus, Olivier's shrew virus 1, which was first detected in Olivier's shrews (*Crocidura olivieri guineensis*) in 2016 in Guinea [68]. Additionally, six unclassified species of shrew arteriviruses were detected in *Crocidura shantungensis* and *Anourosorex squamipes* in China in recent years [10,16].

3.2. Astroviruses Detected in Shrews

The family *Astroviridae* currently comprises 2 genera and 22 species, which infect mammals (19 species) or birds (3 species). Some astroviruses, such as human astrovirus and duck astrovirus, can cause diseases in humans or domestic animals [83]. Infection with astroviruses often involves damage to livers, kidneys, or immune system. Duck astrovirus causes highly fatal hepatitis in ducklings [84].

Three unclassified species in the genus *Bastrovirus*, three unclassified species in the genus *Mamastrovirus*, and one species in an unclassified genus of *Astroviridae* were detected in shrews, such as *Anourosorex squamipes*, *Crocidura attenuata*, *Chodsigoa smithii*, *Episoriculus leucops* (long-tailed brown-toothed shrews), and *Suncus murinus* in China in recent years.

3.3. Caliciviruses Detected in Shrews

The family *Caliciviridae* currently comprises 11 genera, each including one or two species, which infect mammals, birds, or fish [85]. Some viruses in this family, such as norovirus and feline calicivirus, can cause diseases in humans or domestic animals.

Rabbit hemorrhagic disease virus in the genus *Lagovirus* of this family is highly pathogenic to rabbits. A survey identified this virus in a dead Mediterranean pine vole and two white-toothed shrews [70], which suggested that shrews could be victims of this virus.

3.4. Coronaviruses Detected in Shrews

The family *Coronaviridae* currently comprises 4 subfamilies, 6 genera, and 54 species, which infect vertebrates (mammals, birds, amphibians, and fish) [86]. Some coronaviruses, such as severe acute respiratory syndrome-related coronavirus, infectious bronchitis virus, and porcine epidemic diarrhea virus, can cause diseases in humans or domestic animals.

Three unclassified shrew coronaviruses in the genus *Alphacoronavirus* of this family were detected in *Sorex araneus* and *Suncus murinus* captured in recent years in China [10,73]. Another unclassified shrew coronavirus in the genus *Alphacoronavirus* was detected in *Sorex araneus* captured in the United Kingdom between 2008 and 2015 [71]. These shrew viruses have been found only in shrews with unknown pathogenicity.

3.5. Flaviviruses Detected in Shrews

The family *Flaviviridae* currently comprises 4 genera and 97 species, which infect various mammals [87,88]. Most members of the genus *Orthoflavivirus* are arthropod-borne. Some flaviviruses, such as yellow fever virus, Dengue virus, Zika virus, human hepatitis C virus, Japanese encephalitis virus, West Nile virus, tick-borne encephalitis virus, and Tambusu virus, can cause diseases in humans or domestic animals.

Tick-borne encephalitis virus (TBEV) in the genus *Orthoflavivirus* of this family can infect humans and cause meningitis, encephalitis, or meningoencephalitis. It circulates between ticks and small mammals. Hedgehogs, shrews, moles, and certain rodents are hosts of ticks and reservoirs of this virus. This virus was detected in 1967 in *Sorex araneus* in Slovakia [74].

Usutu virus in *Orthoflavivirus* is a *Culex*-associated mosquito-borne flavivirus that was also found in *Crocidura* sp. captured between 2012 and 2013 in Senegal. This virus was initially discovered in 1959 and is associated with mosquitoes as its vectors. Currently, the virus has been isolated from birds, arthropods, and humans in Europe and Africa, and subsequently detected in bats and horses [77].

Powassan virus 2 in *Orthoflavivirus* can lead to encephalitis in humans and is primarily transmitted by arthropod vectors. This virus has been reported not only in humans and other animals but also detected in *Blarina brevicauda* captured between 2018 and 2020 in Massachusetts and Rhode Island of the United States. Early studies suggested that groundhog ticks and squirrel ticks naturally maintain this virus with woodchucks and other medium-sized mammals, such as skunks and raccoons, or squirrels. These animals likely serve as the reservoir of the virus [76].

Shrews could be minor reservoirs or accidental hosts of the aforementioned TBEV, Usutu virus, and Powassan virus.

Rat pegivirus in *Pegivirus* was initially identified in the sera of desert wood-rats in 2013, which was also discovered in *Suncus murinus* captured between 2013 and 2015 in China. This virus primarily targets lymphocytes and causes asymptomatic infections in humans and other animals [78].

An unclassified shrew flavivirus in the genus *Pegivirus*, three unclassified shrew flaviviruses in the genus *Hepacivirus*, and an unclassified shrew flavivirus in the genus *Pestivirus* were detected in *Suncus murinus*, *Chodsigoa smithii*, or *Crocidura attenuata* in China in recent years through HTS [10] [78,89].

3.6. Hepeviruses Detected in Shrews

The family *Hepeviridae* currently comprises 6 genera and 39 species, five genera and ten species, which infect mammals, birds and fish [90]. Some viruses in this family, such as human hepatitis E virus and rabbit hepatitis E virus, can cause diseases in humans and/or domestic animals.

Three unclassified shrew hepeviruses species were detected in *Crocidura attenuata*, *Chodsigoa smithii*, and *Crocidura attenuata* in China in recent years through HTS [10,16]. An unclassified shrew hepeviruses species was detected in *Crocidura russula* in Germany in recent years through HTS as suggested by a sequence reported to GenBank.

3.7. Nodaviruses Detected in Shrews

The family *Nodaviridae* currently comprises two genera, each including four or more species, which infect insects (*Alphanodavirus*) or fish (*Betanodavirus*) [91]. Some viruses in this family, such as striped jack nervous necrosis virus, can cause diseases in animals.

Multiple unclassified species in this family with unknown pathogenicity were detected in *Anourosorex squamipes*, *Chodsigoa smithii*, *Crocidura attenuata*, and *Crocidura shantungensis* in China in recent years through HTS [10].

3.8. Picornaviruses Detected in Shrews

The family *Picornaviridae* currently comprises 63 genera and 147 species, which infect vertebrates (at least six of the seven classes) [92]. Numerous viruses in this family, such as poliovirus and foot-and-mouth disease virus, can cause diseases in humans or domestic animals.

An unclassified species in the genus of *Dicpivirus* of this family was detected in lungs of *Crocidura lasiura* in China using HTS [16].

Hepatitis A virus (HAV) in the genus *Hepatovirus* of *Picornaviridae* is an ancient and ubiquitous human pathogen recovered previously only in primates. Human HAV could originate from the virus in rodents [80]. Although HAV-like viruses have been identified in *Sorex araneus* in Germany and in *Suncus murinus* in China, shrew HAVs are distinct from human HAV, and its pathogenicity to humans remains unknown [10,80]. Patterns of shrew HAV infection in shrews mimicked those of human HAV in hepatotropism, fecal shedding, acute nature, and extinction of the virus in a closed host population [74].

One unclassified species in the genus *Parechovirus* of *Picornaviridae* was detected in *Crocidura leucodon* and *Sorex antinorii* (Valais shrews) in Italy using RT-PCR [81]. Two unclassified species in the genus *Mischivirus*, one unclassified species in the genus *Parobovirus*, and eight species of unclassified genera in *Picornaviridae* from *Anourosorex squamipes*, *Blarinella griselda*, *Chodsigoa smithii*, *Crocidura attenuata*, *Crocidura lasiura*, *Sorex caecutiens*, and *Suncus murinus* in China were detected using the HTS [10,16]. The pathogenicity of these shrew viruses remains unknown.

4. dsDNA Viruses Detected in Shrews

Table 4 shows that seven species of dsDNA viruses in shrews have been detected.

Table 4. Detection of dsDNA viruses in shrews using high-throughput sequencing (HTS), PCR, RT-PCR, serological methods, and/or virus isolation.

Virus family and genus	Virus species and name	Host species	Country	Detection and references
<i>Adenoviridae</i> , <i>Mastadenovirus</i>	unclassified species (Asian house shrew adenovirus)	<i>Suncus murinus</i> ; <i>Sorex araneus</i>	China	feces_PCR [93]
	unclassified species (shrew adenovirus)	<i>Sylvisorex</i> sp.	Cameroon	feces_PCR [94]
<i>Hepadnaviridae</i> , <i>Orthohepadnavirus</i>	<i>Orthohepadnavirus soricisinensis</i> (shrew hepatitis B virus)	<i>Anourosorex squamipes</i> ; <i>Crocidura attenuata</i> ; <i>Crocidura lasiura</i>	China	liver_PCR [95]
		<i>Crocidura</i> spp.	Cameroon and Congo	liver+spleen_PCR [72]
<i>Polyomaviridae</i> , <i>Alphapolyomavirus</i>	unclassified <i>Alphapolyomavirus</i> * (human polyomavirus 12)	<i>Sorex araneus</i> ; <i>Sorex coronatus</i> ; <i>Sorex minutus</i>	Germany and Norway	diverse samples_PCR [96]
	<i>Orthopoxvirus monkeypox</i> * (monkeypox virus)	<i>Crocidura</i> spp.	Zambia and Congo	sera+spleen_PCR+serology [97]
<i>Poxviridae</i> , <i>Orthopoxvirus</i>	unclassified species (shrew orthopoxvirus)	<i>Sorex araneus</i>	Norway	lung_PCR [52]

4.1. Adenoviruses Detected in Shrews

The family *Adenoviridae* currently comprises 6 genera and 109 species, which infect mammals, birds, reptiles, amphibians, or fish [98]. Some adenoviruses, such as human adenovirus 1 and canine adenovirus 1, can cause diseases in humans or domestic animals.

