

Review

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Review

# The Oral Microbiota, Microbial Metabolites, and Immuno-Inflammatory Mechanisms in Cardiovascular Disease

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**Abstract:** Cardiovascular diseases (CVD) remain a leading cause of global morbidity and mortality. Recent advancements in high-throughput omics techniques have enhanced our understanding of the human microbiome's role in the development of CVD. Although the relationship between the gut microbiome and CVD has attracted considerable research attention and has been rapidly evolving in recent years, the role of the oral microbiome remains less understood, with most prior studies focusing on periodontitis-related pathogens. In this review, we summarized previously reported associations between the oral microbiome and CVD, highlighting known CVD-associated such as Porphyromonas gingivalis, Fusobacterium nucleatum, and Aggregatibacter actinomycetemcomitans. We also discussed the interactions between the oral and gut microbes. The potential mechanisms by which the oral microbiota can influence CVD development include oral and systemic inflammation, immune responses, cytokine release, translocation of oral bacteria into the bloodstream, and the impact of microbial-related products such as microbial metabolites (e.g., short-chain fatty acids [SCFAs], trimethylamine oxide [TMAO], hydrogen sulfide [H2S], nitric oxide [NO]) and specific toxins (e.g., lipopolysaccharide [LPS]). The processes driven by these mechanisms may contribute to atherosclerosis, endothelial dysfunction, and other cardiovascular pathologies. Integrated multi-omics methodologies, along with large-scale longitudinal population studies and intervention studies, will facilitate a deeper understanding of the metabolic and functional roles of the oral microbiome in cardiovascular health. This fundamental knowledge will support the development of targeted interventions and effective therapies to prevent or reduce the progression from cardiovascular risk to clinical CVD events.

**Keywords:** oral microbiome, gut microbiome, cardiovascular diseases, inflammatory markers, microbial metabolites

#### 1. Introduction

Cardiovascular diseases (CVD) remain a primary cause of mortality worldwide, responsible for an estimated 17.9 million deaths each year (data from WHO), and also a leading cause of morbidity and disability [1]. The global prevalence of CVD nearly doubled from 1990 to 2019, reaching approximately 523 million cases. During this same period, global trends for the burden of CVD-associated events, along with the years of life lost due to CVD, showed a marked increase [1].

Over the past decade, advances in sequencing and multi-omics technologies (such as metabolomics and proteomics) have significantly enhanced our understanding of human

microbiome-host interactions, including the potential role of the human microbiome in the development of CVD [2,3] The microbiome plays a crucial role in human metabolic function and is essential for the proper development of the immune system [4,5]. Microbiome dysbiosis, including the alterations of microbiome composition, distribution, microbial related function, or metabolic activities [6], have been linked to several CVD-related conditions, such as atherosclerosis, heart failure, hypertension, and type 2 diabetes [7,8].

The oral microbiota is the second-largest microbial community in the human body, surpassed only by the gut microbiota. More than 772 microbial species have been identified in the oral cavity [9]. The oral cavity contains several ecological niches, including saliva, the tongue, buccal mucosa, palate, dental surfaces, gingiva, and both subgingival and supragingival sites. Each of these niches hosts distinct microbial species with varying activities, and their susceptibility to disease is also different. The oral microbiota is involved in the preliminary digestion of food and produces a variety of primary and secondary metabolites [10,11]. The circulatory system provides a route for oral bacteria and their byproducts—including microbial metabolites, endotoxins, and microbial-related pro-inflammatory cytokines— to enter the bloodstream and trigger systemic inflammation that can impact various organs and tissues throughout the body [12]. Increasing evidence from epidemiological studies, clinical investigations, and basic science studies supports the interactions between oral bacteria, oral microbial products, and the development of cardiovascular inflammation and CVD [13,14].

This manuscript reviews the existing evidence on the relationship between the oral microbiome and the progression of primary CVD. We discuss the potential mechanisms by which the oral microbiome can affect cardiometabolic health, especially through oral microbiota-induced inflammation and immune responses, and the role of oral microbial metabolites. We then discuss future directions and oral microbiome-related strategies for CVD prevention and management.

#### 2. Oral Microbiome and CVD

# 2.1. The Interaction Among Periodontitis, Oral Microbiota, and CVD

Previous studies have highlighted the role of specific oral bacteria in periodontal infections, and both epidemiological and mechanistic evidence have established links between periodontitis and CVD (especially atherosclerosis and thromboembolic events) [15] .

The bacterial biofilms on teeth and gums, commonly known as dental plaque, was known as primary etiological factors for the Development of Periodontitis, a chronic inflammatory condition characterized by the progressive destruction of gingival connective tissue, periodontal ligaments, and alveolar bone. These biofilms are predominantly composed of viridans group streptococci [16]. Additionally, several anaerobic bacteria such as *Porphyromonas gingivalis*, *Treponema denticola*, and *Tannerella forsythia*, play significant roles in chronic periodontitis [17,18]. Periodontitis serves as a reservoir for a wide range of microorganisms, which can enter the bloodstream through ulcerated, inflamed crevices and the pocket epithelium, allowing access to the adjacent gingival microcirculation.[16]. The severity of bacteraemia in patients with chronic periodontitis is directly related to the level of gingival inflammation [19]. The levels of *Streptococcus mutans* and *Porphyromonas gingivalis* have been observed to be elevated in conjunction with the development of periodontal disease and systemic inflammation [20,21].

Meta-analyses of large population studies have identified periodontitis as a risk factor for both peripheral and carotid atherosclerotic CVD[22–24]. Studies have demonstrated that periodontitis significantly increases the risk of atherosclerotic plaque formation. A prospective study of the US population found that individuals with periodontitis had a 25% increased risk of developing atherosclerotic plaques [25]. Furthermore, data from the Atherosclerosis Risk in Communities (ARIC) study revealed that periodontal disease is significantly associated with an increased incidence of stroke, with a hazard ratio (HR) of 2.6 for cardioembolic stroke and 2.2 for thrombotic stroke [26]. Periodontitis was also linked to heart failure, coronary heart disease, and hypertension [27,28].

Previous evidence has summarized the role of oral microbiota in the context of periodontitis. While the interactions between some specific oral pathogenic bacteria and periodontitis, as well as the association between periodontitis and CVD, were well-documented, the direct relationship between the overall oral microbiota composition and CVD remains under-investigated. More research is needed to elucidate the precise mechanisms connecting oral microbiota to cardiovascular disease outcomes [29].

# 2.2. Specific oral bacterial Taxa and CVD

The relative abundance of oral *Porphyromonas, Fusobacterium, Aggregatibacter*, and *Campylobacter* has been found to be associated with CVD in several human studies (**Table 1**). Many of these pathogenic bacteria share common characteristics: they are anaerobic, have outer membrane vesicles, possess extracellular proteolytic activity, engage in the amino acids anaerobic fermentation, and produce harmful metabolites. These properties enable the bacteria to degrade periodontal tissue and infiltrate the bloodstream. Their tissue-destructive capabilities may explain the presence of these bacteria or the detection of bacterial DNA in serum and arterial walls. And many of these bacteria are also associated with periodontitis.

Bacterial material, such as DNA, has been identified in multiple cardiovascular structures, particularly within atherosclerotic plaques. Based on data from 1,791 individuals across 63 studies, researchers identified 23 unique oral commensal bacteria present within the atherosclerotic plaques in individuals undergoing interventional procedures[30]. The in vivo identification of bacterial material within atherosclerotic plaques has drawn considerable interest. More compelling evidence comes from the cultivation of periodontal pathogenic bacteria from atheromatous plaques. Kozarov et al. found the presence of vital *Porphyromonas gingivalis* and *Aggregatibacter actinomycetemcomitans* inside atheromatous tissue cultured with primary human coronary endothelial cells [31]. Furthermore, the in vitro ability of periodonto-pathogens to invade cardiovascular cells (including aortic and heart endothelial cells) has been established [32].

**Table 1.** Human studies examining the association between cardiovascular disease and the oral microbiome.

Location	Sample size	Oral sample site	Methods	Findings	Speci
					fic
					Quali
					ty
					featu
					res
Mexican	Patients	supragingival	16S, V3-V4	A positive and significant	N
	with	dental plaque		correlation of blood TMAO	
	myocardial			levels with oral Porphyromonas	
	infarction			was identified in patients with	
	(n=16)			myocardial infarction.	
India	Patients	subgingival	16S,V3-V4	22 bacterial genera were shared	N
	with CAD	plaque		between subgingival and	
	(n=12)			atherosclerotic plaques, with	
				Acinetobacter being dominant.	
Germany	Patients	Subgingival plaq	16S,V3-V4	One biomarker of Saccharibacteria	N
	with CVD	ue		phylum (class: TM7-3, order:	
	(n=102)			CW040, family: F16) was	
	Mexican	Mexican Patients with myocardial infarction (n=16) India Patients with CAD (n=12)  Germany Patients with CVD	Mexican Patients supragingival with dental plaque myocardial infarction (n=16) India Patients subgingival with CAD plaque (n=12)  Germany Patients Subgingival plaq with CVD ue	Mexican Patients supragingival 16S, V3-V4 with dental plaque myocardial infarction (n=16)  India Patients subgingival 16S,V3-V4 with CAD plaque (n=12)  Germany Patients Subgingival plaq 16S,V3-V4 with CVD ue	Mexican       Patients with with dental plaque       supragingival       16S, V3-V4 and significant correlation of blood TMAO levels with oral Porphyromonas infarction (n=16)         India       Patients subgingival with CAD (n=12)       16S,V3-V4 atherosclerotic plaques, with Acinetobacter being dominant.         Germany       Patients Subgingival plaq with CVD       16S,V3-V4 ue       One biomarker of Saccharibacteria phylum (class: TM7-3, order:

					associated with the incidence of a secondary CV event	
Leskelä et	Germany	Controls	Saliva	Targeted	Specific oral bacteria A.	Cases
al., 2020.		(n=100),		qPCR	actinomycetemcomitans	matc
[49]		Patients		sequencing	concentration↑ in ischemic	hed
		with stroke			stroke cases than controls. IgG	contr
		(n=98), total			against A. actinomycetemcomitans	ols on
		(n=198)			is one of the main determinants	age
					of LPS neutralizing capacity.	and
						sex.
Perry et	New	Patients	Saliva	Targeted	Acute stroke patients were at	N
al., 2020.	Zealand.	with		qPCR	increased risk of colonisation	
[125]		atherosclero			from respiratory pathogens. The	
		sis (n=100)			presence of these pathogens in	
					saliva at one month was	
					associated with adverse	
					respiratory events.	
Nikolaev	Russia	Patients	Oral plaque	Targeted	In acute myocardial infarction	N
a et al.,		with angina		16S	patients, the frequency	
2019. [36]		pectoris		sequencing	of P.gingivalis, T. forsythia,	
		(n=15),			and A.actinomycetemcomitans det	
		with acute			ection was significantly higher	
		myocardial			than participants without CVD.	
		infarction				
		(n=15),				
		with chest				
		pain but no				
		CAD (n=15),				
		total (n=45)				
Author, yea	r Location	Sample size	e Oral	Methods I	indings	Specific