Two unclassified species with unknown pathogenicity in the genus *Mastadenovirus* of this family were detected in *Suncus murinus* in China and *Sylvisorex* sp. in Cameroon, respectively, using PCR [93,94].

4.2. Hepadnaviruses Detected in Shrews

The family *Hepadnaviridae* currently comprises 5 genera 18 species, which infect mammals, birds, and fish [85]. Some viruses in this family, such as human hepatitis B virus and duck hepatitis B virus, can cause diseases in humans or domestic animals.

Shrew hepatitis B virus with unknown pathogenicity in the genus *Orthohepadnavirus* of this family has been detected in *Anourosorex squamipes*, *Crocidura attenuata*, and *Crocidura lasiura*, but not in *Suncus murinus*, captured in China in recent years [95]. Phylogenetic analysis revealed that shrew HBVs were closely related to a bat hepatitis B virus with a similar genome structure.

4.3. Orthoherpesviruses Detected in Shrews

The family *Orthoherpesviridae* currently comprises 17 genera and 118 species, which infect mammals, birds and reptiles [99]. Some viruses in this family, such as Kaposi's sarcoma-associated herpesvirus and pseudorabies virus, can cause diseases in humans or domestic animals.

Four shrew species with unknown pathogenicity in an unclassified genus of this subfamily were detected in *Crocidura* spp. in Cameroon and Congo [72].

4.4. Polyomaviruses Detected in Shrews

The family *Polyomaviridae* currently comprises 6 genera and 112 species, which infect Mammals, birds, and fish [100]. Some viruses in this family, such as JC virus and SV40, can cause diseases in humans or domestic animals.

An unclassified polyomavirus with unknown pathogenicity in *Sorex araneus* and *Sorex coronatus* (Millet's shrews) was detected in Germany and Norway through PCR [96].

4.5. Poxviruses Detected in Shrews

The family *Poxviridae* currently comprises 22 genera and 83 species, which infect vertebrates and arthropods [101]. Some viruses in this family, such as variola virus and chicken poxvirus, can cause diseases in humans and/or domestic animals.

Monkeypox virus in the genus *Orthopoxvirus* of this family is zoonotic and has caused human infections in numerous countries in recent years. A survey in Zambia suggested that 14 of 42 shrews had antibodies against this virus [97]. However, the role of shrews in the epidemiology of this virus remains enigmatic. An unclassified species in this genus was detected in lung samples of *Sorex araneus* in Norway using PCR [52].

5. ssDNA Viruses Detected in Shrews

Table 5 shows that four species of ssDNA viruses in shrews have been detected.

Table 5. Detection of ssDNA viruses in shrews using high-throughput sequencing (HTS), PCR, RT-PCR, serological methods, and/or virus isolation.

Virus family and genus	Virus species and name	Host species	Country	Detection and references
<i>Anelloviridae</i> , <i>Rhotorquevirus</i>	<i>Rhotorquevirus murid 1</i> (Torque teno rodent virus)	<i>Suncus murinus</i>	China	throat+sera_PCR [102]
<i>Parvoviridae</i> , <i>Protoparvovirus</i>	<i>Rodent protoparvovirus 1</i> (Mpulungu bufavirus)	<i>Crocidura hirta</i>	Zambia	spleen+intestinal contents_HTS+ PCR [103]
<i>Parvoviridae</i> , <i>Bocaparvovirus</i>	unclassified species* (porcine bocavirus G4)	<i>Suncus murinus</i>	China	feces+sera_PCR [104]
<i>Parvoviridae</i> , <i>Dependoparvovirus</i>	unclassified species (shrew adeno-associated virus)	<i>Suncus murinus</i>	China	feces+sera_PCR [105]

5.1. Anelloviruses Detected in Shrews

The family *Anelloviridae* currently comprises 34 genera and 173 species, which infect mammals, birds, or reptiles [106]. Some viruses in this family, such as Torque teno virus and Torque teno mini virus, are prevalent in humans and/or domestic animals. They could be associated with liver or respiratory diseases, hematological disorders, cancer, and immune suppression.

The species *Rhotorquevirus murid 1* (Torque teno rodent virus) was detected in *Suncus murinus* captured in China in recent years through PCR [102].

5.2. Parvoviruses Detected in Shrews

The family *Parvoviridae* currently comprises two subfamilies (*Parvovirinae* and *Densovirinae*), 13 genera, and >75 species. Viruses in *Parvovirinae* infect vertebrates, including mammals, birds, and reptiles. Viruses in *Densovirinae* infect insects, crustacea, and echinoderms [107]. Some parvoviruses, such as parvovirus B19 and canine parvovirus, can cause diseases in humans or domestic animals.

The species *Rodent protoparvovirus 1* (Mpulungu bufavirus) in the genus *Protoparvovirus* of this family was detected in lesser red musk shrews (*Crocidura hirta*) captured between 2011 and 2013 in Zambia using HTS and PCR [103].

A new genotype of porcine bocavirus in the genus *Bocaparvovirus* of *Parvoviridae* can cause respiratory and gastrointestinal diseases in pigs. This virus was detected in *Suncus murinus* captured between 2015 and 2017 in China, but shrews are unlikely its reservoir [104].

Adeno-associated virus in the genus *Dependoparvovirus* of *Parvoviridae* can infect humans and multiple domestic and wild animals, including pigs, cows, goats, chickens, snakes, bats, and rodents. It was detected in *Suncus murinus* captured between 2015 and 2017 in China [105].

6. dsRNA Viruses Detected in Shrews

Table 6 shows that 26 species of dsRNA viruses in shrews have been detected.

Table 6. Detection of dsRNA viruses in shrews using high-throughput sequencing (HTS), PCR, RT-PCR, serological methods, and/or virus isolation.

Virus family and genus	Virus species and name	Host species	Country	Detection and references
<i>Picobirnaviridae</i> <i>unclassified</i> <i>genus</i>	unclassified species (shrew picobirnavirus 1)	<i>Crocidura lasiura</i> ; <i>Crocidura shantungensis</i>	China	Lung_HTS [16]
<i>Sedoreoviridae</i> , <i>Orbivirus</i>	<i>Orbivirus caerulinguae</i> * (Bluetongue virus)	<i>unknown shrews</i>	Unknown (Africa)	sera_serology [108]
	unclassified species (Wufeng shrew orbivirus)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	<i>Rotavirus alphagastroenteritidis</i> * (rotavirus A)	<i>Suncus murinus</i> ; <i>Chodsigoa smithii</i>	China	feces_RT-PCR [109]; multiple organs and feces_HTS [10]
	unclassified species (Jingmen_shrew rotavirus_1)	<i>Crocidura shantungensis</i>	China	multiple organs and feces_HTS [10]
<i>Sedoreoviridae</i> , <i>Rotavirus</i>	unclassified species (Wufeng shrew_rotavirus 1)	<i>Blarinella griselda</i> ; <i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew_rotavirus 2)	<i>Anourosorex squamipes</i> ; <i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Crocidura shantungensis rotavirus 9)	<i>Crocidura shantungensis</i>	China	Lung_HTS [16]
	(Crocidura shantungensis rotavirus 10)	<i>Crocidura shantungensis</i>	China	Lung_HTS [16]
	unclassified species (Wufeng shrew reo-like virus 2)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
<i>Sedoreoviridae</i> , <i>unclassified</i> <i>genera</i>	unclassified species (Wufeng shrew reo-like virus 8)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew reo-like virus 3)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew cypovirus 1)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
<i>Spinareoviridae</i> , <i>Cypovirus</i>	unclassified species (Wufeng shrew cypovirus 2)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
<i>Spinareoviridae</i> , <i>Orthoreovirus</i>	unclassified species (Wenzhou shrew orthoreovirus 1)	<i>Suncus murinus</i>	China	multiple organs and feces_HTS [10]

Spinareoviridae, unclassified genera	unclassified species (mammalian orthoreovirus)	<i>Crocidura shantungensis</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew reovirus 1)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wenzhou shrew reovirus 1)	<i>Suncus murinus</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wenzhou shrew reovirus 2)	<i>Suncus murinus</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew reo- like virus 1)	<i>Chodsigoa smithii</i> ; <i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew reo- like virus 4)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew reo- like virus 6)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew reo- like virus 7)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew reo- like virus 10)	<i>Crocidura attenuata</i>	China	multiple organs and feces_HTS [10]
	unclassified species (Wufeng shrew reo- like virus 5)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]
Reovirales, unclassified family	unclassified species (Wufeng shrew_reo- like_virus_9)	<i>Chodsigoa smithii</i>	China	multiple organs and feces_HTS [10]

6.1. Picobirnaviruses Detected in Shrews

The family *Picobirnaviridae* currently comprises one genus and three species, which infect vertebrates and invertebrates [110]. Some picobirnaviruses are widely distributed geographically among humans and other mammals. They have been mainly identified in fecal specimens and in raw sewage samples. The pathogenicity of picobirnaviruses has not been established. An unclassified picobirnavirus was identified in lungs of *Crocidura lasiura*, and *Crocidura shantungensis* in China using HTS [16].

6.2. Sedoreoviruses Detected in Shrews

The family *Sedoreoviridae* currently comprises 6 genera and 39 species, which infect mammals, birds, crustaceans, arthropods, algae, and plants [111]. Some viruses in this family, such as rotavirus A, bluetongue virus, and African horse sickness virus, can cause diseases in humans or domestic animals.

Bluetongue virus in the genus *Orbivirus* of this family primarily infects sheep and various ruminant animals, with a high fatality rate in sheep. This virus can also infect rodents and carnivores and can be transmitted by certain mosquitoes. This virus was found in shrews in Africa in the 1990s [108]. The role of shrews in the epizootiology of bluetongue needs elucidation.

Rotavirus A in the genus *Rotavirus* of this family can infect humans and diverse domestic or wild animals. This virus and five unclassified species in this genus in *Anourosorex squamipes*, *Blarinella griselda*, *Chodsigoa smithii*, *Crocidura shantungensis*, *Crocidura attenuata*, and *Suncus murinus* captured in China in recent years were detected through HTS [10,109]. Additionally, three unclassified species of shrew sedoreoviruses in *Chodsigoa smithii* and *Crocidura attenuate* captured in China, which were distinct from known genera of this family, were detected in recent years through HTS [10,109].

6.3. Spinareoviruses Detected in Shrews

The family *Spinareoviridae* currently comprises 9 genera and 58 species, which infect mammals, aquatic animals (fish, mammals, crustaceans, and molluscs), birds, reptiles, arthropods, fungi, and plants [112]. Some spinareoviruses, such as human orthoreovirus and turkey orthoreovirus, can cause diseases in humans and domestic animals.

Two unclassified species in the genus *Cypovirus*, two unclassified species in the genus *Orthoreovirus*, and eight unclassified species distinct from known genera of this family were detected in *Chodsigoa smithii*, *Crocidura attenuata*, and *Suncus murinus* captured in China in recent years through HTS [10].

7. Discussion and Outlook

This review shows that shrew populations host at least 32 families and 190 species of viruses. Two recent publications regarding large-scale detection of shrew viruses in China greatly expanded the knowledge about shrew virome or shrew viral diversity [10,16].

Dozens of other species of viruses have also been identified in shrews, but they were plant viruses or invertebrate viruses, and hence have not been described in this review [10,16]. They were detected in shrew samples possibly due to their existence in shrew food or shrew surroundings.

Shrews have been identified with compelling evidence as the reservoirs of the zoonotic Langya virus, TBEV, SFTSV, and BDV-1. Shrews could be intermediate or incidental hosts or victims of Ebola virus, bluetongue virus, bufavirus, Usutu virus, tick-borne encephalitis virus, monkeypox virus, avian influenza virus, and RHDV, which are pathogenic to humans or domestic animals. Nevertheless, these infections have demonstrated that shrews are significant in the epidemiology and control of zoonosis and pose considerable threats to public health and animal health because shrews share overlapping habitats with humans and livestock. This highlights the surveys of shrew viruses for public health and animal welfare.

Dozens of virus species, such as Thottapalayam virus, Seewis virus, Thiafora virus, Wencheng shrew virus, *Suncus murinus* hepacivirus, and Olivier's shrew virus 1, have been identified only in shrews. This suggests that shrews are likely the reservoirs of these species of viruses.

At least 24 species of hantaviruses, 13 species of paramyxoviruses, 17 species of phenuiviruses, 11 species of phaboviruses, 9 species of flaviviruses, 13 species of nodaviruses, 13 species of nairoviruses, 11 species of sedoreoviruses, and 12 species of spinareoviruses in shrews have been identified (Figure 1). This suggests that shrews are likely the reservoirs of some species of these virus families, as it is unlikely that multiple species of the same virus family all occasionally infect shrews.

The counts of identified virus species were significantly different among shrew genera and shrew species (Figure 2). The shrew genus *Crocidura* hosts more shrew species than other shrew genera, and the identified virus species were more from this shrew genus than from other shrew genera. *Chodsigoa smithii*, *Crocidura attenuate*, *Suncus murinus*, *Anourosorex squamipes*, *Crocidura shantungensis*, *Sorex araneus*, and *Crocidura lasiura* have been identified as being infected with more species of viruses than other shrew species. The factors, such as the abundance and distribution of these shrew genera or species, as well as the difficulty of capturing shrews of these genera or species, can affect the counts of identified virus species.

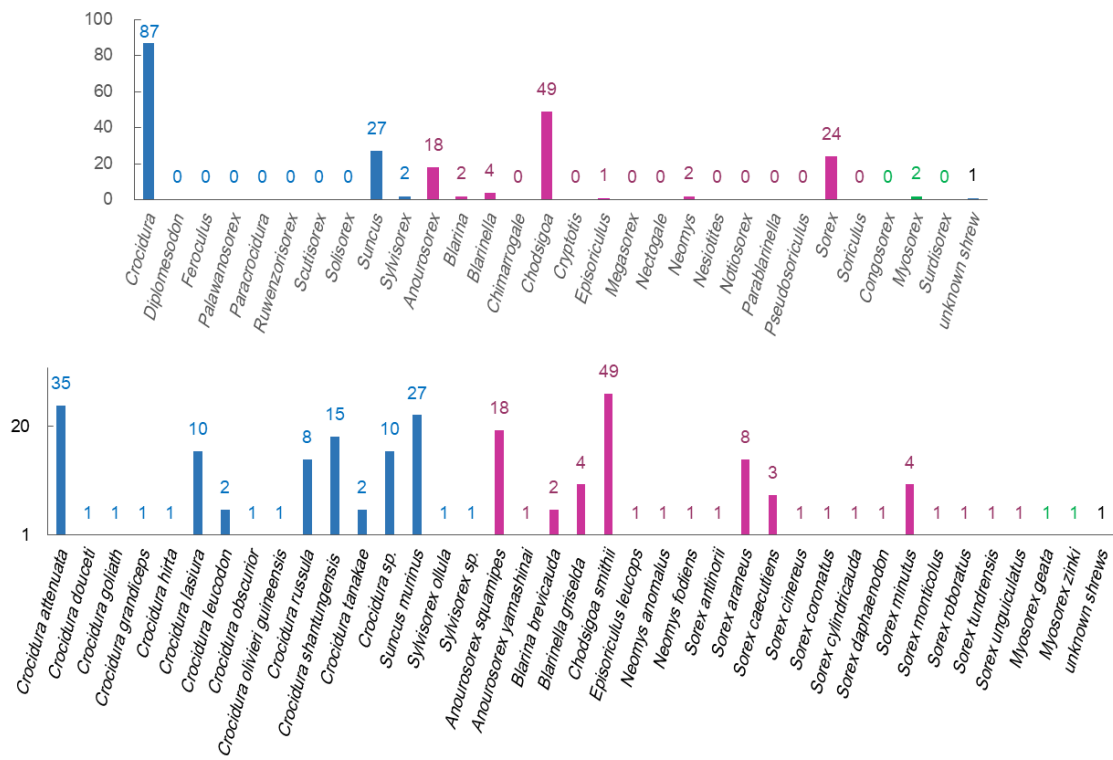


Figure 2. Numbers of virus species that have been identified in certain shrew genera and shrew species. The blue, purple, and green color represent genera or species in the shrew subfamilies of Crocidurinae, Soricinae, and Myosoricinae, respectively.

A database showed that 237, 201, and 204 species of viruses have been detected in *Mus musculus*, *Sus scrofa*, and *Bos taurus*, respectively [113]. With the consideration of the genetic diversity of shrews, most shrew viruses have yet to be identified so far. Meanwhile, much information regarding shrew viruses is based on viral sequences rather than viral isolates. The paucity of knowledge regarding virus diversity and viral isolates from shrews has hampered progress in demonstrating their pathogenic potential.

Shrews' small size and ease of handling make them ideal for studying the viruses of shrews and other animals, including humans. Their biological traits, like reproductive cycles, lifespan, and immune systems, are valuable for investigating the transmission, pathogenicity, and potential therapies of viral diseases. In this sense, it is desirable to develop efficient methods for breeding them in laboratories [114].

Future research should assess the transmission risks of shrew viruses from shrews to humans and domestic animals and conduct broader surveys to delve into the diversity, evolution, and potential pathogenicity of these viruses. This will enhance our capacity to respond swiftly to the potential outbreaks of shrew viruses in humans or domestic animals with effective measures.