Author, year	Location	Sample size	Oral	Methods	Findings	Specific
			sample site			Quality
						features
Su et al.,	Japan	Total sample	Tongue	Targeted	Subjects with medical histories	N
2019. [40]		(n=70)	dorsum	PCR	of stroke and heart disease	
				sequencing	exhibited a trend toward	
					higher P. gingivalispositive rates	
					on the tongue dorsum than those	
					without such disorders.	
Liljestrand et	Finland	Control	Subgingiva	DNA	periodontal pathogens such as	Multivar
al., 2018. [50]		(n=123),	l plaque	hybridizati	A.actinomycetemcomitans and the	iable
		stable CAD		on	antibody levels to these	adjustm
		(n=184),			pathogens associated with	ent for
		ACS				age,

		(n=169),			coronary artery disease and	gender,
		ACS-like no			acute coronary syndrome	
					acute coronary syndrome	and
		CAD (n=29),				CVD
		total (n=505)				risk
						factors.
Ziebolz, Rost	Germany	Patients	Subgingiva	Targeted	Demonstrate the presence of	N
et al., 2018.		undergoing	l plaque	PCR	periodontal bacteria DNA in	
[39]		surgery for		sequencing	human cardiac tissue. Identified	
		aortic valve			correlations of inflammatory	
		stenosis			proteins and infection markers	
		(n=10)			with valvular heart disease.	
Ziebolz, Jahn	Germany	Patients	Subgingiva	Targeted	Periodontal pathogens (e.g. P.	N
et al., 2018.		undergoing	l plaque	PCR	gingivalis ,C.rectus, P. intermedia,	
[45]		surgery for		sequencing	F. nucleatum) DNA found in	
		aortic valve			atrial and myocardial tissue, and	
		stenosis			linked to tissue inflammation.	
		(n=30)				
Kannoch et	Serbia	Patients	Subgingiva	Targeted	Detect presence of	N
al., 2018. [37]		with	l plaque	16S	periopathogenic bacteria in	
		atherosclero		sequencing	subgingival and atherosclerotic	
		sis (n=100)			plaques(P.gingivalis, P.intermedia,	
					T. forsythensis) Patient's age can	
					influence the findings	
Mahalakshm	India	Patients	Subgingiva	Targeted	Statistical significance was	N
i et al., 2017.		with	l plaque	16S	observed for the prevalence of	
[35]		atherosclero		sequencing	16S rRNA of P. gingivalis, T.	
		sis (n=65),			forsythia, T. denticola and P.	
		with			nigrescens both in subgingival	
		periodontitis			plaque and atheromatous plaque	
		but no			. Significant odds and risk ratio	
		systemic			to atherosclerosis were observed	
		disease			for these bacteria.	
		(n=59),				
		Control				
		(n=100),				
		total (n=224)				
Author, year	Loca San	nple size Ora	l sample N	Methods I	Findings	Specific
•	tion	site	-			Quality
						features

Boaden,	UK	Patients	Saliva, buccal	16S,V1–V9	Described the bacterial profile of	N
2017. [126]		with stroke	mucosa,		the oral flora during the first 2	
		(n=50)	tongue,		weeks following a stroke,14 of	
			gingiva, hard		the 20 most common bacterial	
			palate		phylotypes found in the oral	
					cavity were Streptococcal species	
					with <i>S.salivarius</i> being the most	
					common. The condition of the	
					oral cavity worsened during the	
					study period.	
Mougeot et	USA	Patients	Coronary	16S,V3-V4	The most abundant species	N
al., 2017.		with CVD	artery tissue		were P. gingivalis, E. faecalis,	
[127]		(n=42)	and femoral		and <i>F.magna</i> .	
			artery tissue			
Fåk et al.,	Swe	Asymptoma	Whole mouth	16S,V1–V2	Abundance of Anaeroglobus in	Cases
2015. [38]	den	tic	swab		the oral cavity could be	matched
		atherosclero			associated with symptomatic	controls on
		sis (n=35),			atherosclerosis.	age and
		symptomati				sex.
		С				
		atherosclero				
		sis (n=27),				
		control				
		(n=30), total				
		(n=92)				
Serra e Silva	Braz	Patients	Subgingival	16S,V1-V9	Periodontal pockets and	N
Filho et al.,	il	with	plaque		atheromatous plaques of	
2014. [34]		periodontitis			cardiovascular disease patients	
		and			can present similarities in the	
		atherosclero			microbial diversity. 17 identical	
		sis (n=18)			phylotypes(including <i>P</i> .	
					gingivalis, T.vincentii,	
					F.nucleatum) were found in	
					atheroma and subgingival	
					samples, indicating possible	
					bacterial translocation.	
Koren et al.,	Swe	Controls	Oral cavity	16S,V1-V2	16S rRNA Sequencing Identified	The
2011. [42]	den	(n=15),	swab		Chryseomonas, Veillonella and	control
		patients			Streptococcus in atherosclerotic	subjects
		with			plaque samples. The combined	were
		atherosclero			abundances of Veillonella and	matched to
		sis (n=15),			Streptococcus in atherosclerotic	the patient
		total (n=30)				

plaques correlated with their group by abundance in the oral cavity. sex.

For these oral bacteria, their ability to live in anaerobic tissue and their relationship with inflammation are considered important. However, their role in the development of CVD and the possible biological mechanisms are not fully understood.

# 2.2.1. Porphyromonas Gingivalis

Porphyromonas gingivalis, a Gram-negative oral anaerobe, is widely recognized as a major etiologic agent responsible for the onset and progression of severe periodontitis [33]. Porphyromonas gingivalis has been detected in both subgingival samples and coronary artery atherosclerotic plaques [34,35]. Elevated levels of Porphyromonas gingivalisin oral samples have been observed in patients with coronary heart disease [34–37], coronary artery disease [38], valvular heart disease [39], and in individuals with CVD history [40]. Evidence also suggested that Porphyromonas gingivalis is linked to the citrullination of host self-antigens, contributing to the pathogenesis of rheumatoid heart disease [16]. Animal studies have indicated that Porphyromonas gingivalis can accelerate atheroma plaque formation and hold the ability to induce the development of fatty streaks in the aorta of rabbits [41].

Porphyromonas gingivalis is also known for its ability in disrupting innate immune function and triggering inflammatory responses [33]. The lipopolysaccharide of *P. gingivalis* can specifically activate host immune defenses, leading to heightened inflammatory pathways. *Porphyromonas gingivalis* exhibits convertase-like enzymatic activity and manipulates complement-TLR interactions to subvert host immune defenses, enabling it to evade immune clearance. This allows the pathogen to persist within the host, contributing to a shift in the periodontal microbiota toward a dysbiotic state, ultimately leading to inflammatory periodontitis [33].

# 2.2.2. Fusobacterium Nucleatum

Fusobacterium is a Gram-negative anaerobic genus commonly residing in both oral cavity and gastrointestinal tract [42,43]. In the oral cavity, it is particularly abundant in dental plaque [44]. Elevated levels of Fusobacterium have been reported in subgingival samples of patients with valvular [39,45]. Additionally, multiple species of Fusobacterium have been detected in subgingival plaque, as well as in coronary artery atherosclerotic plaques [34], and carotid artery plaque tissues [30,42,46]. Both gut and oral Fusobacterium might be potential sources of Fusobacterium in carotid artery plaque [47,48]. As the most representative and thoroughly studied species in this genus, Fusobacterium nucleatum has been associated with an increased risk of carotid artery atherosclerotic plaque [47,48]. Recent integrated omics studies using gut microbiome and serum proteomics data indicated that. Fusobacterium nucleatumwas positively associated with several serum proteomic markers which might be involved in host inflammation and immune activation related to bacterial infection (e.g., CXCL9, TNFRSF9), and the association between Fusobacterium nucleatumand carotid artery plaque can be partially explained by these serum inflammatory markers[47,48]. Furthermore, Fusobacterium is proteolytic and form cytotoxic end products. It has been associated with multiple plasma metabolites, such as lysophosphatidylcholines (LPCs), lysophosphatidylethanolamines (LPEs), and diacylglycerols (DGs), which have been further linked to an increased risk of carotid artery plaque [47].

## 2.2.3. Aggregatibacter Actinomycetemcomitans

Aggregatibacter actinomycetemcomitans is a facultative anaerobic, Gram-negative rod that takes on a coccoid shape. It is frequently detected in the oral cavity, particularly in subgingival plaque from individuals with aggressive periodontitis [44]. This bacterium has also been identified in samples

from both subgingival plaque and atherosclerotic lesions in coronary arteries [37], indicating its potential involvement in cardiovascular pathology.

The relative abundance of *Aggregatibacter actinomycetemcomitans* has been reported to increase in saliva samples from patients with ischemic stroke [49]. Similarly, higher levels of *Aggregatibacter actinomycetemcomitans* have been detected in subgingival or periodontal pocket samples from individuals with acute coronary syndrome, coronary artery disease [50], and valvular heart disease [39,45]. The subspecies of *Aggregatibacter actinomycetemcomitans* exhibit varying characteristics, such as the ability to induce leukocyte and monocyte cytotoxicity. Additionally, these bacteria can produce immunoglobulin proteases and collagenase, which contribute to their pathogenic potential and tissue destruction [44].