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org. Table S1. Detailed information regarding the viruses having been identified in shrews

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References

1. Jürgens, K. D., Etruscan shrew muscle: The consequences of being small. *J Exp Biol* **2002**, 205, (15), 2161-2166.
2. Chen, Z.; Hu, J.; He, K.; Zhang, B.; Zhang, Y.; Chu, J.; Zhao, K.; Onditi, K. O.; Jiang, X., Molecular and morphological evidence support a new species of Asiatic short-tailed shrew (Eulipotyphla: Soricidae). *Journal of Mammalogy* **2023**, 104, (6), 1455-1467.
3. Zhang, Z., Animal biodiversity: An outline of higher-level classification and survey of taxonomic richness (Addenda 2013). *Zootaxa* **2013**, 3703(1), 1-82.
4. Chung, D. J.; Madison, G. P.; Aponte, A. M.; Singh, K.; Li, Y.; Pirooznia, M.; Bleck, C. K. E.; Darmani, N. A.; Balaban, R. S., Metabolic design in a mammalian model of extreme metabolism, the North American least shrew (*Cryptotis parva*). *J Physiol* **2022**, 600, (3), 547-567.
5. Bedford, J. M.; Mock, O. B.; Goodman, S. M., Novelities of conception in insectivorous mammals (*Lipotyphla*), particularly shrews. *Biol Rev Camb Philos Soc* **2004**, 79, (4), 891-909.
6. Kowalski, K.; Rychlik, L., Venom use in *Eulipotyphlans*: An evolutionary and ecological approach. *Toxins (Basel)* **2021**, 13, (3).
7. Liao, Z.; Tang, X.; Chen, W.; Jiang, X.; Chen, Z.; He, K.; Li, Q.; Duan, Z.; He, X.; Kamau, P. M.; Lv, L.; Zhang, Z.; Rong, M.; Lv, Q.; Lai, R., Shrew's venom quickly causes circulation disorder, analgesia and hypokinesia. *Cell Mol Life Sci* **2022**, 79, (1), 35.
8. Chai, S.; Tian, R.; Rong, X.; Li, G.; Chen, B.; Ren, W.; Xu, S.; Yang, G., Evidence of echolocation in the common shrew from molecular convergence with other echolocating mammals. *Zool Stud* **2020**, 59, (4), 1-11.
9. Siemers, B. M.; Schauerermann, G.; Turni, H.; von Merten, S., Why do shrews twitter? Communication or simple echo-based orientation. *Biol Lett* **2009**, 5, (5), 593-6.
10. Chen, Y. M.; Hu, S. J.; Lin, X. D.; Tian, J. H.; Lv, J. X.; Wang, M. R.; Luo, X. Q.; Pei, Y. Y.; Hu, R. X.; Song, Z. G.; Holmes, E. C.; Zhang, Y. Z., Host traits shape virome composition and virus transmission in wild small mammals. *Cell* **2023**, 186, (21), 4662-4675.e12.
11. Guo, L.; Liu, S.; Song, J.; Han, L.; Zhang, H.; Wu, C.; Wang, C.; Zhou, H.; Wang, J., Seroprevalence of Wenzhou virus in China. *Biosaf Health* **2020**, 2, (3), 152-156.
12. Li, K.; Lin, X. D.; Wang, W.; Shi, M.; Guo, W. P.; Zhang, X. H.; Xing, J. G.; He, J. R.; Wang, K.; Li, M. H.; Cao, J. H.; Jiang, M. L.; Holmes, E. C.; Zhang, Y. Z., Isolation and characterization of a novel arenavirus harbored by rodents and shrews in Zhejiang province, China. *Virology* **2015**, 476, 37-42.
13. Ushijima, Y.; Abe, H.; Ozeki, T.; Ondo, G. N.; Mbadinga, M.; Bikangui, R.; Nze-Nkogue, C.; Akomo-Okoue, E. F.; Ella, G. W. E.; Koumba, L. B. M.; Nso, B.; Mints-Nguema, R.; Makouloutou-Nzassi, P.; Makanga, B. K.; Nguelet, F. L. M.; Zadeh, V. R.; Urata, S.; Mbouna, A. V. N.; Massinga-Loembe, M.; Agnandji, S. T.; Lell, B.; Yasuda, J., Identification of potential novel hosts and the risk of infection with lymphocytic choriomeningitis virus in humans in Gabon, Central Africa. *Int J Infect Dis* **2021**, 105, 452-459.
14. Wang, B.; Cai, C. L.; Li, B.; Zhang, W.; Zhu, Y.; Chen, W. H.; Zhuo, F.; Shi, Z. L.; Yang, X. L., Detection and characterization of three zoonotic viruses in wild rodents and shrews from Shenzhen city, China. *Virol Sin* **2017**, 32, (4), 290-297.
15. Hilbe, M.; Herrsche, R.; Kolodziejek, J.; Nowotny, N.; Zlinszky, K.; Ehrensperger, F., Shrews as reservoir hosts of borna disease virus. *Emerg Infect Dis* **2006**, 12, (4), 675-7.

16. Zhang, J. T.; Hu, Z. Y.; Tang, F.; Liu, Y. T.; Tan, W. L.; Ma, X. F.; Zhang, Y. F.; Si, G. Q.; Zhang, L.; Zhang, M. Q.; Peng, C.; Fu, B. K.; Fang, L. Q.; Zhang, X. A.; Liu, W., Decoding the RNA viromes in shrew lungs along the eastern coast of China. *NPJ Biofilms Microbiomes* **2024**, 10, (1), 68.
17. Morvan, J. M.; Deubel, V.; Gounon, P.; Nakouné, E.; Barrière, P.; Murri, S.; Perpète, O.; Selekon, B.; Coudrier, D.; Gautier-Hion, A.; Colyn, M.; Volechkov, V., Identification of Ebola virus sequences present as RNA or DNA in organs of terrestrial small mammals of the Central African Republic. *Microbes Infect* **1999**, 1, (14), 1193-201.
18. Yashina, L. N.; Abramov, S. A.; Zhigalin, A. V.; Smetannikova, N. A.; Dupal, T. A.; Krivopalov, A. V.; Kikuchi, F.; Senoo, K.; Arai, S.; Mizutani, T.; Suzuki, M.; Cook, J. A.; Yanagihara, R., Geographic distribution and phylogeny of soricine shrew-borne Seewis virus and Altai virus in Russia. *Viruses* **2021**, 13, (7), 1286.
19. Arai, S.; Kang, H. J.; Gu, S. H.; Ohdachi, S. D.; Cook, J. A.; Yashina, L. N.; Tanaka-Taya, K.; Abramov, S. A.; Morikawa, S.; Okabe, N.; Oishi, K.; Yanagihara, R., Genetic diversity of Artybash virus in the Laxmann's shrew (*Sorex caecutiens*). *Vector Borne Zoonotic Dis* **2016**, 16, (7), 468-75.
20. Radosa, L.; Schlegel, M.; Gebauer, P.; Ansorge, H.; Heroldová, M.; Jánová, E.; Stanko, M.; Mošanský, L.; Fričová, J.; Pejčoch, M.; Suchomel, J.; Purchart, L.; Groschup, M. H.; Krüger, D. H.; Ulrich, R. G.; Klempa, B., Detection of shrew-borne hantavirus in Eurasian pygmy shrew (*Sorex minutus*) in Central Europe. *Infect Genet Evol* **2013**, 19, 403-10.
21. Gu, S. H.; Nicolas, V.; Lalis, A.; Sathirapongsasuti, N.; Yanagihara, R., Complete genome sequence and molecular phylogeny of a newfound hantavirus harbored by the Doucet's musk shrew (*Crocidura douceti*) in Guinea. *Infect Genet Evol* **2013**, 20, 118-23.
22. Guo, W. P.; Lin, X. D.; Wang, W.; Tian, J. H.; Cong, M. L.; Zhang, H. L.; Wang, M. R.; Zhou, R. H.; Wang, J. B.; Li, M. H.; Xu, J.; Holmes, E. C.; Zhang, Y. Z., Phylogeny and origins of hantaviruses harbored by bats, insectivores, and rodents. *PLoS Pathog* **2013**, 9, (2), e1003159.
23. Arai, S.; Gu, S. H.; Baek, L. J.; Tabara, K.; Bennett, S. N.; Oh, H. S.; Takada, N.; Kang, H. J.; Tanaka-Taya, K.; Morikawa, S.; Okabe, N.; Yanagihara, R.; Song, J. W., Divergent ancestral lineages of newfound hantaviruses harbored by phylogenetically related crocidurine shrew species in Korea. *Virology* **2012**, 424, (2), 99-105.
24. Kang, H. J.; Arai, S.; Hope, A. G.; Cook, J. A.; Yanagihara, R., Novel hantavirus in the flat-skulled shrew (*Sorex roboratus*). *Vector Borne Zoonotic Dis* **2010**, 10, (6), 593-7.
25. Sun, X. F.; Zhao, L.; Zhang, Z. T.; Liu, M. M.; Xue, Z. F.; Wen, H. L.; Ma, D. Q.; Huang, Y. T.; Sun, Y.; Zhou, C. M.; Luo, L. M.; Liu, J. W.; Li, W. Q.; Yu, H.; Yu, X. J., Detection of Imjin virus and Seoul virus in crocidurine shrews in Shandong province, China. *Vector Borne Zoonotic Dis* **2017**, 17, (6), 425-431.
26. Song, J. W.; Gu, S. H.; Bennett, S. N.; Arai, S.; Puorger, M.; Hilbe, M.; Yanagihara, R., Seewis virus, a genetically distinct hantavirus in the Eurasian common shrew (*Sorex araneus*). *Virol J* **2007**, 4, 114.
27. Schlegel, M.; Radosa, L.; Rosenfeld, U. M.; Schmidt, S.; Triebenbacher, C.; Löhr, P. W.; Fuchs, D.; Heroldová, M.; Jánová, E.; Stanko, M.; Mošanský, L.; Fričová, J.; Pejčoch, M.; Suchomel, J.; Purchart, L.; Groschup, M. H.; Krüger, D. H.; Klempa, B.; Ulrich, R. G., Broad geographical distribution and high genetic diversity of shrew-borne Seewis hantavirus in Central Europe. *Virus Genes* **2012**, 45, (1), 48-55.
28. Kang, H. J.; Gu, S. H.; Yashina, L. N.; Cook, J. A.; Yanagihara, R., Highly divergent genetic variants of soricid-borne Altai virus (*Hantaviridae*) in Eurasia suggest ancient host-switching events. *Viruses* **2019**, 11, (9), 857.
29. Gu, S. H.; Arai, S.; Yu, H. T.; Lim, B. K.; Kang, H. J.; Yanagihara, R., Genetic variants of Cao Bang hantavirus in the Chinese mole shrew (*Anourosorex squamipes*) and Taiwanese mole shrew (*Anourosorex yamashinai*). *Infect Genet Evol* **2016**, 40, 113-118.
30. Arai, S.; Bennett, S. N.; Sumibcay, L.; Cook, J. A.; Song, J. W.; Hope, A.; Parmenter, C.; Nerurkar, V. R.; Yates, T. L.; Yanagihara, R., Phylogenetically distinct hantaviruses in the masked shrew (*Sorex cinereus*) and dusky shrew (*Sorex monticolus*) in the United States. *Am J Trop Med Hyg* **2008**, 78, (2), 348-51.
31. Arai, S.; Song, J. W.; Sumibcay, L.; Bennett, S. N.; Nerurkar, V. R.; Parmenter, C.; Cook, J. A.; Yates, T. L.; Yanagihara, R., Hantavirus in northern short-tailed shrew, United States. *Emerg Infect Dis* **2007**, 13, (9), 1420-3.
32. Gu, S. H.; Markowski, J.; Kang, H. J.; Hejduk, J.; Sikorska, B.; Liberski, P. P.; Yanagihara, R., Boginia virus, a newfound hantavirus harbored by the Eurasian water shrew (*Neomys fodiens*) in Poland. *Virol J* **2013**, 10, 160.

33. Kang, H. J.; Kadjo, B.; Dubey, S.; Jacquet, F.; Yanagihara, R., Molecular evolution of Azagny virus, a newfound hantavirus harbored by the West African pygmy shrew (*Crocidura obscurior*) in Côte d'Ivoire. *Virol J* **2011**, 8, 373.
34. Zuo, S. Q.; Gong, Z. D.; Fang, L. Q.; Jiang, J. F.; Zhang, J. S.; Zhao, Q. M.; Cao, W. C., A new hantavirus from the stripe-backed shrew (*Sorex cylindricauda*) in the People's Republic of China. *Virus Res* **2014**, 184, 82-6.
35. Song, J. W.; Kang, H. J.; Gu, S. H.; Moon, S. S.; Bennett, S. N.; Song, K. J.; Baek, L. J.; Kim, H. C.; O'Guinn, M. L.; Chong, S. T.; Klein, T. A.; Yanagihara, R., Characterization of Imjin virus, a newly isolated hantavirus from the Ussuri white-toothed shrew (*Crocidura lasiura*). *J Virol* **2009**, 83, (12), 6184-91.
36. Carey, D. E.; Reuben, R.; Panicker, K. N.; Shope, R. E.; Myers, R. M., Thottapalayam virus: a presumptive arbovirus isolated from a shrew in India. *Indian J Med Res* **1971**, 59, (11), 1758-60.
37. Kang, H. J.; Stanley, W. T.; Esselstyn, J. A.; Gu, S. H.; Yanagihara, R., Expanded host diversity and geographic distribution of hantaviruses in sub-Saharan Africa. *J Virol* **2014**, 88, (13), 7663-7.
38. Chastel, C.; Main, A. J.; Richard, P.; Le Lay, G.; Legrand-Quillien, M. C.; Beaucournu, J. C., Erve virus, a probable member of *Bunyaviridae* family isolated from shrews (*Crocidura russula*) in France. *Acta Virol* **1989**, 33, (3), 270-80.
39. Ozeki, T.; Abe, H.; Ushijima, Y.; Nze-Nkogue, C.; Akomo-Okoue, E. F.; Ella, G. W. E.; Koumba, L. B. M.; Nso, B.; Mintsu-Nguema, R.; Makouloutou-Nzassi, P.; Makanga, B. K.; Nguet, F. L. M.; Ondo, G. N.; Mbadinga, M.; Igasaki, Y.; Okada, S.; Hirano, M.; Yoshii, K.; Lell, B.; Bonney, L. C.; Hewson, R.; Kurosaki, Y.; Yasuda, J., Identification of novel orthonairoviruses from rodents and shrews in Gabon, Central Africa. *J Gen Virol* **2022**, 103, (10).
40. Walker, P. J.; Widen, S. G.; Firth, C.; Blasdel, K. R.; Wood, T. G.; Travassos da Rosa, A. P.; Guzman, H.; Tesh, R. B.; Vasilakis, N., Genomic characterization of Yogue, Kasokero, Issyk-Kul, Keterah, Gossas, and Thiafora viruses: Nairoviruses naturally infecting bats, shrews, and ticks. *Am J Trop Med Hyg* **2015**, 93, (5), 1041-51.
41. Low, D. H. W.; Ch'ng, L.; Su, Y. C. F.; Linster, M.; Zhang, R.; Zhuang, Y.; Kwak, M. L.; Borthwick, S. A.; Hitch, A. T.; Smith, G. J. D.; Mendenhall, I. H., Cencurut virus: A novel *Orthonairovirus* from Asian house shrews (*Suncus murinus*) in Singapore. *One Health* **2023**, 16, 100529.
42. Lee, S. H.; Kim, K.; Kim, J.; No, J. S.; Park, K.; Budhathoki, S.; Lee, S. H.; Lee, J.; Cho, S. H.; Cho, S.; Lee, G. Y.; Hwang, J.; Kim, H. C.; Klein, T. A.; Uhm, C. S.; Kim, W. K.; Song, J. W., Discovery and genetic characterization of novel paramyxoviruses related to the genus *Henipavirus* in *Crocidura* species in the republic of Korea. *Viruses* **2021**, 13, (10), 2020.
43. Zhang, X. A.; Li, H.; Jiang, F. C.; Zhu, F.; Zhang, Y. F.; Chen, J. J.; Tan, C. W.; Anderson, D. E.; Fan, H.; Dong, L. Y.; Li, C.; Zhang, P. H.; Li, Y.; Ding, H.; Fang, L. Q.; Wang, L. F.; Liu, W., A zoonotic henipavirus in febrile patients in China. *N Engl J Med* **2022**, 387, (5), 470-472.
44. Vanmechelen, B.; Meurs, S.; Horemans, M.; Loosen, A.; Joly Maes, T.; Laenen, L.; Vergote, V.; Koundouno, F. R.; Magassouba, N.; Konde, M. K.; Condé, I. S.; Carroll, M. W.; Maes, P., The characterization of multiple novel paramyxoviruses highlights the diverse nature of the subfamily Orthoparamyxovirinae. *Virus Evol* **2022**, 8, (2), 1-12.
45. Horemans, M.; Van Bets, J.; Joly Maes, T.; Maes, P.; Vanmechelen, B., Discovery and genome characterization of six new orthoparamyxoviruses in small Belgian mammals. *Virus Evol* **2023**, 9, (2), 1-10.
46. Chen, J. J.; Zhang, X. A.; Fan, H.; Jiang, F. C.; Jin, M. Z.; Dai, K.; Wang, N.; Zhang, P. H.; Li, X. K.; Li, H.; Shi, W.; Yang, Z. C.; Fang, L. Q.; Zhou, H. S.; Wei, Y. H.; Liu, W., Distribution and characteristics of Beilong virus among wild rodents and shrews in China. *Infect Genet Evol* **2020**, 85, (2020), 104454.
47. Liu, J. W.; Wen, H. L.; Fang, L. Z.; Zhang, Z. T.; He, S. T.; Xue, Z. F.; Ma, D. Q.; Zhang, X. S.; Wang, T.; Yu, H.; Zhang, Y.; Zhao, L.; Yu, X. J., Prevalence of SFTSV among Asian house shrews and rodents, China, January-August 2013. *Emerg Infect Dis* **2014**, 20, (12), 2126-8.
48. Shope, R. E.; Murphy, F. A.; Harrison, A. K.; Causey, O. R.; Kemp, G. E.; Simpson, D. I.; Moore, D. L., Two African viruses serologically and morphologically related to rabies virus. *J Virol* **1970**, 6, (5), 690-2.
49. Radoshitzky, S. R.; Buchmeier, M. J.; Charrel, R. N.; Gonzalez, J. J.; Günther, S.; Hepojoki, J.; Kuhn, J. H.; Lukashevich, I. S.; Romanowski, V.; Salvato, M. S.; Sironi, M.; Stenglein, M. D.; Torre, J. C., ICTV Virus Taxonomy Profile: Arenaviridae 2023. *J Gen Virol* **2023**, 104, (9).
50. Radoshitzky, S. R.; Buchmeier, M. J.; Charrel, R. N.; Clegg, J. C. S.; Gonzalez, J. J.; Günther, S.; Hepojoki, J.; Kuhn, J. H.; Lukashevich, I. S.; Romanowski, V.; Salvato, M. S.; Sironi, M.; Stenglein, M. D.; de la Torre, J. C.; Ictv Report, C., ICTV Virus Taxonomy Profile: Arenaviridae. *J Gen Virol* **2019**, 100, (8), 1200-1201.