#### 2.2.4. Prevotella Intermedia

Prevotella intermedia is a slow-growing, anaerobic, Gram-negative rod that exhibits a coccoid shape [44]. Prevotella intermedia, along with other Prevotella species such as Prevotella nigrescens and Prevotella loescheii, have been detected in both oral samples (e.g., subgingival plaque) and coronary artery atherosclerotic lesions [51]. Other studies have reported positive associations between various Prevotella species (both in whole mouth samples [38] and subgingival samples [34,35]) and coronary artery disease. Additionally, Prevotella species have been associated with valvular heart disease [39,45], highlighting the broader role of this genus in cardiovascular pathology. Among the various Prevotella species, Prevotella intermedia is one of the most extensively studied in relation to CVD.

Prevotella intermedia produces lipase during bacterial invasion of the periodontium and exhibits a proteolytic metabolism, breaking down proteins to support its growth and virulence within host tissues. This enzymatic activity plays a key role in contributing to inflammatory processes and tissue degradation.

# 2.2.5. Treponema Denticola

*Treponema denticola* is an oral anaerobic bacterial species associated with chronic periodontitis [52]. The main site for *T. denticola* habitation in the oral cavity is the gingival crevice. In patients with coronary artery disease, the relative abundance of *Treponema* species were found to be increased in subgingival plaque and whole mouth samples [38]. Of note, *Treponema denticola* has been detected in both coronary artery plaque samples and subgingival samples [35–37], further suggesting the potential involvement of *Treponema denticola* in coronary artery disease.

*Treponema denticola* is known as an invasive spirochete. Previous animal studies indicated that it can penetrate gingival tissues and circulate through blood vessels, with the potential to invade the heart and cardiovascular endothelium in medium to large arteries, including the aorta, coronary, and carotid arteries [53]. A key virulence factor of T. denticola is the chymotrypsin-like proteinase (Td-CTLP), which activates matrix metalloproteinases, particularly pro-MMP-8 and pro-MMP-9, facilitating the degradation of extracellular matrix components. This enzymatic activity plays a pivotal role in inflammation and tissue remodeling. Moreover, Td-CTLP has been found to degrade important proteinase inhibitors, including TIMP-1, TIMP-2, and  $\alpha$ -1-antichymotrypsin, as well as the complement component C1q, further contributing to immune evasion and tissue damage.

Several other oral microbial species, including *Tannerella forsythia*, *Campylobacter rectus*, *Parvimonas micra*, and *Eubacterium timidum*, have also been linked to oral dysbiosis and cardiovascular disease. However, most current research on the relationship between oral microbiome and CVD has concentrated on bacteria taxa specifically associated with periodontitis [54,55]. This limited focus highlights the need for broader investigations into the role of the entire oral microbiome (e.g.,at community level) in cardiovascular health. Further studies are also warranted to explore the role of other microbial taxa of the oral microbiome community in CVD and to elucidate the potential biological mechanisms involved.

#### 3. Oral Microbiome and Gut Microbiome

Similarities have been detected between the compositions of the oral and gut microbiota [56]. It is estimated that approximately 45% of the microbial species may overlap between these two ecosystems [57]. The oral microbiota primarily consists of five dominant phyla: *Proteobacteria, Firmicutes, Bacteroidetes, Actinobacteria, and Fusobacteriota* [58]. Some of these phyla, such as Firmicutes, Bacteroidetes, and Actinobacteria, are also predominant in the gut microbiota. The predominant genera in the oral cavity may vary across populations from different geographical locations. For example, *Neisseria* is the dominant genus in the Chinese population, *Veillonella* in the Canadian population, and Prevotella in the Qatari population [59,60].

Currently, the mechanisms of oral-gut microbiota interactions in the progression of CVD remain not fully understood. Evidence suggests that oral microbiota can trigger gut dysbiosis either through direct translocation to the gut or via other systemic pathways. In a large-scale study analyzing oral and gut microbiota across participants from five countries, bioinformatic analyses demonstrated that approximately 10% of the oral microbiota successfully transfers and establishes itself in the gut [61]. Our previous research on the gut microbiome identified that the well-known oral bacteria, Fusobacterium nucleatum, an opportunistic pathogen, can also be found in the gut. Fusobacterium nucleatum in the gut has been positively associated with arterial plaque formation [47,48].

Most current studies are animal-based, typically involving the oral administration of microbiota to mice and then investigating subsequent changes in gut microbiota. For example, after introducing *Fusobacterium nucleatum* orally to healthy mice, researchers observed changes in the fecal microbiota, including an increased abundance of *Fusobacterium nucleatum* and elevated levels of autophagy markers in colorectal tissues. This effect was mitigated or eliminated by the administration of antibiotics, such as metronidazole [62].

The transfer of opportunistic pathogens has been observed to occur more frequently among individuals with underlying disease conditions. The presence of certain specific pathogens, such as *Fusobacterium nucleatum* subspecies, may also promote this transfer and subsequently worsen the severity of the disease [62].

Other examples highlighting the link between the oral and gut microbiomes in relation to CVD include the oral-mediated alterations in the proportions of gut *Bacteroidetes* and *Firmicutes*, which serve as important biomarkers in patients with coronary heart disease and stroke [63,64]. In animal studies, the oral administration of *Porphyromonas gingivalis* has been shown to influence gut microbial composition, particularly altering the proportions of *Bacteroidetes* and *Firmicutes*. It was shown that introducing *Porphyromonas gingivalis* leads to a clear separation of gut microbiota composition, with an elevation in *Bacteroidetes* and a reduction in *Firmicutes*, coupled with enhanced intestinal permeability and compromised barrier function [65]. In C57BL/6 mice, oral administration of *Porphyromonas gingivalis* resulted in a decrease in the relative abundance of *Bacteroidetes* in gut [66].

Thus, we hypothesize that the oral microbiota may influence the gut microbiota and contribute to the progression of CVD, with significant therapeutic potential. For example, oral microbiota transplantation (OMT) has shown promise in mitigating detrimental alterations in both oral and gut bacteria[55], suggesting a potentially positive impact on CVD management.

#### 4. Potential Mechanisms by Which the Oral Microbiota Affect CVD

#### 4.1. Oral Microbial Translocation

One plausible mechanism by which oral microbiota dysbiosis affects the development of CVD is through the translocation of oral bacteria, particularly the invasion of pathogenic bacteria into the circulation [16]. Oral inflammation enhances the permeability of periodontal blood vessels, facilitating the entry of oral bacteria into the systemic circulation, where they colonize atherosclerotic plaques and intensify inflammatory responses.

Previous human and animal studies have provided substantial evidence supporting this mechanism. Traces of DNA, RNA, or antigens from oral commensal bacteria, including Porphyromonas gingivalis, Actinobacillus actinomycetemcomitans [67], and Veillonella species

[42,68],, have been identified in atheromatous plaques, suggesting that various pathogenic species are capable of reaching and colonizing these affected sites. Importantly, in 2005, Kozarov et al. found the presence of vital *Porphyromonas gingivalis* and *Actinobacillus actinomycetemcomitans* inside atheromatous tissue cultured with primary human coronary endothelial cells, providing strong evidence for the oral bacteria translocation hypothesis [31].

A previous study has identified 23 unique oral commensal bacterial present within the atherosclerotic plaques, in individuals undergoing interventional procedures[30].

Evidence from animal models highlights the causal role of *Porphyromonas gingivalis* invasion in arterial endothelial cells, contributing to atherosclerotic plaque formation. In *Porphyromonas gingivalis* inoculated mice, administration of the antibiotic metronidazole completely prevented the development of atherosclerotic lesions in mice fed a normal diet, and notably diminished the severity and extent of lesions in those on a high-fat diet.[69].

In addition, the oral microbiota may contribute to gut dysbiosis through oral-gut microbiota transfer, which increases gut permeability and promotes bacterial translocation. These changes are associated with chronic inflammation [16]. This process allows endotoxins to breach compromised gut membranes and directly enter the circulation, ultimately contributing to the development of CVD [70].

## 4.2. Inflammation and Immune Responses

Evidence has demonstrated that alterations in the oral microbiota significantly contribute to both local and systemic inflammation, impacting cardiometabolic health and accelerating the development of CVD [71]. Oral microbiota dysbiosis and elevated levels of periodontal pathogens induce immune responses, including the activation of neutrophils and macrophages. These immune responses also engage cells such as dendritic and gamma delta cells, which subsequently release proinflammatory mediators [72]. The persistence of this inflammatory response is closely linked to oral microbiota imbalance, perpetuating a harmful feedback loop[72].

Oral inflammatory lesions and elevated levels of specific oral bacteria lead to the release of proinflammatory cytokines into the bloodstream, promoting atherosclerotic plaque formation through the propagation of these inflammatory mediators [16]. This systemic inflammatory state is featured by increased levels of acute-phase proteins, pro-inflammatory cytokines and chemokines such as interleukin (IL)-6, and fibrinogen [73]. Additionally, oral bacteria entering the bloodstream and disruptions to gut microbiota through oral-gut microbial transfer may intensify systemic inflammation and contribute to the progression of cardiovascular disease[61,65].

#### 4.2.1. Lipopolysaccharide

Lipopolysaccharide (LPS), commonly known as endotoxin, is a component of the outer membrane of Gram-negative bacteria. Previous studies have highlighted the significant role of LPS in promoting inflammation which in turn may contribute to the progression of atherosclerosis. LPS acts as an immunostimulator or immunomodulator that can translocate from the oral cavity into the circulation, causing endotoxaemia and endothelial dysfunction [74]. Once in the circulation, LPS undergoes a transfer cascade, first binding to lipopolysaccharide binding protein (LBP), then to CD14, and finally to toll-like receptor 4 (TLR4). This binding sequence activates an inflammatory response involving the release of pro-inflammatory cytokines from targeted cells [75]. TLR4 has been detected in human atherosclerotic plaques and has been shown to facilitate atherosclerosis development in mouse models [76,77]. Moreover, elevated serum levels of LPS or LBP have been linked to an increased risk of cardiovascular disease [78,79].

Previous reports indicated that the keystone periodontal pathogen Porphyromonas gingivalis secretes LPS, which interferes with polymorphonuclear leukocyte function by interacting with adhesion molecules like IL-8, ICAM-1, and E-selectin. This interaction disrupts leukocyte recruitment and hampers the immune response[80,81].

# 4.2.2. Cytokines

Pro-inflammatory cytokines are critical mediators that link the oral microbiome to inflammatory atherosclerosis. Epidemiological evidence has demonstrated that elevated levels of cytokines like IL-1, IL-6, and TNF are associated with increased cardiovascular risk[82,83]. Many of these pro-inflammatory cytokines are linked with known oral pathogens. For example, in human studies using 16S sequencing data, oral *Porphyromonas* was positively associated with salivary cytokines such as IL-1 $\beta$ , IL-2, IL-8, and IL-13, while *Fusobacterium* was associated with IL-1 $\beta$  [84]. Additionally, oral *Streptococcus* species showed positive correlations with multiple cytokines, including IL-1 $\beta$ , IL-2, IL-4, IL-6, IL-7, IL-9, IL-12, and IL-17 [85].

The well-studied oral pathogen *Porphyromonas gingivalis* expresses autoinducer 2, which induces the secretion of IL-8 in oral epithelial cells [86]. This cytokine, along with IL-6 and MCP1, is linked to endothelial dysfunction, characterized by increased procoagulant properties, mononuclear cell adhesion, and elevated expression of cell adhesion molecules [87,88]. Additionally, both *Porphyromonas gingivalis* and *Fusobacterium nucleatum* can act on macrophages, neutrophils, and monocytes to induce the production of TNF- $\alpha$ , IL-6, and IL-8 [85]. An animal study indicated that in mice, infection with *Porphyromonas gingivalis* induces the accumulation of macrophages and inflammatory mediators like CD40, IFN- $\gamma$ , IL-1 $\beta$ , IL-6, and TNF- $\alpha$  in atherosclerotic lesions, with these responses being milder in immunodeficient mice [89].

Elevated concentrations of bacterial surface molecules stimulate the production of various inflammatory mediators and cytokines, fueling both local and systemic inflammatory responses[90,91]. These processes are driven through the activation of several inflammatory pathways, including matrix metalloproteinase 9 (MMP9), Nuclear factor kappa-B (NF- $\kappa$ B), and Basic Helix-Loop-Helix ARNT-Like 1 (BMAL1) [21,92]. Under the influence of pro-inflammatory agents such as TNF- $\alpha$ , IL-6, and transforming growth factor  $\beta$  (TGF $\beta$ ), epithelial and immune cells are prompted to produce reactive oxygen species (ROS), reactive nitrogen species, and matrix metalloproteinases, which in turn activate the NF- $\kappa$ B signaling pathway, further amplifying the inflammatory response [93].

In addition to the aforementioned specific pathogens, other oral bacteria are also capable of inciting harmful inflammation that engages both the innate and adaptive immune systems [94]. The mechanisms underscore the interaction between pro-inflammatory cytokines driven by the oral microbiota, which still requires further studies.

# 4.3. Modulation of Platelet Aggregation

One potential mechanism by which the oral microbiota influences the progression of CVD is through modulation of platelet aggregation, which can exacerbate inflammatory responses and contribute to the development of atherosclerosis and thromboembolic events.

Oral bacteria can stimulate platelets either by direct interaction and activation or indirectly by releasing platelet-activating factors. For instance, viridans group streptococci (VGS), such as *Streptococcus sanguinis, Streptococcus gordonii, Streptococcus mutans*, and *Streptococcus mitis*, have been shown to promote platelet adhesion and aggregation in vitro via various surface proteins, including platelet aggregation-associated protein (PAAP), serine-rich glycoproteins, adhesins, and glucosyltransferases [95–98]. Moreover, the immune response triggered by oral bacteria can induce platelet activation[96]. This process contributes to localized thrombus formation, depletion of platelets, and an increase in the secretion of pro-inflammatory cytokines and mediators by the activated platelets, thus playing a role in promoting inflammation, atherogenesis, and thrombosis [99].

#### 4.4. Oral Microbial Metabolites

#### 4.4.1. Trimethylamine N-Oxide

Trimethylamine N-oxide (TMAO) is recognized for its detrimental role in promoting the progression of cardiovascular disease. TMAO affects cardiovascular health by inhibiting reverse cholesterol transport, reducing bile acid synthesis [100], promoting platelet reactivity and thrombosis potential [101,102], and instigating vascular inflammation [103] TMAO is generated through microbial and host biochemical reactions, starting with the production of trimethylamine (TMA) from dietary choline and carnitine. While the gut microbiome is the primary source of TMA, emerging evidence suggests that oral bacteria, especially *Prevotella* and *Fusobacterium*, can also produce TMA. Studies have shown that patients with periodontal disease have increased TMAO levels, linking oral pathogens to increased cardiovascular risk[104]. Additionally, a recent study highlighted the association between the oral microbiome and TMAO levels in patients with myocardial infarction[105].

# 4.4.2. Short-Chain Fatty Acids

SCFAs are important metabolites produced by the microbiota, playing a vital role in the host's regulation of gluconeogenesis, lipid metabolism, and inflammatory response [106].. Bacterial species in the oral cavity, such as *Streptococcus*, *Actinomyces*, *Lactobacillus*, *Propionibacterium*, and *Prevotella* [107], can utilize carbohydrate-active enzymes to degrade carbohydrates into SCFAs, which sustain their energy requirements[108]. Dietary patterns, particularly high sugar intake, significantly influence the oral microbiota composition and SCFA production levels [109]. There is conflicting evidence regarding the effects of SCFAs in the oral cavity and the circulatory system. On one hand, SCFAs have anti-inflammatory effects in the plasma by inhibiting the NF-kB and Akt signaling pathways and lowering cytokine levels[64]. Moreover, they inhibit histone deacetylases (HDACs) and engage specific G protein-coupled receptors (GPRs), offering cardiovascular protection [110]. On the other hand, SCFAs can also affect the expression of connexins and adhesion proteins, which may compromise oral epithelial cell function [111], while still exerting systemic anti-inflammatory benefits that contribute to cardiovascular health.

#### 4.4.3. Nitric Oxide

Nitric oxide (NO) can play a beneficial role in vascular function due to its vasodilatory effects. It helps lower blood pressure, protect endothelial cells, and counteract the progression of atherosclerosis. Additionally, NO contributes to inflammation regulation, providing cardiovascular benefits by reducing inflammatory responses in blood vessels [112]. A deficiency in NO has been strongly linked to the development of cardiovascular disease, making it a significant marker for predicting cardiovascular events [113]. The oral microbiota contributes substantially to the production of NO, serving as a vital reservoir for NO within both the bloodstream and tissues [114]. Insufficient synthesis of NO by endogenous nitric oxide synthase (NOS) can be compensated by nitrite (NO2-) produced by oral microbiota. Current evidence indicate that alterations in the oral microbiota can significantly impact NO levels, thereby affecting the progression of CVD[115].

# 4.4.4. Hydrogen Sulfide

Hydrogen sulfide (H2S) is a crucial endogenous gaseous signaling molecule involved in cardiovascular function [116]. It exerts antioxidant and anti-inflammatory effects, improves insulin resistance, and helps regulate blood pressure and atherosclerosis [117]. Elevated H2S levels have been linked to protection against hypertension and diabetic cardiomyopathy [118]. It may interact with nitric oxide (NO) to provide additional cardiovascular benefits [119].

However, a paradox arises with H2S in the oral cavity. While systemic H2S provides protective effects, increased H2S levels in the oral environment have been linked to heightened oral inflammation [120]. The oral microbiota, particularly proteolytic bacteria such as Prevotella and

Porphyromonas, hold the ability to produce H2S from sulfur-containing amino acids[116]. In healthy individuals, H2S levels are low, but in disease states where microbial loads increase, H2S production is significantly elevated [121]. The impact of oral microbiota-derived H2S on systemic H2S levels and cardiovascular health requires further investigation [122].

## 5. Research Gaps and Future Directions

While substantial evidence links the human microbiome to multiple CVD outcomes, most research has focused on the gut microbiome, with less known about the relationship between the oral microbiome and CVD, despite growing interest in this field. The oral environment differs significantly from the intestinal environment, so mechanisms linking the gut microbiome and its products to CVD may not fully apply to the oral cavity. Additionally, the oral microbiota, positioned upstream of the digestive tract, can also influence gut microbiota composition and function. Distinct mechanisms may be involved in how the gut and oral microbiomes contribute to the progression of CVD, which warrants further investigation.

Importantly, most current studies on the link between oral bacteria and CVD are cross-sectional and often focus on specific bacteria, especially pathogens associated with periodontitis or other oral diseases. Two important aspects remain underexplored. First, how do alterations in the overall oral microbiome community, as well as oral bacteria other than periodontal pathogens, contribute to the development of CVD? Second, what are the mechanisms by which oral dysbiosis or oral microbiome alterations, rather than periodontal disease, influence the development of CVD? Further investigations, including large-scale prospective population studies and intervention studies, are needed to establish causal relationships between the oral microbiome and CVD and determine the biological validity of the proposed pathophysiological mechanisms discussed in this review.

The integrative analysis of multi-omics techniques (e.g., metagenomics, metatranscriptomics, metabolomics, and proteomics) may improve the current understanding. Integrated multi-omics studies, along with standardized and comparable methodologies, will facilitate a deeper and comprehensive understanding of the metabolic and functional roles of the oral microbiome in cardiovascular health. This approach will help clarify some of the hypothesized inflammatory mechanisms connecting oral microbiome alterations to CVD development. Moreover, it will contribute to the development of targeted interventions and effective therapies aimed at preventing or reducing the progression from cardiovascular risk to clinical CVD events.