51. Rubbenstroth, D.; Briesse, T.; Dürrwald, R.; Horie 堀江真行, M.; Hyndman, T. H.; Kuhn, J. H.; Nowotny, N.; Payne, S.; Stenglein, M. D.; Tomonaga 朝長啓造, K.; Ictv Report, C., ICTV Virus Taxonomy Profile: Bornaviridae. *J Gen Virol* **2021**, 102, (7).
52. Sandvik, T.; Tryland, M.; Hansen, H.; Mehl, R.; Moens, U.; Olsvik, O.; Traavik, T., Naturally occurring orthopoxviruses: Potential for recombination with vaccine vectors. *J Clin Microbiol* **1998**, 36, (9), 2542-7.
53. Nobach, D.; Bourg, M.; Herzog, S.; Lange-Herbst, H.; Encarnação, J. A.; Eickmann, M.; Herden, C., Shedding of Infectious Borna Disease Virus-1 in Living Bicolored White-Toothed Shrews. *PLoS One* **2015**, 10, (8), e0137018.
54. Kuhn, J. H.; Dheilly, N. M.; Junglen, S.; Paraskevopoulou, S.; Shi, M.; Di Paola, N., ICTV Virus Taxonomy Profile: Jingchuvirales 2023. *J Gen Virol* **2023**, 104, (12).
55. Biedenkopf, N.; Bukreyev, A.; Chandran, K.; Di Paola, N.; Formenty, P. B. H.; Griffiths, A.; Hume, A. J.; Mühlberger, E.; Netesov, S. V.; Palacios, G.; Pawęska, J. T.; Smither, S.; Takada, A.; Wahl, V.; Kuhn, J. H., ICTV Virus Taxonomy Profile: Filoviridae 2024. *J Gen Virol* **2024**, 105, (2).
56. Hussein, H. A., Brief review on ebola virus disease and one health approach. *Heliyon* **2023**, 9, (8), e19036.
57. Bradfute, S. B.; Calisher, C. H.; Klempa, B.; Klingström, J.; Kuhn, J. H.; Laenen, L.; Tischler, N. D.; Maes, P., ICTV Virus Taxonomy Profile: Hantaviridae 2024. *J Gen Virol* **2024**, 105, (4).
58. Song, J. W.; Kang, H. J.; Song, K. J.; Truong, T. T.; Bennett, S. N.; Arai, S.; Truong, N. U.; Yanagihara, R., Newfound hantavirus in Chinese mole shrew, Vietnam. *Emerg Infect Dis* **2007**, 13, (11), 1784-7.
59. Kuhn, J. H.; Alkhovsky, S. V.; Avšič-Županc, T.; Bergeron, É.; Burt, F.; Ergünay, K.; Garrison, A. R.; Marklewitz, M.; Mirazimi, A.; Papa, A.; Pawęska, J. T.; Spengler, J. R.; Palacios, G., ICTV Virus Taxonomy Profile: Nairoviridae 2024. *J Gen Virol* **2024**, 105, (4).
60. Uribe, M.; Rodríguez-Posada, M. E.; Ramirez-Nieto, G. C., Molecular Evidence of Orthomyxovirus Presence in Colombian Neotropical Bats. *Front Microbiol* **2022**, 13, 845546.
61. Liu, S.; Zhuang, Q.; Wang, S.; Jiang, W.; Jin, J.; Peng, C.; Hou, G.; Li, J.; Yu, J.; Yu, X.; Liu, H.; Sun, S.; Yuan, L.; Chen, J., Control of avian influenza in China: Strategies and lessons. *Transbound Emerg Dis* **2020**, 67, (4), 1463-1471.
62. Kuhn, J. H.; Abe, J.; Adkins, S.; Alkhovsky, S. V.; Avšič-Županc, T.; Ayllón, M. A.; Bahl, J.; Balkema-Buschmann, A.; Ballinger, M. J.; Kumar Baranwal, V.; Beer, M.; Bejerman, N.; Bergeron, É.; Biedenkopf, N.; Blair, C. D.; Blasdel, K. R.; Blouin, A. G.; Bradfute, S. B.; Briesse, T.; Brown, P. A.; Buchholz, U. J.; Buchmeier, M. J.; Bukreyev, A.; Burt, F.; Büttner, C.; Calisher, C. H.; Cao, M.; Casas, I.; Chandran, K.; Charrel, R. N.; Kumar Chaturvedi, K.; Chooi, K. M.; Crane, A.; Dal Bó, E.; Carlos de la Torre, J.; de Souza, W. M.; de Swart, R. L.; Debat, H.; Dheilly, N. M.; Di Paola, N.; Di Serio, F.; Dietzgen, R. G.; Digiaro, M.; Drexler, J. F.; Duprex, W. P.; Dürrwald, R.; Easton, A. J.; Elbeaino, T.; Ergünay, K.; Feng, G.; Firth, A. E.; Fooks, A. R.; Formenty, P. B. H.; Freitas-Astúa, J.; Gago-Zachert, S.; Laura García, M.; García-Sastre, A.; Garrison, A. R.; Gaskin, T. R.; Gong, W.; Gonzalez, J. J.; de Bellocq, J.; Griffiths, A.; Groschup, M. H.; Günther, I.; Günther, S.; Hammond, J.; Hasegawa, Y.; Hayashi, K.; Hepojoki, J.; Higgins, C. M.; Hongō, S.; Horie, M.; Hughes, H. R.; Hume, A. J.; Hyndman, T. H.; Ikeda, K.; Jiāng, D.; Jonson, G. B.; Junglen, S.; Klempa, B.; Klingström, J.; Kondō, H.; Koonin, E. V.; Krupovic, M.; Kubota, K.; Kurath, G.; Laenen, L.; Lambert, A. J.; Li, J.; Li, J. M.; Liu, R.; Lukashevich, I. S.; MacDiarmid, R. M.; Maes, P.; Marklewitz, M.; Marshall, S. H.; Marzano, S. L.; McCauley, J. W.; Mirazimi, A.; Mühlberger, E.; Nabeshima, T.; Naidu, R.; Natsuaki, T.; Navarro, B.; Navarro, J. A.; Neriya, Y.; Netesov, S. V.; Neumann, G.; Nowotny, N.; Nunes, M. R. T.; Ochoa-Corona, F. M.; Okada, T.; Palacios, G.; Pallás, V.; Papa, A.; Paraskevopoulou, S.; Parrish, C. R.; Pauvolid-Corrêa, A.; Pawęska, J. T.; Pérez, D. R.; Pfaff, F.; Plemper, R. K.; Postler, T. S.; Rabbidge, L. O.; Radoshitzky, S. R.; Ramos-González, P. L.; Rehanek, M.; Resende, R. O.; Reyes, C. A.; Rodrigues, T. C. S.; Romanowski, V.; Rubbenstroth, D.; Rubino, L.; Runstadler, J. A.; Sabanadzovic, S.; Sadiq, S.; Salvato, M. S.; Sasaya, T.; Schwemmle, M.; Sharpe, S. R.; Shi, M.; Shimomoto, Y.; Kavi Sidharthan, V.; Sironi, M.; Smither, S.; Song, J. W.; Spann, K. M.; Spengler, J. R.; Stenglein, M. D.; Takada, A.; Takeyama, S.; Tatara, A.; Tesh, R. B.; Thornburg, N. J.; Tian, X.; Tischler, N. D.; Tomitaka, Y.; Tomonaga, K.; Tordo, N.; Tu, C.; Turina, M.; Tzanetakis, I. E.; Maria Vaira, A.; van den Hoogen, B.; Vanmechelen, B.; Vasilakis, N.; Verbeek, M.; von Bargen, S.; Wada, J.; Wahl, V.; Walker, P. J.; Waltzek, T. B.; Whitfield, A. E.; Wolf, Y. I.; Xia, H.; Xylogianni, E.; Yanagisawa, H.; Yano, K.; Ye, G.; Yuan, Z.; Zerbini, F. M.; Zhang, G.; Zhang, S.; Zhang, Y. Z.; Zhao, L.; Økland, A. L., Annual (2023) taxonomic update of RNA-directed RNA polymerase-encoding negative-sense RNA viruses (realm Riboviria: kingdom Orthornavirae: phylum Negarnaviricota). *J Gen Virol* **2023**, 104, (8).

63. Horemans, M.; Van Bets, J.; Joly Maes, T.; Maes, P.; Vanmechelen, B., Discovery and genome characterization of six new orthoparamyxoviruses in small Belgian mammals. *Virus Evol* **2023**, 9, (2), vead065.
64. Sasaya, T.; Palacios, G.; Briese, T.; Di Serio, F.; Groschup, M. H.; Neriya, Y.; Song, J. W.; Tomitaka, Y., ICTV Virus Taxonomy Profile: Phenuiviridae 2023. *J Gen Virol* **2023**, 104, (9).
65. Li, J.; Li, S.; Yang, L.; Cao, P.; Lu, J., Severe fever with thrombocytopenia syndrome virus: a highly lethal bunyavirus. *Crit Rev Microbiol* **2021**, 47, (1), 112-125.
66. Walker, P. J.; Freitas-Astúa, J.; Bejerman, N.; Blasdel, K. R.; Breyta, R.; Dietzgen, R. G.; Fooks, A. R.; Kondo, H.; Kurath, G.; Kuzmin, I. V.; Ramos-González, P. L.; Shi, M.; Stone, D. M.; Tesh, R. B.; Tordo, N.; Vasilakis, N.; Whitfield, A. E.; Ictv Report, C., ICTV Virus Taxonomy Profile: Rhabdoviridae 2022. *J Gen Virol* **2022**, 103, (6).
67. Delmas, O.; Holmes, E. C.; Talbi, C.; Larrous, F.; Dacheux, L.; Bouchier, C.; Bourhy, H., Genomic diversity and evolution of the lyssaviruses. *PLoS One* **2008**, 3, (4), e2057.
68. Vanmechelen, B.; Vergote, V.; Laenen, L.; Koundouno, F. R.; Bore, J. A.; Wada, J.; Kuhn, J. H.; Carroll, M. W.; Maes, P., Expanding the arterivirus host spectrum: Olivier's shrew virus 1, a novel arterivirus discovered in African giant shrews. *Sci Rep* **2018**, 8, (1), 11171.
69. Hu, B.; Chmura, A. A.; Li, J.; Zhu, G.; Desmond, J. S.; Zhang, Y.; Zhang, W.; Epstein, J. H.; Daszak, P.; Shi, Z., Detection of diverse novel astroviruses from small mammals in China. *J Gen Virol* **2014**, 95, (Pt 11), 2442-2449.
70. Calvete, C.; Mendoza, M.; Sarto, M. P.; Bagüés, M. P. J.; Luján, L.; Molín, J.; Calvo, A. J.; Monroy, F.; Calvo, J. H., Detection of rabbit hemorrhagic disease virus GL2/RHDV2/b in the Mediterranean pine vole (*Microtus duodecimcostatus*) and White-Toothed Shrew (*Crocidura russula*). *J Wildl Dis* **2019**, 55, (2), 467-472.
71. Tsoleridis, T.; Onianwa, O.; Horncastle, E.; Dayman, E.; Zhu, M.; Danjitrong, T.; Wachtl, M.; Behnke, J. M.; Chapman, S.; Strong, V.; Dobbs, P.; Ball, J. K.; Tarlinton, R. E.; McClure, C. P., Discovery of novel *Alphacoronaviruses* in European rodents and shrews. *Viruses* **2016**, 8, (3), 84.
72. Ntumvi, N. F.; Mbala Kingebeni, P.; Tamoufe, U.; Kumakamba, C.; Ndze, V.; Ngay Lukusa, I.; LeBreton, M.; Atibu Losoma, J.; Le Doux Diffio, J.; N'Kawa, F.; Takuo, J. M.; Mulembakani, P.; Nwobegahay, J.; Makuwa, M.; Muyembe Tamfum, J. J.; Gillis, A.; Harris, S.; Rimoin, A. W.; Hoff, N. A.; Fair, J. M.; Monagin, C.; Ayukekbong, J.; Rubin, E. M.; Wolfe, N. D.; Lange, C. E., High herpesvirus diversity in wild rodent and shrew species in Central Africa. *Intervirology* **2018**, 61, (4), 155-165.
73. Wu, Z.; Lu, L.; Du, J.; Yang, L.; Ren, X.; Liu, B.; Jiang, J.; Yang, J.; Dong, J.; Sun, L.; Zhu, Y.; Li, Y.; Zheng, D.; Zhang, C.; Su, H.; Zheng, Y.; Zhou, H.; Zhu, G.; Li, H.; Chmura, A.; Yang, F.; Daszak, P.; Wang, J.; Liu, Q.; Jin, Q., Comparative analysis of rodent and small mammal viromes to better understand the wildlife origin of emerging infectious diseases. *Microbiome* **2018**, 6, (1), 178.
74. Nosek, J.; Grulich, I., The relationship between the tick-borne encephalitis virus and the ticks and mammals of the Tribec mountain range. *Bull World Health Organ* **1967**, 36 Suppl, (1), 31-47.
75. Bakhvalova, V. N.; Chicherina, G. S.; Potapova, O. F.; Panov, V. V.; Glupov, V. V.; Potapov, M. A.; Seligman, S. J.; Morozova, O. V., Tick-Borne Encephalitis Virus Diversity in Ixodid Ticks and Small Mammals in South-Western Siberia, Russia. *Vector Borne Zoonotic Dis* **2016**, 16, (8), 541-9.
76. Goethert, H. K.; Mather, T. N.; Johnson, R. W.; Telford, S. R., 3rd, Incrimination of shrews as a reservoir for Powassan virus. *Commun Biol* **2021**, 4, (1), 1319.
77. Diagne, M. M.; Ndione, M. H. D.; Di Paola, N.; Fall, G.; Bedekelabou, A. P.; Sembène, P. M.; Faye, O.; Zannotto, P. M. A.; Sall, A. A., Usutu virus isolated from rodents in Senegal. *Viruses* **2019**, 11, (2), 181.
78. Gao, Y. W.; Wan, Z. W.; Wu, Y.; Li, X. F.; Tang, S. X., PCR-based screening and phylogenetic analysis of rat pegivirus (RPGV) carried by rodents in China. *J Vet Med Sci* **2020**, 82, (10), 1464-1471.
79. Guan, D.; Li, W.; Su, J.; Fang, L.; Takeda, N.; Wakita, T.; Li, T. C.; Ke, C., Asian musk shrew as a reservoir of rat hepatitis E virus, China. *Emerg Infect Dis* **2013**, 19, (8), 1341-3.
80. Drexler, J. F.; Corman, V. M.; Lukashev, A. N.; van den Brand, J. M.; Gmyl, A. P.; Brünink, S.; Rasche, A.; Seggewiß, N.; Feng, H.; Leijten, L. M.; Vallo, P.; Kuiken, T.; Dotzauer, A.; Ulrich, R. G.; Lemon, S. M.; Drosten, C., Evolutionary origins of hepatitis A virus in small mammals. *Proc Natl Acad Sci U S A* **2015**, 112, (49), 15190-5.
81. Fevola, C.; Rossi, C.; Rosso, F.; Girardi, M.; Rosà, R.; Manica, M.; Delucchi, L.; Rocchini, D.; Garzon-Lopez, C. X.; Arnoldi, D.; Bianchi, A.; Buzan, E.; Charbonnel, N.; Collini, M.; Ďureje, L.; Ecke, F.; Ferrari, N.; Fischer, S.; Gillingham, E. L.; Hörnfeldt, B.; Kazimírová, M.; Konečný, A.; Maas, M.; Magnusson, M.; Miller, A.;