#### 6. Conclusions

In summary, current evidence indicates the significant role of the oral microbiome in the development and progression of CVD. This review highlighted several well-established CVD-associated oral bacteria (e.g. Porphyromonas gingivalis, Fusobacterium nucleatum, and Aggregatibacter actinomycetemcomitans), and discussed the mechanisms underlying these associations. The potential mechanisms by which the oral microbiota can influence CVD progression include: oral and systemic inflammation; immune responses and cytokine release; oral bacteria translocation; and through microbial-related products, such as metabolites (e.g. TMAO, SCFAs, NO, H2S etc.) and toxins (e.g. LPS). Despite substantial progress in understanding these mechanisms, considerable gaps in knowledge remain, particularly regarding the impact of overall oral microbiome alterations beyond periodontal pathogens. Future research should aim to unravel these complex interactions through large-scale, prospective studies and multi-omics approaches. These efforts will be essential for developing targeted interventions and effective therapies to mitigate the risk and progression of CVD, leading to more effective strategies for CVD prevention and management.

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#### References

- 1. Roth, G.A.; Mensah, G.A.; Johnson, C.O.; Addolorato, G.; Ammirati, E.; Baddour, L.M.; Barengo, N.C.; Beaton, A.Z.; Benjamin, E.J.; Benziger, C.P. *et al.* Global Burden of Cardiovascular Diseases and Risk Factors, 1990-2019: Update from the GBD 2019 Study. J Am Coll Cardiol **2020**, *76*, 2982-3021.
- Witkowski, M.; Weeks, T.L.; Hazen, S.L. Gut Microbiota and Cardiovascular Disease. Circ Res 2020, 127, 553-570.
- 3. The Integrative Human Microbiome Project: Dynamic Analysis of Microbiome-Host Omics Profiles during Periods of Human Health and Disease. Cell Host Microbe **2014**, *16*, 276-289.
- 4. Honda, K.; Littman, D.R. The Microbiota in Adaptive Immune Homeostasis and Disease. Nature **2016**, 535, 75-84.
- 5. Shreiner, A.B.; Kao, J.Y.; Young, V.B. The Gut Microbiome in Health and in Disease. Curr Opin Gastroenterol **2015**, *31*, 69-75.
- 6. DeGruttola, A.K.; Low, D.; Mizoguchi, A.; Mizoguchi, E. Current Understanding of Dysbiosis in Disease in Human and Animal Models. Inflamm Bowel Dis **2016**, 22, 1137-1150.
- 7. Tang, W.H.W.; Kitai, T.; Hazen, S.L. Gut Microbiota in Cardiovascular Health and Disease. Circ Res **2017**, *120*, 1183-1196.
- 8. Fan, Y.; Pedersen, O. Gut Microbiota in Human Metabolic Health and Disease. Nat Rev Microbiol **2021**, 19, 55-71.
- 9. Verma, D.; Garg, P.K.; Dubey, A.K. Insights into the Human Oral Microbiome. Arch Microbiol 2018, 200, 525-540.
- 10. Lamont, R.J.; Koo, H.; Hajishengallis, G. The Oral Microbiota: Dynamic Communities and Host Interactions. Nat Rev Microbiol **2018**, *16*, 745-759.
- 11. Kinane, D.F.; Stathopoulou, P.G.; Papapanou, P.N. Periodontal Diseases. Nat Rev Dis Primers **2017**, *3*, 17038.
- 12. Bui, F.Q.; Almeida-da-Silva, C.L.C.; Huynh, B.; Trinh, A.; Liu, J.; Woodward, J.; Asadi, H.; Ojcius, D.M. Association between Periodontal Pathogens and Systemic Disease. Biomed J **2019**, *42*, 27-35.
- 13. Nikitakis, N.G.; Papaioannou, W.; Sakkas, L.I.; Kousvelari, E. The Autoimmunity-Oral Microbiome Connection. Oral Dis **2017**, *23*, 828-839.
- 14. 14. Kholy, K.E.; Genco, R.J.; Van Dyke, T.E. Oral Infections and Cardiovascular Disease. Trends Endocrinol Metab **2015**, *26*, 315-321.
- 15. Sanz, M.; Marco Del Castillo, A.; Jepsen, S.; Gonzalez-Juanatey, J.R.; D'Aiuto, F.; Bouchard, P.; Chapple, I.; Dietrich, T.; Gotsman, I.; Graziani, F. *et al.* Periodontitis and Cardiovascular Diseases: Consensus Report. J Clin Periodontol **2020**, *47*, 268-288.
- 16. Tonelli, A.; Lumngwena, E.N.; Ntusi, N.A.B. The Oral Microbiome in the Pathophysiology of Cardiovascular Disease. Nat Rev Cardiol **2023**, *20*, 386.
- 17. Socransky, S.S.; Haffajee, A.D. Evidence of Bacterial Etiology: A Historical Perspective. Periodontol 2000 **1994**, *5*, 7-25.
- 18. Socransky, S.S.; Haffajee, A.D.; Cugini, M.A.; Smith, C.; Kent, R.L. Microbial Complexes in Subgingival Plaque. J Clin Periodontol **1998**, *25*, 134-144.
- 19. Forner, L.; Larsen, T.; Kilian, M.; Holmstrup, P. Incidence of Bacteremia After Chewing, Tooth Brushing and Scaling in Individuals with Periodontal Inflammation. J Clin Periodontol **2006**, *33*, 401-407.
- 20. 20. Lucchese, A. Streptococcus Mutans Antigen I/II and Autoimmunity in Cardiovascular Diseases. Autoimmun Rev **2017**, *16*, 456-460.
- 21. Zie, M.; Tang, Q.; Nie, J.; Zhang, C.; Zhou, X.; Yu, S.; Sun, J.; Cheng, X.; Dong, N.; Hu, Y. *et al.* BMAL1-Downregulation Aggravates Porphyromonas Gingivalis-Induced Atherosclerosis by Encouraging Oxidative Stress. Circ Res **2020**, *126*, e15-e29.
- 22. Zeng, X.; Leng, W.; Lam, Y.; Yan, B.P.; Wei, X.; Weng, H.; Kwong, J.S.W. Periodontal Disease and Carotid Atherosclerosis: A Meta-Analysis of 17,330 Participants. Int J Cardiol **2016**, 203, 1044-1051.
- 23. Wang, J.; Geng, X.; Sun, J.; Zhang, S.; Yu, W.; Zhang, X.; Liu, H. The Risk of Periodontitis for Peripheral Vascular Disease: A Systematic Review. Rev Cardiovasc Med **2019**, *20*, 81-89.
- 24. 24. Kaschwich, M.; Behrendt, C.; Heydecke, G.; Bayer, A.; Debus, E.S.; Seedorf, U.; Aarabi, G. The Association of Periodontitis and Peripheral Arterial Occlusive Disease-A Systematic Review. Int J Mol Sci **2019**, *20*, 2936.
- 25. DeStefano, F.; Anda, R.F.; Kahn, H.S.; Williamson, D.F.; Russell, C.M. Dental Disease and Risk of Coronary Heart Disease and Mortality. BMJ 1993, 306, 688-691.

- 26. Sen, S.; Giamberardino, L.D.; Moss, K.; Morelli, T.; Rosamond, W.D.; Gottesman, R.F.; Beck, J.; Offenbacher, S. Periodontal Disease, Regular Dental Care use, and Incident Ischemic Stroke. Stroke 2018, 49, 355-362.
- 27. Conroy, R.M.; Pyörälä, K.; Fitzgerald, A.P.; Sans, S.; Menotti, A.; De Backer, G.; De Bacquer, D.; Ducimetière, P.; Jousilahti, P.; Keil, U. *et al.* Estimation of Ten-Year Risk of Fatal Cardiovascular Disease in Europe: The SCORE Project. Eur Heart J **2003**, *24*, 987-1003.
- 28. 28. Hollander, W. Role of Hypertension in Atherosclerosis and Cardiovascular Disease. Am J Cardiol 1976, 38, 786-800.
- 29. Li, Y.; Zhu, M.; Liu, Y.; Luo, B.; Cui, J.; Huang, L.; Chen, K.; Liu, Y. The Oral Microbiota and Cardiometabolic Health: A Comprehensive Review and Emerging Insights. Front Immunol 2022, 13, 1010368.
- 30. Chhibber-Goel, J.; Singhal, V.; Bhowmik, D.; Vivek, R.; Parakh, N.; Bhargava, B.; Sharma, A. Linkages between Oral Commensal Bacteria and Atherosclerotic Plaques in Coronary Artery Disease Patients. npj Biofilms and Microbiomes **2016**, 2.
- 31. Kozarov, E.V.; Dorn, B.R.; Shelburne, C.E.; Dunn, W.A.; Progulske-Fox, A. Human Atherosclerotic Plaque Contains Viable Invasive Actinobacillus Actinomycetemcomitans and Porphyromonas Gingivalis. Arterioscler Thromb Vasc Biol **2005**, *25*, 17.
- 32. Deshpande, R.G.; Khan, M.B.; Genco, C.A. Invasion of Aortic and Heart Endothelial Cells by Porphyromonas Gingivalis. Infect Immun 1998, 66, 5337-5343.
- 33. Mysak, J.; Podzimek, S.; Sommerova, P.; Lyuya-Mi, Y.; Bartova, J.; Janatova, T.; Prochazkova, J.; Duskova, J. Porphyromonas Gingivalis: Major Periodontopathic Pathogen Overview. J Immunol Res **2014**, 2014, 476068.
- 34. Serra e Silva Filho, Wagner; Casarin, R.C.V.; Nicolela, E.L.; Passos, H.M.; Sallum, A.W.; Gonçalves, R.B. Microbial Diversity Similarities in Periodontal Pockets and Atheromatous Plaques of Cardiovascular Disease Patients. PLoS One **2014**, *9*, e109761.
- 35. Mahalakshmi, K.; Krishnan, P.; Arumugam, S.B. "Association of Periodontopathic Anaerobic Bacterial Co-Occurrence to Atherosclerosis" A Cross-Sectional Study. Anaerobe **2017**, *44*, 66-72.
- Nikolaeva, E.N.; Tsarev, V.N.; Tsareva, T.V.; Ippolitov, E.V.; Arutyunov, S.D. Interrelation of Cardiovascular Diseases with Anaerobic Bacteria of Subgingival Biofilm. Contemp Clin Dent 2019, 10, 637-642
- 37. Kannosh, I.; Staletovic, D.; Toljic, B.; Radunovic, M.; Pucar, A.; Matic Petrovic, S.; Grubisa, I.; Lazarevic, M.; Brkic, Z.; Knezevic Vukcevic, J. *et al.* The Presence of Periopathogenic Bacteria in Subgingival and Atherosclerotic Plaques an Age Related Comparative Analysis. J Infect Dev Ctries **2018**, *12*, 1088-1095.
- 38. 38. Fåk, F.; Tremaroli, V.; Bergström, G.; Bäckhed, F. Oral Microbiota in Patients with Atherosclerosis. Atherosclerosis **2015**, 243, 573-578.
- 39. Ziebolz, D.; Rost, C.; Schmidt, J.; Waldmann-Beushausen, R.; Schöndube, F.A.; Mausberg, R.F.; Danner, B.C. Periodontal Bacterial DNA and their Link to Human Cardiac Tissue: Findings of a Pilot Study. Thorac Cardiovasc Surg **2018**, *66*, 83-90.
- 40. 40. Su, C.; Shigeishi, H.; Nishimura, R.; Ohta, K.; Sugiyama, M. Detection of Oral Bacteria on the Tongue Dorsum using PCR Amplification of 16S Ribosomal RNA and its Association with Systemic Disease in Middle-Aged and Elderly Patients. Biomed Rep 2019, 10, 70-76.
- 41. 41. Schenkein, H.A.; Loos, B.G. Inflammatory Mechanisms Linking Periodontal Diseases to Cardiovascular Diseases. J Clin Periodontol **2013**, 40 Suppl 14, 51.
- 42. 42. Koren, O.; Spor, A.; Felin, J.; Fåk, F.; Stombaugh, J.; Tremaroli, V.; Behre, C.J.; Knight, R.; Fagerberg, B.; Ley, R.E. *et al.* Human Oral, Gut, and Plaque Microbiota in Patients with Atherosclerosis. Proc. Natl. Acad. Sci. U. S. A. **2011**, *108*, 4592-4598.
- 43. 43. Huh, J.-.; Roh, T.-. Opportunistic Detection of Fusobacterium Nucleatum as a Marker for the Early Gut Microbial Dysbiosis. BMC Microbiol. **2020**, *20*.
- 44. Lund Håheim, A.L. Oral Anaerobe Bacteria-a Common Risk for Cardiovascular Disease and Mortality and some Forms of Cancer? Front Oral Health **2024**, *5*, 1348946.
- 45. Ziebolz, D.; Jahn, C.; Pegel, J.; Semper-Pinnecke, E.; Mausberg, R.F.; Waldmann-Beushausen, R.; Schöndube, F.A.; Danner, B.C. Periodontal Bacteria DNA Findings in Human Cardiac Tissue is there a Link of Periodontitis to Heart Valve Disease? Int J Cardiol **2018**, *251*, 74-79.
- 46. 46. Figuero, E.; Sánchez-Beltrán, M.; Cuesta-Frechoso, S.; Tejerina, J.M.; Del Castro, J.A.; Gutiérrez, J.M.; Herrera, D.; Sanz, M. Detection of Periodontal Bacteria in Atheromatous Plaque by Nested Polymerase Chain Reaction. J. Periodontol. **2011**, *82*, 1469-1477.
- 47. 47. Wang, Z.; Peters, B.A.; Usyk, M.; Xing, J.; Hanna, D.B.; Wang, T.; Post, W.S.; Landay, A.L.; Hodis, H.N.; Weber, K. *et al.* Gut Microbiota, Plasma Metabolomic Profiles, and Carotid Artery Atherosclerosis in HIV Infection. Arterioscler. Thromb. Vasc. Biol. **2022**, 101161ATVBAHA121317276.

- 48. 48. Wang, Z.; Peters, B.A.; Bryant, M.; Hanna, D.B.; Schwartz, T.; Wang, T.; Sollecito, C.C.; Usyk, M.; Grassi, E.; Wiek, F. *et al.* Gut Microbiota, Circulating Inflammatory Markers and Metabolites, and Carotid Artery Atherosclerosis in HIV Infection. Microbiome **2023**, *11*, 119.
- 49. Leskelä, J.; Pietiäinen, M.; Safer, A.; Lehto, M.; Metso, J.; Malle, E.; Buggle, F.; Becher, H.; Sundvall, J.; Grau, A.J. *et al.* Serum Lipopolysaccharide Neutralizing Capacity in Ischemic Stroke. PLoS One **2020**, *15*, e0228806.
- 50. Liljestrand, J.M.; Paju, S.; Pietiäinen, M.; Buhlin, K.; Persson, G.R.; Nieminen, M.S.; Sinisalo, J.; Mäntylä, P.; Pussinen, P.J. Immunologic Burden Links Periodontitis to Acute Coronary Syndrome. Atherosclerosis 2018, 268, 177-184.
- 51. Maki, K.A.; Ganesan, S.M.; Meeks, B.; Farmer, N.; Kazmi, N.; Barb, J.J.; Joseph, P.V.; Wallen, G.R. The Role of the Oral Microbiome in Smoking-Related Cardiovascular Risk: A Review of the Literature Exploring Mechanisms and Pathways. J Transl Med 2022, 20, 584.
- 52. Nieminen, M.T.; Listyarifah, D.; Hagström, J.; Haglund, C.; Grenier, D.; Nordström, D.; Uitto, V.; Hernandez, M.; Yucel-Lindberg, T.; Tervahartiala, T. *et al.* Treponema Denticola Chymotrypsin-Like Proteinase may Contribute to Orodigestive Carcinogenesis through Immunomodulation. Br J Cancer **2018**, 118, 428-434.
- 53. Chukkapalli, S.S.; Rivera, M.F.; Velsko, I.M.; Lee, J.; Chen, H.; Zheng, D.; Bhattacharyya, I.; Gangula, P.R.; Lucas, A.R.; Kesavalu, L. Invasion of Oral and Aortic Tissues by Oral Spirochete Treponema Denticola in ApoE(-/-) Mice Causally Links Periodontal Disease and Atherosclerosis. Infect Immun 2014, 82, 1959-1967.
- 54. Wu, L.; Zeng, T.; Deligios, M.; Milanesi, L.; Langille, M.G.I.; Zinellu, A.; Rubino, S.; Carru, C.; Kelvin, D.J. Age-Related Variation of Bacterial and Fungal Communities in Different Body Habitats Across the Young, Elderly, and Centenarians in Sardinia. mSphere 2020, 5, 558.
- 55. Xiao, H.; Fan, Y.; Li, Y.; Dong, J.; Zhang, S.; Wang, B.; Liu, J.; Liu, X.; Fan, S.; Guan, J. et al. Oral Microbiota Transplantation Fights Against Head and Neck Radiotherapy-Induced Oral Mucositis in Mice. Comput Struct Biotechnol J **2021**, *19*, 5898-5910.
- 56. Donia, M.S.; Cimermancic, P.; Schulze, C.J.; Wieland Brown, L.C.; Martin, J.; Mitreva, M.; Clardy, J.; Linington, R.G.; Fischbach, M.A. A Systematic Analysis of Biosynthetic Gene Clusters in the Human Microbiome Reveals a Common Family of Antibiotics. Cell **2014**, *158*, 1402-1414.
- 57. Segata, N.; Haake, S.K.; Mannon, P.; Lemon, K.P.; Waldron, L.; Gevers, D.; Huttenhower, C.; Izard, J. Composition of the Adult Digestive Tract Bacterial Microbiome Based on Seven Mouth Surfaces, Tonsils, Throat and Stool Samples. Genome Biol **2012**, *13*, R42.
- 58. Cheung, M.K.; Chan, J.Y.K.; Wong, M.C.S.; Wong, P.Y.; Lei, P.; Cai, L.; Lan, L.; Ho, W.C.S.; Yeung, A.C.M.; Chan, P.K.S. *et al.* Determinants and Interactions of Oral Bacterial and Fungal Microbiota in Healthy Chinese Adults. Microbiol Spectr **2022**, *10*, e0241021.
- 59. Nearing, J.T.; DeClercq, V.; Van Limbergen, J.; Langille, M.G.I. Assessing the Variation within the Oral Microbiome of Healthy Adults. mSphere **2020**, *5*, 451.
- 60. Murugesan, S.; Al Ahmad, S.F.; Singh, P.; Saadaoui, M.; Kumar, M.; Al Khodor, S. Profiling the Salivary Microbiome of the Qatari Population. J Transl Med **2020**, *18*, 127.
- 61. Schmidt, T.S.; Hayward, M.R.; Coelho, L.P.; Li, S.S.; Costea, P.I.; Voigt, A.Y.; Wirbel, J.; Maistrenko, O.M.; Alves, R.J.; Bergsten, E. *et al.* Extensive Transmission of Microbes Along the Gastrointestinal Tract. Elife **2019**, *8*, e42693.
- 62. Dong, J.; Li, Y.; Xiao, H.; Zhang, S.; Wang, B.; Wang, H.; Li, Y.; Fan, S.; Cui, M. Oral Microbiota Affects the Efficacy and Prognosis of Radiotherapy for Colorectal Cancer in Mouse Models. Cell Rep **2021**, *37*, 109886.
- 63. Yang, T.; Santisteban, M.M.; Rodriguez, V.; Li, E.; Ahmari, N.; Carvajal, J.M.; Zadeh, M.; Gong, M.; Qi, Y.; Zubcevic, J. *et al.* Gut Dysbiosis is Linked to Hypertension. Hypertension **2015**, *65*, 1331-1340.
- 64. Tsai, H.; Tsai, W.; Hung, W.; Hung, W.; Chang, C.; Dai, C.; Tsai, Y. Gut Microbiota and Subclinical Cardiovascular Disease in Patients with Type 2 Diabetes Mellitus. Nutrients **2021**, *13*, 2679.
- 65. Kato, T.; Yamazaki, K.; Nakajima, M.; Date, Y.; Kikuchi, J.; Hase, K.; Ohno, H.; Yamazaki, K. Oral Administration of Porphyromonas Gingivalis Alters the Gut Microbiome and Serum Metabolome. mSphere 2018, 3, 460.
- 66. Nakajima, M.; Arimatsu, K.; Kato, T.; Matsuda, Y.; Minagawa, T.; Takahashi, N.; Ohno, H.; Yamazaki, K. Oral Administration of P. Gingivalis Induces Dysbiosis of Gut Microbiota and Impaired Barrier Function Leading to Dissemination of Enterobacteria to the Liver. PLoS One **2015**, *10*, e0134234.
- 67. Haraszthy, V.I.; Hariharan, G.; Tinoco, E.M.; Cortelli, J.R.; Lally, E.T.; Davis, E.; Zambon, J.J. Evidence for the Role of Highly Leukotoxic Actinobacillus Actinomycetemcomitans in the Pathogenesis of Localized Juvenile and Other Forms of Early-Onset Periodontitis. J Periodontol 2000, 71, 912-922.
- 68. Koren, O.; Spor, A.; Felin, J.; Fåk, F.; Stombaugh, J.; Tremaroli, V.; Behre, C.J.; Knight, R.; Fagerberg, B.; Ley, R.E. *et al.* Human Oral, Gut, and Plaque Microbiota in Patients with Atherosclerosis. Proc Natl Acad Sci U S A **2011**, *108 Suppl 1*, 4592-4598.