- Niemimaa, J.; Nordström, Å.; Obiegala, A.; Olsson, G.; Pedrini, P.; Piątek, J.; Reusken, C. B.; Rizzolli, F.; Romeo, C.; Silaghi, C.; Sironen, T.; Stanko, M.; Tagliapietra, V.; Ulrich, R. G.; Vapalahti, O.; Voutilainen, L.; Wauters, L.; Rizzoli, A.; Vaheri, A.; Jääskeläinen, A. J.; Henttonen, H.; Hauffe, H. C., Geographical distribution of Ljungan virus in small mammals in Europe. *Vector Borne Zoonotic Dis* **2020**, 20, (9), 692-702.
82. Brinton, M. A.; Gulyaeva, A. A.; Balasuriya, U. B. R.; Dunowska, M.; Faaberg, K. S.; Goldberg, T.; Leung, F. C. C.; Nauwynck, H. J.; Snijder, E. J.; Stadejek, T.; Gorbalenya, A. E., ICTV Virus Taxonomy Profile: Arteriviridae 2021. *J Gen Virol* **2021**, 102, (8).
 83. Wilson, P.; Rosenstengle, C.; Spak, C., Underrecognized cause diarrhea in solid organ transplant: a report of astroviridae enteritis in liver transplant. *Transpl Infect Dis* **2024**, 26, (2), e14257.
 84. Johnson, C.; Hargest, V.; Cortez, V.; Meliopoulos, V. A.; Schultz-Cherry, S., Astrovirus Pathogenesis. *Viruses* **2017**, 9, (1).
 85. Vinjé, J.; Estes, M. K.; Esteves, P.; Green, K. Y.; Katayama, K.; Knowles, N. J.; L'Homme, Y.; Martella, V.; Vennema, H.; White, P. A.; Ictv Report, C., ICTV Virus Taxonomy Profile: Caliciviridae. *J Gen Virol* **2019**, 100, (11), 1469-1470.
 86. Woo, P. C. Y.; de Groot, R. J.; Haagmans, B.; Lau, S. K. P.; Neuman, B. W.; Perlman, S.; Sola, I.; van der Hoek, L.; Wong, A. C. P.; Yeh, S. H., ICTV Virus Taxonomy Profile: Coronaviridae 2023. *J Gen Virol* **2023**, 104, (4).
 87. Postler, T. S.; Beer, M.; Blitvich, B. J.; Bukh, J.; de Lamballerie, X.; Drexler, J. F.; Imrie, A.; Kapoor, A.; Karganova, G. G.; Lemey, P.; Lohmann, V.; Simmonds, P.; Smith, D. B.; Stapleton, J. T.; Kuhn, J. H., Renaming of the genus Flavivirus to Orthoflavivirus and extension of binomial species names within the family Flaviviridae. *Arch Virol* **2023**, 168, (9), 224.
 88. Simmonds, P.; Becher, P.; Bukh, J.; Gould, E. A.; Meyers, G.; Monath, T.; Muerhoff, S.; Pletnev, A.; Rico-Hesse, R.; Smith, D. B.; Stapleton, J. T.; Ictv Report, C., ICTV Virus Taxonomy Profile: Flaviviridae. *J Gen Virol* **2017**, 98, (1), 2-3.
 89. Guo, H.; Cai, C.; Wang, B.; Zhuo, F.; Jiang, R.; Wang, N.; Li, B.; Zhang, W.; Zhu, Y.; Fan, Y.; Chen, W.; Chen, W.; Yang, X.; Shi, Z., Novel *Hepacivirus* in Asian house shrew, China. *Sci China Life Sci* **2019**, 62, (5), 701-704.
 90. Purdy, M. A.; Drexler, J. F.; Meng, X. J.; Norder, H.; Okamoto, H.; Van der Poel, W. H. M.; Reuter, G.; de Souza, W. M.; Ulrich, R. G.; Smith, D. B., ICTV Virus Taxonomy Profile: Hepeviridae 2022. *J Gen Virol* **2022**, 103, (9).
 91. Sahul Hameed, A. S.; Ninawe, A. S.; Nakai, T.; Chi, S. C.; Johnson, K. L.; Ictv Report, C., ICTV Virus Taxonomy Profile: Nodaviridae. *J Gen Virol* **2019**, 100, (1), 3-4.
 92. Zell, R.; Delwart, E.; Gorbalenya, A. E.; Hovi, T.; King, A. M. Q.; Knowles, N. J.; Lindberg, A. M.; Pallansch, M. A.; Palmenberg, A. C.; Reuter, G.; Simmonds, P.; Skern, T.; Stanway, G.; Yamashita, T.; Ictv Report, C., ICTV Virus Taxonomy Profile: Picornaviridae. *J Gen Virol* **2017**, 98, (10), 2421-2422.
 93. Zheng, X. Y.; Qiu, M.; Ke, X. M.; Guan, W. J.; Li, J. M.; Huo, S. T.; Chen, S. W.; Zhong, X. S.; Zhou, W.; Xiong, Y. Q.; Ge, J.; Chen, Q., Detection of novel adenoviruses in fecal specimens from rodents and shrews in southern China. *Virus Genes* **2016**, 52, (3), 417-21.
 94. Diffo, J.; Ndze, V. N.; Ntumvi, N. F.; Takuo, J. M.; Mouiche, M. M. M.; Tamoufe, U.; Nwobegahay, J.; LeBreton, M.; Gillis, A.; Schneider, B. S.; Fair, J. M.; Monagin, C.; McIver, D. J.; Joly, D. O.; Wolfe, N. D.; Rubin, E. M.; Lange, C. E., DNA of diverse adenoviruses detected in Cameroonian rodent and shrew species. *Arch Virol* **2019**, 164, (9), 2359-2366.
 95. Nie, F. Y.; Tian, J. H.; Lin, X. D.; Yu, B.; Xing, J. G.; Cao, J. H.; Holmes, E. C.; Ma, R. Z.; Zhang, Y. Z., Discovery of a highly divergent hepadnavirus in shrews from China. *Virology* **2019**, 531, 162-170.
 96. Gedvilaite, A.; Tryland, M.; Ulrich, R. G.; Schneider, J.; Kurmauskaitė, V.; Moens, U.; Preugschas, H.; Calvignac-Spencer, S.; Ehlers, B., Novel polyomaviruses in shrews (*Soricidae*) with close similarity to human polyomavirus 12. *J Gen Virol* **2017**, 98, (12), 3060-3067.
 97. Orba, Y.; Sasaki, M.; Yamaguchi, H.; Ishii, A.; Thomas, Y.; Ogawa, H.; Hang'ombe, B. M.; Mweene, A. S.; Morikawa, S.; Saijo, M.; Sawa, H., Orthopoxvirus infection among wildlife in Zambia. *J Gen Virol* **2015**, 96, (Pt 2), 390-394.
 98. Benkő, M.; Aoki, K.; Arnberg, N.; Davison, A. J.; Echavarría, M.; Hess, M.; Jones, M. S.; Kaján, G. L.; Kajon, A. E.; Mittal, S. K.; Podgorski, I.; San Martín, C.; Wadell, G.; Watanabe, H.; Harrach, B.; Ictv Report, C., ICTV Virus Taxonomy Profile: Adenoviridae 2022. *J Gen Virol* **2022**, 103, (3).
 99. Magnius, L.; Mason, W. S.; Taylor, J.; Kann, M.; Glebe, D.; Dény, P.; Sureau, C.; Norder, H.; Ictv Report, C., ICTV Virus Taxonomy Profile: Hepadnaviridae. *J Gen Virol* **2020**, 101, (6), 571-572.

100. Moens, U.; Calvignac-Spencer, S.; Lauber, C.; Ramqvist, T.; Feltkamp, M. C. W.; Daugherty, M. D.; Verschoor, E. J.; Ehlers, B.; Ictv Report, C., ICTV Virus Taxonomy Profile: Polyomaviridae. *J Gen Virol* **2017**, *98*, (6), 1159-1160.
101. McInnes, C. J.; Damon, I. K.; Smith, G. L.; McFadden, G.; Isaacs, S. N.; Roper, R. L.; Evans, D. H.; Damaso, C. R.; Carulei, O.; Wise, L. M.; Lefkowitz, E. J., ICTV Virus Taxonomy Profile: Poxviridae 2023. *J Gen Virol* **2023**, *104*, (5).
102. Xiong, Y. Q.; Mo, Y.; Chen, M. J.; Cai, W.; He, W. Q.; Chen, Q., Detection and phylogenetic analysis of torque teno virus (TTV) carried by murine rodents and house shrews in China. *Virology* **2018**, *516*, 189-195.
103. Sasaki, M.; Orba, Y.; Anindita, P. D.; Ishii, A.; Ueno, K.; Hang'ombe, B. M.; Mweene, A. S.; Ito, K.; Sawa, H., Distinct Lineages of Bufavirus in Wild Shrews and Nonhuman Primates. *Emerg Infect Dis* **2015**, *21*, (7), 1230-3.
104. Xiong, Y. Q.; You, F. F.; Chen, X. J.; Chen, Y. X.; Wen, Y. Q.; Chen, Q., Detection and phylogenetic analysis of porcine bocaviruses carried by murine rodents and house shrews in China. *Transbound Emerg Dis* **2019**, *66*, (1), 259-267.
105. Xiong, Y. Q.; Zhang, M. Y.; Zhou, J. H.; Li, Y. Z.; You, F. F.; Wen, Y. Q.; He, W. Q.; Chen, Q., A molecular epidemiological investigation of carriage of the Adeno-Associated virus in murine rodents and house shrews in China. *Intervirology* **2018**, *61*, (3), 143-148.
106. Varsani, A.; Kraberger, S.; Opriessnig, T.; Maggi, F.; Celer, V.; Okamoto, H.; Biagini, P., Anelloviridae taxonomy update 2023. *Arch Virol* **2023**, *168*, (11), 277.
107. Cotmore, S. F.; Agbandje-McKenna, M.; Canuti, M.; Chiorini, J. A.; Eis-Hubinger, A. M.; Hughes, J.; Mietzsch, M.; Modha, S.; Ogliastro, M.; Péntzes, J. J.; Pintel, D. J.; Qiu, J.; Soderlund-Venermo, M.; Tattersall, P.; Tijssen, P.; Ictv Report, C., ICTV Virus Taxonomy Profile: Parvoviridae. *J Gen Virol* **2019**, *100*, (3), 367-368.
108. Alexander, K. A.; MacLachlan, N. J.; Kat, P. W.; House, C.; O'Brien, S. J.; Lerche, N. W.; Sawyer, M.; Frank, L. G.; Holekamp, K.; Smale, L.; et al., Evidence of natural bluetongue virus infection among African carnivores. *Am J Trop Med Hyg* **1994**, *51*, (5), 568-76.
109. Li, K.; Lin, X. D.; Huang, K. Y.; Zhang, B.; Shi, M.; Guo, W. P.; Wang, M. R.; Wang, W.; Xing, J. G.; Li, M. H.; Hong, W. S.; Holmes, E. C.; Zhang, Y. Z., Identification of novel and diverse rotaviruses in rodents and insectivores, and evidence of cross-species transmission into humans. *Virology* **2016**, *494*, 168-77.
110. Delmas, B.; Attoui, H.; Ghosh, S.; Malik, Y. S.; Mundt, E.; Vakharia, V. N.; Ictv Report, C., ICTV virus taxonomy profile: Picobirnaviridae. *J Gen Virol* **2019**, *100*, (2), 133-134.
111. Matthijssens, J.; Attoui, H.; Bányai, K.; Brussaard, C. P. D.; Danthi, P.; Del Vas, M.; Dermody, T. S.; Duncan, R.; Fāng 方勤, Q.; Johne, R.; Mertens, P. P. C.; Mohd Jaafar, F.; Patton, J. T.; Sasaya 笹谷孝英, T.; Suzuki 鈴木信弘, N.; Wei 魏太云, T., ICTV Virus Taxonomy Profile: Sedoreoviridae 2022. *J Gen Virol* **2022**, *103*, (10).
112. Matthijssens, J.; Attoui, H.; Bányai, K.; Brussaard, C. P. D.; Danthi, P.; Del Vas, M.; Dermody, T. S.; Duncan, R.; Fāng, Q.; Johne, R.; Mertens, P. P. C.; Mohd Jaafar, F.; Patton, J. T.; Sasaya, T.; Suzuki, N.; Wei, T., ICTV Virus Taxonomy Profile: Spinareoviridae 2022. *J Gen Virol* **2022**, *103*, (11).
113. Mihara, T.; Nishimura, Y.; Shimizu, Y.; Nishiyama, H.; Yoshikawa, G.; Uehara, H.; Hingamp, P.; Goto, S.; Ogata, H., Linking Virus Genomes with Host Taxonomy. *Viruses* **2016**, *8*, (3), 66.
114. Geyer, B.; Erickson, N. A.; Müller, K.; Grübel, S.; Hueber, B.; Hetz, S. K.; Brecht, M., Establishing and maintaining an etruscan shrew colony. *J Am Assoc Lab Anim Sci* **2022**, *61*, (1), 52-60.

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