- 69. Amar, S.; Wu, S.; Madan, M. Is Porphyromonas Gingivalis Cell Invasion Required for Atherogenesis? Pharmacotherapeutic Implications. J Immunol **2009**, *182*, 1584-1592.
- 70. Francisqueti-Ferron, F.V.; Nakandakare-Maia, E.T.; Siqueira, J.S.; Ferron, A.J.T.; Vieira, T.A.; Bazan, S.G.Z.; Corrêa, C.R. The Role of Gut Dysbiosis-Associated Inflammation in Heart Failure. Rev Assoc Med Bras (1992) 2022, 68, 1120-1124.
- 71. 71. Pietiäinen, M.; Liljestrand, J.M.; Kopra, E.; Pussinen, P.J. Mediators between Oral Dysbiosis and Cardiovascular Diseases. Eur J Oral Sci 2018, 126 Suppl 1, 26-36.
- 72. Abusleme, L.; Dupuy, A.K.; Dutzan, N.; Silva, N.; Burleson, J.A.; Strausbaugh, L.D.; Gamonal, J.; Diaz, P.I. The Subgingival Microbiome in Health and Periodontitis and its Relationship with Community Biomass and Inflammation. ISME J 2013, 7, 1016-1025.
- 73. Buhlin, K.; Hultin, M.; Norderyd, O.; Persson, L.; Pockley, A.G.; Rabe, P.; Klinge, B.; Gustafsson, A. Risk Factors for Atherosclerosis in Cases with Severe Periodontitis. J Clin Periodontol **2009**, *36*, 541-549.
- 74. Moran, A.P.; Prendergast, M.M.; Appelmelk, B.J. Molecular Mimicry of Host Structures by Bacterial Lipopolysaccharides and its Contribution to Disease. FEMS Immunol Med Microbiol **1996**, *16*, 105-115.
- 75. Fitzgerald, K.A.; Kagan, J.C. Toll-Like Receptors and the Control of Immunity. Cell **2020**, *180*, 1044-1066.
- 76. Edfeldt, K.; Swedenborg, J.; Hansson, G.K.; Yan, Z. Expression of Toll-Like Receptors in Human Atherosclerotic Lesions: A Possible Pathway for Plaque Activation. Circulation **2002**, *105*, 1158-1161.
- 77. Michelsen, K.S.; Wong, M.H.; Shah, P.K.; Zhang, W.; Yano, J.; Doherty, T.M.; Akira, S.; Rajavashisth, T.B.; Arditi, M. Lack of Toll-Like Receptor 4 Or Myeloid Differentiation Factor 88 Reduces Atherosclerosis and Alters Plaque Phenotype in Mice Deficient in Apolipoprotein E. Proc Natl Acad Sci U S A **2004**, *101*, 10679-10684.
- 78. Asada, M.; Oishi, E.; Sakata, S.; Hata, J.; Yoshida, D.; Honda, T.; Furuta, Y.; Shibata, M.; Suzuki, K.; Watanabe, H. *et al.* Serum Lipopolysaccharide-Binding Protein Levels and the Incidence of Cardiovascular Disease in a General Japanese Population: The Hisayama Study. J Am Heart Assoc **2019**, *8*, e013628.
- 79. Pastori, D.; Carnevale, R.; Nocella, C.; Novo, M.; Santulli, M.; Cammisotto, V.; Menichelli, D.; Pignatelli, P.; Violi, F. Gut-Derived Serum Lipopolysaccharide is Associated with Enhanced Risk of Major Adverse Cardiovascular Events in Atrial Fibrillation: Effect of Adherence to Mediterranean Diet. J Am Heart Assoc 2017, 6, e005784.
- 80. Bo. Darveau, R.P. Periodontitis: A Polymicrobial Disruption of Host Homeostasis. Nat Rev Microbiol **2010**, *8*, 481-490.
- 81. Hajishengallis, E.; Hajishengallis, G. Neutrophil Homeostasis and Periodontal Health in Children and Adults. J Dent Res **2014**, *93*, 231-237.
- 82. Amin, M.N.; Siddiqui, S.A.; Ibrahim, M.; Hakim, M.L.; Ahammed, M.S.; Kabir, A.; Sultana, F. Inflammatory Cytokines in the Pathogenesis of Cardiovascular Disease and Cancer. SAGE Open Med **2020**, *8*, 2050312120965752.
- 83. Kaptoge, S.; Seshasai, S.R.K.; Gao, P.; Freitag, D.F.; Butterworth, A.S.; Borglykke, A.; Di Angelantonio, E.; Gudnason, V.; Rumley, A.; Lowe, G.D.O. *et al.* Inflammatory Cytokines and Risk of Coronary Heart Disease: New Prospective Study and Updated Meta-Analysis. Eur Heart J **2014**, *35*, 578-589.
- 84. Pushalkar, S.; Paul, B.; Li, Q.; Yang, J.; Vasconcelos, R.; Makwana, S.; González, J.M.; Shah, S.; Xie, C.; Janal, M.N. *et al.* Electronic Cigarette Aerosol Modulates the Oral Microbiome and Increases Risk of Infection. iScience **2020**, *23*, 100884.
- 85. So. Joshi, V.; Matthews, C.; Aspiras, M.; de Jager, M.; Ward, M.; Kumar, P. Smoking Decreases Structural and Functional Resilience in the Subgingival Ecosystem. J Clin Periodontol **2014**, *41*, 1037-1047.
- 86. Yee, M.; Kim, S.; Sethi, P.; Düzgüneş, N.; Konopka, K. Porphyromonas Gingivalis Stimulates IL-6 and IL-8 Secretion in GMSM-K, HSC-3 and H413 Oral Epithelial Cells. Anaerobe **2014**, *28*, 62-67.
- 87. Roth, G.A.; Moser, B.; Roth-Walter, F.; Giacona, M.B.; Harja, E.; Papapanou, P.N.; Schmidt, A.M.; Lalla, E. Infection with a Periodontal Pathogen Increases Mononuclear Cell Adhesion to Human Aortic Endothelial Cells. Atherosclerosis **2007**, *190*, 271-281.
- 88. Roth, G.A.; Moser, B.; Huang, S.J.; Brandt, J.S.; Huang, Y.; Papapanou, P.N.; Schmidt, A.M.; Lalla, E. Infection with a Periodontal Pathogen Induces Procoagulant Effects in Human Aortic Endothelial Cells. J Thromb Haemost 2006, 4, 2256-2261.
- 89. Hayashi, C.; Madrigal, A.G.; Liu, X.; Ukai, T.; Goswami, S.; Gudino, C.V.; Gibson, F.C.; Genco, C.A. Pathogen-Mediated Inflammatory Atherosclerosis is Mediated in Part Via Toll-Like Receptor 2-Induced Inflammatory Responses. J Innate Immun 2010, 2, 334-343.
- 90. Ramos, H.C.; Rumbo, M.; Sirard, J. Bacterial Flagellins: Mediators of Pathogenicity and Host Immune Responses in Mucosa. Trends Microbiol **2004**, *12*, 509-517.
- 91. Zindel, J.; Kubes, P. DAMPs, PAMPs, and LAMPs in Immunity and Sterile Inflammation. Annu Rev Pathol **2020**, *15*, 493-518.
- 92. Isola, G.; Polizzi, A.; Ronsivalle, V.; Alibrandi, A.; Palazzo, G.; Lo Giudice, A. Impact of Matrix Metalloproteinase-9 during Periodontitis and Cardiovascular Diseases. Molecules **2021**, *26*, 1777.

- 93. Zhang, Y.; Wang, X.; Li, H.; Ni, C.; Du, Z.; Yan, F. Human Oral Microbiota and its Modulation for Oral Health. Biomed Pharmacother **2018**, *99*, 883-893.
- 94. Hajishengallis, G.; Abe, T.; Maekawa, T.; Hajishengallis, E.; Lambris, J.D. Role of Complement in Host-Microbe Homeostasis of the Periodontium. Semin Immunol **2013**, *25*, 65-72.
- 95. Serrigan, S.W.; Cox, D. Platelet-Bacterial Interactions. Cell Mol Life Sci 2010, 67, 513-523.
- 96. Kerrigan, S.W.; Jakubovics, N.S.; Keane, C.; Maguire, P.; Wynne, K.; Jenkinson, H.F.; Cox, D. Role of Streptococcus Gordonii Surface Proteins SspA/SspB and Hsa in Platelet Function. Infect Immun 2007, 75, 5740-5747.
- 97. Plummer, C.; Wu, H.; Kerrigan, S.W.; Meade, G.; Cox, D.; Ian Douglas, C.W. A Serine-Rich Glycoprotein of Streptococcus Sanguis Mediates Adhesion to Platelets Via GPIb. Br J Haematol **2005**, *129*, 101-109.
- 98. Taniguchi, N.; Nakano, K.; Nomura, R.; Naka, S.; Kojima, A.; Matsumoto, M.; Ooshima, T. Defect of Glucosyltransferases Reduces Platelet Aggregation Activity of Streptococcus Mutans: Analysis of Clinical Strains Isolated from Oral Cavities. Arch Oral Biol **2010**, *55*, 410-416.
- 99. Fitzgerald, J.R.; Foster, T.J.; Cox, D. The Interaction of Bacterial Pathogens with Platelets. Nat Rev Microbiol **2006**, *4*, 445-457.
- 100. 100. Koeth, R.A.; Wang, Z.; Levison, B.S.; Buffa, J.A.; Org, E.; Sheehy, B.T.; Britt, E.B.; Fu, X.; Wu, Y.; Li, L. *et al.* Intestinal Microbiota Metabolism of L-Carnitine, a Nutrient in Red Meat, Promotes Atherosclerosis. Nat. Med. **2013**, *19*, 576-585.
- 101. 101. Skye, S.M.; Zhu, W.; Romano, K.A.; Guo, C.; Wang, Z.; Jia, X.; Kirsop, J.; Haag, B.; Lang, J.M.; DiDonato, J.A. *et al.* Microbial Transplantation with Human Gut Commensals Containing CutC is Sufficient to Transmit Enhanced Platelet Reactivity and Thrombosis Potential. Circ Res **2018**, *123*, 1164-1176.
- 102. 102. Zhu, W.; Gregory, J.C.; Org, E.; Buffa, J.A.; Gupta, N.; Wang, Z.; Li, L.; Fu, X.; Wu, Y.; Mehrabian, M. *et al.* Gut Microbial Metabolite TMAO Enhances Platelet Hyperreactivity and Thrombosis Risk. Cell **2016**, 165, 111-124.
- 103. 103. Seldin, M.M.; Meng, Y.; Qi, H.; Zhu, W.; Wang, Z.; Hazen, S.L.; Lusis, A.J.; Shih, D.M. Trimethylamine N-Oxide Promotes Vascular Inflammation through Signaling of Mitogen-Activated Protein Kinase and Nuclear Factor-κB. J Am Heart Assoc 2016, *5*, e002767.
- 104. 104. Janeiro, M.H.; Ramírez, M.J.; Milagro, F.I.; Martínez, J.A.; Solas, M. Implication of Trimethylamine N-Oxide (TMAO) in Disease: Potential Biomarker Or New Therapeutic Target. Nutrients **2018**, *10*, 1398.
- 105. 105. Hernández-Ruiz, P.; Escalona Montaño, A.R.; Amezcua-Guerra, L.M.; González-Pacheco, H.; Niccolai, E.; Amedei, A.; Aguirre-García, M.M. Potential Association of the Oral Microbiome with Trimethylamine N-Oxide Quantification in Mexican Patients with Myocardial Infarction. Mediators Inflamm 2024, 2024, 3985731.
- 106. Zhuang, P.; Li, H.; Jia, W.; Shou, Q.; Zhu, Y.; Mao, L.; Wang, W.; Wu, F.; Chen, X.; Wan, X. et al. Eicosapentaenoic and Docosahexaenoic Acids Attenuate Hyperglycemia through the Microbiome-Gut-Organs Axis in Db/Db Mice. Microbiome 2021, 9, 185.
- 107. 107. Sanders, M.E.; Merenstein, D.J.; Reid, G.; Gibson, G.R.; Rastall, R.A. Probiotics and Prebiotics in Intestinal Health and Disease: From Biology to the Clinic. Nat Rev Gastroenterol Hepatol **2019**, *16*, 605-616.
- 108. 108. Inui, T.; Walker, L.C.; Dodds, M.W.J.; Hanley, A.B. Extracellular Glycoside Hydrolase Activities in the Human Oral Cavity. Appl Environ Microbiol **2015**, *81*, 5471-5476.
- 109. 109. Nyvad, B.; Takahashi, N. Integrated Hypothesis of Dental Caries and Periodontal Diseases. J Oral Microbiol 2020, 12, 1710953.
- 110. 110. Liu, H.; Wang, J.; He, T.; Becker, S.; Zhang, G.; Li, D.; Ma, X. Butyrate: A Double-Edged Sword for Health? Advances in Nutrition 2018, 9, 21-29.
- 111. 111. Magrin, G.L.; Strauss, F.J.; Benfatti, C.A.M.; Maia, L.C.; Gruber, R. Effects of Short-Chain Fatty Acids on Human Oral Epithelial Cells and the Potential Impact on Periodontal Disease: A Systematic Review of in Vitro Studies. Int J Mol Sci 2020, 21, 4895.
- 112. 112. Cosby, K.; Partovi, K.S.; Crawford, J.H.; Patel, R.P.; Reiter, C.D.; Martyr, S.; Yang, B.K.; Waclawiw, M.A.; Zalos, G.; Xu, X. *et al.* Nitrite Reduction to Nitric Oxide by Deoxyhemoglobin Vasodilates the Human Circulation. Nat Med **2003**, *9*, 1498-1505.
- 113. Bondonno, C.P.; Croft, K.D.; Hodgson, J.M. Dietary Nitrate, Nitric Oxide, and Cardiovascular Health. Crit Rev Food Sci Nutr **2016**, *56*, 2036-2052.
- 114. 114. Koch, C.D.; Gladwin, M.T.; Freeman, B.A.; Lundberg, J.O.; Weitzberg, E.; Morris, A. Enterosalivary Nitrate Metabolism and the Microbiome: Intersection of Microbial Metabolism, Nitric Oxide and Diet in Cardiac and Pulmonary Vascular Health. Free Radic Biol Med 2017, 105, 48-67.
- 115. 115. Gangula, P.; Ravella, K.; Chukkapalli, S.; Rivera, M.; Srinivasan, S.; Hale, A.; Channon, K.; Southerland, J.; Kesavalu, L. Polybacterial Periodontal Pathogens Alter Vascular and Gut BH4/nNOS/NRF2-Phase II Enzyme Expression. PLoS One **2015**, *10*, e0129885.
- 116. 116. Hampelska, K.; Jaworska, M.M.; Babalska, Z.Ł; Karpiński, T.M. The Role of Oral Microbiota in Intra-Oral Halitosis. J Clin Med 2020, *9*, 2484.

- 118. 118. Hsu, C.; Hou, C.; Chang-Chien, G.; Lin, S.; Tain, Y. Maternal Garlic Oil Supplementation Prevents High-Fat Diet-Induced Hypertension in Adult Rat Offspring: Implications of H2S-Generating Pathway in the Gut and Kidneys. Mol Nutr Food Res **2021**, *65*, e2001116.
- 119. 119. Tian, D.; Dong, J.; Jin, S.; Teng, X.; Wu, Y. Endogenous Hydrogen Sulfide-Mediated MAPK Inhibition Preserves Endothelial Function through TXNIP Signaling. Free Radic Biol Med **2017**, *110*, 291-299.
- 120. 120. Basic, A.; Serino, G.; Leonhardt, Å; Dahlén, G. H2S Mediates Increased Interleukin (IL)-1β and IL-18 Production in Leukocytes from Patients with Periodontitis. J Oral Microbiol **2019**, *11*, 1617015.
- 121. 121. Dilek, N.; Papapetropoulos, A.; Toliver-Kinsky, T.; Szabo, C. Hydrogen Sulfide: An Endogenous Regulator of the Immune System. Pharmacol Res **2020**, *161*, 105119.
- 122. 122. Xie, L.; Gu, Y.; Wen, M.; Zhao, S.; Wang, W.; Ma, Y.; Meng, G.; Han, Y.; Wang, Y.; Liu, G. *et al.* Hydrogen Sulfide Induces Keap1 S-Sulfhydration and Suppresses Diabetes-Accelerated Atherosclerosis Via Nrf2 Activation. Diabetes **2016**, *65*, 3171-3184.
- 123. 123. Rao, A.; Lokesh, J.; D'Souza, C.; Prithvisagar, K.S.; Subramanyam, K.; Karunasagar, I.; Kumar, B.K. Metagenomic Analysis to Uncover the Subgingival and Atherosclerotic Plaque Microbiota in Patients with Coronary Artery Disease. Indian J Microbiol 2023, 63, 281-290.
- 124. 124. Schulz, S.; Reichert, S.; Grollmitz, J.; Friebe, L.; Kohnert, M.; Hofmann, B.; Schaller, H.; Klawonn, F.; Shi, R. The Role of Saccharibacteria (TM7) in the Subginival Microbiome as a Predictor for Secondary Cardiovascular Events. Int J Cardiol **2021**, *331*, 255-261.
- 125. 125. Perry, S.E.; Huckabee, M.; Tompkins, G.; Milne, T. The Association between Oral Bacteria, the Cough Reflex and Pneumonia in Patients with Acute Stroke and Suspected Dysphagia. J Oral Rehabil 2020, 47, 386-394
- 126. 126. Boaden, E.; Lyons, M.; Singhrao, S.K.; Dickinson, H.; Leathley, M.; Lightbody, C.E.; McLoughlin, A.; Khan, Z.; Crean, S.; Smith, C. *et al.* Oral Flora in Acute Stroke Patients: A Prospective Exploratory Observational Study. Gerodontology **2017**, *34*, 343-356.
- 127. 127. Mougeot, J.-.C.; Stevens, C.B.; Paster, B.J.; Brennan, M.T.; Lockhart, P.B.; Mougeot, F.K.B. Porphyromonas Gingivalis is the most Abundant Species Detected in Coronary and Femoral Arteries. J Oral Microbiol **2017**, *9*, 1281562.

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